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Dynamics of growth and phytomass allocation in seedlings of *Pistacia atlantica* desf. versus *Pistacia vera* L. under salt stress

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Abstract

Salinity represents a major environmental constraint which can severely limit plant establishment in the arid and semi-arid areas in Tunisia. The present work was carried out to study the response of *Pistacia vera* L. and *Pistacia atlantica* Desf. seedlings to salt stress. Evaluation of salinity effects on both species was investigated using five increasing NaCl concentrations (0, 20, 40, 60 and 80 mM) during 60 days. The exposure to NaCl at seedlings stages, affects the majority of the studied parameters. Morphological parameters, such as height of shoot, number of green leaves, leaf area and consequently, phytomass allocation were significantly decreased. Additionally, *P. vera* accelerated leaf senescence. Both species showed a preferential allocation of the resources in favor of the roots for concentrations of NaCl lower than 40 mM. Whereas, only *P. atlantica* maintained a root growth for higher concentrations. Under salt stress, the specific leaf area decreased in *P. atlantica* (p<0.001) and translate a mechanism of tolerance to salinity. It appeared that both species tolerate concentrations lower than 40 mM of NaCl. However, for higher concentrations, *P. atlantica* seedlings are more tolerant.

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Introduction

The problem of salinity becomes more and more extensive in the majority of the countries in the process of development, mainly due to water scarcity (Rengasamy, 2010). Approximately 20% of the world's cultivated land and nearly half of all irrigated lands are affected by salinity (Zhu, 2001; Yamaguchi and Blumwald, 2005; Sun et al., 2009; Wu et al., 2010). In Tunisia, the saline lands are relatively frequent, occupying a surface of 1.5 million hectares, about 25% of the total cultural soils of the country (Jbira et al., 2001; Ben Ahmed et al., 2008). Salt stress affects plant growth through a double osmotic and ionic constraint followed by nutriment imbalance. Firstly, high concentrations of salts in the soil solution impose an osmotic stress on cell water relations which leads to reduction in water availability to plants and cellular dehydration (Sairam et al., 2002; Chartzoulakis, 2005). Secondly, an ionic toxicity caused by ions accumulation (mainly Na+ and Cl⁻) at high levels (Tavakkoli et al., 2011). Besides, nutriment deficiencies occurred due to the interaction of these ions with other nutritive elements (Tejera et al., 2006; Rejili, 2007; Evelin et al., 2009). The disturbance of potassium (K⁺) nutrition resulting from potassium-sodium interaction is often reported (Cramer et al., 1987). The salinity of soil and water of irrigation counts among the principal factors which limit plant growth and productivity (Zahran, 1999; Flowers, 2004; Parida and Das, 2005). Species develop morphological and physiological changes in the dynamics of growth, such as leaf number, plant height and leaf area reductions, increases in root growth and changes in dry phytomass allocation, as increases in leaf senescence, a decrease in specific leaf area and greater biomass allocation to roots. Plant growth reduction is often accompanied by inducing the senescence of leaves, mechanism adopted by the pistachio tree and other Mediterranean species (Colla 2006).The leaf et al., expansion decreases immediately following an increase in the salt concentration and the duration of the treatment (Munns, 2002; Bartels and Sunkar, 2005). The seedlings of pistachio tree reduce their growth when they are subjected to high salinity (Chelli-Chaabouni *et al.*, 2010). In spite of the direct exposure of the root to salinity, its growth is less affected by salt than the shoot (Parida and Das, 2005). It was reported that species surviving under optimal conditions present high specific leaf area (SLA), high relative growth rate (RGR) and high allocation of biomass to shoots, while species with low SLA and low RGR, allocate more biomass to roots in unfavourable habitat (Poorter and Garnier, 1999). The early stage of seedling growth is a critical phase in arid and semi-arid lands where plants are exposed to a series of abiotic and biotic stress. Plants exposed to salt stress are negatively affected with reduction in establishment and growth.

Pistacia vera and Pistacia atlantica are two pistachio species, cultivated and spontaneous respectively, belonging to the Anacardiaceae family. They are species of economic, ecological and medicinal interests (Sari et al., 2010; Tomaino et al., 2010). They are characteristic of the arid and semi-arid area. In Tunisia, according to Chaieb and Boukhris (1998), they are cultivated on all the territory, with prevalence in the arid and semi-arid areas. P. vera, which produces edible fruits, can survive in areas where annual rainfall is lower than 200 mm (Chaieb and Boukhris, 1998). This species supports the dryness better than the olive, the almond and the fig tree, considered as a typical xerophytes tree, with the advantage to resist to salinity (Behboudian et al., 1986; Rieger, 1995) and to tolerate extreme temperature, until -30°C in winters and 50°C during the summer. But for others, it is classified as a medium tolerant to salinity (Ebert, 2000).

