



Microbial cell as bio-factory for bioactive compounds: review on microbial nutraceuticals

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Abstract

Now a days it is a challenging and very serious issue to meet an adequate supply of quality food for growing population which is exploding at a geometric progression. Nutrient deficient diet is one of the major cause of constantly increasing health related problems. To resolve the health issues caused by malnourishments, nutraceutical like polysaccharides, multi-vitamins and enzymes have been produced from plants since many years. However, use of plant-derived nutraceuticals is limited due to low growth rate, seasonal variation, plant diseases or other environmental influences. Instead, microorganisms can be used for rapid, ecofriendly and economically nutraceuticals production in less space and generation time. Scientists are focusing on cultivation of useful microbial strains that have the potential to produce metabolites possessing pharmaceutical as well as nutrition value (nutraceuticals). This modern approach can compensate for food demand (quality diets) and disease curing. Various bacteria, fungi, algae and yeast have been cultivated for the production of the bioactive compounds. Such useful microbial extracted compounds are considered to be food graded healthy nutrition or nutraceuticals. This advance research can bring revolution in the field of medicine and nutrition.

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Introduction

Proper nutrition is an indispensable requirement for growth, development and normal body functions of human beings. Though, getting sufficient nutrition is of great concern even one is taking enough food. Here and now, lack of proper nutrition is a global issue as one third population suffers from malnutrition (Avula, 2015). In the world, due to malnutrition, significant number of precious life is lost annually with about 3 million young lives every year (UNICEF, 2016). Malnutrition may leads to poor growth and development of children, skin and bone disease, overweight, high level of sugar, fats or cholesterol and salt in blood, minerals and vitamin deficiency, or serious complications in immunocompromised people. Intensification of various diseases due to malnutrition and poor diet are becoming the major risks for health throughout the world. Policies have been established at world level to improve nutrition and to completely finish the malnutrition till 2030

(UNICEF, 2016). To overcome the problems of the malnutrition, different phytochemicals have been used as nutraceuticals. The terms “nutrition and pharmaceuticals” were put together to derive the word ‘nutraceuticals’ by Dr Stephen de Felice which categorically describe the food or products of the food, used to improve health, prevent and treat various health (Biesalski, 2001). In the course of recent years various new diet ingredients have been introduced to nutrition and pharmaceutical market as nutraceuticals. Bacteria, yeast and micro algae can act as producers or catalysts for the synthesis of food ingredients, compounds like enzymes and nutraceuticals (Hugenholtz and Smid, 2002).

A potential nutraceutical is one that benefits the health as shown in table. 1, and for this purpose various studies have been carried out to analyze the potency of nutraceuticals for biological advantages and provide protection against chronic diseases.

Table 1. Beneficial metabolites produced from microorganisms.

Microorganism	Nutraceuticals produced	Effects on Humans	References
Protein			
<i>Lactobacillus acidophilus</i>	Leptinacidolin, Acidophilin, Lactocidin and Bacteriocin	Anti-obesity, improvements in the digestive system	Sousa <i>et al.</i> , 2008; Gupta <i>et al.</i> , 2000
<i>Lactobacillus rhamnosus</i> GG (LGG)	Specific protein called p75 and p40	Prevention of inflammatory bowel disorders (IBD)	Gupta <i>et al.</i> , 2000; Kassinen <i>et al.</i> , 2007
<i>Escherichia.Coli</i>	GLP-1	Triggers cells in the pancreas to make insulin	Bronzwaer, 2008; Cani <i>et al.</i> , 2009
<i>Escherichia intermedia</i> , <i>Ervinia herbicola</i>	L-DOPA	Brain neurotransmitter	Kumagai <i>et al.</i> , 1970; Kumagai <i>et al.</i> , 1972
Carbohydrates			
<i>Agrobacterium radiobacter</i>	Sulphated derivatives of curdlan	Antitumor and antiviral	Zhan <i>et al.</i> , 2012
<i>Aerobacter levanicum</i> , <i>Zymomonas mobilis</i> , <i>Lactobacillus sanfranciscensis</i> , <i>Bacillus subtilis</i> , <i>Bacillus polymyxa</i> , <i>Acetobacter xylinum</i> , <i>Gluconoacetobacter xylinus</i> ,	Levan	Anti-obesity, hypolipidaemic, antitumor and anti-radiation protective	Kang <i>et al.</i> , 2004; Yoo <i>et al.</i> , 2004; Bekers <i>et al.</i> , 2005; Combie, 2006
<i>Elsinoe leucospila</i>	Elsinan	Cholesterol-lowering and Antitumor	Shirasugi and Misaki, 1992; Misaki, 2004
<i>Agaricus bisporus</i> , <i>Auricularia auricula</i> , <i>Flammulina velutipes</i> , <i>Lentinus edodes</i>	Edible mushroom glucans	Antioxidant	He <i>et al.</i> , 2012
<i>S. cerevisiae</i>	Zyosan	Anti-inflammatory	Ohno <i>et al.</i> , 2001; Goodridge <i>et al.</i> , 2009
<i>Azotobacter vinelandii</i> , <i>Azotobacter chroococcum</i>	Bacterial alginate	Anti-inflammatory (Anti-ulcer)	Sabra <i>et al.</i> , 2001
Fatty acids			
<i>Lactobacillus species</i>	Conjugated linoleic acid (CLA), Conjugated linolenic acid (CLnA) and 10-Hydroxy-cis-12-octadecenoate (HYA)	Reduces adiposity, Improves insulin sensitivity, Anti-cancer, Reduces atherosclerosis,	Belury, 2000; Bergamo <i>et al.</i> , 2014

