



Effect of lead stress on polyphenols, flavonoids, and proline contents in radish (*Raphanus sativus* L.)

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Abstract

Radish (*Raphanus sativus* L.) is a suitable model plant for eco-toxicological studies due to its tolerance to various stresses, such as metallic and saline stress. Lead is a toxic and exogenous metal to plants, and its presence causes several disturbances in plant. This work consists to determine the effect of lead stress applied during two weeks (0, 500, 1000, 2000 ppm) after 45 days of the plant's growth. The parameters analyzed in the plant are polyphenols, flavonoids, and proline. The assay is performed using a JENWAY 6505 UV-Visible Spectrophotometer. The results obtained show a decrease in proline, but polyphenols and flavonoids show a marked increase. The high levels of proline (0.117 mg/g dry matter in the control) and polyphenols (0.248 mg/g dry matter at the 2000 ppm dose of lead) are recorded in the root system. However, the highest content of flavonoid (8,586 mg/g dry matter in the control) is recorded in the aerial part of the plant. Radish (*Raphanus sativus* L.) is a tolerant plant to lead stress and has a phytoremediator power for lead.

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Introduction

The increase of heavy metals levels in the soil presents a risk for the environment, mainly due to human activities of over exploitation of the soil. Lead can be found in the environment, such as through discharges around lead treatment sites or through automobile pollution, which contaminate soil, vegetation, animals and humans (Ben Ghanya *et al.*, 2006). Lead is a non-essential element for plants, and its phytotoxicity can inhibit the enzymatic activity, disrupt the mineral nutrition, unbalance the water, modify the hormonal status and alter the membrane permeability (Sharma and Dubey, 2005). Thus, these ions can intensify the production processes of reactive oxygen species (ROS) leading to oxidative stress (Prasad *et al.*, 1999; Cuypers *et al.*, 1999). In the face of this disturbance, plants develop several defense mechanisms, including the use of biochemical compounds as stress regulators such as amino acids and phenolic compounds. Proline is an amino acid known as osmolyte (Ashraf and Foolad, 2007), it is involved in the adaptation of plants to environmental constraints by stabilizing proteins, free radical scavenging and regulation of cellular redox potential (Kilani *et al.*, 2012). Polyphenols are among the metabolites that have important antioxidant capacities (Gomez-Caravaca *et al.*, 2006; Xiuzhen *et al.*, 2010), and a high chelating power of metals which represents an important biological interest (Ravichandran *et al.*, 2014; Selvaraj *et al.*, 2014; Kasprzak *et al.*, 2015). According to Symonowicz and Kolanek (2012) flavonoids have an ability to detoxify free radicals, and their ability to chelate heavy metals has been demonstrated by (Brown *et al.*, 1998). These antioxidant mechanisms have been exhaustively described (Gill and Tuteja, 2010; Dat *et al.*, 2000; Apel and Hirt, 2004).

The plant material used in this study is radish (*Raphanus sativus* L.) from the Brassicaceae family, which was selected for its use in the laboratory as a model plant for eco-toxicology studies of various pollutants (Sun *et al.*, 2010), and for its improved germination rate, rapid growth and high biomass (Bitour, 2012).

The objective of this study is to evaluate the effects of increased lead levels in the soil on the antioxidant activity of radish (*Raphanus sativus* L.) by the determination of antioxidants such as total polyphenols, flavonoids, and proline synthesis.

Materials and methods

Plant material

The plant material used in this study is the radish (*Raphanus sativus*L.), the variety "National".

Methods of culture

The radish seeds (*Raphanus sativus* L.) are disinfected with 2% sodium hypochlorite for 10 min, then washed several times with distilled water and germinated in alveoli containing compost for 15 days. The seedlings are transplanted into the cylinders (30 cm high and 20 cm in diameter) containing a mixture of sand and compost (2V/V) for a month and a half (45 days). Watering was done 3 times a week, twice with distilled water and once with a nutrient solution of Hoagland (1938). The plants are harvested after a period of 60 days of cultivation, and transported to the laboratory in order to be cut in two aerial and root parts, then they were dried at 80°C during 48 hours.

Application of stress

Radish stress is applied after 45 days of cultivation twice for two weeks, the lead doses used are 0, 500, 1000 and 2000 ppm, with three repetitions for each dose.

Analyzed parameters

Preparation of methanolic extracts

The plant material used in the preparation of the extracts was macerated in a methanol/water mixture (7:3 V/V) at a ratio of 1/10 (W/V), under gentle agitation overnight at room temperature (Talbi *et al.*, 2015).

