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Suspended algal communities in high altitude rice wetlands of Apatani Plateau in Eastern Himalaya

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Abstract

The suspended algal life forms (SALF) were studied in the high altitude rice fields (WRF) covering five randomly selected sites of Apatani Plateau in Arunachal Pradesh, India. The samples were collected fortnightly from WRF in three inundated phases *viz.* water accumulation (April- May), flooding (June-July) and early recession (August –September) during the rice growing season of 2013. A total of 62 taxa of SALF belonging to the classes Chlorophyceae, Bacillariophyceae and Cyanophyceae were recorded. *Closterium* sp., *Spondylosium* sp., *Scenedesmus* sp., *Ulothrix* sp., *Zygnema* sp., *Spirogyra* sp., *Docidium* sp. of Chlorophyceae, *Navicula* sp., *Pinnularia* sp., *Rhizosolenia* sp., *Tabellaria* sp. of Bacillariophyceae and *Oscillatoria* sp., *Nostoc* sp., *Phormidium* sp. of Cyanophyceae were in high abundance in WRF. The study also revealed that Chlorophyceae was the most dominant and diverse class of SALF followed by Bacillariophyceae and Cyanophyceae which was ensued for concurrent existence of rice stems as the affixing substrate of attached algal life forms (AALF) in WRF.

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Introduction

Wet rice fields (WRF) form sufficiently rich natural habitat for diversified aquatic organisms (Heckman, 1979; Fernando, 1993). Generally, WRF remain flooded temporarily for about 4-5 months in a year due to accumulation of monsoon water (Fernando, 1996; Saikia and Das, 2004; Mondal *et al.*, 2005). Therefore, by forming a shallow marsh like environment, it becomes highly productive and provides suitable ground for growth of various producers (Fernando, 1993). In general, such rice fields practically show four phases *viz.*, the water accumulation, flooding, recession and drying phases in the context of water depth and rainfall patterns. The algal communities in a typical WRF depict two life forms either as 'phytoplankton' being suspended in water or as 'periphyton' being attached with submerged rice stems (Hansson, 1988; Stevenson, 1996; Das *et al.*, 2007). Hence, being free floating in the water, the suspended algal life forms (SALF) have first access to light, whereas the attached algal life forms (AALF) have the first access to nutrients from the field waters (Wetzel, 1979).

Thus, resource competition between these two life-forms regulates the structure of algal assemblages in shallow WRF (Smith and Kalff, 1983; Hansson, 1988). Moreover, SALF plays the pivotal role in maintaining trophic structure and providing food sources to zooplankton and fishes in WRF (Tas and Gonulol, 2007; Pawar and Sonawane, 2011).

The reviews of literature (Das *et al.*, 2011; Hosmani and Mruthunjaya, 2012) indicated that Chlorophyceae and Cyanophyceae are the most dominant classes compared to Bacillariophyceae within the SALF of WRF and lakes. On the other hand, Senthilkumar and Sivakumar (2008), Bahaar and Bhatt (2011) reported that the class Bacillariophyceae is dominant over Chlorophyceae in paddy fields. Various authors also observed that Chlorophyceae is higher in total counts than the other taxa of planktonic algae in rice fields, ponds, streams and lakes (Ayoade, 2009; Balasingh, 2010; Singh,

2011; Negi and Rajput, 2011; Jemi and Balasingh, 2011; Ssanyu *et al.*, 2011; Nwezei and Ude, 2013). However, perennial and deep water wetlands having less submerged vegetations or substrates hardly possess the scope of assemblages of SALF compared to the transparent shallow water rice field which allow sufficient lights to strike the bottom of the field.

The benthic algae along with AALF colonize on rice stems being suitable substrate for their growth in WRF (Lucinska, 1979; Moss, 1998; Stanley *et al.*, 2003; Saikia and Das, 2009). Accordingly, Baba *et al.* (2011), Reza and Arash (2012), and Duncka *et al.* (2013) confirmed that Bacillariophyceae was the most dominant class among AALF in Sindh river of Kashmir valley and in some wetlands of Brazil and Iran. However, the information regarding the occurrences and richness of SALF particularly in mountain WRF are very meager till date.

With this backdrop, an investigation was carried out in mountain WRF during water accumulation phase (WAP), flooding phase (FP) and early recession phase (ERP) of the rice growing season, 2013. The standard sampling method was adopted for enumeration of SALF from the field water. This article is thus intended to discuss on the richness and diversity of algae into field water as well to appraise the reasons behind their occurrences and variations in such a seasonal ecosystem.

Materials and methods

Study site

The study was carried out in high altitude wet rice fields of Apatani plateau, located in Lower Subansiri district of Arunachal Pradesh, India (Fig. 1). Geographically, the area was located at 26°50' - 98°21' N latitude and 92°40' - 94°21' E longitude and with altitude about 5000 ft. above mean sea level. A total of 15 rice fields were randomly selected from the five villages namely, Mudang tage, Dutta, Nenchalya, Pine grobe and Tajang. These high altitude flooded rice fields have an average water depth of 16-20 cm under emergent rice canopy.