P. atlantica is a very powerful woody and spontaneous species, adapted to the constraining pedo-climatic conditions of the arid and semi-arid areas. However, it is observed only on the degraded forests as aged and dispersed individuals, not exceeding 1500 individuals in Tunisia (Ghorbel et al., 1998), being threatened with extinction. Their rusticities in front of edaphic and climatic and their constraining factors low natural regeneration rates made very interesting to study these species. The reintroduction of tolerant species in the degraded lands could limit the extension of the marginal zones (Melgar et al., 2006; Tattini et al., 2009). P. atlantica is also useful to receive the graft of fruit pistachio (Gijón et al., 2010), improving vigor and production. However, the mechanism of improvement still not understood. In woody crops, rootstocks are mainly used in order to improve some of the tree characteristics as tree vigor, crop production or production quality (Olien and Lakso, 1986; Cohen and Naor, 2002; Weibel et al., 2003), thus, the graft may influence the movement of substances in the xylem such as ions (Jones, 1971; Tavallali and Rahemi, 2007), water status (Olien and Lakso, 1986) and plant-growth regulating hormones (Kamboj et al., 1999). In this context, Atkinson et al. (2003) and Solari et al. (2006) showed that the tree hydraulic conductance associated with specific rootstocks are determinant of growth potential in grafted trees. The used pistachio rootstocks are Pistacia vera L, Pistacia integerrima L., Pistacia terebinthus L. and Pistacia atlantica Desf (Karimi and Kafkas, 2011). The most used rootstock in Tunisia are P. atlantica and P. vera, but Germana (1997) reported that P. atlantica is more susceptible to drought stress than P. terebinthus. The present study was aimed to: a) assess comparatively the effect of increasing concentrations of NaCl on the dynamics of growth and phytomass production in Pistacia vera and Pistacia atlantica at the seedling stage, b) identify the principal physiological mechanisms of salt stress tolerance for the endangered Pistacia atlantica versus Pistacia vera and c) find a potential relationship between the improvement of P. vera vigor since grafted on P. atlantica and salinity tolerance.

Materials and methods

Materials and culture conditions

The objective of this experiment is to study the response of *Pistacia vera* and *Pistacia atlantica* to the salinity. The seeds of *Pistacia vera* (Mateur variety) were sampled in the region of Sidi Aïch (Tunisian South-East zone), while the seeds of *P. atlantica* were collected from wild trees in Meknassy (Tunisian West-Central zone) in 2009.

Seedlings production

The production of the seedlings was made in the laboratory at a temperature of 22°C and a photoperiod of 10h/14h light/obscurity. In order to avoid tegumentary inhibition, the seeds of P. atlantica underwent a mechanical scarification (Removing the pulp). The external hulls of P. vera were also removed. Germination took place in germinators filled with a mixture of peat (2/3) and sand (1/3). Transplantation was carried out after one month of growth. The seedlings at the stage of four to five leaves were then, transplanted in plastic pots, at a rate of an individual per pot. This later is of conical form, 15 cm in diameter and 13 cm in depth, whose capacity is two liters. The contents of the pots were pure sand, really washed, in order to avoid the interference of the trophic factor. The whole pots received the same amount of watering (200 ml) at a rate of two irrigations per week until obtaining vigorous and healthy individuals aged of 70 days.

Treatments applied

Since the sodium chloride (NaCl) represents the major source of salt in the ground and water of irrigation (Turkan and Demiral, 2009), it was used as the source of salt throughout this experimentation. Five increasing concentrations of NaCl were applied: 0, 20, 40, 60 and 80 mM of NaCl, respectively 0, 1.17, 2.34, 3.51 and 4.68 g of salt per liter of nutritive solution, at a frequency of 2 waterings per week. The individuals received a Hoagland solution. From 1st May 2010, the saline treatment was applied during 60 days. The test related to two species, five increasing concentrations of NaCl and 12 repetitions for each one, giving a totally of 120 pots. The culture was placed under a shelter protected by netting at an ambient temperature between 27 and 30°C, a relative humidity of 70% and a photoperiod of 12 h/12 h light/obscurity.

Measurement parameters

During the experiment, several parameters were measured. The green leaf number was estimated once per week by counting the number of green leaves per individual which appear along the experiment (n=6).

The senescent leaves were assessed as soon as the leaf senescence appeared in the treated individuals until the end of the test. A leaf is considered senescent when its reserves are completely impoverished, being thus, dropped from the individual. The Height of shoot was measured using a graduated ruler on an average of six plants for each treatment. This parameter was measured once per week (Expressed in cm). The leaf area by individual was given after the scanning of the leaves by a scanner and analyses of the photographs by the software of image processing Mesurim pro. 8 (Expressed in cm² by plant). This parameter was carried out at the end of the test (n=6). The shoot and root phytomass were obtained by separating the roots from shoots under water. Then, shoot phytomass (SP, in g per plant) and root phytomass (RP, in g per plant) were determined by weighing after a drying of the vegetable biomasses in a drying oven at 80°C during 48 h. This sampling was carried out at the end of the test (n=6). The RP/SP ratio was calculated. It is the relationship between root phytomass (RP) and shoot phytomass (SP). It is given as: Rate=(RP/SP)*100. The specific leaf area (SLA) is the relationship between the leaf area of the leaves (LA) and the corresponding dry mass (DL), expressed in cm²/g (n=6). It is given as: SLA=(LA /DL).

