



Antibacterial activity of *Trichosanthes cucumerina* seed lectin and study of its structural stability by fluorescence spectroscopy

Md. Golam Kibria, Md. Rezaul Karim, Imtiaj Hasan, A.K.M. Asaduzzaman, Md. Belal Uddin, Syed Rashel Kabir*

Department of Biochemistry and Molecular Biology, Faculty of Science, Rajshahi University, Rajshahi-6205, Bangladesh

Key words: Fluorescence spectroscopy; bacterial agglutination; bacterial growth inhibition.

<http://dx.doi.org/10.12692/ijb/9.6.187-192>

Article published on December 14, 2016

Abstract

A *Trichosanthes cucumerina* seed lectin (TCSL) was purified previously that showed potent inhibitory effects against Ehrlich ascites carcinoma (EAC) cells *in vivo* in mice. In the present study, the lectin was treated with guanidine-HCl for 2 and 4h in the presence and absence of Ca²⁺ and changes in the tryptophan fluorescence shift were monitored by fluorescence spectroscopy. It was found that the lectin stability was increased in the presence of Ca²⁺. Although the denaturant changed the environment of tryptophan residue, it did not affect the binding sites of TCSL as red blood cells became agglutinated after the treatment with EDTA. Besides the agglutination of three pathogenic bacterial species, the lectin also partially inhibited the growth of *Salmonella enteritidis* and *Staphylococcus aureus*.

*Corresponding Author: Dr. Syed Rashel Kabir ✉ rashelkabar@ru.ac.bd

Introduction

Belonging to the family Cucurbitaceae, Snake gourd (*Trichosanthes cucumerina*) is found in the wild across much of South and Southeast Asia, including India, Bangladesh, Nepal, Pakistan, Sri Lanka, Indonesia, Malaysia, Myanmar (Burma), and southern China (Guangxi and Yunnan). It is an important summer vegetable in Bangladesh, but it may grow throughout the year except in extreme winter (Khatun *et al.*, 2010).

Plant lectins are carbohydrate-binding proteins with different biological properties, such as agglutination, toxicity, anti-proliferation of cancer cells, anti-fungal and anti-bacterial activities (Sitohy *et al.*, 2007; Tian *et al.*, 2008; Liu *et al.*, 2010; Kabir *et al.*, 2011a; Kabir *et al.*, 2011b; Kabir *et al.*, 2013; Kabir and Reza 2014; Rafiq *et al.*, 2013). Previously, a novel lectin was purified from Snake gourd seeds cultivated in Bangladesh with the molecular weight of 56 ± 2 kD containing two sub units 34 ± 1 and 22 ± 1 (Kabir *et al.*, 2012). This lectin was different from the one purified from the Indian Snake gourd seeds (Padma *et al.*, 1999). The lectin showed broad range of pH optima with the heat stability up to 70°C. TCSL agglutinated Human blood (A, B, O & AB), mouse, chicken, cow and some pathogenic bacteria. It showed toxicity against brine shrimp nauplii. The lectin decreased the EAC cells growth 28% and 72% at a dose of 1 and 2 mg/ml concentrations, respectively. Moreover the lectin increased the blood parameters of the mice towards the normal level with a decrease of tumor burden and increased their life span. After observing this antitumor activity of TCSL, its structural stability, bacterial agglutination and antibacterial activity against the pathogenic bacteria was studied in this manuscript.

Material and methods

Purification of TCSL

Lectin was purified from Snake gourd seeds (Kabir *et al.*, 2012) and the protein concentration was determined by Lowry's methods (Lowry *et al.*, 1951) where BSA was used as the standard protein. Agglutination of red blood cells was studied according

to the method described by Kabir *et al.* (Kabir *et al.*, 2012).