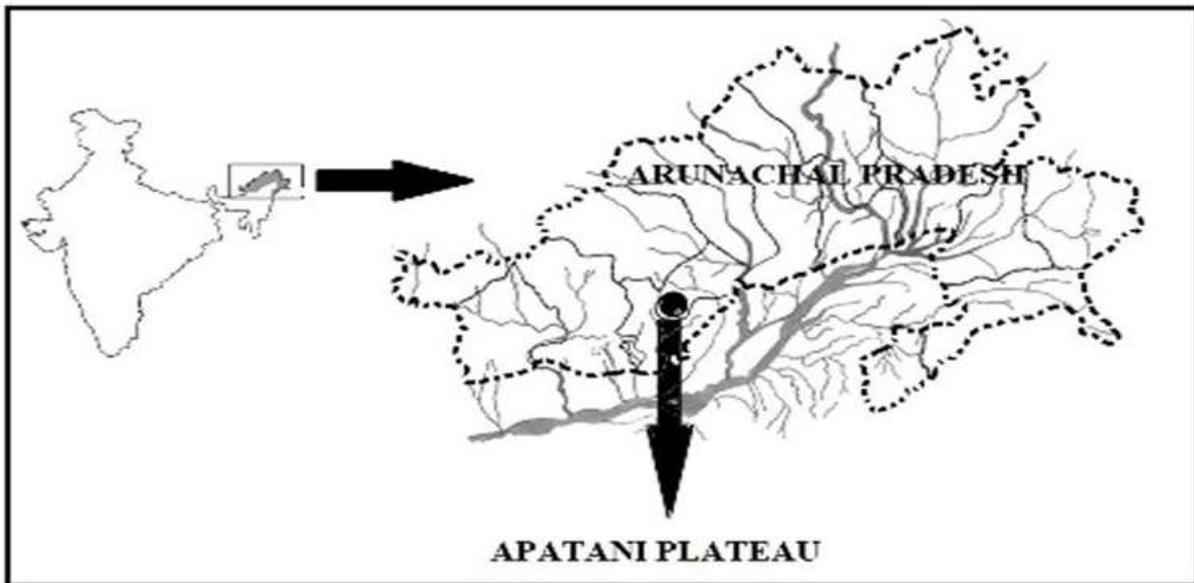


Fig. 1. Location map of sampling site in Arunachal Pradesh.

Analysis of SALF and water qualities

The SALF samples were collected fortnightly by sieving 25 L of rice field water through plankton net made up of bolting silk (mesh size 60 μ m) starting from the month of WAP till to ERP. The samples were preserved in 4% neutralized formalin and were further analyzed both qualitatively and quantitatively under a binocular light microscope (Nikon, ECLIPSE E200) using 10X and 40X objectives respectively. The quantitative analyses of SALF were done with the help of drop count method (Lackey, 1938) and values were expressed as individual per cent litre. Standard identification keys and monographs were used for confirmation of genera or species level identity of individuals (Belcher and Swale, 1976; Turner, 1978; Tonapi, 1980; Penticost, 1984; Desikachary, 1989; Edmonson, 1992). Physico-chemical parameters of field water like alkalinity, dissolved oxygen (DO), biological oxygen demand (BOD), free dissolved carbon di oxide (FDCO₂), chloride, Ca hardness (CaH), total hardness (TH), nitrate - Nitrogen and phosphate - Phosphorus were also analyzed (APHA, 2012) from each of the study fields. The water temperature (WT), water depth (except refuge trenches), pH, specific conductivity (SC) and total dissolved solid (TDS) were recorded on spot at each field trip using standard equipment like thermometer (ZPHI-9100 Zico), centimeter scale, digital water

analyzer (Systronics 371) respectively. Besides, random sampling for estimating rice stand densities were undertaken following quadrat count method (Catling, 1987).

Statistical analysis

The variation in total SALF was estimated by two-way ANOVA assuming classes and phases as two factors. Shannon and Simpson indices were employed for estimating diversity, while evenness and species richness were computed using Buzas Gibson evenness index and Margelef index respectively. To compare the diversities of different communities the estimated diversity indices were subjected to pair-wise permutation tests in two ways (i.e. within class between seasons and between classes within season). All the analyses were performed using PAST software, 3.02 version.

Results

The high precipitation of actual rainfall data of 2013 was peak in the month of May and August during the rice growing period. Similarly, the average rainfall pattern of last ten years (2002-12) of the plateau area depicted the high precipitation from April to September in general (**Fig. 2**). Actual rainfall and the pattern of rainfall indicated clearly the constant accumulation of water in rice fields of the study area.

The results unfolded that the stand density ($8.97m^{-2}$) and water depth (3.7 cm) were rising from WAP which attained maximum level of $136.94 m^{-2}$ and 16.9 cm in FP, and again declined a little in the ERP (Fig. 3). A significant positive correlation (Fig. 4) between stand density and SALF ($r=0.998$, $p<0.01$) was evident. Variation of physico-chemical parameters of water (Table 1) were analyzed fortnightly in three different phases of WRF. The mean value of water

physico – chemical parameters like WT, pH, SC, TDS, DO, BOD, Nitrate – N, Phosphate –P, Alkalinity, TH, CaH, FDCO₂, Chloride were ranged $22.00-29.00^{\circ}C$, $6.38-6.62$, $583.27-520.10 \mu Scm^{-1}$, $352.95-292.15 mg L^{-1}$, $11.55-4.32 mg L^{-1}$, $3.38-10.23 mg L^{-1}$, $2.06-2.70 mg L^{-1}$, $0.03-0.21 mg L^{-1}$, $24.85-39.82 mg L^{-1}$, $15.85-29.00 mg L^{-1}$, $9.85-12.03 mg L^{-1}$, $11.67-21.63mg L^{-1}$, $26.92-39.80 mg L^{-1}$ respectively during study period.