Statistical analyses

The results were subjected to the variance analysis (ANOVA) according to a factorial model with fixed factors, by using the software of statistics SPSS (Version 11.5). Then, the results were presented in the shape of the curves, joining average values framed by their standard deviations. The software SigmaPlot (Version 11.0) was used to prepare the various features.

Results

Effect of salt stress on growth of seedlings

In control seedlings, the number of green leaves increased significantly (p<0.001). Indeed, the average of the number of leaves increased reaching at the end of the experiment 18 leaves in *P. atlantica* and 13 leaves in *P. vera* (Fig. 1). Following the salt

concentrations, probably high on plant, particularly concentration 80 mM, in *P. vera*, not only new leaves didn't appear but also, the formed leaves were desiccated gradually during the experiment. At the end, 56 days of the treatment, the individual was completely senescent (Fig. 1). Under the same conditions of stress, 80 mM also induced an inhibition of the neo-formation of the leaves throughout the test without leaf senescence (Fig. 1). At this level of concentration, the salt stress marked a reduction of only 57%. Thus, the differences between the two species are highly significant (p<0.001).

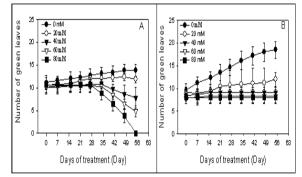


Fig. 1. Salinity effect (0, 20, 40, 60, 80 mM NaCl) on the number of leaves of *Pistacia vera* (A) and *Pistacia atlantica* (B) over a period of 60 days. Each value is the average of 6 replicates. Vertical bars indicate the standard error.

Salt stress marked a highly significant difference between the two species (p<0.001) for the leaf senescence. Contrary to *P. atlantica* seedlings, which maintained their leaves alive under salt stress throughout the experiment and even for the highest tested NaCl concentration, the seedlings of *P. vera* triggered after 35 days of the treatment, a premature leaf senescence, largely dependent on the applied NaCl concentration (Fig. 2). Indeed, the number of senescent leaves increased gradually throughout the experiment. The senescent leaves represented 15, 38, 61 and 100% for 20, 40, 60 and 80 mM of NaCl, respectively.

In control seedlings, the differences between the two species are highly significant for the leaf area (p<0.001). Indeed, at the end of the test, it reached 12 and 5.2 cm² in *P. vera* and *P. atlantica*, respectively (Fig. 3). Thus, the effect of the saline stress on the leaf

area is highly significant (p<0.001) and the excess of salt in the soil (80 mM) reduced the leaf area (p<0.001) by 80 and 89% for *P. atlantica* and *P. vera* seedlings, respectively.

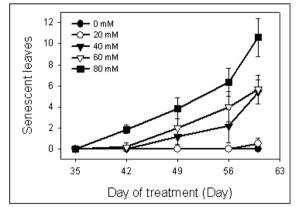


Fig. 2. Salinity effect (0, 20, 40, 60, 80 mM NaCl) on senescent leaves in *Pistacia vera* (A) and *Pistacia atlantica* (B) plants under salinity treatment. Each value is the average of six replicates and vertical bars represent standard errors.

Under salt stress, the specific leaf area showed differences largely depending on the species as well as the tested NaCl concentrations (Fig. 4). Indeed, in P. atlantica, the SLA decreased while the salinity increased. The variation of shoot height along the experiment reveals differences between the applied treatments (p<0.001) and the species (p<0.001) (Fig. 5). In control seedlings, the height of P. atlantica moved from 3.90 to 6.85 cm at the end of the test. On the contrary, the height of *P. vera* remained variable between 9.30 and 10.75 cm, whereas, for the seedlings subjected to salt stress concentration 80 mM induced an inhibition of the growth in height for both species. However the height decreased in P. vera seedlings against a non significant effect in P. atlantica seedlings under 20 and 40 mM of NaCl. At the end of the experiment, the reduction in height reaches 51% in P. atlantica against only 17% in P. vera for 80 mM.

Effect of salt stress on the phytomass allocation

The analysis of Fig. 6 shows that shoot phytomass was reduced in the seedlings subjected to the various NaCl concentrations as a result of the reduction of the number of green leaves, the height of the individuals, the leaf area of both species and the increase in the number of senescent leaves in *P. vera*. This reduction is marked in *P. vera* for the various concentrations of NaCl and particularly at 80 mM. Whereas in *P. atlantica*, even for high NaCl concentration, the individuals reduced SP by 45%, compared to *P. vera* (88%) that increased leaf senescence.

