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RESEARCH PAPER

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Seroprevalence of *Toxoplasma gondii* Infection in Domestic Animals of District Charsadda, Khyber Pakhtunkhwa, Pakistan

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Abstract

This study was conducted in to find out the seroprevalence of toxoplasmosis in domestic animals of District Charsadda. Toxoplasmosis is an infection caused by a unicellular parasite *Toxoplasma gondii* which is cosmopolitan in distribution among the animals including domestic animals and human beings. Very little information is available about the infection rate of toxoplasmosis in domestic animals of District Charsadda, Pakistan. A total of 374 blood samples were collected from cows, buffaloes, sheep and goats and were tested by Latex agglutination test, out of which (79.7%) were found seropositive. Out of 260 females, (82.69%) were found infected while in 114 males (78.80%) were found infective. A high prevalence of (81.65%) was detected in age group above one year. Individual sex wise prevalence also revealed that females were more positive as compared to males. In male buffaloes, (76.92%) were positive while in females, (75.67%) were detected positive. In male goats (72.41%) were positive while in case of females, (84.78%). In male cows (69.23%) were detected seropositive while in females cows the infection rate was (73.77%). This increased rate of infection may be due to free access of cat to drinking water and other food sources as well as warm and humid conditions of the area.

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Introduction

Toxoplasma gondii which is an intracellular parasite distributed worldwide (Shah et al., 2013a) and is responsible for toxoplasmosis disease (Aldebret et al., 2011). The discovery of cats as definitive host for T. gondii was done in 1960 (Innes, 2010). Life cycle of T. gondii is completed in two stages, sexual stage starts in definitive host, which include cats and asexual stage is completed in warm blooded animals (Afonso, 2008). Intermediate hosts are infected by ingesting oocysts, which change into tachyzoites through asexual reproduction and then change into bradyzoites. Once again the felines are infected by eating bradyzoites-infected meat (Webster, 2007). About (33-60%) of the human worldwide population have been infected by this parasite (Holliman, 1997; Shah et al., 2014), but this rate changes according to the geographical, climatic and nutritional factors, socio- cultural habits and routes of transmission (Shah et al., 2013b). Prevalence rate of T. gondii varies in different countries.

The infection rate of (28.44%) was recorded in Mohmand Agency, Pakistan (shah et al., 2014). The seroprevelence of T. gondii was 63% in Punjab, 48% in Azad Kashmir and 38% in Khyber Pukhtunkhwa (Tenter et al., 2000). Prevalence rate of (17.4%) was found in young school children in Islamabad, Pakistan (Sadaruddin et al., 1991). The prevalence rate in Dera Ghazi Khan, Pakistan was detected (29.5%) (Tasawar et al., 2011), Infection rate has declined recently due to awareness in people by not using under cooked meat and advancement in animal husbandry work (Shah et al., 2017). The seroprevalence of T. gondii in domestic animals and meat producing animals, such as goats, and sheep, has found the same infection all the time. In District Mardan (Pakistan) the prevalence rate of toxoplasmosis in goats was recorded 42.28% whereas in sheep the infection rate was 44.13% (Shah et al., 2013b). In Mohmand Agency, Pakistan, the prevalence rate of toxoplasmosis in goats was recorded 53.84% while in sheep it was 36.00% (Shah et al., 2013a). In farmed sheep, the prevalence in Europe is related with age, increasing from lambs (1722%) to adult (65 -89%) (Halos, 2010). Viable T. gondii have been detected in about 67% of sheep samples. In Southern European countries infected meat of Sheep is the main source of infection. Seropositivity for goats varies from 4 to 77% (Dubey, 2011). The seroprevalence of this infection in sheep in Newzeland was reported to be 30-90% and in UK 77% prevalence was reported in goats while 29% was reported in sheep (De Bhur, 2008). In Mohmand Agency, Pakistan, the seroprevalence of toxoplasmosis reported in cows was 20% whereas 16% was recorded in buffaloes (Shah *et al.*, 2013a).