Table 1. Average range of water physico – chemical parameters in high altitude rice wetlands of Apatani Plateau during rice growing season.

Parameters	WAP (Mean ± SD)	FP (Mean ± SD)	ERP (Mean ± SD)
WT (°C)	22.37±1.72	27.55±0.70	29.62±1.80
pH	6.38±0.42	6.48±0.21	6.62±0.22
SC (μScm^{-1})	583.27±63.57	557.80±55.49	520.10±89.62
TDS (mg L ⁻¹)	352.95±13.85	292.15±23.01	295.80±33.80
DO (mg L ⁻¹)	11.55±1.12	5.31±1.58	4.32±2.12
BOD (mg L ⁻¹)	10.23±0.89	3.38±0.85	3.46±1.49
Nitrate-N (mg L ⁻¹)	2.70±0.66	2.25±1.13	2.06±0.68
Phosphate P (mg L ⁻¹)	0.03±0.02	0.16±0.25	0.21±0.23
Alkalinity (mg L ⁻¹)	24.85±0.39	30.37±7.24	39.82±2.85
TH (mg L ⁻¹)	21.35±6.62	15.85±2.08	29.00±4.59
Ca H (mg L ⁻¹)	11.71±1.24	9.85±0.31	12.03±1.64
Free CO ₂ (mg L ⁻¹)	21.63±1.81	13.53±2.63	11.67±1.90
Chloride (mg L ⁻¹)	26.92±7.80	31.63±14.50	39.80±5.01

The study revealed the presence of total 62 taxa of SALF belonging to three major classes namely, Chlorophyceae (31), Bacillariophyceae (25) and Cyanophyceae (6). It was demonstrated that *Closterium* sp., *Spondylosium* sp., *Scenedesmus* sp., *Ulothrix* sp., *Zygnema* sp., *Spirogyra* sp., *Docidium* sp. under Chlorophyceae, *Navicula* sp., *Pinnularia* sp., *Rhizosolenia* sp., *Tabellaria* sp. under Bacillariophyceae and *Oscillatoria* sp., *Nostoc* sp., *Phormidium* sp. under Cyanophyceae were the dominant genera (Table 2) during the study period. Two-way ANOVA revealed that phases do not have any significant effect ($p>0.05$) on the variation of SALF. Hence, only class-wise total individuals are presented within each phase, where Chlorophyceae always showed the highest values (Fig. 5). The diversity and richness values of the classes were also calculated within each phase. The diversity values showed insignificant results ($p>0.05$, data are not presented) when analyzed between seasons within

class. So, the values between classes within season are presented separately (Table 3).

There was significant variation for all the measured indices ($p<0.0001$), except Buzas Gibson evenness index. In case of WAP and FP, Buzas Gibson evenness was found to be significant in only two instances viz., between Chlorophyceae and Bacillariophyceae, and between Chlorophyceae and Cyanophyceae, where Chlorophyceae always had lower evenness (Table 3). On the other hand, in ERP, significant results were obtained in all the three classes where Chlorophycean algae had the lowest and Cyanophycean algae had the highest evenness value. Margalef index revealed that the species richness was in the order of Chlorophyceae>Bacillariophyceae>Cyanophyceae. Finally, Shannon and Simpson indices precisely explained that Chlorophyceae was the most diverse class, while Cyanophyceae was the least in all the three phases during this study.

Table 2. Individuals of SALF (ind CL⁻¹) in high altitude rice wetlands of Apatani Plateau during rice growing season.