Content of polyphenols

The determination of total polyphenols is performed by a colorimetric method of (Folin-Ciocalteu) (Singleton *et al.*, 1999; Heilerova *et al.*, 2003). The

absorbances are read from the UV-Visible Spectrophotometer at 760 nm.

Content of flavonoids

The estimation of the total flavonoid content is made by the method of (Bahorun *et al.*, 1996) the absorbance is read at 415 nm using a UV-Visible Spectrophotometer.

Content of proline

The method used to measure proline is that of (Troll and Lindsley, 1955) modified by (Dreier and Goring, 1974) and then by (Monneveux and Nemmar, 1986). The assay was performed by a UV-Visible Spectrophotometer JENWAY 6505 at a wavelength of 528 nm. The content is expressed in mg/g dry matter.

Statistical analyzes

The results obtained were analyzed statistically using STATBOX software. Version 6.4. The analyzes

performed are the variance (ANOVA) and the NEWMAN-KEULS test with a significance level ($P = 5\%$).

Results

Polyphenols content of the aerial and root part of Radish stressed by the lead

The total polyphenols content increases in both aerial and root parts of the radish. This content is higher in the root part (between 0.202 mg/g dry matter in the control and 0.248 mg/g dry matter in the 2000 ppm dose of lead) compared to the aerial part (between 0.140 mg/g dry matter in the control and 0.091 mg/g dry matter in the 2000 ppm dose of lead). The level of total polyphenols recorded in plants stressed with lead is higher than that of the control in the root part and lower than the control in the aerial part (Fig.1). The variation of the total polyphenols content in both the aerial and root parts of radish under lead stress is not significant ($P > 0.05$).

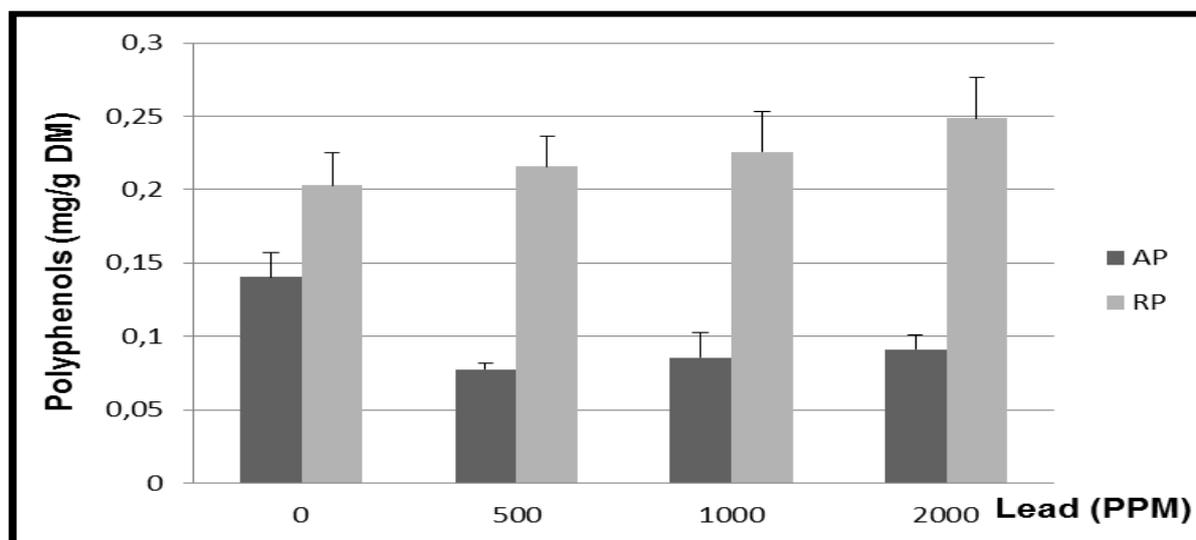


Fig.1. The polyphenols content (mg/g dry matter) in the aerial (AP) and root part (RP) of radish (*Raphanus sativus* L.) under lead stress.

Flavonoids content of the aerial and root part of Radish stressed by the lead

The total flavonoid content in radish decreases in the aerial part and increases in the root part gradually and proportionally with increasing lead dose. The level of flavonoids recorded in the aerial part (between 8.586 mg/g dry matter in the control and 6.287 mg/g dry matter in the 2000 ppm dose of lead) is higher than that recorded in the root part (between

2.490 mg/g dry matter in the control and 4.751 mg/g dry matter in the 2000 ppm dose of lead). The 500 and 1000 ppm doses of lead mark almost equal levels of flavonoids in both parts of the plant (Fig.2). The ANOVA statistical analysis shows that the difference in flavonoids content recorded in the aerial and root part of radish under the lead effect is not significant ($p > 0.05$).

