



RESEARCH PAPER

OPEN ACCESS

Seedling quality, biomass promotion and nutrient uptake potential of AMF, *Azotobacter* and *Pseudomonas* in *Azadirachta indica* under nursery condition

K. K. Chandra

Department of Forestry, Wildlife and Environmental Sciences, Guru Ghasidas Vishwavidyalaya, Bilaspur -495009 India

Article published on April 25, 2013

Key words: Microbial inoculants, root infection, seedling quality, nutrient uptake.

Abstract

The efficacy potential of Arbuscular-Mycorrhizal fungi (*Glomus interaradics* and *Acaulospora scrobiculata*), Phosphorus Solubilizing Bacteria (*Pseudomonas straita*) and *Azotobacter* (*Azotobacter chroococcum*) were determined in *Azadirachta indica* seedling. Fifty gram AMF, 10g *Pseudomonas* and 10g *Azotobacter* were inoculated in rhizosphere of newly recruited seedlings individually or in integration. Seedling quality index enhanced significantly recorded maximum by 62.56% and seedling vigour by 63.85 percent with tripartite inoculation of AMF+PSB+*Azotobacter* over uninoculated seedling. Similarly, the seedling biomass was also enhanced substantially up to 72.42% with the same treatment. The biomass increment was also observed in seedlings inoculated with AMF+*Azotobacter* (60.0%), AMF+PSB (24.78%), and PSB+*Azotobacter* (17.60%) in comparison to control plants. The seedling dependency to microbial inoculants was found highest of 72.13% on AMF+PSB+*Azotobacter* followed with AMF+*Azotobacter* (60%). The microbial tripartite gave best results in production of superior quality planting material than with bipartite inoculation. The percent root infection by AMF recorded maximum 66.0% in seedling with AMF+*Azotobacter* followed with AMF+*Azotobacter* compared to control seedlings (7.66%). Similarly the integration of AMF+PSB+*Azotobacter* was found most effective in the uptake of nitrogen, phosphorus and potassium content which were increased by 42%, 105% and 137% respectively. The highest content of Fe (10.71 µg/g), Zn (0.18 µg/g) Mg (16.74 µg/g), Mn (0.98 µg/g) and Cu (0.12 µg/g) were analysed on inoculation with AMF, PSB and *Azotobacter*. The combined inoculation of AMF+PSB+*Azotobacter* was given best results in improving the quality of seedling, biomass and nutrient uptake of *Azadirachta indica* which has opened the new implications for their use in healthy seedlings production in nursery for the supply of ever increasing demands for afforestation and reforestation.

*Corresponding Author: k. K. Chandra ✉ kkchandra_31@rediffmail.com

Introduction

Neem (*Azadiracta indica* A. Juss) is considered as sacred species by Indian's from ancient time. It is projected as an elegant species because of its pesticidal and medicinal virtues planted in large scale by community development and social forestry department. It is known to clean the environment of industrial and polluted area, conserve soil, augment aesthetic value, provide shelter in summer and improves physical and chemical properties of soil. The leaves and seed kernel of the species is being used as major ingredient in formulations against several insects of vegetable and field crops (Bramhachari, 2004). Moreover the leaves have good fodder value and the tender shoots are commonly used as tooth brush due to medicinal values of the species. In addition, it is worshiped by rural people represent goddess Durga and planted in front of their house. Due to these, there is an interest in society for raising Neem tree on bund of agriculture field, degraded forest area, near village, bund of ponds and on wastelands.

Since the tree is important for human and animal (Ambe, 2001) large scale afforestation is under taken every year to achieve plantation target but just after one season the majority of plants did not survive due to no use of fertilizers, use of poor quality planting materials, dependence on monsoon rain and non adoption of scientific method at the time of planting. Querejeta *et al.* (2003) also identified nutrient deficiency and water stress as major limiting factors for plant survival and growth after transplanting in tropical soils. Moreover presently the soil is becoming fragile by increased use of pesticides, pressure of population, management system and pollution unable to maintain its fertility and support plant growth. In this situation either regular fertilizer application and irrigation are required to maintain the growth or survival of plant or the production of seedlings of superior quality is required for reversing the current degradation of soil and forests (Pindi, 2011).