Bacterial Agglutination Activity

Listeria monocytogenes, *Salmonella enteritidis*, *Shigella flexneri*, *Staphylococcus aureus*, *Shigella boydii* and *Pseudomonas aeruginosa* were used to examine the bacterial agglutination activities of TCSL. The bacteria were grown at 37°C for 18 h in liquid nutrition medium and centrifuged at 1,027 g for 5 min and washed with 10 mM Tris-HCl buffer saline, pH 7.8. The bacteria were re-suspended in the same buffer with a turbidity of 2.3 at A_{630} . Then TCSL (0.8 mg/ml) was serially diluted with a hemagglutination buffer in the presence and absence of 0.8 mM of lactose and 50 μ l of each bacterial suspension was mixed to a final volume of 100 μ l in a 96-well microtiter plates. The plate was agitated for 2 min and was kept at room temperature for 60 min. Finally, light microscope was used to monitor the bacterial agglutinating activity.

Bacterial Growth Inhibition

In the presence and absence of different concentrations of TCSL (30 μ g/ml - 240 μ g/ml) in bacterial nutrient broth, the bacterial growth inhibition was studied according to the method previously described (Kabir *et al.*, 2011). Three species of bacteria (*Pseudomonas aeruginosa*, *Salmonella enteritidis* and *Staphylococcus aureus*) were used for this study. The bacteria were grown overnight in the nutrient broth at 37°C and the absorbance was adjusted to 0.18-0.2 at A_{630} by adding the liquid nutrient medium. Then TCSL was serially diluted in the nutrient broth in a 96-well microtiter plate and 50 μ l of each bacterial suspension was mixed to a final volume of 100 μ l. Four wells without the lectin for each bacterium were used as control. The reading was taken at A_{630} after the plates were agitated for 8 h at 32°C by using temperature controlled titer plate shaker. According to the following formula the percentage of bacterial growth inhibition was determined:

% inhibition = $\{(Absorbance\ of\ control - Absorbance$

of test) /Absorbance of control} × 100.

Fluorescence spectroscopy

The fluorescence measurement of TCSL (40 and 50 µg/ml) in tris buffer saline (TBS) has been performed at 30°C temperature by using a Shimadzu Spectrofluorometer RF-5301 PC. Fluorescence spectrum of 40 µg/ml TCSL was taken in the presence and absence of 1mM of CaCl₂. TCSL (50 µg/ml) was treated with 0.5 M of Guanidine-HCl for 2 h and 4 h in the presence of 1 mM of CaCl₂. All samples were placed in a 1×1×4.5 cm quartz cuvette, excited at 280

nm and the emission was recorded in the range of 300-400 nm Widths for the excitation and emission monochromators were maintained at 5 nm.

Results and discussion

Bacterial Agglutination Assay

Out of six, TCSL agglutinated three bacterial species including both gram-positive (*Staphylococcus aureus*) and gram-negative bacteria (*Salmonella enteritidis* and *Shigella flexneri*) and the minimum agglutination concentration for each bacterial species was 50 µg/ml (as summarized in Table 1).

Table 1. Bacterial agglutination by TCSL.

Bacterium	TCSL Concentration (µg/ml)
<i>Listeria monocytogenes</i>	No agglutination (NA)
<i>Salmonella enteritidis</i>	50
<i>Shigella flexneri</i>	50
<i>Staphylococcus aureus</i>	50
<i>Shigella boydii</i>	NA
<i>Pseudomonas aeruginosa</i>	NA

In the previous study, it was observed that *Bacillus megaterium*, *Bacillus subtilis*, *Salmonella typhi*, *Sarchina lutia*, *Shigella shiga* and *Shigella sonnei* were also agglutinated by this lectin in different concentrations and among those, *Shigella sonnei* was the most sensitive towards TCSL (Kabir *et al.*, 2012). The lectin was unable to agglutinate *Escherichia coli*, *Listeria monocytogenes*, *Pseudomonas aeruginosa*

and *Shigella boydii*. These results indicated that with a broad-spectrum antibiotic activity, TCSL can recognize the bacterial surface carbohydrate molecules on several species of bacteria and agglutinates those possibly due to the presence of cognate glycan antigens on their cell surface (Ghanekar *et al.*, 1980).