T. gondii Infections show mild symptoms or mostly without (Sarkar symptoms et al., 2012). Immunocompetent individuals in acute infection are generally asymptomatic or show some common specific symptoms (Selseleh, 2012), often give flu-like symptoms in the early acute stage (Menotti et al., 2003). While in immunos up pressed patients T. gondii causes encephalitis and leading illness (jones and Roberts, 2013) and infect brain, heart which results in myocarditis, and infection of liver, pancreas, bone marrow, bladder and lymph nodes, kidneys, spleen, and skin (Arnold et al., 1997).

The placental barrier is very strong against the parasite during the first three months of pregnancy, but transmission rate increases as pregnancy proceeds which results in abortions (Dunn, 1999). Congenital toxoplasmosis also causes deafness, mental retardation, micro cephalous, eye lesions, cataract, retinal necrosis (Delair, 2011). Abortion or still birth are common during first three months of pregnancy while Infection acquired in later pregnancy the newborn is usually without symptoms but may cause eye problems (shah et al., 2014) Congenital toxoplasmosis leads to CNS damage, blindness epilepsy and mental growth is retarded with fetal death or spontaneous abortion in about 10% of cases [23]. T. gondii causes abortions, stillbirth and neonatal death in sheep and goats (Engeland et al., 1998).

The distribution of *T. gondii* is related with weather condition of an area and environment where the oocysts survive (Dubey, 2004).

Prevalence of toxoplasmosis varies in different parts of the world and this variation related to life style, age, climatic conditions, nutritional habits and other socio-cultural factors (shah *et al.*, 2014). This study was carried out in the study area with the aim to determine the infection rate in domestic animals, minimize the economic loss in domestic animals and to aware the people about the adverse effects of toxoplasmosis.

Materials and methods

A total of 374 blood samples were collected from domestic animals (cows, goats, sheep and buffaloes) from District Charsadda, Pakistan. About five ml of blood was collected from each animal. The samples were centrifuged for extraction of serum.

Serological examination

The latex regent is a suspension of polystyrene particle which is sensitized with the antigens of *T. gondii*. The distant agglutination pattern when observed after mixing the serum reflects formation of antigen-antibody complexes. When the organism has no infection, then no agglutination is observed. A greater than 4 IU/ml value was considered as standard for positive result.

Test procedure

According to the manufacturer standard protocol (Toxocell Latex- Spain) the test procedure was performed. The samples were diluted in NaCl 0.9% saline solution. One drop or 50ul of diluted sera was mixed with one drop (25ul) of chemical and mixed well by sterile wood stick, then the slide was slowly rotated for 5 minutes and visible agglutination was observed. The result was classified into positive or negative on a slide, using pip stirrers provided. A negative reaction was the indication of the absence of toxoplasma antibodies. A clear positive reaction showed the presence of toxoplasma antibodies equal or greater than 4 IU/ml which showed either an evolving infection or a past infection.

Statistical analysis

For simplification all the results were expressed in percentages. The values between different sex and age groups were recorded and relevantly expressed in percentages. Microsoft Excel (version-10) was utilized by windows-08, (Release 16.0 standard version).

Results

A total of 374 blood samples were collected from domestic animals of three Tehsils (Shabqadar, Tangi and Charsadda) of District Charsadda and were tested for the presence of *T. gondii* infection by using Latex Agglutination Test. Out of these 374 blood samples, 298 (79.7%) were found positive for toxoplasmosis. A total of 100 blood samples were collected from cows in which 72 (72%) were found positive and 28 (28%) were negative, out of 121 goats, 99 (81.8%) were positive and 22 (18.2%) were negative. Similarly, 103 samples were collected from Sheep, out of which 89 (86.4%) were positive and 14(13.6%) were found negative whereas out of 50 samples from buffalo, 38(76%) were found positive and 12(24%) were detected negative (Table 1).

Table 1. Comparative seroprevalence of *T. gondii*infection in domestic animals.