The microbial technology is emerging as new tools provides guarantee for production of quality nursery stock and highest survival in fields by reducing the transplanting sock and increase the nutrients uptake and water (Khalafallah and Abo-Ghalia, 2008). The technology consists plant growth promoting rhizotrophic microorganism bacteria and fungi that actively colonize plant root, fix atmospheric nitrogen, solubilize insoluble phosphate in soil and secretes some growth promoting substances for plants and capable of accelerating microbial transformation to facilitate more availability of nutrients, its uptake and enhance physiological process.

Naturally the plant interacts with many microorganisms AMF (Arbuscular Mycorrhizal Fungi), phosphorus solubiling microorganism, rhizobia and with other free living microorganisms (Miransari, 2011) to fulfill the requirements of nutrients and water apparent result is not seen because of the presence of low and ineffective microorganism in soil. Thus it is imperative to use the potential of soil microorganisms in production system by selecting their best combinations. It becomes thus possible to encourage healthy cultural systems without using chemical fertilizers in forestry and to sustain a better productivity and ecosystem conservation. From screening of literature it is also revealed that research on comparative study of different microbial inoculants in forest species is also not much available. In this context the present study was conducted to determine plant growth promoting potential of AMF, *Azotobacter* and Phosphorus solubilizing microorganism on the biomass and nutrient uptake of *Azadiracta indica* under nursery condition.

Material and methods

Study site

The nursery soil used in experiment was sandy loam collected form pasture stand of State Forest research Institute campus, Jabalpur (India) lies between 23° 10'N and 79° 59'E. The soil was passed with 2mm size

sieve then filled in polythene bag of size 13cm X 23cm size.

Seed of *Azadirachta indica* collected from Katni provenance were sown in bags.

Isolation and identification of AMF

AMF (*Glomus interradicis* Schenck and Smith and *Acaulospora scrobiculata* (Trappe) Morton) isolated from rhizosphere soil of same provenance was isolated and multiplied in *Panicum maximum* grass for 8 months period in pot culture and used in this experiment.

Inoculation of microbial culture

AMF 50g consisted substrate, spores, infected root fragments and hyphae were placed in root region of recruit in each polythene bag at 5-10cm just below the soil surface. Non inoculated plants received the same amount of autoclaved soil with a 10 ml water extract of the soil by filtration as controls to balance composition of the microbial community (Jansa *et al.*, 2005). The inoculum load calibrated by the MPN method was 0.17×10^4 per 25g of inoculum. Ten gm Phosphorus Solubilizing Bacteria (*Pseudomonas straita*) and 10g *Azotobacter chroococum*. (Inoculum load 2.17×10^6) were also inoculated in seedlings as per experimental design.

Experimental Design

The experiment was arranged in Randomized Block Design with five replications contain ten seedlings in each replicate. The treatment was arranged as T0- Uninoculated control, T1-AMF, T2, PSB, T3- *Azotobacter*, T4-AMF+PSB, T5- AMF+*Azotobacter*, T6- PSB+*Azotobacter*, T7- AMF+PSB+*Azotobacter*.

Data Record and observations

After 150 days of inoculation four seedlings were selected randomly from each replicate for observing the biomass and AMF root infection in *A. indica*. Plant material was dried in an oven at 70°C for 7 days and weighed to determine biomass production of the seedling. The seedling quality is calculated as per formula given by Dickson *et al.*, (1960) while seedling

vigour of the seedling was determined by Qualls and Cooper (1968). The plant dependency to different microorganisms was calculated by expressing the difference between the total dry weight of treated plants by particular microorganism and the dry matter weight of non treated plants as a percentage of the total dry weight of treated plants according to Plenchette *et al.* (1983).

Detection of AMF in soil and roots

Percent root infection by AMF was calculated by cleaning the root fragments in 10% KOH and stained with 0.05% trypan blue according to the method of Phillips and Hayman (1970). The stained roots were examined under microscope to observe the presence of intracellular mycelium, vesicle and arbuscular in root pieces and calculated as length of cortical cells colonized (in mm) by the AMF fungi for each root segment and expressed as a percentage of total root length infected (Mc Gonigle *et al.*, 1990).