		Anti-inflammatory	
<i>Mortierella alpina</i>	Arachidonic acid (ARA)	Infant nutrition	Casterton <i>et al.</i> , 2009
<i>Isochrysis galbana</i>	Eicosapentaenoic acid	Treating obesity, metabolic syndrome, nonalcoholicsteatohepatitis and type-2 diabetes	Baker <i>et al.</i> , 2008; Wen and Chen, 2003
		Vitamins	
<i>Pseudomonas denitrificans</i> and <i>Propionibacteriumshermanii</i>	Vitamin B12	Protection against anemia	Kassinen <i>et al.</i> , 2007
<i>Lactococcus acidophilus</i>	Vitamin K	Prevention of inflammatorybowel disorders and colitis	Gupta <i>et al.</i> , 2000
<i>Blakeslea trispora</i> and <i>Duniella salina</i>	β -carotene (provitamin A)	Antioxidants	Nelis and de Leenheer, 1991

With the evolution and improvement of green technology and production of high yielding cereals varieties, major advancements have been done to food production in last 30 years.

The malnourished proportion of population ranged 46.5% (in the early 1960) to 26.7% (2000) in the under developed countries of the world (Lipton, 2001; Clugston and Smith, 2002).With the present situation of malnutrition and the impact of global increase in population, new polices are indispensable to overcome the food deficiencies and to meet the future demands for world food and nutrients. According to the United Nations (UN) report, there will be a huge change/increase occur in the world's population with growth from 8 billion to 9.4 billion in 2025 and 2050 respectively (93% of the increase in population will occur in the developing countries) (Clugston and Smith, 2002).

Dietary intake of phytochemicals used from decades may promote health benefits, protecting against cancer, cardiovascular, neurodegenerative diseases, oxidative stresses, inflammatory diseases, hypertension, hyperlipidaemia and renal failure (Osawa, 2007; Wojcikowskiet *al.*, 2006), butthe major risks associated with phytochemicals are the lack of availability of required plant parts due to slow growth, seasonal variation and biotic and abiotic factors.

Therefore, alternate ways have been found for the nutraceutical production from beneficial microbes that can replace the plant based phytochemicals due to cost effectiveness, eco environmentally friendly and

short space and generation time (Fig. 1).

Microbial production of substances, for example, amino acids, biocatalysts, proteins, vitamins, anti-microbial and hydrocolloids additionally stays essential to promote health (Dufosse, 2009).

Lactobacillus microbes, specifically *Lactococcus lactis*, have been shown to be perfect cells for the generation of nutraceuticals. Microalgae and cyanobacteria also act like biological machineries for the production of vitamins, ω -3 unsaturated fats, proteins, pigments (Pulz and Gross, 2004).

Various microbial species and aerobic microorganisms like *Corynebacterium glutamicum* is considered one of the best source for the production of antioxidants like Superoxide dismutases (SODs) (El Shafey *et al.*, 2008).

Improvement in the genetic abilities of the food graded microorganisms indicates that the production of certain nutraceuticals can be upgraded through over expression of the genes or mutating the microbial metabolic qualities (Hugenholtz andSmid, 2002).

Bacterial cells are genetically engineered to increase the production of human insulin (Humuline), which is then purified and used for diabetes treatment (Barsh *et al.*, 2000; Bajzer and Seeley, 2006).

Important nutraceuticals as metabolites isolated and purified from various microorganisms in a variety of mechanisms are detailed in the following headings.

Table 2. Bioactive amine molecules produced via various microbial enzymes.