Sl. No.	Cholorophyceae	WAP	FP	ERP
1.	<i>Arthodesmus curvatus</i>	333.33	555.55	-
2.	<i>Closterium longissima</i>	-	222.22	-
3.	<i>Closterium setacoum</i>	-	111.11	111.11
4.	<i>Closterium</i> sp.	1944.44	3555.58	2111.11
5.	<i>Cosmerium obsulatum</i>	111.11	333.33	-
6.	<i>Cosmerium</i> sp.	-	1477.78	333.33
7.	<i>Docidium sceptrum</i>	-	666.70	666.70
8.	<i>Docidium setigorum</i>	222.22	-	-
9.	<i>Docidium swartzii</i>	972.23	2111.11	111.11
10.	<i>Docidum</i> sp.	6444.45	5233.33	7888.90
11.	<i>Euastrum</i> sp.	-	1666.70	2111.11
12.	<i>Euastrum verrucosum</i>	555.56	222.22	333.34
13.	<i>Gonatozygon reticulatum</i>	-	222.22	-
14.	<i>Gonatozygon</i> sp.	1888.90	2444.44	4222.22
15.	<i>Micrasterias denticulata</i>	333.33	-	-
16.	<i>Micrasteras rotata</i>	-	-	222.22
17.	<i>Micrasterias americana</i>	-	999.99	-
18.	<i>Micrasterias</i> sp.	1444.44	1111.11	1444.44
19.	<i>Microspora</i> sp.	1555.55	333.33	1000.00
20.	<i>Odogonium</i> sp.	666.66	1888.89	2444.45
21.	<i>Spondylosium</i> sp.	2833.33	4777.78	12777.78
22.	<i>Pleurotaenium trabecula</i>	-	111.11	111.11
23.	<i>Scenedesmus</i> sp.	3888.89	1444.44	7333.33
24.	<i>Spirogyra inflata</i>	4222.22	6888.89	4677.78
25.	<i>Spirogyra</i> sp.	14222.22	24111.12	32444.45
26.	<i>Spirotaenia</i> sp.	111.11	-	111.11
27.	<i>Spirotaennia truncata</i>	222.22	777.78	-
28.	<i>Triploceras gracile</i>	111.11	66.66	555.55
29.	<i>Triploceras</i> sp.	1777.78	1922.15	888.89
30.	<i>Ulothrix</i> sp.	6555.11	2666.71	11888.89
31.	<i>Zygnema</i> sp.	7888.89	15555.56	10000.00
Sl. No.	Bacillariophyceae			
1.	<i>Achanthes</i> sp.	444.44	222.22	777.78
2.	<i>Cocconeis</i> sp.	222.22	444.44	222.22
3.	<i>Cymbella angustata</i>	111.11	777.78	-
4.	<i>Cymbella lanceolata</i>	-	555.55	111.11
5.	<i>Cymbella longissima</i>	-	555.55	666.67
6.	<i>Cymbella</i> sp.	444.44	555.55	111.11
7.	<i>Diatoma</i> sp.	444.44	444.44	444.44
8.	<i>Fragillaria</i> sp.	-	444.44	111.11
9.	<i>Gyrosigma</i> sp.	777.77	1000.00	666.67
10.	<i>Navicula lanceolata</i>	222.22	-	-
11.	<i>Navicula placenta</i>	-	555.55	1000.00
12.	<i>Navicula</i> sp.	222.22	2444.44	4333.34
13.	<i>Navicula viridula</i>	333.33	-	111.11
14.	<i>Nitzschia</i> sp.	333.33	222.22	-
15.	<i>Pinnularia brevistriata</i>	-	111.11	-
16.	<i>Pinnularia vulgaris</i>	1277.77	6222.23	5666.65
17.	<i>Rhicosphenia</i> sp.	-	-	333.33
18.	<i>Rhizosolenia</i> sp.	1333.33	888.88	1555.55
19.	<i>Rhopalodia</i> sp.	-	222.22	-
20.	<i>Stauroneis</i> sp. <i>phoneiocentron</i>	-	111.11	-
21.	<i>Tabellaria</i> sp.	1333.33	3333.34	888.89
22.	<i>Penium</i> sp.	2555.56	2111.11	9666.66
23.	<i>Netrium digitus</i>	-	222.22	-
24.	<i>Netrium</i> sp.	933.33	666.70	777.80
25.	<i>Mesotaenium</i> sp	999.99	2111.11	5333.33
Sl. No.	Cyanophyceae			
1	<i>Anabaena</i> sp.	333.33	555.55	2000.00
2	<i>Cylindrospermum</i> sp.	444.44	222.22	-
3	<i>Nostoc</i> sp.	2000.00	555.55	1666.67
4	<i>Oscillatoria</i> sp.	888.88	999.99	888.89
5	<i>Phormidium</i> sp.	1777.78	4911.11	5333.33
6	<i>Spirulina</i> sp.	555.55	333.33	999.99

Discussion

In WRF the occurrence and diversity of the SALF are regulated by complex interactions of multiple factors

(Heckman, 1979; Dalkiran and Dere, 2006). The environment itself possesses location specific variability of nutrient concentrations, water

conditions, rainfall patterns and influx of light intensities along with physical surroundings of the locations (Fonge *et al.*, 2012; Kumar and Sahu, 2012). The average ranges of water physico-chemical qualities obtained were suitable for the growth of SALF in high altitude WRF (Saikia and Das, 2010). After field preparation, the decomposition of organic matters may influence the water quality with higher value of FDCO₂, pH, and SC. Increased mean value of

pH in ERP might have caused better growth of certain genera of SALF (Thirugnanamoorthy and Selvaraju, 2009). According to Saikia and Das (2004) location specific high rainfall plays an important role for maintaining the water depth of the WRF of Apatani plateau. The gradual increase of water depth from 3.0-5.0 cm at the WAP up to 20.0 cm or more at FP is favorable for concurrent rice- fish culture (Hora and Pillay, 1962; Saikia and Das, 2010).

Table 3. Diversity indices between three classes of SALF within in high altitude rice wetlands of Apatani Plateau during rice growing season.

Estimated Indices	WAP			FP			ERP		
	Chlorophyceae	Bacillariophyceae	Cyanophyceae	Chlorophyceae	Bacillariophyceae	Cyanophyceae	Chlorophyceae	Bacillariophyceae	Cyanophyceae
Shannon	2.67	2.31	1.59	2.57	2.25	1.17	2.48	2.09	1.37
Simpson	0.89	0.88	0.76	0.87	0.83	0.55	0.88	0.82	0.69
Bujaz Gibson evenness	0.53	0.78	0.81	0.42	0.52	0.53	0.44	0.54	0.79
Margelef	2.96	1.81	0.78	3.31	2.25	0.75	2.77	1.87	0.57

The simultaneous rise of substrate density with more surface area having AALF depressed the growth of SALF by reducing the outflow of dissolved nutrients from the mineralization zone of the substrate surface (Das *et al.*, 2007). On the other hand, SALF reduced

the growth of AALF by diminishing the amount of light reaching to the substrate where light and the nutrient competition were the major factors for the growth of SALF and AALF (Hansson, 1988).