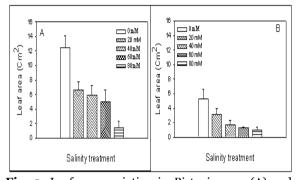


Fig. 3. Leaf area variation in *Pistacia vera* (A) and *Pistacia atlantica* (B) plants under salinity treatment. Each value is the average of six replicates and vertical bars represent standard errors.

Root phytomass (RP) increased for the applied saline treatments (20 and 40 mM) for both species (Fig. 7). However, for higher NaCl concentrations (60 and 80 mM), RP was reduced. This reduction is marked in *P. vera* (87%) at 80 mM, whereas in *P. atlantica*, even for the concentrations of 60 and 80 mM, RP was maintained at higher values compared to the control ones (41%).

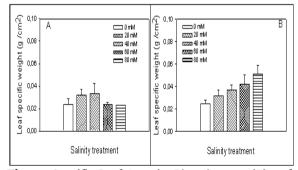


Fig. 4. Specific Leaf Area in *Pistacia vera* (A) and *Pistacia atlantica* (B) plants under salinity treatment. Each value is the average of six replicates and vertical bars represent standard errors.

The RP/SP ratio increased under salt stress, except for 80 mM of NaCl (Fig. 8). Thus, the salinity stimulated the roots growth, especially for *P*. *atlantica* seedlings (p<0.001).

Discussion

Dynamic of growth under salt stress

The NaCl causes a reduction in plant growth (Shannon and Grieve, 1999; Munns, 2002; Parida and Das, 2005; Chelli-Chaabouni et al., 2010; Correia et al., 2010). It is considered as one of the first manifestations of salt stress (Parida and Das, 2005) on morphological and physiological traits. It appears on leaf area, leaf number, mechanism adopted by the pistachio tree and other Mediterranean species (Munns, 2002; Kchaou et al., 2010) to reduce the water losses by transpiration. The reduction in the growth can be explained by the reduction in the photosynthesis and in the leaf area (Parida and Das, 2005). The salt stress causes an immediate reduction in leaf expansion which is inhibited by increasing the salt concentration and duration of treatment (Curtis and Läuchli, 1987; Parida et al., 2003; Abbruzzese et al., 2009), thus, the new leaves develop slowly and the senescence of the old leaves accelerates. This reduction in leaf growth is the result of inhibition of the cellular divisions under abiotic stress, as salt and water stress. Several authors showed that the reduction of the shoots growth in the seedlings of pistachio is related to disturbances of the growth regulators, as the abscissic acid and the cytokinin (Termaat et al., 1985; Kuiper, 1990). In fact, a decrease in the cytokinin hormone was recorded in Pinus sylvestris, Pinus koraiensis and Abies hallophylla (Liu et al., 1998). Also, several studies reported that abscissic acid increased significantly under salinity stress (Mulholland et al., 2003; Zhu et al., 2005; Albacete et al., 2008; Zorb et al., 2013). Moreover, the free auxin decreases according to species (Wang et al., 2001; Ghanem et al., 2008; Salah et al., 2013; Zorb et al., 2013). The reduction of growth can be due also to a nutritional deficiency in stressed plants (Chelli-Chaabouni et al., 2010).

The intensification of salinity is accompanied by reductions in green leaves and their dimension and the stems length, and increases in senescent leaves, thus the morphological parameters are sensitive to the double osmotic and ionic constraints under salt stress (Munns, 2002). The reduction in plant growth of P. vera and P. atlantica is due to a decline in cellular division and elongation (Munns, 2002). Indeed, the reduction of plant height of the seedlings is accompanied by a reduction of the number of leaves, a delay of development of the new leaves and an acceleration of the senescence of old ones. Our results are in agreement with other works (Karimi et al., 2009; Chelli-Chaabouni et al., 2010), and for this reason the morphological changes are adopted as alternate indicators of stress. The senescence of the old leaves contributes to preserving the soil water. This mechanism is considered a recycling program for the redistribution of resources in favor of young leaves, stems or roots (Chaves et al., 2002). In P. *vera*, an acceleration of the leaf senescence at the end of the experiment was observed particularly at 80 mM of NaCl. This phenomenon was preceded by total leaf necrosis, as revealed by Chelli-Chaabouni et al. (2010), that constitute a sign of toxicity by accumulation and excess of salt in the leaves (Ben Ahmed et al., 2008). These symptoms were often associated with a high rate of chloride (Picchioni and Graham, 2001) or of sodium in leaves (Miyamoto et al., 1985; Karakas et al., 2000; Chelli-Chaabouni et al., 2010; Kchaou et al., 2010). Under increasing saline stress, the leaf specific weight increases in P. atlantica, a general response of species adapted to unfavorable habitats (Poorter and Garnier, 1999). The excess of salt in soil reduced more the growth of the shoots in P. vera seedlings throught a significant reductions in the leaf growth and increases in the leaf senescence. However, P. atlantica maintains a growth less affected for the NaCl concentrations.