Analysis of nutrient content in plant

Total N was analyzed by Auto Kjeltach, P by spectrophotometer and K by flame photometer as per Jackson (1973). The content of Zn, Cu, Mg, Fe and Mn were determined by means of an atomic absorption spectrophotometer GBC (John, 1970). All experiment data were statistically analysed by using SPSS Window software.

Results and discussion

Effect on Seedling quality, Vigour index and Biomass

The seedling quality, vigour and biomass production in *A. indica* inoculated with AMF, PSB and *Azotobacter* were compared to the control plants are given in Figure 1. Seedling quality index enhanced significantly and recorded maximum 62.56% with AMF+PSB+*Azotobacter*, 55.29 percent with AMF+*Azotobacter* and 54.21 percent with PSB+*Azotobacter* inoculation than control seedlings (Table -1). The vigour index of seedling was influenced maximum of 63.85 percent with the inoculation of AMF+PSB+*Azotobacter* over uninoculated seedling. The treatment of AMF either individual or in integration with other microbial

culture was found more effective on quality index, vigour index, root shoot ratio and biomass of the seedling as compared to PSB and *Azotobacter* inoculation.

The integration of tripartite microbial inoculants gave maximum result on all the parameter of the present study declined subsequently to bipartite and single microbe inoculation. The rhizotrophic microorganism viz. AMF, PSB and *Azotobacter* inoculated with *A. indica* made congenial environment, protected from diseases (Lax *et al.*, 2011), increase absorptive capacity by AMF mycelium network (Hassan *et al.*, 2012), compensated stress and drought resulted into enhanced uptake of nutrients which converted into higher biomass and improved the quality parameter of the seedlings. The same trend of influence was reflected in seedling biomass as observed in seedling quality index and vigour with AMF+PSB+*Azotobacter*. The biomass enhanced maximum 41.90 percent with AMF+PSB + *Azotobacter* followed with AMF*Azotobacter* and AMF+PSB respectively. Non-inoculated control plants exhibited very weak shoot growth and produced little biomass compared to the inoculated plants. The absence of the AMF in soil and root and other microorganisms is a factor that could limit seriously the growth and the development of control plants. Practically these conditions, the lack of growth with little biomass production in control plants could lead to a higher mortality of plants (Chen *et al.*, 2008). It was also observed that the inoculation of tripartite microbial organisms influenced to higher biomass than bipartite and individual microbial organism.

The synergistic interaction effect of AMF+PSB+*Azotobacter* rendered higher benefit to seedlings rather competing by way of higher N fixation, solubilization of insoluble P and facilitated higher uptake of nutrients to seedling. These findings substantiate the reports of several researchers (Zaidi, 2003, Roy and Srivastava, 2011, Raman, 2012). The dual inoculated plants also had positive effects on

biomass and nutrients uptake as compared to single microbial inoculated plants and non inoculated control plants. *A. indica* was the most dependent tree for juvenile growth to different microbial inoculants ranged 12.31 to 72.13 percent.

However the plant depended maximum 72.13 percent on AMF + PSB + *Azotobacter* followed by AMF + *Azotobacter* (60 percent) and PSB + *Azotobacter* (45.90 percent) respectively (Figure 2).

Table 1. Effect of different microorganisms on quality parameters of *A. indica* seedlings. The data represent ± Standard Error (n=20).

Treatments	Dickson Quality Index	Seedling Vigour index	Root Shoot Ratio
T0	0.76±0.03	32.30±3.22	0.213±0.006
T1	0.93±0.04	52.45±3.50	0.550±0.004
T2	0.88±0.01	46.80± 1.78	0.550±0.010
T3	0.89±0.03	41.50±2.50	0.495±0.004
T4	1.55±0.05	62.00±3.00	0.620±0.006
T5	1.70±0.02	67.00±2.20	0.670±0.009
T6	1.66±0.05	60.25±1.30	0.560±0.006
T7	2.03±0.06	89.35±2.80	0.780±0.010

Table 2. Nutrient contents in *A. indica* seedlings as influenced by microbial inoculants.