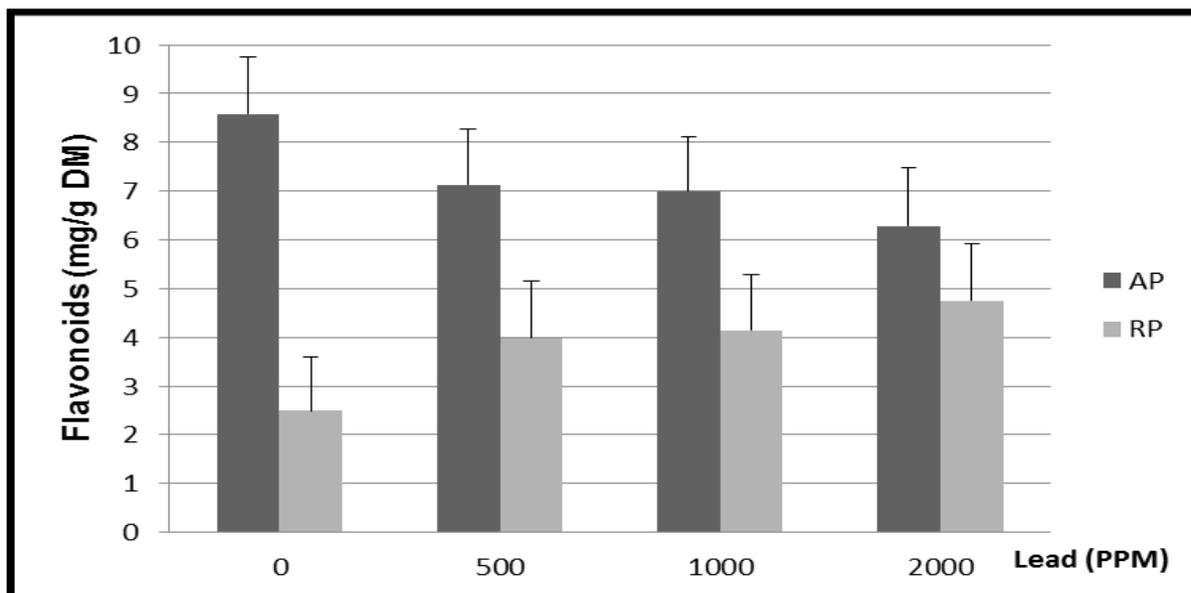


Fig. 2. The flavonoids content (mg/g dry matter) in the aerial (PA) and root part (RP) of radish (*Raphanus sativus* L.) under lead stress.

Proline content of the aerial and root part of Radish stressed by the lead

The proline content decreases proportionally with increasing lead dose in both aerial and root parts of Radish. The proline level is higher in the root part compared to the aerial part, and the difference in proline content between these two parts is 0.025, 0.041, 0.048 and 0.049 mg/g dry matter for the control, for 500, 1000 and 2000 ppm doses of lead respectively. The highest proline content is recorded in the control plant (0.117 and 0.092 mg/g dry matter) in the root and aerial parts respectively. On the other hand, the lowest proline content is recorded in the 2000 ppm dose of lead (0.080 and 0.031 mg/g dry matter) in the root and aerial parts respectively (Fig.3). Statistical analysis of variance shows a non-significant effect ($P > 0.05$) of the lead dose increase on proline content in both aerial and root parts of radish.

Discussion

Polyphenols and flavonoids

Polyphenols increase in the root and aerial part of the radish proportionally with increasing doses of lead. However, flavonoids increase in the root part and decrease in the aerial part of radish as lead dose increases. Stressed plants accumulate polyphenols and flavonoids more than the control in the root part

and less than in the aerial part of the plant. This could be caused by triggered stress levels in the different organs of the plant; which shows that the lead effect is not only dose-dependent, but also organ-dependent. According to Smirnoff, (2005) polyphenols are found in large quantity in different parts of plant under several stresses effects, such as heavy metals (Briat and Lebrum, 1999). A correlation between polyphenol content and antioxidant activity was observed, since phenolic levels contribute directly to antioxidant activity (Duh, 1999). The accumulation of polyphenols can be explained by the stimulation of polyphenol oxidase production which is a key enzyme in the metabolic pathway of polyphenols (Tuna *et al.*, 2008). Flavonoids act as antioxidant molecules that fix ROS (Lovdal *et al.*, 2010). Flavonoids can act in different ways in oxidative stress regulation processes by directly capturing reactive oxygen species, chelating metals and inhibiting the activity of some enzymes responsible for the production of ROS (Morris, 1995).