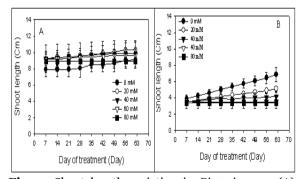


Fig. 5. Shoot length variation in *Pistacia vera* (A) and *Pistacia atlantica* (B) plants under salinity treatment. Each value is the average of six replicates and vertical bars represent standard errors.

Phytomass allocation under salt stress

Salinity is a determining element for the plant growth, particularly in arid and semi-arid areas. It induces a significant reduction of shoots growth, whereas it improves the root growth except for the highest salt concentration (80 mM). Similar effects were observed on the growth of the seedlings of pistachio tree subjected to salt stress in vivo conditions (Ranjbar et al., 2002; Tavallali et al., 2008; Karimi et al., 2009) and in vitro (Benmahioul et al., 2009). The same results were also obtained in other plants as Olea europea (Kchaou et al., 2010). According to Zhu (2001), the reduction of the shoots growth is an adaptive mechanism necessary to the survival of the plants exposed to abiotic stress. Plants protect the shoots against invasion from the toxic ions (Na⁺ and Cl⁻) by its accumulation in roots. In addition, Bouraoui et al. (1998) showed that the passage of the cells from the division to the elongation stage is accompanied by a modification of the respiratory metabolism. The maintenance of the root elongation and the stimulation of respiration on medium salted correspond a mechanism of tolerance to salinity (Bouraoui et al., 1998). Indeed, the leaf area decreased significantly in both species, which led to the development of the root, considered as a criterion for salinity tolerance (Chelli-Chaabouni et al., 2010). This would allow better use of the available water (Benmahioul et al., 2009), suggesting an internal settlement mechanism, thus, the osmoregulation allows the plant to adjust its water potential while promoting root elongation and water supply.

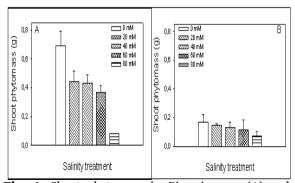
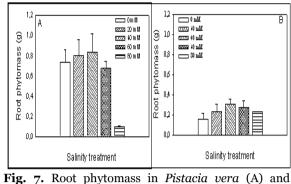


Fig. 6. Shoot phytomass in *Pistacia vera* (A) and *Pistacia atlantica* (B) plants under salinity treatment. Each value is the average of six replicates and vertical bars represent standard errors.



Pistacia atlantica (B) plants under salinity treatment. Each value is the average of six replicates and vertical bars represent standard errors.

Roots are affected by abiotic stresses in many ways. Salinity induces a reduction of root system biomass through a decrease in root length and width. Cotton seedlings growing in hydroponic salt solution produced less and thinner roots with increasing salinity. Root anatomical analysis showed shorter and more nearly iso-diametrical cortical cells than those of control plants (Kurth et al., 1986). Salt stress may lead to root lignifications in pistachio (Walker et al., 1987). Under salinity, the results represent a preferential allowance of the resources in favor of the roots in *P. atlantica* at higher concentrations of NaCl. It maintained a higher root growth and shoots, compared to P. vera for the NaCl concentrations. These responses translate a tolerance of *P. atlantica* to salt stress, thus, the good performance of P. vera grafted on *P. atlantica* can be allotted to the adaptive ecophysiologic characteristics related particularly to root growth.

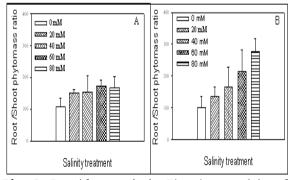


Fig. 8. Root/shoot ratio in *Pistacia vera* (A) and *Pistacia atlantica* (B) plants under salinity treatment. Each value is the average of six replicates and vertical bars represent standard errors.

Potentiality of establishment of P. vera and P. atlantica under salinity stress

Under salinity, a significant reduction in shoots was observed in both species, considered as a mechanism of tolerance necessary for the survival of plants exposed to abiotic stress (Zhu, 2001). However, P. atlantica maintains shoots less affected by NaCl, compared to P. vera. The increases in leaf specific weight and the preferential allowance of the resources in favor of the roots translate a tolerance of P. atlantica to salt stress, against root necrosis and a senescence of leaves in P. vera. This confirms that species from favourable productive habitats present high SLA, high relative growth rate (RGR) and high allocation of biomass to shoots, while species with low SLA and low RGR, allocate more biomass to roots in unfavourable habitat (Poorter and Garnier, 1999). The mechanisms explain the good performance of the seedlings of P. atlantica, compared with P. vera under higher concentration of NaCl (80 mM of NaCl). That joined the literature to explain the agricultural practice based on the use of *P. atlantica* as a rootstock of P. vera for a better rusticity to the abiotic constraints as salinity. The good performance of P. vera grafted on P. atlantica can be allotted to the adaptive ecophysiologic characteristics related particularly to root growth.

References

Abbruzzese G, Beritognolo I, Muleo R, Piazzai M, Sabatti M, Mugnozza GS, Kuzminsky E. 2009. Leaf morphological plasticity and stomatal conductance in three *Populus alba* L. genotypes subjected to salt stress. Environmental and Experimental Botany **66**, 38-388.