Animals	Samples (N)	Positive N (%)	Negative N (%)
Cows	100	72 (72)	28 (28)
Goats	121	99 (81.8)	22 (18.2)
Sheep	103	89 (86.4)	14 (13.6)
Buffaloes	50	38 (76)	12 (24)
Total	374	298	76

A total of 260 females and 114 males were tested for T. gondii infection. Out of which 215 (82.69%) females 83 (78.80%) males were found positive. This also indicated that prevalence among the females was high among these animals. Prevalence of toxoplasmosis was also detected in different sex groups. The highest prevalence of toxoplasmosis was found in females as compared to males. Out of 13 male buffaloes, 10 (76.92%) were positive while out of 37 females, 28 (75.67%) were found positive. In goats out of 29 males, 21 (72.41%) were found infective whereas out of 92 females, 78 (84.78%) were positive. Out of 33 male sheep 25 (84.78%) were found seropositive and out of 70 female sheep (91.42%) were found infective. Out of 39 male cows, 27 (69.23%) were detected infective whereas out of 61 females, 45 (73.77%) were found seropositive (Table 2).

Animals	Samples Male	Positive N (%)	Female	Positive N (%)
Buffaloes	50	13	10 (76.92)	37 28 (75.67)
Goats	121	29	21 (72.41)	92 78 (84.78)
Sheep	103	33	25 (84.78)	70 64 (91.42)
Cows	100	39	27 (69.23)	61 45 (73.77)
Total	374	114	87	260 215

Table 2. Sex wise seroprevalence of toxoplasmosis among goats, sheep, cows and buffaloes.

Seroprevalence of toxoplasmosis was also detected in different age groups. A total of 178 (81.7%) of age above one year and 120 (76.9%) under one year were detected seropositive. Seroprevalence of toxoplasmosis was higher in age group above one year (Table 3).

Table 3. Age wise seroprevalence of toxoplasmosisamong domestic animals.

Age	Sample (N)	Positive (%)	Negative (%)
> One year	218	178 (81.7)	40 (18.3)
<one td="" year<=""><td>156</td><td>120 (76.9)</td><td>36 (23.1)</td></one>	156	120 (76.9)	36 (23.1)
Total	374	298	76

Discussion

Toxoplasmosis is a zoonotic disease arising from close contact of human with felids (kravetz and federman, 2002). Domestic cats play a vital role in the spread of toxoplasmosis because they are the definitive hosts and play an important role in transmission of *T. gondii* infection by shedding oocysts in their faeces (Dubey, 1994).

It varies between, herds, countries and regions, methods of diagnoses and even at different times in the same herd. In our study, infection rate for *T. gondii* in cattle, goats and sheep was extremely high (79.7%) than 32.29% in domestic animals in Mohmand Agency in Pakistan ((shah *et al.*, 2013a)), 9.2% reported from Guangxi and 46.4% from Xinjiang, China (Lv and Cui, 1994).

In present study 81.81% prevalence rate of *T. gondii* was found in goats, which is higher than 51% in goat's population recorded in Saudi Arabia (Sanad and Al-ghabban, 2007), in Brazil 28.9% (Bisson *et al.*, 2000), Thiland 27.9% (Jittapalapong *et al.*, 2005), 25.4% in Pakistan (Ramzan *et al.*, 2009). Toxoplasmosis was also detected in sheep population of Charsadda. The present study, revealed that in sheep toxoplasmosis is very high (86.40%) than reported from Brazil 46.2% (Silva *et al.*, 2013) and

Greece 48.6% (tzanidakis *et al.*, 2012), 31% reported in Turkey (oncel and Vural, 2006) and Northeastern China 4.4% (Yang *et al.*, 2013), 44.13% from Mardan.

Toxoplasma infection in cattle shows a high degree of variation across the globe, ranging virtually from 0% to 99% (Ivana *et al.*, 2006). Seropositivity rate recorded for *T. gondii* in the present study in cows is extremely high (66%) as compared to 9% in Indonesia (Matsuo and Husin, 1996), while seroprevalence for *T. gondii* in Mohmand Agency, Pakistan was found 20% in cows and 16% in buffaloes (shah *et al.*, 2013a), which is lower than reported in the present study but seroprevalence in present study (66%) is lower than (76.3%) in cattle in Serbia (Ivana *et al.*, 2006).

