

International Journal of Biosciences | IJB | ISSN: 2220-6655 (Print) 2222-5234 (Online) http://www.innspub.net Vol. 14, No. 3, p. 477-481, 2019

RESEARCH PAPER

OPEN ACCESS

Screening of mastitis in dairy cattle's of district Khairpur, Sindh, Pakistan

Majeeda Ruk¹, Javed Ahmed Ujan^{*1}, Sham Lal², Majida Parveen Narejo¹, Sadaf Jumani¹

¹Department of Zoology, Shah Abdul Latif University, Khairpur, Sindh, Pakistan ²Department of Microbiology, Shah Abdul Latif University, Khairpur, Sindh, Pakistan

Key words: Cattle, CMT, Staphylococci, Sub-clinical, Khairpur

http://dx.doi.org/10.12692/ijb/14.3.477-481

Article published on March 31, 2019

Abstract

Dairy cattle mastitis is disease that is usually caused by bacterial infections in udder tissue. When the milk production and quality are reduced, the disease causes considerable damage to cattle. Early detection and corrective actions may lead to early cure. Mastitis can be clinical or subclinical. Clinical mastitis can cause udder milk abnormalities and swelling of udder. A clinical mastitis case reaches 240 million annually, causing huge economic losses and other losses in Pakistan. Therefore, this study aimed to screen out mastitis in cattle present in different farms of district Khairpur. Out of 400 milk samples tested, 298samples were positive for mastitis as indicated by using world-recognized somatic cell counting test (SCC), California Mastitis Test (CMT). Bacteria were isolated and identified based on cultural, microscopic and biochemical tests. Our results show that among the animals reared in the backyard showed highest number ((34.5%), and subsequently surrounding areas (33.5%) and other farms (31.75%) cases of mastitis. In addition, one hundred eighty-three bacterial isolates belonging to 6 different genera i.e *Staphylococci* (37%), *Escherichia* (21%), *Streptococci* (17%), *Pseudomonas* (13%), *Klebsiella* (7%) and *Bacillus* (5%) were isolated. The study showed that routine testing of cattles and preventive measures are recommended to reduce the outbreaks of clinical and subclinical mastitis in Khairpur, Sindh, Pakistan.

* Corresponding Author: Dr. Javed Ahmed Ujan \boxtimes javed.ujan@salu.edu.pk

Introduction

The Milk consumed by us depend on its quality, but bacterial invasion to milk producing glands resultantly making this milk unfit for consumers, Mastitis is an infection caused by bacteria commonly known as Staphylococcus aureus. S. aureusis Gram positive bacteria capable to survive in high salt and high temperature range appearing as an opportunistic pathogen and frequent colonizer of the epithelial tissues cause devastating diseases in humans and domestic animals. The organism is of zoonotic importance that becomes more serious in its resistant form against antimicrobial agents (Barkema et al., 2009; Hiramatsu et al., 2011) Mastitis is one of the serous maladies in the cattles causing huge production losses to livestock industry in Pakistan (Ali et al., 2011). Bovine mastitis hence a major problem in dairy cattles of Khairpur, in those who are kept under various farming systems or domesticated both deserves attention due to its potential impact on milk production, safety and its security. Mastitis causes two third losses of the total milk production due to affected quarters of animal (Radostits et al., 2007). Most common pathogens isolated from mastitis milk are contagious (Bilal et al., 2004). According to a report 70-80% of all the bovine mastitis is generally caused by either infected with Staphylococcus aureus or Streptococcus agalactiae (Memon et al., 1994). Bovine mastitis is diagnosed by (CMT) California Mastitis Test, White side test (WST) or Surf field mastitis test and somatic cell counts (Muhammad et al., 2010). The current study observed mastitis in cattle's of three talukas named Gamabt, Theri bypass and Khairpur city of district Khairpur. Previously no any research work has been done on chemical analysis and microbial screening of isolates of mastitis at Khairpur district Sindh, Pakistan. Present study aimed to screened-out 400 cattle's from different farms of district Khairpur, through the world-recognized somatic cell counting test (SCC) / California Mastitis Test (CMT).

Material and methods

Selection of study area and study animals

Two hundred ninety-eight lactating cattle from the three areas (Gambat, Theri bypass and Khairpur city, Sindh,

Pakistan) were observed. A total of 298 milk samples were collected in sterilized screw caped test tubes.