The results obtained in this work are in conformity with several studies showing an increase in polyphenols and flavonoids under lead stress in *Atriplex halimus* at dose (0.1000, 4000, 8000 ppm) (Belarbi, 2018), in *Vallisneria natans* at different lead concentrations (Chao Wang *et al.*, 2011), in *Echuim*

vulgare L. under metallic stress from lead and zinc (Slowomir Dresher *et al.*, 2017), in *Ginsingpnanax* at a dose of 3000 ppm copper (Ali *et al.*, 2006), in *Kandelia obovata* after 7 days of treatment with lead and manganese (Zhongzhenget *al.*, 2011), in *Vaccinium corymbosum* L. under aluminium and cadmium stress (Manquian-cerdaa *et al.*, 2016). An increase in polyphenol content is also marked in *Raphanus sativus* (Tomislav *et al.*, 2008), in *Aeluropus littoralis* under the effect of heavy metals

(Pb, Co, Cd, Ag) (Rastgoo *et al.*, 2011), and in *Viciafaba* subjected to different mercury concentrations (Benahmed f. 2010). An increase in flavonoid content is also estimated in *Taraxacum officinal* under stress of (Cr, Pb, Cu, Ni, and Zn) (Bretzel *et al.*, 2014). The decrease in flavonoids and the increase in polyphenols in the aerial part of radish are also recorded in *Brassica juncia*, a species of the same family of radish under cadmium stress (Dhriti K. *et al.*, 2014).

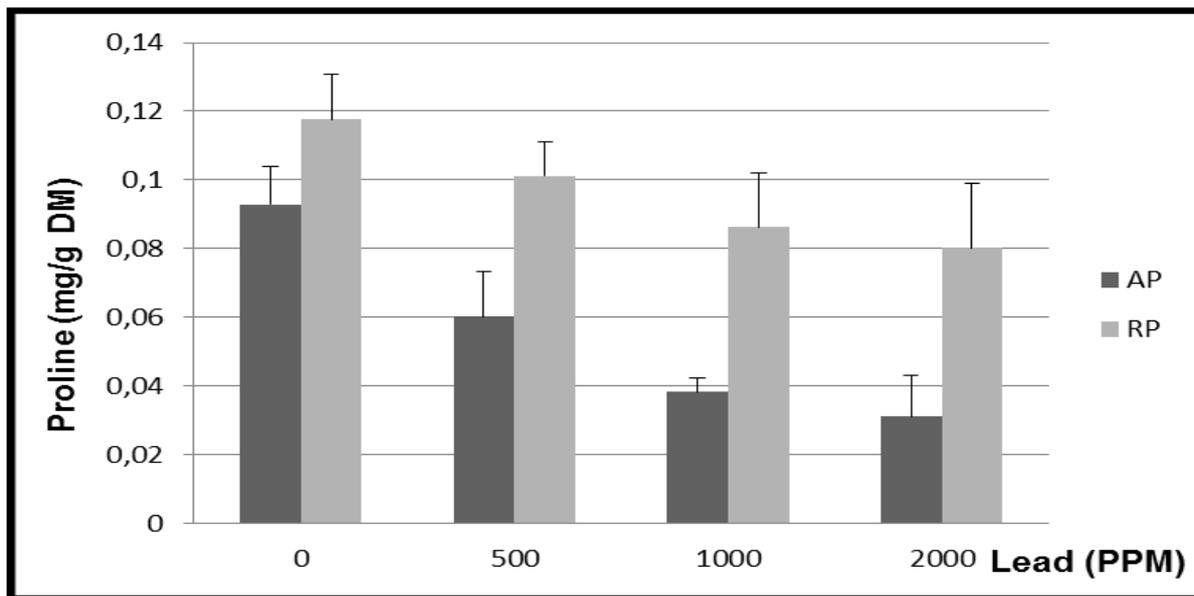


Fig.3. Proline content (mg/g dry matter) in the aerial (AP) and root part (RP) of radish (*Raphanus sativus* L.) under lead stress.