The differences in prevalence reported by all these studies could be accounted on host, age, breed, sex, environmental conditions, farm size and number of cats and management practices (Van Der *et al.,* 2000). During the present study, a high prevalence of 82.7% was reported in females as compared to males (78%), indicating high prevalence among the females as compared to 35.8% in female and 21.1% in male sheep and goats (Ramzan *et al.,* 2009; Van Der *et al.,* 2000). Different studies indicated that females are more easily infected by protozoan parasites as compared to males (Alexander and Stinson, 1998).

In female immunity can be broken due to various factors nutrition, age, pregnancy e.g., and environmental factors. Toxoplasmosis and age relationship in the present study revealed that seroprevalence is high in age group above one year (81.65%) as compared to age group less than one year (76.92%) which are in support with 77.7% in age group of 60 to 75 months and lowest prevalence 39.3% in age group of 12 to 27 months (Jittapalapong et al., 2005).

Older goats were more seropositive as compared to younger ones under one year old (Olivier *et al.,* 2007).

The present work was an attempt to find out the prevalence rate of toxoplasmosis in the study area. The high prevalence rate of toxoplasmosis in the study area may be due to warm and humid climatic conditions of this area. The increased infection rate in study area may also be due to unhygienic conditions because *T. gondii* prevalence ranged from 0-100% in different areas of the world and this variation is due to the life styles of the inhabitant's customs, traditions, weather conditions, age of the animals and husbandry practices (Olivier *et al.*, 2007).

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References

Afonso E. 2008. Spatial distribution of soil contamination by T. gondii in relation tocat defaecation behaviour in an urban area. International Journal of Parasitology **38**, 1017-1023. https://doi. org/10.1016/j.ijpara. 2008.01.004.

Aldebret D, Hypolite M, Cavaillaes P, Touque B, Flori P, Loeuillet C, Cesbron-Delauw MF. 2011. Development of High-Throughput methods to quantify cysts of T. gondii. Cytometry Part A **79(11)**, 952-958.

https://doi.org/10.1002/cyto.a.21138.

Alexander J, Stinson WH. 1988. Sex hormones and the course of parasitic infection. Parasitology Today 4, 189-193.

https://doi.org/10.1016/0169-4758(88)9

Arnold SJ, Kinney MC, Mccormick MS, Dummer S, Scott MA. 1997. Disseminated toxoplasmosis. Unusual presentations in the immunocompromised host. Archives of Pathology and Laboratory Medicine **12(8)**, 869-873.

Bisson A, Maley S, Rubaire-Akiiki CM, Watling JM. 2000. The seroprevalence of antibodies to T. gondii in domestic goats in Uganda. Acta Tropica **76**, 33-38. **De Bhur K.** 2008. T. gondii seroprevalence –current results in German swine herds. Archive Für Lebensmittelhygiene **59**, 5-8.

Delair E. 2011. Clinical manifestations of ocular toxoplasmosis. Ocular Immunology and Inflammation **19**, 91-102.

Dubey JP. 1994. Toxoplasmosis. J. American Veterinary Medical Association **205**, 1593-1598.

Dubey JP. 2004. Toxoplasmosis – a waterborne zoonosis. Veterinary Parasitology, **126**, 57-72. https://doi.org/10.1016/j.vetpar.2004.09.005.

Dubey JP. 2011. High prevalence and genotypes of Toxoplasma gondii isolated from goats, from a retail meat store, destined for human consumption in the USA. Internatinal Journal for Parasitology **41**, 827-833. https://doi.org/10.1016/j. ijpara.2011.03.006.

Dunn D. 1999. Mother-to-child transmission of toxoplasmosis risk estimates for clinical counselling. Lancet **353**, 1829-1833. https://doi.org/10.1016/S01.