Amino acids/Peptides/Derivatives	Enzymes	References
Glutamylpeptides	Glutamylcysteine synthetase	Nakayama <i>et al.</i> , 1981
Glutamylmethyl amide and theanine	Glutamine synthetase	Yamamoto <i>et al.</i> , 2005
Aspartic acid	Aspartase	Sato and Tosa, 1993
Alanine	L-aspartic acid β -decarboxylase	Chibata <i>et al.</i> , 1965
DOPA	Tyrosine Phenol-Lyase	Kumagai <i>et al.</i> , 1970;
Hydroxyphenylglycine	Hydantoinase	Yamada <i>et al.</i> , 1978
Hydroxy-L-proline	Proline 4-hydroxylase	Shibasaki <i>et al.</i> , 2000
Aspartame	Thermolysin	O'Donnell, 2006
Glutamyl-L-tryptophan	Glutamyltranspeptidase	Suzuki <i>et al.</i> , 2004

*Microbial biosynthesis**Amino acids and derivatives*

Amino acids are the monomer or precursor for the proteins. There are twenty amino acids and several of their derivatives have been identified for their role in maintenance of health. Body will face a number of disorders upon missing any of the amino acids.

Amino acids are considered to be the building block of life that have long been performed significant role in the nutrition and health maintenance of both animal and human (Bercovician and Fuller, 1995).

Nitrogen-bearing molecules, for example, oligopeptides, amino acids and its derivatives have gotten unusual consideration by influencing human physiology in different ways as shown in Table 1.

Microorganisms possess wide range of useful properties and such microbial strains can be used for the human wellbeing. In 1957, Shukuo Kinoshita and his partners obtained amino acids from direct fermentation technique using *Corynebacterium glutamicum* also known as *Micrococcus glutamicus* (Kinoshita *et al.*, 1957a).

According to an estimation more than 2.3 million tons of amino acids were produced in 2000 by using microorganisms or bacterial enzymes (evaluated by the Essential Amino Acid Association, Japan).

As some amino acids can't be prepared in enough amount by just simple fermentation, enzymatic strategies utilizing microbial catalysts were discovered.

Table 3. Food graded polysaccharides produced and isolated from microorganisms.

Microorganisms	Products	References
<i>Xanthomonas campestris</i>	Xanthan	Kennedy and Bradshaw, 1984
<i>Acetobacter xylinum</i>	Acetan or xylinan	Van Kranenburg <i>et al.</i> , 1999
<i>Leuconostoc mesenteroides</i> , <i>Lactobacillus curvatus</i>	Commercial dextran	Leathers, 2002; Ruhmkorf <i>et al.</i> , 2012
<i>Sphingomonas paucimobilis</i>	Gellan	Giavasis <i>et al.</i> , 2006
<i>Agrobacterium</i> and <i>Rhizobium</i> species	Curdlan	Sutherland, 1998
<i>Alcaligenes faecalis</i> and <i>Pseudomonas</i> species	Succinoglycan	Freitas <i>et al.</i> , 2011; Moosavi-Nasab <i>et al.</i> , 2012
<i>Lactobacillus hilgardii</i>	Kefiran	Ghasemlou <i>et al.</i> , 2011
<i>Pullularia pullulans</i> or <i>Aureobasidium</i>	Pullulan	Leathers, 2003
<i>Sclerotium glaucanicum</i> and <i>Sclerotium rolfsii</i>	Scleroglucan	Giavasis <i>et al.</i> , 2002
<i>Schizophyllum commune</i> and <i>Lentinus edodes</i>	Schizophyllan and lentinan	Giavasis <i>et al.</i> , 2002; Giavasis and Biliaderis, 2006
<i>S. cerevisiae</i>	Glycan	Xu <i>et al.</i> , 2009

Table No. 2 enlists various mode of action of Microorganisms that they perform to produce various peptides, amino acids and their derivatives via the enzymatic action. Usage of an auxotroph for amino acids production requires the expansion of an insignificant quantity of substrates to help the development of the bacterium as Kinoshita and his partners reported that some arginine or citrulline auxotrophs utilized L-ornithine in a medium

supplemented with limited amount of L-arginine for growth and multiplication (Kinoshita *et al.*, 1957b). As some amino acids can't be delivered productively by fermentation, enzymatic strategies utilizing microbial catalysts were created. Applying immobilized aspartase enzyme produced from *E. coli*, Sato and Tosa (1993) built up a continuous enzymatic production technique to make L-aspartic acid from fumaric acid.

Table 4. Microbial transformation pathway to produce fatty acids.