Albacete A, Ghanem ME, Martinez-Andujar C, Acosta M, Sanchez-Bravo J, Martinez V, Lutts S, Dodd IC, Perez-Alfocea F. 2008. Hormonal changes in relation to biomass partitioning and shoot growth impairment in salinized tomato (*Solanum lycopersicum* L.) plants. Journal of Experimental Botany **59**, 4119- 4131.

Atkinson CJ, Else MA, Taylor L, Dover CJ.

2003. Root and stem hydraulic conductivity as determinants of growth potential in grafted trees of apple (*Malus pumila* Mill.) Journal of Experimental Botany **54**, 1221-1229.

Bartels D, Sunkar R. 2005. Drought and Salt Tolerance in Plants. Critical Reviews in Plant Sciences **24**, 23-58.

Behboudian M, Walker R, Torokfalvy. 1986. Effect of water stress and salinity on photosynthesis of pistachio. Scientiae Horticulturae **29**, 251-261.

Ben Ahmed C, Ben Rouina B, Boukhris M, 2008. Changes in water relations, photosynthetic activity and proline accumulation in one-year-old olive trees (*Olea europaea* L. cv.Chemlali) in response to NaCl salinity. Acta Physiologiae Plantarum **30**, 553-560.

Benmahioul B, Daguin F, Meriem Kaid-Harche M. 2009. Effet du stress salin sur la germination et la croissance *in vitro* du pistachier (*Pistacia vera* L.). Comptes rendus Biologies **332**, 752-758.

Bouraoui N, Grignon C, Zid E. 1998. Effet de NaCl sur la croissance et la respiration racinaire du triticale (*X-Triticosecale* wittmack). Cahiers Agriculture**7**, 372- 376.

Chaieb M, Boukhris M. 1998. Flore succincte et illustrée des zones arides sahariennes des zones aride de Tunisie. ATPNE, 145.

Chartzoulakis KS. 2005. Salinity and olive: Growth, salt tolerance, photosynthesis and yield. Agricultural Water Management **78**, 108-121.

Chaves MM, Pereira JS, Maroco J, Rodrigues ML, Ricardo CPP, Osório ML., Carvalho I, Faria T, Pinheiro C. 2002. How plants cope with water stress in the field. Photosynthesis and growth. Annals of Botany **89**, 907-916.

Chelli-Chaabouni A, Ben Mosbah A, Maalej M, Gargouri K, Gargouri-Bouzid R, Drira N. 2010. In vitro salinity tolerance of two pistachio rootstocks: *Pistacia vera* L. and *P. atlantica* Desf. Environmental and Experimental Botany **69**, 302- 312.

Cohen S, Naor A. 2002. The effect of three rootstocks on water use, canopy conductance and hydraulic parameters of apple trees and predicting canopy from hydraulic conductance. Plant, Cell and Environment **25**, 17-28.

Colla G, Roupahel Y, Cardarelli M. 2006. Effect of salinity on yield, fruit quality, leaf gas exchange, and mineral composition of grafted water melon plants. Hortscience **41**, 622- 627.

Correia PJ, Gamaa F, Pestana M, Martins-Louciaño MA. 2010. Tolerance of young (*Ceratonia siliqua* L.) carob rootstock to NaCl. Agricultural Water Management **97**, 910- 916.

Cramer GR, Lynch J, Laüchli A, Epstein E. 1987. Influx of Na⁺, K⁺, and Ca ²⁺ into roots of salt stressed cotton seedlings: effects of supplemental Ca²⁺. Plant Physiology **83**, 510- 516.

Curtis PS, Läuchli A, 1987. The effect of moderate salt stress on leaf anatomy in *Hibiscus cannabinus* (Kenaf) and its relation to leaf area. American Journal of Botany **74**, 538- 542.

Ebert G. 2000. Salinity problems in sub-tropical fruit production. Acta Horticulturae **531**, 99-105.

Evelin H, Kapoor R, Giri B. 2009. Arbuscular mycorrhizal fungi in alleviation of salt stress: A review. Annals of Botany **104**, 1263-1280.

Flowers TJ. 2004. Improving crop salt tolerance. Journal of Experimental Botany **55**, 307-319.

Germana C. 1997. The response of pistachio trees to water stress as affected by two different rootstocks. Acta Horticulturae **449**, 513- 519.

Ghanem ME, Albacete A, Martinez-Andujar C, Acosta M, Romero-Aranda R, Dodd IC, Lutts S, Perez-Alfocea F. 2008. Hormonal changes during salinity-induced leaf senescence in tomato (*Solanum lycopersicum* L.). Journal of Experimental Botany **59**, 3039- 3050.

Ghorbel A, Ben Salem-Fnayou A, Chatibi M, Twey A. 1998. Genetic resources of *Pistacia* in Tunisia. Toward a comprehensive documentation and use of *Pistacia* genetic diversity in central and west Asia, North Africa and Europe. In: Padulosi S, Hadj Hassan A. eds. Report of the IPGRI workshop, 14-17 December 1998, Irbid Jordan.