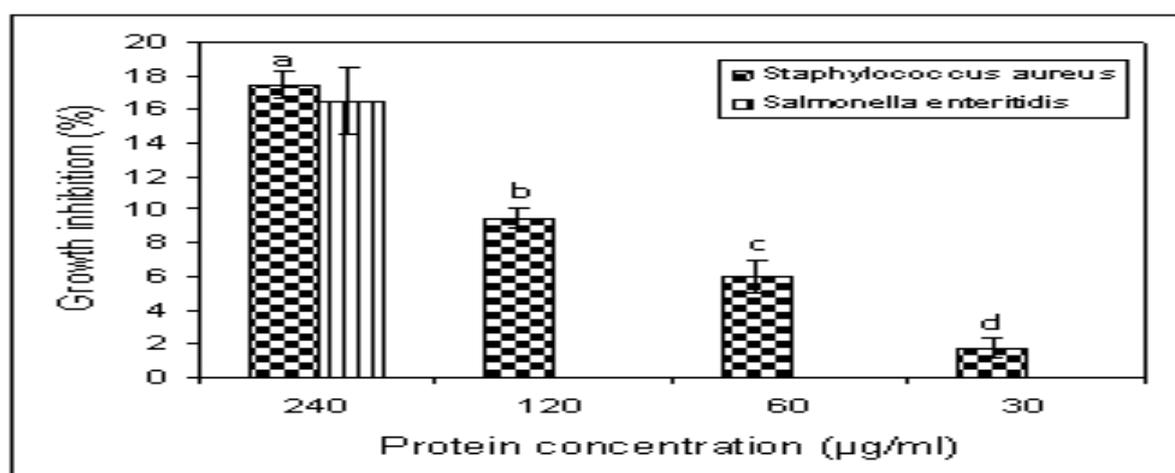


Fig. 1. Percentage of growth inhibition of *S. enteritidis* and *S. aureus* in presence of different concentrations of TCSL. (n = 3, mean ±S.D.). Values followed by different lowercase letters on top of column bar are significantly different at P<0.05.

Bacterial Growth Inhibition

TCSL partially inhibited the growth of *Salmonella enteritidis* and *Staphylococcus aureus* and did not show any inhibitory effect against *Pseudomonas aeruginosa*. Maximum growth inhibition was observed at a concentration of 240 $\mu\text{g/ml}$ for both

bacteria (Fig. 1). The inhibitory effect against *Staphylococcus aureus* was decreased gradually with the decrease of protein concentration, while the growth of *Salmonella enteritidis* was not affected at 120 $\mu\text{g/ml}$ and in smaller protein concentrations.