Treatments	N (%)	P (mg kg ⁻¹)	K (mg kg ⁻¹)
T0	0.152±0.005	10.60±2.50	137.12±6.50
T1	0.182±0.004	20.20±3.00	179.97±5.56
T2	0.176±0.004	19.82±0.55	171.40±6.35
T3	0.193±0.002	19.18±0.40	175.69±4.30
T4	0.209±0.00	20.10±0.80	257.10±9.00
	4		
T5	0.210±0.003	21.20±0.28	287.10±6.50
T6	0.201±0.004	19.90±0.12	201.40±6.50
T7	0.216±0.005	21.78±0.11	325.60±6.11

The data represent ± Standard Error (n=5).

Effect on AMF development in roots: The mycorrhizal development in root observed higher in

inoculated seedlings than non inoculated seedling. The percentage AMF root infection was highest with AMF+*Azotobacter* followed in seedlings with AMF+PSB+*Azotobacter* (Figure 3).

The rate of AMF colonization was statistically at par for above treatments however it was lower as compared to bipartite inoculated seedling might be due the higher availability of soil P due to *Pseudomonas strait* as also profounded by other researchers (Smith and Smith, 2008, Ren *et al.*, 2012)

that the increasing status of soil P limit the development of AMF in root.

The AMF inoculated seedling exhibited maximum of 87.87 percent improvement with AMF+*Azotobacter* followed with AMF+PSB+*Azotobacter* and AMF+PSB. AMF inoculated seedling exhibited 53 percent root infection as compared to uninoculated seedlings (7.5 percent) due to the presence of indigenous endophytes.

Table 3. Nutrient contents in *A.indica* seedlings influenced by microbial inoculants.

Treatments	Nutrient content (µg dry weight g-1)				
	Fe	Mg	Mn	Cu	Zn
T0	6.56±0.10	15.60±0.04	0.72±0.01	0.089±0.001	0.090±0.003
T1	8.53±0.30	16.16±0.05	0.85±0.02	0.110±0.003	0.110±0.003
T2	7.80±0.07	15.80±0.05	0.76±0.02	0.104±0.002	0.090±0.002
T3	7.91±0.22	16.54±0.08	0.85±0.03	0.108±0.001	0.102±0.002
T4	8.25±0.07	16.60±0.01	0.86±0.01	0.100±0.003	0.101±0.003
T5	9.66±0.13	16.10±0.05	0.92±0.01	0.121±0.002	0.101±0.003
T6	7.45±0.05	16.01±0.02	0.83±0.02	0.081±0.002	0.170±0.003
T7	10.71±0.11	16.74±0.10	0.98±0.02	0.121±0.003	0.180±0.002

The data represent ± Standard Error (n=5).

Effect on Nutrients uptake

The nutrients contents of major and micro nutrients in seedlings were influenced significantly by interactive potential of microbial inoculants compared to the non-inoculated seedlings (Table 2). Seedlings inoculated with tripartite of AMF+PSB+*Azotobacter* exhibited highest improvement in N (42.0%), P (105%) and K (137%) contents followed by AMF+ *Azotobacter* (N 38.15%, P 100 % and K 109.37%) over uninoculated seedlings. The increment in NPK contents by AMF alone found 19.7%, 90.56% and 31.35% respectively. The inoculation of AMF and its interaction with other microbial inoculants was proved to be most effective in nutrients uptake in *A. indica* seedling in the present study. The higher nutrient content in inoculated seedlings might be due the increased extra N fixation by *Azotobacter*, higher P availability

(Raman, 2012) and mobility of nutrients via mycelium network of the AMF resulted in to better nutrient and water uptake as also reported by Li *et al.* (2009), Kahiluoto *et al.* (2012). Similarly, the Fe content increased maximum of 63.26%, Mn (36.11%), Cu (35.95%) and Zn (100%) in seedlings inoculated with AMF+PSB+*Azotobacter* over uninoculated seedlings (Table 3). The AMF inoculated seedling developed strong symbiosis between host and fungus resulted in to mycorrhizal roots take up nutrients more efficiently than uncolonized root system. Similar findings were also reported by Smith and Read (2008), Kahiluoto *et al.* (2012).