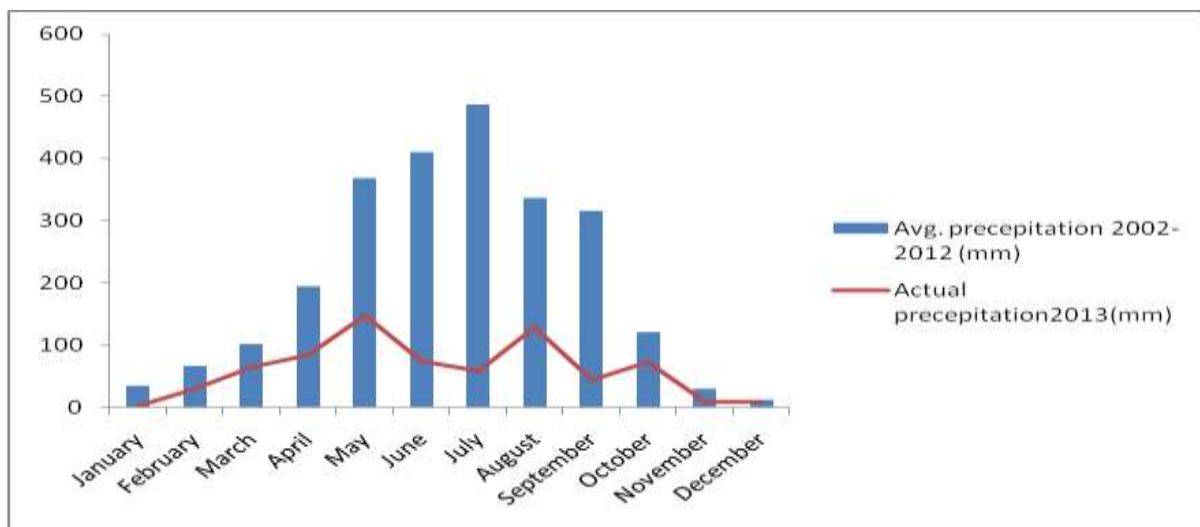


Fig. 2. Average rainfall pattern in last ten years (2002-12) and actual rainfall pattern in high altitude wet rice fields of eastern Himalaya during rice growing season, 2013

Interestingly, among AALF Bacillariophyceae was more dominant on submerged rice because of getting sufficient surface area for growth in shallow WRF. Moreover, high amount of silica in the rice stems

influenced the growth of Bacillariophyceae (Lukaw *et al.*, 2012) because it was essential nutrient for the formation of frustules. Morphological and physiological nature of rice stand might have also

influenced the development of AALF, as the submerged rice had the ability to secrete mucilage to form stalks or mucilaginous matrices, allowing attachment to substrates (Fritsch, 1945; Round,

1991). However, in suspended condition, algal communities have shown comparatively less population where Chlorophyceae was the most dominant as well as diverse class (Das *et al.*, 2007).

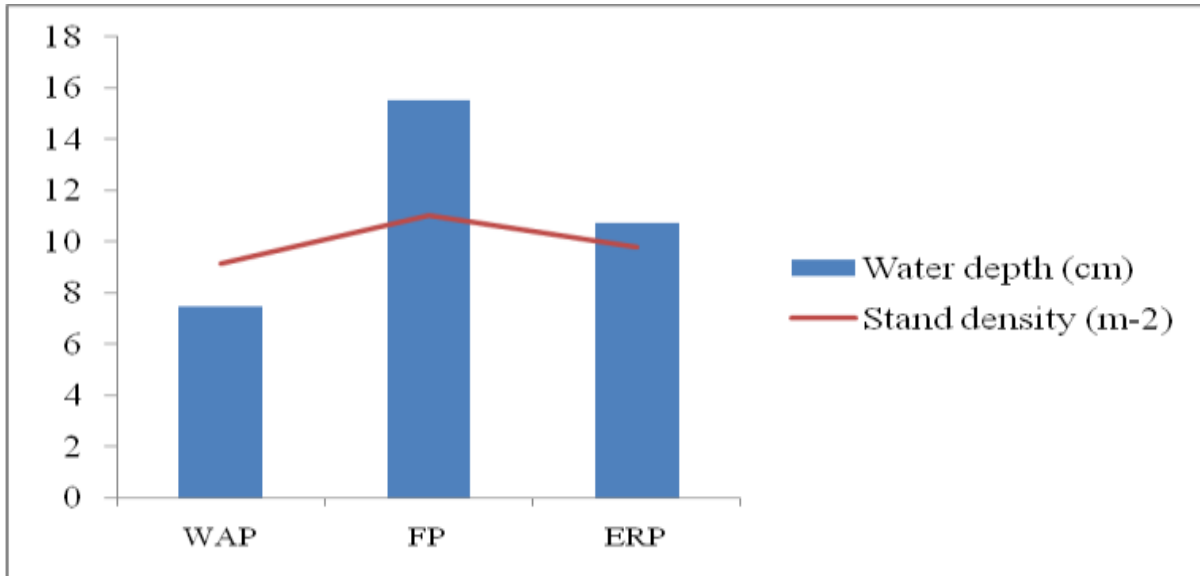


Fig. 3. Variation of water depth and stand density in wet rice fields of eastern Himalaya during rice growing season, 2013.