Proline

The proline content recorded in *Raphanus sativus* plants subjected to different lead concentrations (0, 500, 1000, 2000 ppm) decreases progressively with increasing of the above doses of lead. According to Lemzeri (2007), the species most sensitive to stress react by accumulating proline more rapidly. In contrast, the tolerant ones show relative stability or low proline accumulation compared to the sensitive ones. This could justify the low proline accumulation in radish under lead stress by the tolerance of this plant to metallic stress. These results are consistent with several researchers demonstrating a decrease of proline in *Raphanus sativus* stressed by zinc at doses of 0 and 100 ppm (Tihana *et al.*, 2008), in *Brassica juncia* subjected to high doses of lead and cadmium (John *et al.*, 2009), and in *Atriplex*

canescens stressed by lead at doses of 1000 and 5000 ppm (Babou, 2014). According to Chen, 2004 proline also decreases in rice roots under metallic stress.

The highest levels of proline and polyphenols are superior in the root part of radish, and the flavonoid content increases gradually with the increase of the lead dose in the root part. Proline accumulation is higher in the root part than the aerial part of plant (Fig.3). These results are identical to those obtained by (Tihana *et al.*, 2008; Tomislav *et al.*, 2008), confirming that the proline content is higher in hypocotyl than in leaves of *Raphanus sativus* under lead stress. According to (Biteur, 2012), proline synthesis increases in roots more than leaves after ten weeks of lead stress in radish.

According to (Jones and Clement 1972; Khan and Frankland, 1983), it is generally known that the roots act as a barrier to the movement of toxic heavy metal through the soil-plant system. Several authors confirm the accumulation of lead in radish roots such as Pahlsson (1989) which showed that lead accumulates mainly in the root system of *R. sativus*, where it binds to the root surface and cell walls. Kristen *et al.* (2015) also confirm that the level of lead in the aerial parts of radish is inferior to the roots. A study conducted by (Asadi Kapourchal *et al.*, 2009) showed that the measured lead accumulated in the roots was 208 ppm whereas it was 27.25 ppm in the radish shoots harvested. These results may justify the estimation of high levels of proline, polyphenols and flavonoids in the root part of radish. Asadi Kapourchal and colleagues (2009) also report that there was no toxicity for radish up to 1000 ppm lead in soil. Consequently, they considered radish as an accumulator plant of lead and that it can be seeded up to five times a year in the same soil and its yield can reach 20 t / ha, so it can be used to remediate the polluted soils by lead.

Conclusion

The increased lead dose causes in radish (*Raphanus sativus* L.) a decrease in proline content and an increase in total polyphenols and flavonoids. The higher levels of proline and total polyphenols are recorded in the root part of the plant, unlike the high levels of flavonoids in the aerial part. The statistical analysis of lead stress's effect on the accumulation of proline, polyphenols and flavonoids in radish is not significant, which shows that *Raphanus sativus* is not affected by the increase in lead dose at the point where stress causes significant differences on the antioxidant (polyphenols, flavonoids) and proline content of this plant. From these results, we can deduce that *Raphanus sativus* is a tolerant plant to the lead stress and it has a phytoremediator power for this metal.

References

Ali MB, Singh N, Shohaël AM, Hahn EJ, Paek KY. 2006. Phenolics metabolism and lignin synthesis

in root suspension cultures of *Panaxginseng* in response to copper stress. *Journal of Plant Sciences* **171**, 147-154.

<http://dx.doi.org/10.1016/j.plantsci.2006.03.005>

Apel K, Hirt H. 2004. Reactive oxygen species: Metabolism, Oxidative Stress, and Signal Transduction. *Annual Review of Plant Biology* **55(3)**, 73-

99.<http://dx.doi.org/10.1146/annurev.arplant.55.031903.141701>

Asadi Kapourchal So., Asadi Kapourchal Sa., Pazira E., Homae M. 2009. Assessing radish (*Raphanus sativus* L.) potential for phytoremediation of lead-polluted soils resulting from air pollution. *Plant, Soil and Environment* **55(5)**, 202–206.

<http://doi.org/10.17221/8/2009-PSE>

Ashraf M, Foolad MR. 2007. Roles of Glycine betaine and proline in improving plant abiotic stress resistance. *Environmental and Experimental Botany* **59**, 206-216.

<http://dx.doi.org/10.1016/j.envexpbot.2005.12.006>

Babou FZ 2014. Lead action on *Atriplex halimus* and *Atriplex canescens* (Push) Nutt resistance markers. *Memory of Magister, Oran University, Algeria* 27-28.