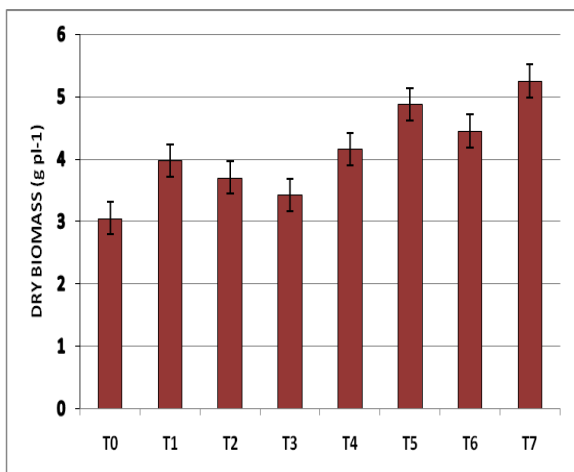


Fig. 1. Effects of AMF,PSB and *Azotobacter* on seedling biomass of *A. indica* seedlings under nursery condition.

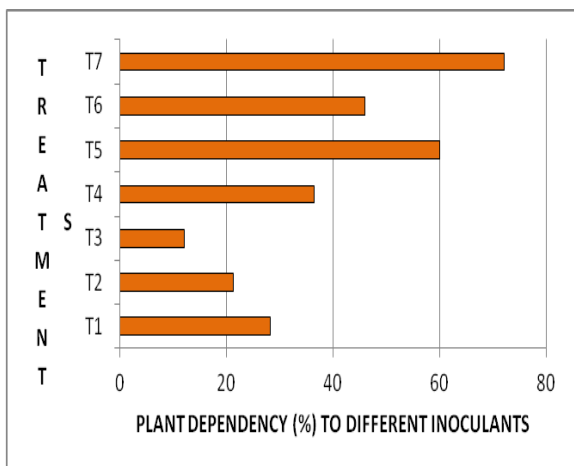


Fig. 2. Percentage Dependency of *A. indica* on different microorganisms under nursery condition.

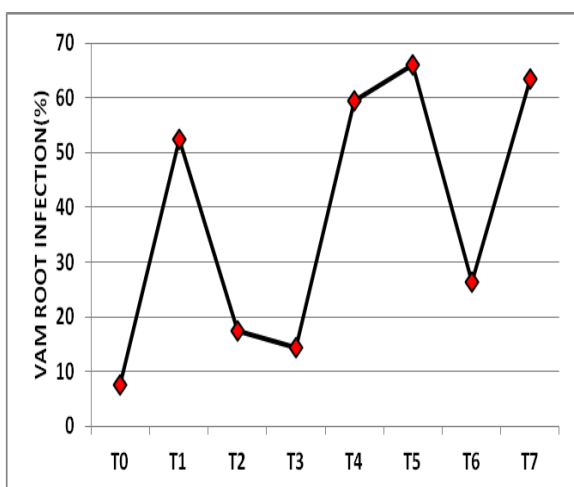


Fig. 3. AMF development in roots of *A. indica* as influenced by AMF, PSB and *Azotobacter* inoculation.

Conclusion

The inoculation of tripartite microbial inoculants proved to be most effective in production of quality nursery stock with superior vigour and higher root shoot ratio. These seedlings attend good height with higher biomass due to more efficient uptake of soil nutrients due to AMF root development. *Azotobacter* and PSB also help seedling in mediating higher N and P availability in rhizosphere which the seedling uptake readily. The seedlings treated with bipartite and individual microbial organisms also found better as compared to uninoculated seedlings. Thus the present finding can be utilized in present production system for higher survival rate of plantations.

References

Ambe AG. 2001. Les fruitiers squvages comestibles des savanes guineannes de cote dilvoire etate des connssances par une population locale les malinke. Biotechnology, Agronomical Society and Environment **5(1)**, 43-58.

Bramhachari G. 2004. Neem: A omnipotent plant: A retrospection. Chemical Biochemistry, **5**,408-421.