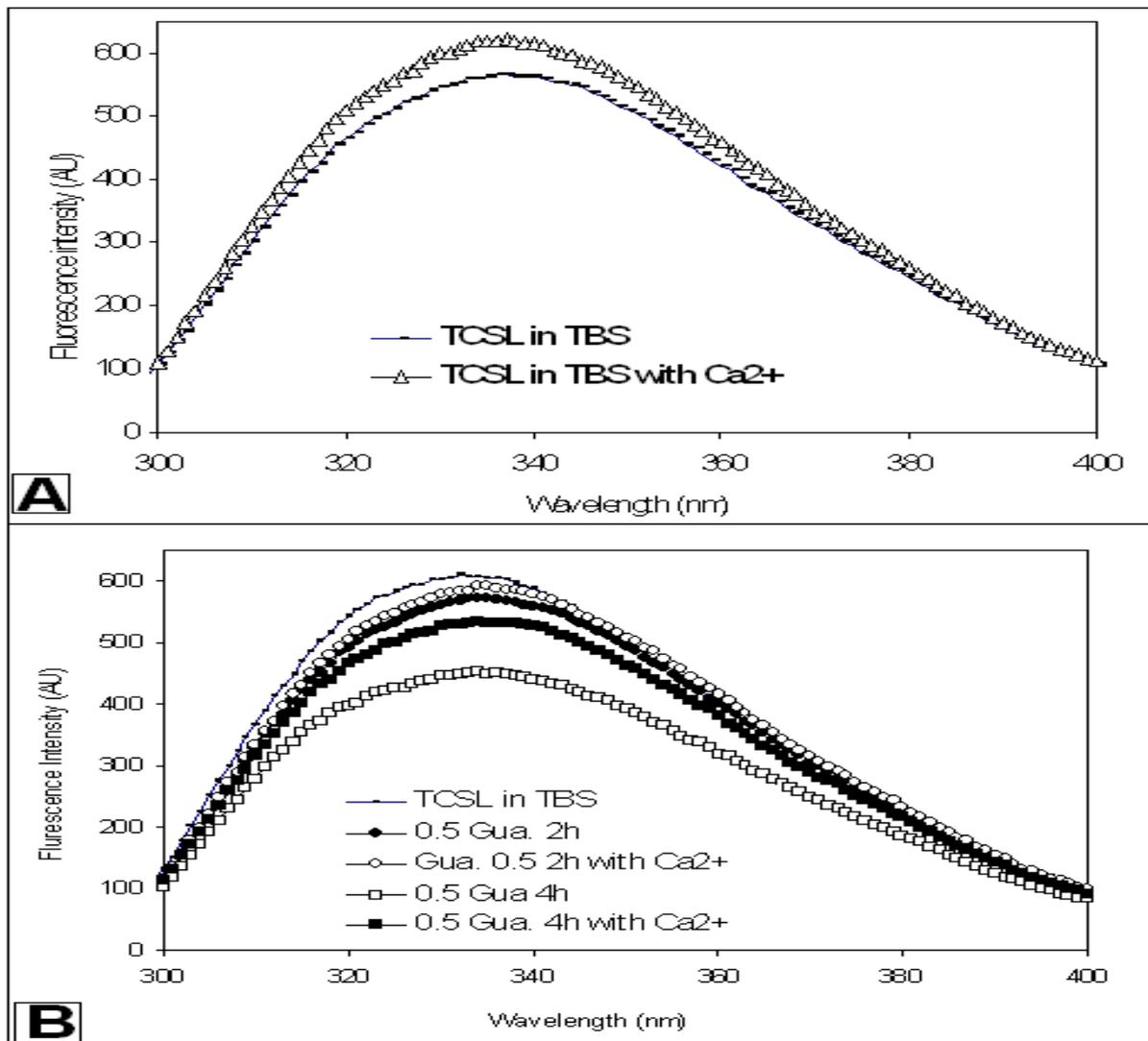


Fig. 2. Fluorescence spectra of TCSL under different conditions. (A) Fluorescence emission spectra of 40 $\mu\text{g/ml}$ TCSL in TBS (-), TBS containing 0.5 mM CaCl_2 (- Δ -). (B) Fluorescence emission spectra of 50 $\mu\text{g/ml}$ TCSL (-). TCSL incubated in 0.5 M guanidine-HCL for 2 h with Ca^{2+} (- \circ -) and without Ca^{2+} (- \square -) and 4 h with Ca^{2+} (- \square -) and without Ca^{2+} (- \circ -).

Two distinct levels of antibacterial activity were demonstrated: a low percentage of inhibition, possibly suggesting bacteriostatic activity and a higher activity that may be bactericidal (Hubert *et al.*, 1996). In fact, observations under the microscope revealed that some bacteria were deformed or showed decreased mobility during the antibacterial assay

when compared to the control (Tunkijjanukij & Olafsen, 1998). TCSL showed this type of low percentage of inhibition towards *Staphylococcus aureus* and *Salmonella enteritidis* whereas high antibacterial activity was exhibited by *Holothuria scabra* lectin (Gowda *et al.*, 2008) inhibiting the complete growth of both gram-positive bacteria

(*Staphylococcus sp*) and gram negative bacteria (*Shigella sp*, *E. coli*, *Proteus sp* and *Serratia sp*).