Physical examination of milk samples

Afterwards the sample collection, an on-farm screening was performed, where samples were subjected to physical examination with naked eyes for detection of any change in color of milk, order, atrophy, any swelling in nodes, consistency or presence of any blood clotting.

Diagnostic test used

Mastitis was detected by California mastitis test (CMT), which is most common type of the test used for diagnosis of clinical and sub clinical mastitis (Qunin *et al.*, 1994).

Transportation of milk samples

Strongly positive results (+++) in CMT were transferred to Microbiological laboratory of Shah Abdu latif University Khairpur in thermopile box.

Bacterial isolation from Milk

Before incubation, the sample was allowed in normal temperature, then 100ul of milk sample was mixed with an equal volume of nutrient broth with the help of pipette and allowed for incubation on 37°C for 24hrs.After incubation, one loop of incubated sample was streaked on nutrient agar and again incubated on37°C for 24hrs. as described by (Lafi and Hailat, 1998). The bacterial colonies were then identified based on cultural and morphological characters (Jhon., 2000).

The prevalence

The prevalence was expressed in percent by using the following formula:

Prevalence (%) = No. of Animals Positive / No. of Animals Tested×100

Results and discussion

Mastitis is an Inflammatory Infections (IMI) usually caused *by Staphylococcus aureus*, which has a wide range host spectrum and can cause serious infection and economic loss (Kenar *et al.*, 2017). In this study, then countered bacterial genera including Staphylococci (37%), Escherichia (21%), Streptococci (17%), Pseudomonas (13%), Klebsiella (7%) and Bacillus (5%) (Giraudo., et al., 1997). The bovine mastitis and its control may be depending on detection of various species found during study and elimination of different host which are associated with the risk factor in mastitis. Amongst multiple reasons of mastitis, sustained level of wide variety of microbes, particularly different bacterial species is the most vital one; it is observed that Staphylococcus aureus, is the most dominating microbial bacteria casing bovine mastitis. A cell count of somatic cells usually in milk is called somatic cell count (SCC), The greater the SCC, the higher level of inflammation in udder tissue. It is because diapedesis of leukocytes is localized, then only udder tissue quarter that is infected will have significant increase in the concentration of leukocytes (SCC). Table 1 shows the prevalence of mastitis in cattle's of different cities of district Khairpur.

Table 1. Prevalence of mastitis screened by (CMT) Test.

Area	No of positive sample (CMT)	Positive Bacterial Growth	Percentage (100%)	No. of Isolated Bacteria
Kairpur city	108	61	36.2	6
Their bypass	92	55	30.8	4
Gambat	98	59	32.8	5
Total	298		100	

Table 2. Above table Showed the prevalence of mastitis screened by California mastitis test (CMT) out of 298 samples majorly, Khairpur city possess highest no 108CMT positive and 92isolated bacteria, followed by Their bypass with 92 positive CMT and Gambat taluka possess 98 positive samples.

Table 3. Form wise prevalence of mastitis in cattlesscreened by (CMT) Test.

Type of form	Total no of cattles	Khairpur	Thehri bypass	Gambat	Prevalence Percent
Backyard	134	48	43	51	(34.5%)
Surrounding areas	133	44	46	44	(33.5%)
Other farms	133	44	44	39	(31.75%)
Total	400				

Table 3 in this table the forms which were under observation specially Backyard showed highest number positive samples in cattle's that were about 34.5%, followed by surround areas that was 33.5 positive cattle's for mastitis it was 31.75% other forms were positive for mastitis.

Table 4. Bacterial genus wise prevalence of mastitisfrom three areas of Khairpur district.

Bacterial spp.	No	(%)
Staphylococci	37	(37%),
Escherichia	21	(21%),
Streptococci	17	(17%)
Pseudomonas	13	(13%),
Klebsiella	7	(7%)
Bacillus	5	(5%)