Metabolites	Pathways/ Mechanisms	References
Acetate	Acetyl Coenzyme A (acetyl-CoA)	Thauer <i>et al.</i> , 1989
Propionate	Succinate pathway	Reichardt <i>et al.</i> , 2014
Butyrate	Butyryl-CoA: Acetate CoA transferase	Duncan <i>et al.</i> , 2002
Trimethylamine and Trimethylamine N-oxide	Cleavage of choline to TMA (Trimethylamine) and acetaldehyde	Craciun <i>et al.</i> , 2014
Indole	Tryptophanase catabolize tryptophan	Kim and Park, 2013
Para-cresol	Fermentation of tyrosine or its metabolite hydroxyphenylacetate	Dawson <i>et al.</i> , 2011
4-Ethylphenol	Tyrosine metabolite para-coumaric acid via cinnamate decarboxylase and vinyl phenol reductase	Yasuda <i>et al.</i> , 2001
Conjugated linoleic acid (CLA), conjugated linolenic acid (CLnA) and 10-hydroxy-cis-12- octadecenoate (HYA)	Transformation of linoleic acid (LA) to CLA, linolenic acid to CLnA and intermediate from linoleic acid pathway	Yang <i>et al.</i> , 2014

The biosynthesis of the amino acid derivatives such as selenocysteines and selenomethionines have been produced from both *Lactobacilli* (Lamberti *et al.*, 2011) and yeasts (Porto *et al.*, 2015). Lysin and ornithine can be metabolized by *E. coli* to produce cadaverine and putrescine respectively (Applebaum *et al.*, 1975). Similarly another bacteria such as *proteus* can harvest putrescine from ornithine as a communication signal (Visick and Fuqua, 2005).

Single cell proteins (SCP) have been produced from the cultures of different microbes, moulds, yeast and algae. SCP produced from microorganisms have over 80% protein despite the fact that they are poor in S-amino acids and have high nucleic acids constituents (Kurbanoglu, 2001).

Polysaccharides and derivatives

Polysaccharides are the carbohydrates, formed by polymerization of saccharide monomers.

Polysaccharides were the first known long term ago for its bioactivities and due to vital role in medicines. Microbial polysaccharides are thought to be ideal candidate for use in functional foods or nutraceuticals because of having significant immunomodulating properties like anti-tumor, anti-inflammatory, antimicrobial or hypo-cholesterolemic and hypo-glycemic properties (Giavasis and Biliaderis, 2006). The global market for this kind of health promoting nutrients is right now extending and scientific interest for this field is developing, as user understand the significances of such food graded medicine to solve health issues (Hardy, 2000). Table 3 contains data of various microbial polysaccharides which have some application in food industries, isolated from fungi, yeasts or bacteria.

Few Fungal strains of the phylum basidiomycetes, some Gram negative (*Xanthomonas*, *Pseudomonas*, *Alcaligenes*, etc) and gram positive

Lactic acid bacteria (LAB) bacteria are the common producers of microbial polysaccharides. Polysaccharides in significant amount are also synthesized by the members of the genus *Saccharomyces*, mainly by yeasts (Giavasis and Biliaderis, 2006). Some of extracellular

polysaccharides (EPS) have been harvested in fermented dairy items from a large number of LAB, especially from *Lactobacillus*, *Streptococcus*, *Lactococcus*, *Pediococcus*, as well as *Bifidobacterium sp.* and *Weissella* strains (Notararigo *et al.*, 2012).

Table 5. Shows vitamins production via various microorganisms.

Water soluble vitamins	Fat soluble vitamins	Microorganisms	References
–	Vitamin A and pro-vitamin A	Microalga <i>Dunaliella</i> and <i>Blakeslea trispora</i>	Borowitzka, 1989
–	Vitamin D	<i>S. cerevisiae</i>	He <i>et al.</i> , 2007
–	Vitamin E	<i>Dunaliella</i> and <i>Spirulina</i>	Valentin and Qi, 2005
–	Vitamin K	<i>Bacillus subtilis</i> and <i>Propionibacterium freudenreichii</i>	Furuichi <i>et al.</i> , 2006; Sato <i>et al.</i> , 2001
Vitamin B1	–	<i>Bacillus subtilis</i>	Goese <i>et al.</i> , 2006
Vitamin B2	–	<i>Ashbya gossypii</i> and <i>Candida famata</i>	Kato and Park, 2012; Dmytruk <i>et al.</i> , 2011
Vitamin B3	–	<i>Rhodococcus rhodochrous</i>	Nagasawa and Yamada, 1989
Vitamin B5	–	<i>E. coli</i> and <i>Corynebacterium glutamicum</i>	Moriya <i>et al.</i> , 1997; Sahn and Eggeling, 1999
Vitamin B6	–	<i>Flavobacterium sp.</i> and <i>Rhizobium meliloti</i>	Tani, 1989
Vitamin B7	–	<i>E. coli</i> , <i>B. sphaericus</i> and <i>B. subtilis</i>	Sanyal <i>et al.</i> , 1996; Ohshiro <i>et al.</i> , 1996; Bower <i>et al.</i> , 1996
Vitamin B9	–	<i>B. subtilis</i> and <i>Ketogulonigenium vulgare</i>	Zhu <i>et al.</i> , 2005; Cai <i>et al.</i> , 2012
Vitamin B12	–	<i>Pseudomonas</i> and <i>Propionibacterium</i>	Eggersdorfer <i>et al.</i> , 1996
Vitamin C	–	<i>Chlorella pyrenoidosa</i>	Running <i>et al.</i> , 1994