This phenomenon could be explained by the fact that, better habitat conditions like slightly acidic nature of water with shallow depth and greater quantity of dissolved organic matters particularly the excreta of pig and other domestic animals might have

influenced the higher diversity of Chlorophyceae in all the three phases (Saravanakumar *et al.*, 2008; Thirugnanamoorthy and Selvaraju, 2009; Ponce *et al.*, 2010; Mustapha, 2010; Nweze and Ude, 2013).

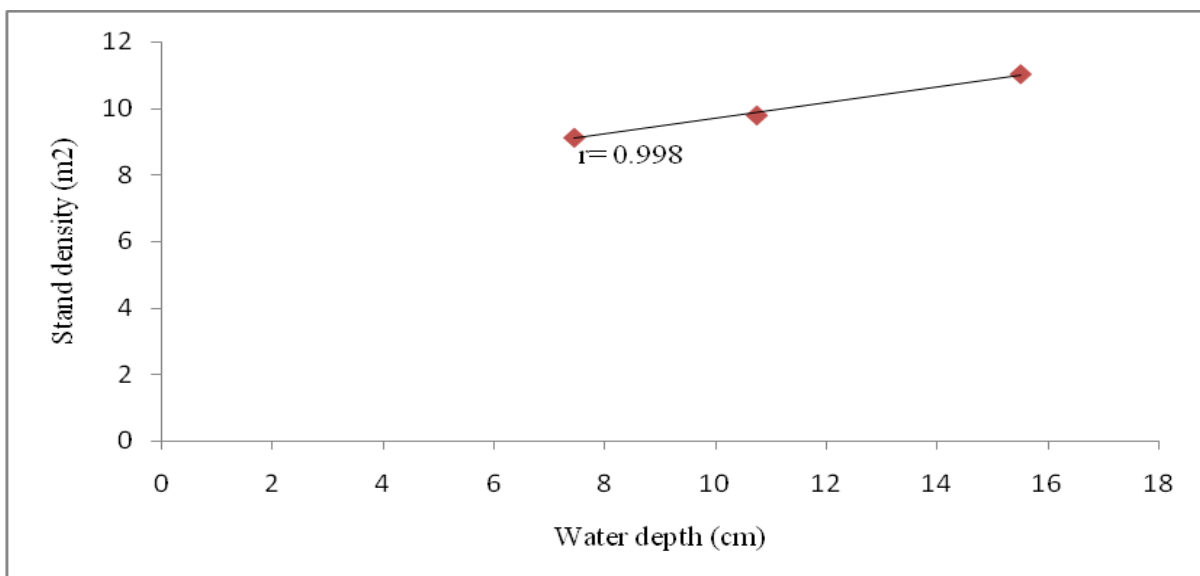


Fig. 4. Relationship between stand density and water depth in high altitude rice wetlands of Apatani Plateau during rice growing season.

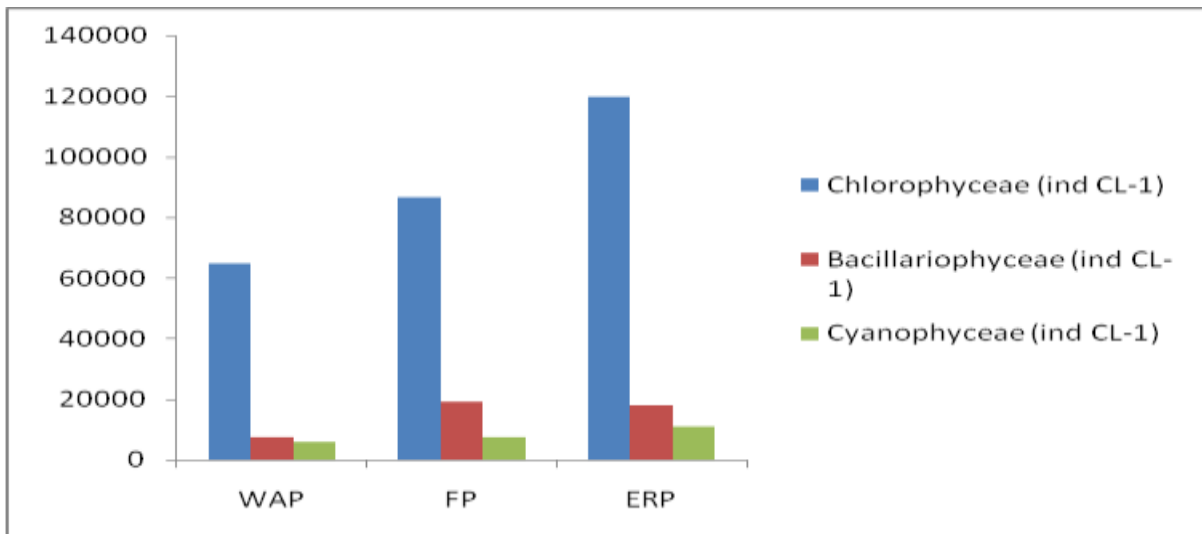


Fig. 5. Variations among the three major classes of SALF in high altitude wet rice fields of eastern Himalaya during rice growing season, 2013.