Fluorescence Measurement of TCSL at different conditions

Structural changes of TCSL were measured upon binding with Ca^{2+} , by measuring fluorescence emission spectra with excitation at 280 nm as shown in Fig. 2A. The fluorescence spectrum of TCSL reached to the peak at 330 nm, mainly from tryptophan residues in the protein molecule. Ca^{2+} stabilization of TCSL was examined by incubating the lectin solution with 0.5M of guanidine-HCl in the presence and absence of 1 mM CaCl_2 during a shorter time period (2 h) and a longer time period (4 h) at 30°C as shown in the figure. When TCSL was incubated with 0.5M of guanidine-HCl for 2 h, a drop in the spectrum was found but when the lectin became treated with 0.5 M of guanidine-HCl for 4 h, a large drop in the spectrum was observed. On the other hand, when the lectin was incubated in 0.5 M of guanidine-HCl for 2 h in the presence of Ca^{2+} , the peak of the spectrum reached higher than that of the peak obtained in the absence of Ca^{2+} . A significant increase in the peak level was observed when TCSL was treated with Ca^{2+} for 4 h (Fig. 2B).

In the previous study, it was found that the hemagglutination activity of TCSL was not affected by the pH values from 3 to 12 and also by the divalent cations, guanidine-HCl, urea and EDTA (Kabir *et al.*, 2012).

In the present study, the fluorescence intensity of TCSL at 330 nm became remarkably increased in the presence of Ca^{2+} salt. When TCSL in TBS was incubated with 0.5 M guanidine-HCl for 2 h and 4 h in the presence and absence of 1 mM CaCl_2 a remarkable change was observed. This result indicated that the change in environment of tryptophan residue(s) does not affect the binding sites. From this result it can be concluded that guanidine-HCl induced a structural change that did not affect the lectin binding sites. On the other hand, Ca^{2+} induced a conformational change in the TCSL

molecule that stabilized the lectin structure.

Conclusion

TCSL inhibited the growth of a number of pathogenic bacteria. Though the denaturant changed the environment around its tryptophan residues, binding sites of TCSL did not become affected. The lectin stability increased in the presence of Ca^{2+} salt that would be very important for the formulation of the lectin as an antibacterial agent.

References

- Ghanekar A, Perombelon MCM.** 1980. Interactions between potato lectin and some phyto-bacteria in relation to potato tuber decay caused by *Erwinia carotovora*. *Journal of Phytopathology-Phytopathologische Zeitschrift* **98**, 137-149.
<http://dx.doi.org/10.1111/j.14390434.1980.tb03726.x>
- Gowda NM, Goswami U, Khan MI.** 2008. T-antigen binding lectin with antibacterial activity from marine invertebrate, sea cucumber (*Holothuria scabra*): possible involvement in differential recognition of bacteria. *Journal of Invertebrate Pathology* **99**, 141-145.
<http://dx.doi.org/10.1016/j.jip.2008.04.003>
- Hubert F, van Der Knaap W, Noel T, Roch P.** 1996. Cytotoxic and antibacterial properties of *Mytilus galloprovincialis*, *Ostrea edulis* and *Crassostrea gigas* (bivalve molluscs) hemolymph. *Aquatic Living Resources* **9**, 115-124.
<http://dx.doi.org/10.1051/alr:1996015>
- Kabir SR, Nabi MM, Haque A, Zaman RU, Mahmud ZH, Reza MA.** 2013. Pea lectin inhibits growth of Ehrlich ascites carcinoma cells by inducing apoptosis and G₂/M cell cycle arrest in vivo in mice. *Phytomedicine* **20**, 1288-1296.
<http://dx.doi.org/10.1016/j.phymed.2013.06.010>
- Kabir SR, Reza MA.** 2014. Antibacterial activity of *Kaempferia rotunda* rhizome lectin and its induction of apoptosis in Ehrlich ascites carcinoma cells. *Applied Biochemistry and Biotechnology* **172**, 2866-

2876.

<http://dx.doi.org/10.1007/s12010-013-0720-2>.

Kabir SR, Hossen MA, Zubair MA, Alom MJ, Islam MF, Hossain MA, Kimura YA. 2011a. A New Lectin from the Tuberos Rhizome of *Kaempferia rotunda*: Isolation, Characterization, Antibacterial and Antiproliferative Activities. *Protein And Peptide Letters* **18**, 1140-1149.