The most well-known bacterial polysaccharide, to be the first industrially produced is the dextran, used in many food ingredients as well as in pharmaceuticals as important substrates (Leathers, 2002). Polysaccharides are produced as extracellular substances by the most LAB (*Lactobacillales* or lactic acid bacteria) using glycosyltransferases or intracellularly by the enzymes glycosyltransferases from sucrose and sugar nucleotide precursors respectively (Ruas-Madiedo *et al.*, 2002). A mushroom *Agaricus blazei*, discovered in Brazil, having edible and medicinal value such as polysaccharides in its fruit body possess an anti-tumour activity (Mizuno *et al.*, 1990).

Mizuno (2002) reported the synthesis of polysaccharides with somewhat different medicinal properties in submerged cultures of *A. blazei* as compared to those polysaccharides contained in fruit

body of mushroom.

Fatty acid and structural lipids

Lipids contain the most important biological molecules, called fatty acids, found every in living world i.e. in organism's body structures, foods and play important role in cell membrane composition by providing integrity. According to the report of Nagy and Tiuca (2017), lipid bodies are the rich energy sources and are very crucial for the structures and functions of all living organisms (signaling, biosynthesis, nutrition and health) as well as, having important applications in the field of nanotechnology, food industries and cosmetics (Mashaghi *et al.*, 2013). Table 4 indicates a significant number of the metabolites (lipid bodies) characterized by these report, resulting from the metabolism of particular dietary supplements by selected types of organisms

that express the important enzymes to ferment the ingredients in the provided supplements and enhancing the production pathways. In this way, the variation of microorganisms using these diet-dependent metabolic pathways might be critical to understanding the variable host reaction to particular dietary components (Zeevi *et al.*, 2015). Den Besten *et al.* (2013) investigated the biochemical pathways prompting the formation of the short chain unsaturated fats (SCUF) by microorganisms, having the potential to hydrolyze sugar molecules. Poly unsaturated fatty acids (PUFA) playing an important role in the development of human body. Because, cell membranes of all living organisms are composed such fatty acids as a major components of the phospholipids due to which membrane fluidity is regulated as well as interaction of other

protein/enzymes to the membrane promoting signals transduction and other important metabolic pathways (Li *et al.*, 2003; Gogus and Smith, 2010). Fatty acids produced by various microorganisms possess some potential medicinal properties as mentioned in Table 1, which reflects its use as quality diet. Oil produced by the microorganisms are commercially available for human use and have been given the name of single cell oils (SCO). *Mucor circinelloides*, *Thraustochytrium* species and microalgae *Isochrysis galbana* have been studied for the commercially available SCO like Gamma-linolenic acid, Docosahexaenoic acid and Eicosapentaenoic acid production respectively (Wen and Chen, 2003; Ratledge, 2005; Burja *et al.*, 2006).