Bahorun T, Gressier B, Trotin F, Brunet C, Dine T, Luyckx M, Vasseur J, Cazin M, Cazin JC, Pinkas M. 1996. Oxygen species scavenging activity of phenolic extracts from hawthorn fresh plant organs and pharmaceutical preparations. *ArzneimForsch* **46**, 1086-1089.

Belarbi A, 2018. Physiological and phytochemical study of tolerance to heavy metals Pb, Cr (III) and Cr (II) by *Atriplex halimus* L. PhD thesis, Mostaganem University, Algeria 179-189.

Ben ghnaya A, Gilbert C, Jeannette B, Michel B. 2006. Phytoremediation: in vitro selection of Rapeseed (*Brassica napus* L.) tolerant of toxic metals.

International Journal of Tropical Ecology and Geography **30(2)**, 69-86.

Benahmed, F. 2010. Oxidative stress in *Vicia faba* L. seedlings subjected to various abiotic stresses (saline stress, hydric stress and heavy metal stress). Memory of Magister, Oran University, Algeria p 35-36.

Biteur N. 2012. Tests for the use of Radish (*Raphanus sativus* L.) in phytoremediation (biodepollution) in soil contaminated by heavy metals (Lead): Study of oxidative stress and some enzymatic parameters. PhD thesis, Oran University, Algeria p 61.

Bretzel, Francesca Benvenuti, Stefano Pistelli, Laura 2014. Metal contamination in urban street sediment in Pisa (Italy) can affect the production of antioxidant metabolites in *Taraxacum officinale* Weber. Environmental Science and Pollution Research **21(3)**, 2325-2333.

<http://dx.doi.org/10.1007/s11356-013-2147-2>

Briat JF, Lebrun H. 1999. Plant responses to metal toxicity. Comptes Rendus de l'Académie des Sciences - Series III **322**, 43-54.

<https://www.ncbi.nlm.nih.gov/pubmed/10047953>

Brown JE, Khodr H, Hider RC, Rice-Evans CA. 1998. Structural dependence of flavonoid interactions with Cu²⁺ ions: implications for their antioxidant properties. Biochemical Journal **330**, 1173-1178.

Chao Wang, Jie Lu, Songhe Zhang, Pei Fang, Wang Jun Hou, Jin Qian. 2011. Effects of Pb stress on nutrient uptake and secondary metabolism in submerged macrophyte *Vallisneria spiralis*. Ecotoxicology and Environmental Safety **74(5)**, 1297-1303.

<http://doi.org/10.1016/j.ecoenv.2011.03.005>

Chen CT, Chen TH, Lo KF, Chiu CY. 2004. Effects of proline on copper transport in rice seedlings under excess copper stress. Plant Science **166**, 103-111.

Cuyppers A, Vangronsveld J, Clijsters H. 1999. The chemical behaviour of heavy metals plays a

prominent role in the induction of oxidative stress. Free Radical Research **31**, 539-543.

Dat J, Vandenabeele S, Vranová E, Van Montagu M, Inzé D, Van Breusegem F. 2000. Dual action of the active oxygen species during plant stress responses. Cellular and Molecular Life Sciences **57**, 779-95.

<http://dx.doi.org/10.1007/s000180050041>

Dhriti K, Satwinderjeet K, Renu B. 2014. Physiological and Biochemical Changes in Brassica juncea Plants under Cd-Induced Stress. BioMed Research International, Article ID 726070.

<http://dx.doi.org/10.1155/2014/726070>

Dreier W, Göring M. 1974. Dereim slushohersolz kongentrasyon en aiesverschideu physiologeshe parameter van-maiswrzelnwiss. Z. Drh. Berlin Nath. Naturwiss R **23**, 641-4.

Duh PD. 1999. Antioxydant activity of water extract of four Harnng Jyur (*Chrysanthemum morifolium* Ramat) varieties in soybean oil emulsion. Food Chemistry **4(66)**, 471-476.

Gill SS, Tuteja N. 2010. Reactive oxygen species and antioxidant machinery in abiotic stress tolerance in crop plants. Plant Physiology and Biochemistry **48(12)**, 909-930.

<http://dx.doi.org/10.1016/j.plaphy.2010.08.016>

Gomez-Caravaca AM, Gomez-Romero M, Arraez-Roman D, Segura-Carretero A, Fernandez-Gutierrez A. 2006. Advances in the analysis of phenolic compounds in products derived from bees. Journal of Pharmaceutical and Biomedical Analysis **41**, 1220-1234.