Conclusion

From the above study it can be concluded that in suspended forms, shallow rice wetlands have the highest dominance and diversity of Chlorophycean algae. But the concurrent presence of rice stems might have caused anchorage of various algal life forms reducing the counts of suspended population in the field water. So, it may be considered that the less diversity of Bacillariophyceae compared to Chlorophyceae among SALF probably resulted because of available additional surface area leading to spontaneous assemblages on the submerged portion of rice stems.

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References

APHA. 2012. Standard methods for the examination

of water and waste water. San Francisco, USA: American Public Health Association.

Ayoade AA. 2009. Changes in physico-chemical features and plankton of two regulated high altitude rivers Garhwal Himalaya, India. *European Journal of Scientific Research* **27**, 77-92.

Baba AI, Sofi, Bhatt SU, Pandit AK. 2011. Periphytic algae of river Sindh in the Sonamarg Area of Kashmir Valley. *Journal of Phytology* **3**, 1-12.

Bahaar SWN, Bhatt GA. 2011. Aquatic biodiversity in the paddy fields of Kashmir Valley (J and K) India. *Asian Journal of Agricultural Research* **5**, 269-276. <http://dx.doi.org/10.3923/ajar.2011.269.276>

Balsingh GS. 2010. Studies on Phytoplankton diversity and seasonal abundance of a perennial pond in Kanyakumari – Tamilnadu, India. *Journal of basic and applied biology* **4**, 188-193.

Belcher H, Swale E. 1976. A beginner's guide to freshwater algae. Her Majesty's stationary office, London.

Catling HD. 1987. Pest management field methods in deepwater rice. Calcutta: International Rice

Research Institute, Government of India.

Dalkiran N, Dere S. 2006. Factors Affecting the phytoplankton diversity and richness in a shallow eutrophic lake in Turkey. *Journal of Freshwater Ecology* **21**, 575-581.

<http://dx.doi.org/10.1080/02705060.2006.9664118>

Das DN, Saikia SK, Das AK. 2007. Periphyton in rice–fish culture system: A case study from Arunachal Pradesh, India. *Renewable Agriculture and Food Systems* **22**, 316–319.

<http://dx.doi.org/10.1017/S1742170507001901>

Das DR, Haque MR, Chowdhary, BBP, Haque, MA, Alam MN. 2011. Study on monthly variations of plankton in relation to the physico-chemical condition of rice-fish fields in boro season. *International Journal Sustainable crop production* **6**, 43-49.

Desikachary TV. 1989. Atlas of Diatom. Madras: Madras Science foundation.

Duncka B, Nogueirab IS, Felisbertob SA. 2013. Distribution of periphytic algae in wetlands (Palm swamps, *Cerrado*) Brazil. *Brazilian Journal Biology* **73**, 331-346.

<http://dx.doi.org/10.1590/S151969842013000200013>.

Edmonson WT. 1992. *Freshwater Biology*. New Delhi: International books and periodicals supply service.

Fernando CH. 1993. Rice field ecology and fish culture an overview. *Hydrobiologia* **259**, 91-113.

Fernando CH. 1996. Perspectives in Asian fisheries. In de Silva, S. S. (ed) *Ecology of rice fields & its bearing o fisheries & fish culture*, 1st edn. Asian fisheries society, Manila, 217-237 p.

Fonge BA, Tening AS, Egbe EA, Yinda GS,

Fongod AN, Achu RM. 2012. Phytoplankton diversity and abundance in Ndop wetland plain, Cameroon. *African Journal of Environmental. Science and Technology* **6**, 247-257.

<http://dx.doi.org/10.5897/AJEST12.025>

Fritsch FE. 1945. *The Structure and Reproduction of the Algae*. Cambridge University Press, Cambridge.

Hansson LA. 1988. Effects of competitive interactions on the biomass development of planktonic and periphytic algae in lakes. *Limnology and Oceanography* **1**, 121-128.

Heckman CW. 1979. *Rice field ecology in Northeastern Thailand*. London: Monographiae Biologicae, Dr W. Junk Publishers.

Hora SL, Pilley TVR. 1962. *Handbook on fish culture in the Indo pacific region*. FAO, Rome.

Hosmani S, Mruthunjaya TB. 2012. Distribution of phytoplankton in lakes of Tirumalkudal Narasipura of Mysore district. *Journal of Research in Science & Technology* **1**, 28- 32.

Jemi RJ, Balasingh GS. 2011. Seasonal variations of phytoplankton in the freshwater temple pond of Munchirai, Vilavncode Taluk, Kanyakumari district. *Journal of Basic & Applied Biology* **5**, 94-99.

Kumar A, Sahu R. 2012. Diversity of Algae (Cholorophyceae) in paddy fields of Lalgutwa area, Ranchi, Jharkhand. *Journal of Applied Pharmaceutical Science* **2**, 92-95.

<http://dx.doi.org/10.7324/JAPS.2012.21116>

Lackey JB. 1938. The manipulation and counting of river plankton and changes in the some organisms due to formalin preservation. *Public health reports* **53**, 2080-2098.

Lucinska M. 1979. Structure of epiphytic algal communities of the Lobelia lakes in the region of BoryTucholskie (Poland). *Algological Studies* **24**,

324–338.

Lukaw YS, Ladu JLC, Kenyi D. 2012. Seasonal influence of physicochemical variables on phytoplankton abundance in Jebel Aulia reservoir KhartovmSudan. *Nature and Science* **10**, 168-175.