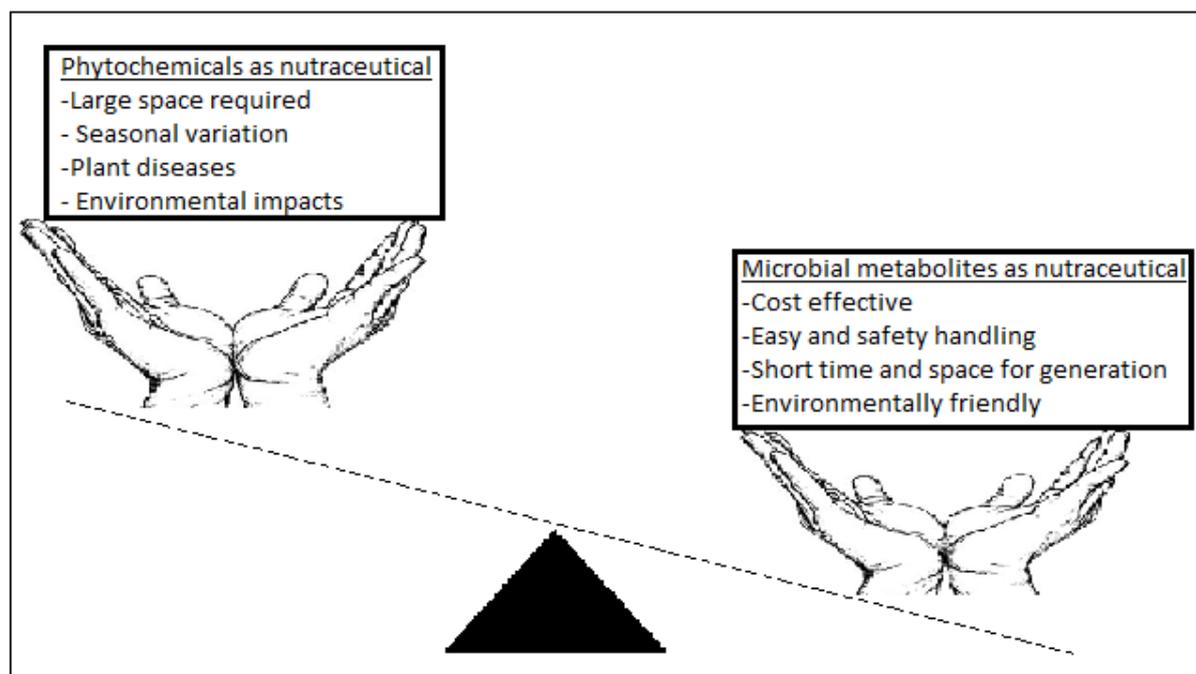


Fig. 1. Scale advantages of nutraceuticals from microbes over the phytochemicals.

Vitamins

Vitamins are characterized as natural compounds that are fundamental for normal development and nutrition and are required in little amounts in the diet since they can't be produced by the body. Thus, biological synthesis of vitamins has been investigated, by identifying the microbial flora, finding the optimized culture conditions, scaling up production methods and upgrading downstream procedures to

remove the other by-products. Vitamin A produced from *Arthrospira* concluded that it has high quality and a valuable impact on human health as compared to artificially produced vitamin A (Annapurna *et al.*, 1991). *Arthrospira* is also known to be a source of vitamin B12. Market available *Arthrospira* contains up to 244 µg vitamin B12 per g dry weight (Watanabe, 2007). Recently, a few metabolic engineering techniques have been discovered keeping in mind the

end goal to produce carotenoids (vitamin A precursor) in various biological pathways (Ye and Bhatia, 2012).

The growth pattern of various fungi like, *Trichoderma*, *Cephalosporium* and *Fusarium* have been researched with respect to their ability to yield ergosterol (vitamin D precursor)(Shimizu, 2008). The two fungal strains, *Eremothecium ashbyii* and *Ashbya gossypii*, belongs phylum ascomycete and commonly used for vitamins production on industrial scale(Stahmann *et al.*, 2000). Table 5 describes biological industrial production of each vitamin within a group, depending on its chemical nature: water-soluble or fat-soluble. Nicotinamide and nicotinic acid have been produced by chemical means as well as through the techniques of the biotechnology (fermentation) (Eggersdorfer *et al.*, 1996). Some other researchers have reported successful production of vitamin B9 by mixed type of microbial fermentation (Miyata and Yonehara, 1999).

Conclusion

Microbial nutraceuticals have the advantages due to well controlled production procedures, in an extensive upscale production inside a relatively constrained space and less generation time, stable harvested bioactive compounds and unhindered accessibility in the market, rather than plant sources whose accessibility, annual production and substance qualities frequently change. These nutraceuticals exhibits medicinal properties like anti-carcinogenic, immuno modulating properties, antidiabetic, anti-obesity, anti-inflammatory, anti-allergic and cholesterol lowering properties, as well as these nutraceuticals also act as prebiotics, promoting the growth of the producer strain or other LAB (Lactic acid bacteria) which could be highly significant in synthesis of novel foods with bioactivity and health promoting properties. However, sometimes during production of microbial nutraceutical, high production costs, low nutraceuticals yields, and tiresome downstream procedures required for isolation and purification are still the matters of consideration for microbial culturing procedures,

therefore, applicable approaches for the optimization of bioprocess can be implemented to improve the nutraceutical production process.

Conflict of interest

Author 1 and author 2 whose names are listed declare that they have no conflict of interest or any kind of financial or organizational involvement.