<http://dx.doi.org/10.1016/j.jpba.2006.03.002>

Heilerova L, Buekova M, Trapeik P, Silhar S, Lbuda J. 2003. Comparaison of antioxydative activity data for aqueous extracts of Lemon Blam (*Melissa officinalis* L.); Oregano (*Origanum vulgare* L.); Thyme (*Thymus vulgaris* L.) and

Agrimony (*Agrimonia eupatoria* L.) obtained by conventional method and the DNA-Based biosensor. *Journal of Food Science* **21(2)**, 78-84.

Hoagland DR, Arnon DI. 1938. The water-culture method for growing plants without soil. Berkeley, California: College of Agriculture, University of California. Circular **347**, 1-39.

John R, Ahmad P, Gadgil K, Sharma S. 2009. Heavy metal toxicity: effect on plant growth, biochemical parameters and metal accumulation by *Brassica juncea* L. *International Journal of Plant Production* **3**, 65-75.

Jones LHP, Clemen CR. 1972. Lead uptake by plants and its significance for animals. In *Lead Environment*, ed., P Hepple, 29-33 p. Applied Science Publishers, Barking, Essex.

Kasprzak MM, Erxleben A, Ochocki J. 2015. Properties and applications of flavonoid metal complexes. *Royal Society of Chemistry Advances* **5**, 45853-45877.
<http://doi:10.1039/C5RA05069C>

Khan DH, Frankland B. 1983. Effects of cadmium and lead on radish plants with particular reference to movement of metals through soil profile and plant. *Plant and Soil* **70**, 335-345.
<http://doi.org/10.1007/BF02374890>

Kilani BR, Chedly A, Arnould S. 2012. Proline, a multifunctional amino-acid involved in plant adaptation to environmental constraints. *Biologie Aujour d'hui* **206(4)**, 291-299.
<http://doi.org/10.1051/jbio/2012030>

Kristen Hladun R, David Parker R, John Trumble T. 2015. Cadmium, Copper, and Lead Accumulation and Bioconcentration in the Vegetative and Reproductive Organs of *Raphanus sativus*: Implications for Plant Performance and Pollination. *Journal of Chemical Ecology* **41(4)**, 386-395.
<http://dx.doi.org/10.1007/s10886-015-0569-7>

Lemzeri H. 2007. Ecophysiological responses of three forest species of the genus *Acacia*, *Eucalyptus* and *Schinus* (*A. cyanophylla*, *E. gomphocephala* and *S. mölle*) under saline stress. *Memory of Magister*, Constantine University, Algeria 180p.

Løvdal T, Olsen KM, Sliemstad R, Verheul M, Lillo C. 2010. Synergetic effects of nitrogen depletion, temperature, and light on the content of phenolic compounds and gene expression in leaves of tomato. *Phytochemistry Journal* **71(5)**, 605-613.
<http://dx.doi.org/10.1016/j.phytochem.2009.12.014>

Manquían-Cerdaa K, Crucesb E, Escudeyac M, Zúñigaa G, Calderónd R. 2016. Interactive effects of aluminum and cadmium on phenolic compounds, antioxidant enzyme activity and oxidative stress in blueberry (*Vaccinium corymbosum* L.) plantlets cultivated in vitro. *Ecotoxicology and Environmental Safety* **150**, 320-326.
<http://doi.org/10.1016/j.ecoenv.2017.12.050>

Monneveux PH, Nemmar M. 1986. Contribution to the study of drought resistance in soft wheat (*Triticum aestivum* L.) and durum wheat (*Triticum durum* Desf.): study of proline accumulation during the development cycle. *Agronomy, Édition Diffusion Presse Sciences* **6(6)**, 583-590.
<https://hal.archives-ouvertes.fr/hal-00884913>

Morris CJ, Earl JR, Trenam CW, Blake DR. 1995. Reactive oxygen species and iron a dangerous partnership in inflammation. *International Journal of Biochemistry & Cell Biology* **27**, 109-122.

Påhlsson AB. 1989 Toxicity of heavy metals (Zn, Cu, Cd, Pb) to vascular plants. *Water, Air, & Soil Pollution* **47**, 287-319.
<http://dx.doi.org/10.1007/BF00279329>

Rastgoo Leila, Alemzadeh, Abbas. 2011. Biochemical Responses of Gouan (*Aeluropus littoralis*) to Heavy Metals Stress. *Australian Journal of Crop Science* **5(4)**, 375-383.