Table 4 showed prevalence of bacteria as Staphylococci 37%, Escherichia 21%, Streptococci 17% Pseudomonas (13%), Klebsiella (7%) Bacillus (5%) at district Khairpur. Prevalence of mastitis was assessed from population of cattles raised under forms of district Khairpur (CMT) test. The prevalence was higher in Khairpur followed Gambat (%) and Theri bypass 45(%) forms. It shows difference due to hygienic and management poor conditions. Cumulative percentage of mastitis in cattles observed was 44% which is much lower than 92% recorded by (Lafi and hailat., 1998) Similarly, an incidence of 54.7, 32.85 and 23.18% has been reported by (Getahun et al., 2007; Pitkalae et al., 2004; Iqbal, et al., 2004) respectively. Variation in prevalence of mastitis may be due the region climatic condition, environment and the management condition. The area wise prevalence of bovine mastitis is presented in Table 2. The results showed highest prevalence in Gambat city. Prevalence recorded was the highest (36%) at Gambat followed by their bypass (55%) and Kairpur city (45%) least no of mastitis found in Khairpur. It may be due to availability of veterinary hospitals in Khairpur city and quality medicine provided to different forms. Mastitis is caused by different bacterial genera as Staphylococcus, Streptococcus and Escherichia bovine mastitis (Allore, 1993; Ahmad, 2001). Bacterial genera found during this research work were Staphylococci 37%, Escherichia 21%, Streptococci 17% Pseudomonas 13%, Klebsiella 7%, Bacillus 5%. The data of bacterial isolates is presented in Table 3. In the recent study, bacterial growth was observed in milk samples. These identified Bacteria were of seven different genera.

(Pitkala *et al.*, 2004) reported microbial growth in 21-33% of milk samples, whereas, (Iqbal *et al.*, 2004) reported only 15.16% in dairy buffaloes. This variation may be due to season, management conditions at the farm, area, transportation conditions, difference in sample handling in the laboratory and use of antibiotic.

In present study, S. aureus (37%) was isolated as topranking pathogen from positive cases for mastitis. In previous studies, it was also reported as major pathogen (Kapur et al., 1992; Allore., 1993; Rabello et al, 2005; Arshad et al., 2006; Ebrahimi et al., 2007; Ali et al., 2008; Botrel et al., 2009; Ebrahimi et al., 2007) reported 8.33% Streptococcus agalactiae and 9.44% E. coli isolates from subclinical bovine mastitis milk samples while Ali et al. (2008) obtained 30% growth of Strep. agalactiae and Strep. dysgalactiae Contaminated and 42.6% Staph. aureus. environment of farm is a main source of coliforms and mostly cause clinical infections. (Ebrahimi et al., 2007) obtained 3.88% Coagulase-negative Staphylococci (CNS), 8.33% Streptococci other than agalactiae and 9.44% E. coli. (Botrel et al., 2009) isolated 30.2% CNS, 13.7% coagulase negative Staphylococci and 9.3% Strep. dysgalactiae from subclinical mastitis milk sample. The table 1. Displays three areas variation in which Gambat had a highest no of positive mastitis cases including 61 isolated bacteria. It showed prevalence of mastitis in three areas of Gambat, Theri bypass and Khairpur city. Table 2. Showed mastitis screened by California mastitis test (CMT) at Khairpur city, where no. of 108 mastitis positive cases gave 61 no. samples yielded bacterial growth. While Gambat bypass 98 no. positive sample yielded 80 bacterial growth. Table 3 no. of positive mastitis animals according to the different forms visited during research work. Backyard showed 133 mastitis positive its about 54%. While surrounding areas were 42% and other forms 32% respectfully. Bacterial species wise prevalence of mastitis from three area of Khairpur district the Staphylococci was 37%, Bacillus was lowest no. of 5% species found in above areas. There should be mastitis control programs, which can help to control and to evaluate the forming system in the district Khairpur. Keeping in view the animal and human

health hazard anchored with this pathogen, the current study was designed to estimate the outbreak of mastitis in the regions of Khairpur district.

References

Ahmad R. 2001. Studies on mastitis among dairy buffaloes. Pakistan Veterinary Journal **21**, 220-221.

Ali L, Muhammad G, Arshad M, Saqib M, Hassan IJ. 2008. Bacteriology of mastitis in buffaloes in tehsil Samundri of district Faisalabad, Pakistan. Pakistan Veterinary Journal **28(1)**, 31-33.

Ali MA, Ahmad MD, Muhammad K, Anjum AA. 2011. Prevalence of sub clinical mastitis in dairy buffaloes of Punjab, Pakistan. Okara **150**, 63-42.

Allore HG. 1993. A review of the incidence of mastitis in buffaloes and cattle. Pakistan Veterinary Journal **13**, 1-7.

Barkema HW, Green MJ, Bradley AJ, Zadoks RN. 2009. Invited review: The role of contagious disease in udder health. Journal of Dairy Science **92(10)**, 4717-29.