References

- Avula R, Kim SS, Chakrabarti S, Tyagi P, Kohli N, Menon P.** 2015. Delivering for Nutrition in Odisha: Insights from a Study on the State of Essential Nutrition Interventions. Poshan Report 7. New Delhi: International Food Policy Research Institute.
- Annapurna V, Deosthale Y, Bamji M.** 1991. *Spirulina* as a source of vitamin A. Plant Foods for Human Nutrition (formerly Qualitas Plantarum) **41**, 125–34.
- Applebaum D, Sabo DL, Fischer EH, Morris DR.** 1975. Biodegradative ornithine decarboxylase of *Escherichia coli*. Purification, properties, and pyridoxal5'-phosphate binding site. Biochemical **14**, 3675–3681.
- Bajzer M, Seeley R.** 2006. Physiology: obesity and gut flora. Nature **444**, 1009–1010.
- Barsh GS, Farooqi IS, O'Rahilly S.** 2000. Genetics of body-weight regulation. Nature **404**, 644–651.
- Bercovici D, Fuller F.** 1995. Industrial amino acids in nonruminant animal nutrition. In: Wallace, R.J., Chesson A (eds) Biotechnology in animal feeds and animal feeding, VCH, Weinheim, p 93–113.
- Biesalski HK.** 2001. Nutraceuticals: the link between nutrition and medicine. In: Kramer, K., Hoppe, P.P., Packer, L. (eds) Nutraceuticals in Health and Disease Prevention. Marcel Dekker, New York, p 1-26.

- Belury MA.** 2000. Dietary conjugated linoleic acid in health: physiological effects and mechanisms of action. *Annual Review of Nutrition* **22**, 505–31.
- Bergamo P, Luongo D, Miyamoto J, Cocca E, Kishino S, Ogawa J.** 2014. Immunomodulatory activity of a gut microbial metabolite of dietary linoleic acid, 10-hydroxy-cis-12-octadecenoic acid, associated with improved antioxidant/detoxifying defences. *Journal of Functional Foods* **11**, 192–202.
- Bekers M, Upite D, Kaminska E, Laukevics J, Grube M, Vigants A, Linde R.** 2005. Stability of levan produced by *Zymomonas mobilis*. *Process Biochemistry* **40**, 1535–9.
- Borowitzka L, Borowitzka M.** 1989. β -Carotene (provitamin A) production with algae. *Biotechnology of Vitamins, Pigments and Growth Factors*. Vandamme, E.J., (ed.), Elsevier Applied Science. London, p 15–26.
- Bower S, Perkins JB, Yocum RR, Howitt CL, Rahaim P, Pero J.** 1996. Cloning, sequencing, and characterization of the *Bacillus subtilis* biotin biosynthetic operon. *Journal of Bacteriology* **178**, 4122–30.
- Bronzwaer S.** 2008. EFSA scientific forum ‘from safe food to healthy diets’. EU risk assessment – Past, present and Future. *Trends in Food Science and Technology* **19**, S2–S8.
- Burja AM, Radianingtyas H, Windust A, Barrow CJ.** 2006. ‘Isolation and characterisation of polyunsaturated fatty acid producing *Thraustochytrium* species; screening of strains and optimization of omega-3 production’. *Applied Microbiology and Biotechnology* **72**, 1161–9.
- Cai L, Yuan MQ, Li ZJ, Chen JC, Chen GQ.** 2012. Genetic engineering of *Ketogulonigenium vulgare* for enhanced production of 2-keto-L-gulonic acid. *Journal of Biotechnology* **157**, 320–5.
- Canal PD, Possemiers S, Van de Wiele T, Guiot Y, Everard A, Rottier O, Geurts L, Naslain D, Neyrinck A, Lambert DM, Muccioli GG, Delzenne NM.** 2009. Changes in gut microbiota control inflammation in obese mice through a mechanism involving GLP-2-driven improvement of gut permeability. *Gut* **58**, 1091–1103.
- Chibata I, Kakimoto T, Kato J.** 1965. Enzymatic production of l-alanine by *Pseudomonas dacunhae*. *Applied Microbiology* **13**, 638–45.
- Clugston GA, Smith TE.** 2002. Global nutrition problems and novel foods. *Asia Pacific Journal of Clinical Nutrition*. 11(s6).
- Combie J.** 2006. Properties of levan and potential medical uses, in polysaccharides for drug delivery and pharmaceutical applications, ACS Symposium Series. *Journal of the American Chemical Society* **934**, 263–9.
- Casterton PL, Curry LL, Lina BAR, Wolterbeek APM, Kruger CL.** 2009. 90-Day feeding and genotoxicity studies on a refined arachidonic acid-rich oil. *Food and Chemical Toxicology* **47**, 2407–18.
- Craciun S, Marks JA, Balskus EP.** 2014. Characterization of choline trimethylaminelyase expands the chemistry of glyceryl radical enzymes. *ACS Chemical Biology* **9**, 1408–13.
- Dawson LF, Donahue EH, Cartman ST, Barton RH, Bundy J, McNerney R, Minton NP, Wren BW.** 2011. The analysis of para-cresol production and tolerance in *Clostridium difficile* O27 and O12 strains. *BMC microbiology* **11**, 86.
- Den Besten G, Van Eunen K, Groen AK, Venema K, Reijngoud DJ, Bakker BM.** 2013. The role of short-chain fatty acids in the interplay between diet, gut microbiota, and host energy metabolism. *Journal of Lipid Research* **54**, 2325–40.