- Ravichandran R, Rajendran M, Devapiriam D.** 2014. Antioxidant study of quercetin and their metal complex and determination of stability constant by spectrophotometry method. *Food chemistry* **146**, 472-478.
<http://dx.doi.org/10.1016/j.foodchem.2013.09.080>
- Selvaraj S, Krishnaswamy S, Devashya V, Sethuraman S, Krishnan UM.** 2014. Flavonoid-metal ion complexes: a novel class of therapeutic agents. *Medicinal research reviews* **34**, 677-702.
<http://dx.doi.org/10.1002/med.21301>
- Sharma P, Dubey RS.** 2005. Lead toxicity in plants. *Brazilian Journal of Plant Physiology* **17**, 35-52.
<http://dx.doi.org/10.1590/S167704202005000100004>
- Singelton VL, Orthofer R, Lamuela-Raventos RM.** 1999. Analyses of total phenols and other oxidation substrates and antioxidants by means of Folin-Ciocalteu reagent. Packer, L., Ed., *Oxidants and Antioxidants, Part A, Methods in Enzymology* **299**, 152-178 Academic Press, New York.
[http://dx.doi.org/10.1016/S0076-6879\(99\)99017-1](http://dx.doi.org/10.1016/S0076-6879(99)99017-1)
- Sławomir D, Magdalena W, Kosiorb I, Sowab G, StanisławskiaIzabela B, Małgorzata W.** 2017. Effect of short-term Zn/Pb or long-term multi-metal stress on physiological and morphological parameters of metallicolous and non metalli colous *Echium vulgare* L. populations. *Plant Physiology and Biochemistry* **(115)**, 380-389.
<http://doi.org/10.1016/j.plaphy.2017.04.016>
- Smirnoff N, Foyer C, Dietz K, Mittler R, Feierabend J, Grace S, Desikan R, Jones M, Vreeburg R, Logan B, Jospers P.** 2005. Antioxidants and reactive oxygen species in plants. Blackwell publishing. *Cellular and Molecular Life Sciences* **57(5)**, 779-795.
- Sun BY, Kan SH, Zhang YZ, Deng SH, Wu J, Yuan H, Qi H, Yang G, Li L, Zhang XH, Xiao H, Wang YJ, Peng H, Li YW.** 2010. Certain antioxidant enzymes and lipid peroxydation of Radish (*Raphanus sativus* L.) as early warning biomarkers of soil copper exposure. *Journal of Hazardous Materials* **183**, 833-838.
<http://doi:10.1016/j.jhazmat.2010.07.102>
- Symonowicz M, KolanekM.** 2012. Flavonoids and their properties to form chelate complexes. *Biotechnology and Food Science* **76**, 35-41.
- Talbi H, Boumaza A, El-mostafa K, Talbi J, Hilali A.** 2015. Evaluation of antioxidant activity and physico-chemical composition of methanolic and aqueous extracts of *Nigella sativa* L. *Journal of Materials and Environmental Science* **6(4)**, 1111-1117.
- Tihana T, John TH, Meri E, Nada P, Vera C, Hrvoje L, Ivna Š, Drago B.** 2008. Antioxidative responses in radish (*Raphanus sativus* L.) Plants stressed by copper and lead in nutrient solution and soil. *Acta biologica cracoviensia. Series Botanica* **50(2)**, 79-86.
- Tomislav V, Nada P, Hrvoje L, Ivna Š, Tihana T.** 2008. Oxidative stress in radish plants grown on soils with different cu and pb level. *Cereal Research Communications* **(36)**, 1519-1522.
<http://www.jstor.org/stable/90003005>
- Troll W, Lindsley J.** 1955. A photometric method for the determination of proline. *Journal of Biological Chemistry* **215**, 655-660.
- Tuna AL, Kaya C, Higgs H, Murillo-Amador B, Aydemir S, Girgin AR.** 2008. Silicon improves salinity tolerance in wheat plants. *Environmental and Experimental Botany* **62(1)**, 10-16.
<http://dx.doi.org/j.envexpbot.2007.06.006>
- Xiuzhen H, Tao S, Hongxiang L.** 2007. Dietary Polyphenols and Their Biological Significance. *International Journal of Molecular Sciences* **8**, 950-988.

Zhong Zheng Yan, Nora Fung, Yee Tam. 2011. Temporal changes of polyphenols and enzyme activities in seedlings of *Kandelia obovata* under lead

and manganese stresses. *Marine Pollution Bulletin* **63(5-12)**, 438-444.
<http://dx.doi.org/10.1016/j.marpolbul.2011.04.027>