Gijón MC, Gimenez C, Perez-López D, Guerrero J, Couceiro JF, Moriana A. 2010. Rootstock influences the response of pistachio (*Pistacia vera* L. cv. Kerman) to water stress and rehydration. Scientia Horticulturae **125**, 666-671.

Jbira N, Chaïbia W, Ammara S, Jemmalib A, Ayadia A. 2001. Root growth and lignification of two wheat species differing in their sensitivity to NaCl, in response to salt stress. Life Sciences **324**, 863- 868.

Jones OP. 1971. Effects of rootstocks and interstock on the xylem sap composition in apple trees: effects of nitrogen, phosphorus and potassium content. Annals of Botany **35**, 825-836.

Kamboj JS, Blake PS, Quinlan JD, Baker DA. 1999. Identification and quantification by GC-MS of zeatin and zeatin riboside in xylem sap from rootstock and scion of grafted apple trees. Plant Growth Regulation **28**, 199- 205.

Karakas B, Bianco RL, Rieger M. 2000. Association of leaf marginal scorch with sodium accumulation in salt-stressed peach. Hortcultural Science **35**, 83-84.

Karimi HR, Kafkas S. 2011. Genetic relationships among *Pistacia* species studied by SAMPL markers. Plant Systematics and Evolution **297**, 207-

Karimi S, Rahemi M, Maftoun M, Eshghi, Tavallali V. 2009. Effects of Long term Salinity on Growth and Performance of Two Pistachio (*Pistacia* L.) Rootstocks. Australian Journal of Basic and Applied Sciences **3**, 1630- 1639.

Kchaou H, Larbi A, Gargouri K, Chaieb M, Morales F, Msallem M. 2010. Assessment of tolerance to NaCl salinity of five olive cultivars based on growth characteristics and Na⁺ and Cl⁻ exclusion mechanisms. Scientia Horticulturae **124**, 306-315.

Kuiper D, Schuit J, Kuiper PJC. 1990. Actual cytokinin concentrations in plant tissue as an indicator for salt resistance in cereals. Plant Soil **123**, 243-250.

Kurth E, Cramer GR, Lauchli A, Epstein E. 1986. Effects of NaCl and CaCl₂ on cell enlargement and cell production in cotton roots. Plant Physiology 82, 1102-1106.

Liu GF, Liu GJ, Yang CP, Wang HM. 1998. The analysis of hormonal change and salt resistant ability of tree species under salt stress. Journal of Northeast Agricultural University **26**, 1-4.

Melgar JC, Benlloch M, Fernandez-Escobar R. 2006. Calcium increases sodium exclusion in olive plants. Scientiae Horticulturae **109**, 303-305.

Miyamoto S, Gobran GR, Piela K. 1985. Salt effects on seedling growth and ion uptake of three pecan rootstock cultivars. Agronomie 77, 383-388.

Mulholland BJ, Taylor IB, Jackson AC, Thompson AJ. 2003. Can ABA mediate responses of salinity stressed tomato. Environmental and Experimental Botany **50**, 17-28.

Munns R. 2002. Comparative physiology of salt and water stress. Plant, Cell and Environment **25**, 239-250.

Olien WC, Lakso AN. 1986. Effect of rootstock on apple (*Malus domestica*) tree water relations. Physiologia Plantarum. **67**, 421-430.

Parida AK, Das AB, Mittra B. 2003. Effects of NaCl stress on the structure, pigment complex compsition and photosynthetic activity of mangrove *Bruguiera parviflora* chloroplasts. Photosynthetica **41**, 191- 200.

Parida AK, Das AB. 2005. Salt tolerance and salinity effects on plants: A review. Ecotoxicology and Environnemental Safety **60**, 324-349.

Picchioni GA, Graham CJ. 2001. Salinity, growth and ion uptake selectivity of container-growth Crataegus opaca. Scientiae Horticulturae **90**, 151-166.

Poorter H, Garnier E. 1999. Ecological significance of inherent variation in relative growth rate. In: Pugnaire F, Valladares X, eds. Handbook of functional plant ecology. New York, USA: Marcel Dekker, 81- 120.

Ranjbar A, Damme PV, Samson R, Lemeur R. 2002. Leaf water status and photosynthetic gas exchange of *Pistacia khinjuk* and *P. mutica* exposed to osmotic drought stress. Acta Horticulturae **591**, 423-428.

Rejili M, Vadel AM, Guetet A, Neffatti M. 2007. Effect of NaCl on the growth and the ionic balance K+/Na+ of two populations of *Lotus creticus* (L.) (Papilionaceae). South African Journal of Botany **73**, 623-631.

Rengasamy P. 2010. Soil processes affecting crop production in salt-affected soils. Australian Journal of Soil Research **37**, 613- 620.