Chen Y, Yuan J, Yang Z, Xin G, Fan L. 2008. Associations between arbuscular mycorrhizal fungi and *rhynchrelyrum repens* in abandoned quarries in southern China. Plant and Soil **304**,257-266.

Dickson A, Leaf A, Honer, JF. 1960. Quality appraisal of white spruce and White pine seedling stock in nurseries. Forestry Chronicle, **36**,10-13.

Hassan HM, Marschner P, MCNeill A, Tang C. 2012. Growth, P uptake in grain legumes and changes in rhizosphere soil P pools. Boilogy and Fertility of Soil. **48**,151-159.

Jackson ML. 1973. Soil Chemical Analysis. Prentice Hall of India, New Delhi.

John MK. 1970. Colorimetric determination of phosphorus in soil and plant materials with ascorbic acid. *Soil Science* **100**,214-220.

Kahiluoto, Helena, Ketoja E, Vestberg, Mauritz. 2012. Plant available P supply is not the main factor determining the benefit from arbuscular mycorrhiza to crop P nutrition and growth in contrasting cropping systems. *Plant and Soil* **350**, 85-98.

Khalafallah AA, Abo-Ghalia HH. 2008. Effect of arbuscular mycorrhizal fungi on the metabolic products and activity of antioxidant system in wheat plants subjected to short term water stress, followed by recovery at different growth stages. *Journal of Applied Science and Research* **4(5)**,559-569.

Lax P, Becerra AG, Soterias F, Cabello M, Doucet ME. 2011. Effect of the arbuscular mycorrhizal fungus *Glomus interaradices* on the false root knot nematode, *Nacobbus aberrans* in tomato plants. *Biology and Fertility of Soil* **47**,591-598.

Li YF, Ran W, Zhang RP, Sun SB, Xu GH. 2009. Facilitated legume nodulation, phosphate uptake and nitrogen transfer by arbuscular inoculation in an upland rice and mung bean intercropping system. *Plant and Soil* **315**, 285-296.

Miransari, M. 2011. Interactions between AMF and soil bacteria. *Applied Journal of Microbiology and Biotechnology* **89(4)**, 917-930.

Phillip JM, Hayman DS. 1970. Improved procedures for clearing roots and staining parasitic and VAM fungi for rapid assessment of infection. *Transaction of British Mycological Society* **53**,158-160.

Pindi PK. 2011. Mycorrhizal association of some agroforestry tree species in two social forestry nurseries. *African Journal of Biotechnology* **10(51)**,10425-10430.

Plenchette C, Fortin JA, Furlan V. 1983. Growth responses of several plant species to mycorrhizae in a soil of moderate P fertility. I. Mycorrhizal dependency under field conditions. *Plant and Soil* **70**,199-209.

Qualis M, Copper CS. 1968. Germination, growth and respiration rates of birdfoot trefoil at three temperatures during the early non photosynthetic stage of development. *Crop Science* **8**, 758-760.

Querejeta JI, Barea JM, Allen MF, Caravac F, Roldan A. 2003. Differential response of a C13 and water use efficiency to arbuscular mycorrhizal infection in two arid land woody plant species. *Oecologia* **135**,510-515.

Raman J. 2012. Response of *Azotobacter*, *Pseudomonas* and *Trichoderma* on growth of Apple seedlings. IPCBEF, Vol. 40. International Conference on Biological and Life Sciences, Singapore, pg. 83-90.

Lixuan R, Yunsheng L, Ning Z. 2012. Role of phosphorus network in carbon and phosphorus transfer between plants. *Biology and Fertility of Soil*. Published online 01 May 2012.

Roy ML, Srivastava RC. 2011. Plant growth promotion potential of *Azotobacter chroococcum* on growth, biomass, leaf area index and yield parameter of Amon rice in Tripura. *Indian Journal Agricultural Research* **45(1)**, 52-58.

Smith SE, Read DJ. 2008. *Mycorrhizal Symbiosis*, Academic Press London.

Zaidi A. 2003. Interactive effect of rhizotrophic microorganisms on yield and nutrient uptake of Chickpea (*Cicer arietinum* L.). *European Journal of Agronomy* **19(1)**, 15-21.