Kabir SR, Islam MF, Alom MJ, Zubair MA, Absar N. 2012. Purification and characterization of a snake gourd seed lectin with antitumor activity against Ehrlich ascites carcinoma cells *in vivo* in mice. *Protein And Peptide Letters* **19**, 360-368.

<http://dx.doi.org/10.2174/092986612799363154>.

Kabir SR, Nabi MM, Nurujjaman M, Reza MA, Alam AHMK, Zaman RU, Khalid-Bin-Ferdous KM, Amin R, Khan MMH, Hossain MA, Uddin MS, Mahmud ZH. 2015a. *Momordica charantia* seeds lectin: toxicity, bacterial agglutination and antitumor properties. *Applied Biochemistry and Biotechnology* **175**, 2616-2628.

<http://dx.doi.org/10.1007/s12010-014-1449-2>.

Kabir SR, Zubair MA, Nurujjaman M, Haque MA, Hasan I, Islam MF, Hossain MT, Hossain MA, Rakib MA, Alam MT, Shaha RK, Hossain MT, Kimura Y, Absar N. 2011b. Purification and characterization of a Ca²⁺-dependent novel lectin from *Nymphaea nouchali* tuber with antiproliferative activities. *Bioscience Reports* **31**, 465-475.

<http://dx.doi.org/10.1042/BSR20100126>.

Khatun MMG, Rabbani MG, Rahaman EHMS. 2010. Estimate of genetic diversity in snake gourd (*Trichosanthes cucumerina*). *Bangladesh Journal of Agricultural Research* **35**, 95-100.

<http://dx.doi.org/10.3329/bjar.v35i1.5870>.

Liu B, Bian HJ, Bao JK. 2010. Plant lectins: Potential antineoplastic drugs from bench to clinic. Mini-review, *Cancer Letters* **287**, 1-12.

<http://dx.doi.org/10.1016/j.canlet.2009.05.013>.

Lowry OH, Rosebrough NJ, Farr AL, Randall RJ. 1951. Protein measurement with the folin phenol reagent. *Journal of Biological Chemistry* **193**, 265-275.

Padma P, Komath SS, Nadimpalli SK, Swamy MJ. 1999. Purification in high yield and characterization of a new galactose-specific lectin from the seeds of *Trichosanthes cucumerina*. *Phytochemistry* **50**, 363-371.

[http://dx.doi.org/10.1016/S0031-9422\(98\)00544-5](http://dx.doi.org/10.1016/S0031-9422(98)00544-5).

Rafiq S, Majeed R, Qazi AK, Ganai BA, Wani I, Rakhshanda S, Qurishi Y, Sharma PR, Hamid A, Masood A, Hamid R. 2013. Isolation and antiproliferative activity of *Lotus corniculatus* lectin towards human tumour cell lines. *Phytomedicine* **21**, 30-38.

<http://dx.doi.org/10.1016/j.phymed.2013.08.005>.

Sitohy M, Doheim M, Badr H. 2007. Isolation and characterization of a lectin with antifungal activity from Egyptian *Pisum sativum* seeds. *Food Chemistry* **104**, 971-979.

<http://dx.doi.org/10.1016/j.foodchem.2007.01.026>.

Tian Q, Wang W, Miao C, Peng H, Liu B, Leng F, Dai L, Chen F, Bao J. 2008. Purification, characterization and molecular cloning of a novel mannose-binding lectin from rhizomes of *Ophiopogon japonicus* with antiviral and antifungal activities. *Plant Science* **175**, 877-884.

<http://dx.doi.org/10.1016/j.plantsci.2008.09.008>.

Tunkijjanukij S, Olafsen JA. 1998. Sialic acid-binding lectin with antibacterial activity from the horse mussel: further characterization and immunolocalization. *Developmental and Comparative Immunology* **22**, 139-350.

[http://dx.doi.org/10.1016/S0145-305X\(98\)00017-2](http://dx.doi.org/10.1016/S0145-305X(98)00017-2).