Engeland IV, Waldeland H, Andresen O, Løken T, Björkman C, Bjerkås I. 1998. Foetal loss in dairy goats: an epidemiological study in 22 herds. Small Ruminant Research **30**, 37-48. https://doi.org/10.1016/S0921-4488(98)00084-4.

Halos L. 2010. An innovative survey underlining the significant level of contamination by T. gondii of ovine meat consumed in France. International Journal for Parasitology **40**, 193-200. https://doi.org/10.1016/j.ijpara.2009.06.009.

Holliman RE. 1997. Toxoplasmosis, behaviour and personality. Journal of Infection **35**, 105-110. https://doi.org/10.1016/S0163-4453(97)91380-3.

Innes EA. 2010. A brief history and overview of T.gondii. Zoonoses and Public health 57, 1-7. https://doi.org/10.1111/j.1863-2378.2009.01276.x.

Ivana L, Olgica DD, Sofija KK, Aleksandra N. 2006. Cross sectional survey of T. gondii infection in cattle, sheep and pigs in Serbia: seroprevalence and risk factors. Veternary Parasitology **135**, 121-131.

Jittapalapong S, Sangvaranond A, Pinyopanuwat N, Chimnoi W, Khachaeram W, Koizumi S, Maruyama S. 2005. Seroprevalence of T. gondii infection in domestic goats in Satun Province, Thailand. Veternary Parasitology **127**, 17-22.

Jones JL, Roberts JM. 2013. Toxoplasmosis hospitalizations in the United States. 2008, and Trends, 1993-2008. Clinical Infectious Disease **54(7)**, e58-e61.

Khan SN, Khan S, Ayaz S, Jan AH, Jehangir S, Attaullah S, Ali J, Shams S. 2011. Seroprevalance and risk factors of toxoplasmosis among pregnant women in district Kohat, Khyber Pakhtunkhwa Pakistan. World Applied Science Journal **14(7)**,1032-1036.

Kravetz JD, Federman DG. 2002. Cat-associated zoonoses. Archives of international Medicine **162**, 1945-1952. DOI: 10.1056/NEJMcpc059027.

Lilenbaum W. 2013. Immunohistochemical identification of T. gondii in tissues from Modified Agglutination Test positivesheep. Veterinary Parasitology **191**, 347-52.

Lv YC, Cui JZ. 1994. Survey of T. gondii infection in pigs and cattle in Guangxi Province, China. Journal of Animal Science and Veterinary Medicine **3**, 26.

Matsuo K, Husin DA. 1996. Survey of T. gondii antibodies in goats and cattle in Lampung Province, Indonesia.Southeast Asian. Journal of Tropical Medicine and Public Health **27**, 554-5.

Menotti, Gustavo Vilela, Stephane Romand, Yves Jean-Francois Garin, Lionel Ades, Eliane Gluckman, Francis Derouin, Patricia Ribaud. 2003. Comparison of PCR-Enzyme-linked immunosorbent Assay and Real-Time PCR Assay for diagnosis of an unusual case of cerebral Toxoplasmosis in stem cell transplant recipient. Journal of Clinical Microbiology **41**, 5313-5316. **Olivier A, Herbert B, Sava B, Pierre C, John DC, Aline DK.** 2007. Surveillance and monitoring of Toxoplasma in humans, food and animals: a scientific opinion of the panel on biological hazards. The European Food Safety Association Journal **583**, 1-64.

Oncel, Vural T. 2006. Occurance of T. gondii antibodies in sheep in Istanbul, Turkey. Veteninarski Arthiv **76**, 547-557.

Ramzan M, Akhtar M, Muhammad F, Hussain I, Hiszczyńska- Sawicka E, Haq AU, Mahmood MS, Hafeez MA. 2009. Seroprevalence of T. gondii in sheep and goats in Rahim Yar Khan (Punjab), Pakistan. Tropical.Animal Health and Production **41**, 1225-9.

Sadaruddin A, Agha F, Anwar F, Ghafoor A. 1991. Seroepidemiology of T. gondii infection in young school children in Islamabad. Journal of Pakistan Medical Association **41**, 131-134.