Rieger M. 1995. Offsetting effects of reduced root hydraulic conductivity and osmotic adjustment following drought. Tree Physiolgy **15**, 379-385. **Sairam RK, Veerabhadra RK, Srivastava GC.** 2002. Differential response ofwheat genotypes to long term salinity stress in relation to oxidative stress, antioxidant activity and osmolyte concentration. Plant Science **163**, 1037-1046.

Salah IB, Albacete A, Messedi D, Gandour M, Andujar CM, Zribi K, Martinez V, Abdelly C, Perez-Alfocea F. 2013. Hormonal responses of nodulated *Medicago ciliaris* lines differing in salt tolerance. Environmental and Experimental Botany 86, 35-43.

Sari I, Baltaci Y, Bagci C, Davutoglu V, Erel O, Celik H, Ozer O, Nur MD, Aksoy, M. 2010. Effect of pistachio diet on lipid parameters, endothelial function, inflammation, and oxidative status: A prospective study. Nutrition **26**, 399-404.

Shannon MC, Grieve CM. 1999. Tolerance of vegetable crops to salinity. Scientia Horticulturae **78**, 5-38.

Sun J, Chen S, Dai S, Wang R, Li N, Shen X. 2009. NaCl-induced alternations of cellular and tissue ion fluxes in roots of salt-resistant and saltsensitive poplar species. Plant Physiology **149**, 1141-1153.

Tattini M, Traversi ML, Castelli S, Biricolti S, Guidi L, Massai R. 2009. Contrasting response mechanisms to root-zone salinity in three cooccurring Mediterranean evergreens: a physiological and biochemical study. Functional Plant Biology **36**, 551-563.

Tavakkoli E, Fatehi F, Coventry S, Rengasamy P, McDonald GK. 2011. Additive effects of Na⁺ and Cl⁻ ions on barley growth under salinity stress. Journal of Experimental Botany **62**, 2189- 2203.

Tavallali V, Rahemi M. 2007. Effects of Rootstock on Nutrient Acquisition by Leaf, Kernel and Quality of Pistachio (*Pistacia vera* L.). American-Eurasian Journal of Agricultural and Environmental Science **2**, 240-246.

Tavallali V, Rahemi M, Panahi B. 2008. Calcium induces salinity tolerance in pistachio rootstocks. Fruits **63**, 285- 296.

Tejera NA, Soussi M, Luch C. 2006. Physiological and nutritional indicators of tolerance to salinity in chickpea plants growing under symbiotic conditions. Environnemental and Experimental Botany **58**, 17-24.

Termaat A, Passora JB, Munns R. 1985. Shoot turgor does not limit shoot growth of NaCl affected wheat and barley. Plant Physiology **77**, 869-872.

Tomaino A, Martorana M, Arcoraci T, Monteleone D, Giovinazzo C, Saija A. 2010. Antioxidatif activity and phenolic profile of pistachio (*Pistacia vera* L., variety Bronte) seeds and skins. Biochimie **92**, 1115- 1122.

Türkan I, Demiral T. 2009. Recent developments in understanding salinity tolerance, uptake of three pecan rootstock cultivars. Agronomie Journal 77, 383-388.

Walker RR, Törökfalvy E, Behboodian MH. 1987. Uptake and distribution of chloride. Sodium and potassium ions and growth of salt-treated pistachio plants. Australian Journal of Agricultural Research **38**, 383- 394.

Wang Y, Mopper S, Hasenstein KH. 2001. Effects of salinity on endogenous ABA, IAA, JA, and SA in *Iris hexagona*. Journal of Chemical Ecolgy **27**, 327-342.

Weibel A, Johnson RS, DeJong TM. 2003. Comparative vegetative growth responses of two peach cultivars grown on size-controlling versus standard rootstocks. Journal of American Society for Horticultural Science **128**, 463-471.

Wu QS, Zou YN, Liu W, Ye XF, Zai HF, Zhao

LJ. 2010. Alleviation of salt stress in citrus seedlings inoculated with mycorrhiza: changes in leaf antioxidant defense systems. Plant, Soil and Environment. **56**, 470- 475.

Yamaguchi T, Blumwald E. 2005. Developing salt-tolerant crop plants: Challenges and opportunities. Trends Plant Science **10**, 615- 620.

Zahran HH. 1999. *Rhizobium*-legume symbiosis and nitrogen fixation under severe conditions and in an arid climate. Microbiology and Molecular Biology Reviews **63(4)**, 968- 989. Zhu C, Schraut D, Hartung W, Schaffner AR. 2005. Differential responses of maize MIP genes to salt stress and ABA. Journal of Experimental Botany 56, 2971-2981.

Zhu JK. 2001. Plant salt tolerance. Trends in Plant Science **6**, 66-71.

Zorb C, Geilfus CM, Muhling KH, Ludwig-Muller J. 2013. The influence of salt stress on ABA and auxin concentrations in two maize cultivars differing in salt resistance. Journal of Plant Physiology **170**, 220-224.