Mondal MR, Dewan S, Hossain MA, Asaduzzaman M, Islam MA, Rozario UA. 2005. Food and feeding habits of *Puntius gonionotus* (Thai Sarpunti) in rice field. *Pakistan Journal of Biological Sciences* 386-395.

<http://dx.doi.org/10.3923/pjbs.2005.380.395>.

Moss B. 1998. Ecology of freshwater, man and medium, past to future. Wiley-Blackwell, Oxford.

Mustapha MK. 2010. Seasonal influence of limnological variation on plankton dynamics of a small shallow, tropical reservoir. *Asian journal of experimental Biological Science* **1**, 60-79.

Negi RK, Rajput A. 2011. Diversity of phytoplankton in the freshwater streams of Kumaon Himalaya of Uttarakhand State. *The Ecoscan* **5**, 15-19.

Nweze NO, Ude BO. 2013. Algae and physico-chemical characteristics of Adani rice field, Enugu State, Nigeria. *Journal of Pharmacy and Biological Sciences* **8**, 12-18.

Pawar M, Sonawane SR. 2011. Diversity of phytoplankton from three water bodies of Satara district (M.S.) India. *International Journal of Biosciences* **1**, 81-87.

Penticost A. 1984. Introduction to freshwater algae. England: Richmond publishing com. Limited.

Ponce JT, Arredondo JL, Castillo-Vargasmachuca SG, Chávez GR, Valle AB, Regalado de Dios MA, Carrillo FM, Villalobos RN, Gurrola JAG, Lugo PL. 2010. The effect of chemical and organic fertilization onphytoplankton

and fish production in carp (Cyprinidae) polyculture system. *Revista Biociencias* **1**, 44-50.

Reza RG, Arash K. 2012. Attached and epipellic algae of the Miankaleh International Wetland (Northern Iran). *Journal of sciences spring, Environmental spring* **9**, 61-72.

Round FE. 1991. Diatoms in River water monitoring studies. *Journal of Applied Phycology* **3**, 129-145.

Saikia SK, Das DN. 2004. 'Aji gnui assonii' – a practice of organic hill farming among the Apatani tribe of Eastern Himalaya. *International Journal of Sustainable Development & World Ecology* **11**, 211-217.

<http://dx.doi.org/10.1080/13504500409469825>

Saikia SK, Das DN. 2009. Feeding ecology of common carp (*Cyprinus carpio* L.) in a rice–fish culture system of the Apatani Plateau (Arunachal Pradesh, India). *Aquatic Ecology* **43**, 559–568.

<http://dx.doi.org/10.1007/s10452-008-9174>

Saikia SK, Das DN. 2010. Ecology of terrace wet rice-fish environment and role of periphyton. *Journal of Wetlands Ecology* **4**, 102-111.

Saravanakumar A, Rajkumar M, Thivakaran GA, Serebiah JS. 2008. Abundance and seasonal variations of phytoplankton in the creek waters of western mangrove of Kachchh-Gujarat. *Journal of Environmental Biology* **29**, 271-274.

Senthilkumar R, Sivakumar K. 2008. Studies on phytoplankton diversity in response to abiotic factors in Veeranam Lake in the Cuddalore district of Tamil Nadu. *Journal of Environmental Biology* **29**, 747-752.

Singh M. 2011. Study of plankton abundance in freshwater fish pond at Malawar, Etah (U. P.). *Indian Journal of biological studies and research* **1**, 39-44.

- Smith R, Kalff J.** 1983. Competition for phosphorus among co-occurring freshwater phytoplankton. *Limnology and Oceanography* **28**, 448-464.
- Ssanyu GA, Rasowo J, Auma E, Ndunguru M.** 2011. Evaluation of Plankton Community Structure in Fish Refugia acting as *Oreochromis niloticus* propagation and nursery units for rice/fish trials, Uganda. *Aquaculture Research & Development* **2**, 1-6.
<http://dx.doi.org/10.4172/2155-9546.1000116>
- Stanley EH, Johnson MD, Ward AK.** 2003. Evaluating the influence of macrophytes on algal and bacterial production in multiple habitats of a freshwater wetland. *Limnology and Oceanography* **48**, 1101-1111.
- Stevenson RJ.** 1996. The stimulation and drag of current. In: Stevenson RJ, Bothwell ML and Lowe RL, Eds. *Algal ecology*, 1st edn. New York, USA: Freshwater benthic eco-systems Academic Press, 321-340.
- Tas B, Gonulol A.** 2007. An ecologic and taxonomic study on phytoplankton of a shallow lake, Turkey. *Journal of Environmental Biology* **28**, 439-445.
- Thirugnanamoorthy K, Selvaraju M.** 2009. Phytoplankton diversity in relation to physico-chemical parameters of Gnanaprekasam temple pond of Chidambaram in Tamilnadu, India. *Recent Research in Science and Technology* **1**, 235-238.
- Tonapi GT.** 1980. *Freshwater animals in India*. New Delhi: Oxford and IBH publishing Co. Ltd.
- Turner WB.** 1978. *The fresh water algae (Principally Desmidiaceae) of East India*. Dehradun: M/S Bishen, Mahendra Palsingh 23-A, New Connaught place.
- Wetzel R.** 1979. The role of the littoral zone and detritus in Lake metabolism. *Ergeb Limnologie* **13**, 145-161.