Sanad MM, Al-ghabban AJ. 2007. Serological survey on toxoplasmosis among slaughtered sheep and goats in Tabouk, Saudi Arabia. Journal of Egyptian Society of Parasitology **37**, 329-340.

Sarkar MD, Anuradha B, Sharma N, Roy RN. 2012. Seropositivity of toxoplasmosis in antenatal women with bad obstetric history in a tertiary-care hospital of Andhra Pradesh, Indian Journal of Health Population and Nutrition **30**, 87-92.

Selseleh M, Modarressi MH, Ali MM, Shojaee S, Eshragian MR, Selseleh M, Aziz E, Keshavarz H. 2012. Real-Time RT-PCR on SAG1 and BAG1 gene expression during stage conversion in immunosuppressed mice infected with T. gondii, Tehran. Strain. Korean. Journal of Parasitology **3**, 199-205.

https://dx.doi.org/10.3347%2Fkjp.2012.50.3.199.

Shah M, Zahid M, Asmat P, Sthanadar AA. 2013b. Seroprevalence of T. gondii in goats and sheep of district Mardan, Pakistan. International Journal of Bioscience **3**, 90-97.

http://dx.doi.org/10.12692/ijb/3.7.90-97.

Int. J. Biosci.

Shah M, Zahid M, Bibi B, Hussain A, Haroon M, Ali B. 2017. Chromatographic immunoassay based detection of of human toxoplasmosis in District Mardan, Khyber Pakhtunkhawa, Pakistan. Pure and Applied Biology **6**,1297-1305.

Shah M, Zahid M, Sthanadar AA, Ali PA. 2014. Seroprevalence of Toxoplasma gondii Infection in Human Population of Mohmand Agency Khyber Pakhtunkhwa, Pakistan. Pakistan Journal of Zoology **44**, 1169-1172.

Shah M, Zahid M, Sthanadar AA, Pir A, Kausar A, Jan AH. 2013a. Seroprevalence of T. gondii infection in domestic animals of Mohmand agency, Pakistan. Journal of Coastal Life Medicine 1, 70-73.

DOI: 10.12980/JCLM.1.2013C254.

Silva AF, Oliveira FC, Leite JS, Mello MF, Brandão FZ, Leite RI, Frazao-Teixeira E, Tasawar Z, Nawaz S, Lashari MH, Aziz F, Hayat CS. 2011. Seroprevalence of human Toxoplasmosis in Dera Ghazi Khan, Punjab. Gomal Journal of Medical Sciences 9, 82-85.

Tenter AM, Heckeroth AR, Weiss LM. 2000. T. gondii: from animals to humans. International Journal of Parasitology **30**, 1217-1258. https://doi.org/10.1016/S0020-7519(00)00124-7.

Teshale S, Dumètre A, Dardé ML, Merga B, Dorchies P. 2007. Serological survey of caprine toxoplasmosis in Ethiopia: prevalence and risk factors. Parasite **14**, 155-159.

https://doi.org/10.1051/ parasite/2007142155.

Tzanidakis N, Maksimov P, Conraths FJ, Kiossis E, Brozos C, Sotiraki S, Schares G. 2012. T. gondii in sheep and goats: seroprevalence and potential risk factors under dairy husbandry practices. Veterinary Parasitology **190**, 340-348. https://doi.org/10.1016/j.vetpar.2012.07.020.

Van Der Puije Wna, Bosompem KM, Canacoo EA, Wastling, Zakanmoribd JM. 2000. The prevalence of anti-T. gondii antibodies in Ghanaiansheep and goats. Acta Tropica 76, 21-26. https://doi.org/10.1016/S0001-706X(00)00084-X.

Webster JP. 2007. The effect of T. gondii on animal behavior: Playing cat and mouse. Schizophrenia bulletin **33(3)**, 752-756.

Yang N, Li H, He J, Mu M, Yang S. 2013. Seroprevalence of T. gondii infection in domestic sheep in Liaoning Province, northeastern China. Journal of Parasitology **99**, 174-5.