Bilal MQ, Iqbal MU, Muhammad G, Avais M, Sajid MS. 2004. Factors affecting the prevalence of clinical mastitis in buffaloes around Faisalabad district (Pakistan). International journal of agriculture and biology **6**, 185-7.

Botrel MA, Haenni M, Morignat E, Sulpice P, Madec JY, Calavas D. 2010. Distribution and antimicrobial resistance of clinical and subclinical mastitis pathogens in dairy cows in Rhône-Alpes, France. Foodborne pathogens and disease **7(5)**, 479-87.

Ebrahimi A, Kheirabadi KH, Nikookhah F. 2007. Antimicrobial susceptibility of environmental bovine mastitis pathogens in west central Iran. Pakistan journal of biological sciences **10(17)**, 3014-6.

Getahun K, Kelay B, Bekana M, Lobago F. 2008. Bovine mastitis and antibiotic resistance patterns in Selalle smallholder dairy farms, central Ethiopia. Tropical Animal Health and Production **40(4)**, 261-8.

Int. J. Biosci.

Giraudo JA, Calzolari A, Rampone H, Rampone A, Giraudo AT, Bogni C, Larriestra A, Nagel R. 1997. Field trials of a vaccine against bovine mastitis. Evaluation in heifers. Journal of Dairy Science **80(5)**, 845-53.

Hiramatsu K, Cui L, Kuroda M, Ito T. 2001. The emergence and evolution of methicillin-resistant Staphylococcus aureus. Trends in microbiology **9(10)**, 486-93.

Iqbal M, Khan MA, Daraz B, Siddique U. 2004. Bacteriology of mastitic milk and in vitro antibiogram of the isolates. Pakistan Veterinary Journal **24(4)**, 161-4.

John GH. 2000. Bergey's manual of determinative bacteriology. Actinomycetales. 9th ed. Williams and Wilkins, Baltimore.

Kapur MP, Anshusharm, Bahardwal RM. 1992. Bacteriology of clinical mastitis in buffaloes. Buffalo Bull **11**, 32-35.

Kenar B, Bagcigil AF, Kuyucuoglu Y, Kahraman BB, Konak S. 2017. Antimicrobial susceptibility profiles and coagulase gene polymorphism of Staphylococcus aureus isolated from bovine subclinical mastitis. Kafkas Universitesi Veteriner Fakultesi Dergisi **23(4)**.

Lafi SQ, Hailat NQ. 1998. Bovine and ovine mastitis in Dhuleil valley of Jordan. Veterinarski Arhiv **68(2)**, 51-7.

Memon MI, Mirbahar KB, Memon MR, Akhter N, Soomro SA, Dewani P. 1999. A study on the etiology of subclinical mastitis in buffaloes. Pakistan Journal of Agriculture, Agricultural Engineering Veterinary Sciences (Pakistan) **15,** 34-36. **Muhammad G, Naureen A, Asi MN, Saqib M.** 2010. Evaluation of a 3% surf solution (surf field mastitis test) for the diagnosis of subclinical bovine and bubaline mastitis. Tropical animal health and production **42(3)**, 457-64.

Pitkala A, Haveri M, Pyorala M, Myllys S, Buzalski TH. 2004. Bovine Mastitis in Finland, Prevalence, Distribution of Bacteria, and Antimicrobial Resistance. Journal of dairy science 87, 2433-2441.

Quinn PJ, Carter ME, Markey BK, Carter GR. 1994. Clinical Veterinary Microbiology. Wolfe Publishing. London, UK. 648p.

Rabello RF, Souza CR, Duarte RS, Lopes RMM, Teixeira LM, Castro AC. 2005. Characterization of Staphylococcus aureus isolates recovered from bovine mastitis in rio de janeiro, brazil. American Dairy Science Assoc 234-345.

Radostits OR, Blood DC, Gay CC. 2007. Mastitis. Veterinary Medicine: Atextbookof the diseases of cattle, horses, sheep, pigs and goats, 9th Edn., Bailertindall, London **pp.** 563-614

Razzaq A. 1998. Comparative efficacy of Vetimast, Tetra-Delta, and Akamycin-D in mastitis of buffalo in and around Lahore. M.Sc. Thesis, College of Vet. Science, Lahore, University of Agriculture, Faisalabad