- D 'Donnell K.** 2006. Aspartame and neotame. In: sweeteners and augar alternatives in food technology. Mitchell, H.L. (ed). Blackwell Publishing, Oxford, p 86–102.
- Dmytruk KV, Yatsyshyn VY, Sybirna NO, Fedorovych DV, Sibirny AA.** 2011. Metabolic engineering and classic selection of the yeast *Candida famata* (*Candida flareri*) for construction of strains with enhanced riboflavin production. *Metabolic Engineering* **13**, 82–8.
- Dufosse L.** 2009. Pigments. *Microbial Encyclopedia Microbiology* **4**, 457–471.
- Duncan SH, Barcenilla A, Stewart CS, Pryde SE, Flint HJ.** 2002. Acetate utilization and butyryl coenzyme A (CoA):acetate-CoA transferase in butyrateproducing bacteria from the human large intestine. *Applied and Environmental Microbiology*. **68**, 5186–90.
- Eggersdorfer M, Adam G, John MWH, Labler L.** 1996. Vitamins. *Biotechnology* **4**, Pape, H., Rehm, H.J. (eds). VCH, Weinheim, p 114–58.
- El Shafey HM, Ghanem S, Guyonvarch A.** 2009. Cloning of recA gene of *Corynebacterium glutamicum* and phenotypic complementation of *Escherichia coli* recombinant deficient strain. *World Journal of Microbiology and Biotechnology* **25**, 367–373.
- Furuichi K, Hojo K, Katakura Y, Ninomiya K, Shioya S.** 2006. Aerobic culture of *Propionibacterium freudenreichii* ET-3 can increase production ratio of 1,4-dihydroxy-2-naphthoic acid to menaquinone. *Journal of Bioscience and Bioengineering* **101**, 464–70.
- Giavasis I, Biliaderis C.** 2006. Microbial polysaccharides, in functional food carbohydrates, Biliaderis ,C., Izydorczyk, M. (eds). CRC Press, New York, p 167–214.
- Goese MG, Perkins P, Sehyns G.** 2006. Thiamin Production by Fermentation.US Patent Pub No US2006/0127993 A1.
- Goodridge HS, Wolf, AJ, Underhill DM.** 2006. β -glucan recognition by the innate immune system. *Immunological Reviews* **230**, 38–50.
- Gupta P, Andrew H, Kirschner BS, Guandalini S.** 2000. Is *Lactobacillus GG* Helpful in Children with Crohn's Disease? Results of a Preliminary, Open-Label Study. *Journal of Pediatric Gastroenterology and Nutrition* **31**, 453–457.
- Hardy G.** 2000. Nutraceuticals and functional foods: Introduction and meaning. *Nutrition* **16**, 688–9.
- Han YW.** 1990. Microbial levan. *Advances in Applied Microbiology* **35**, 171–94.
- He JZ, Ru QM, Dong DD, Sun PL.** 2012. Chemical characteristics and antioxidant properties of crude water soluble polysaccharides from four common edible mushrooms. *Molecules* **17**, 4373–87.
- He X, Guo X, Liu N, Zhang B.** 2007. Ergosterol production from molasses by genetically modified *Saccharomyces cerevisiae*. *Applied Microbiology and Biotechnology* **75**, 55–60.
- Hughenoltz J, Smid EJ.** 2002. Nutraceutical production with food-grade microorganisms. *Current Opinion in Biotechnology* **13**, 497–507.
- Kang SA, Hong K, Jang KH, Kim S, Lee KH, Chang BI, Kim CH, Choue R.** 2004. Antiobesity and hypolipidemic effects of dietary levan in high fat diet-induced obese rats. *Journal of Microbiology and Biotechnology* **14**, 796–804.
- Kassinen A, Krogius-Kurikka L., Makivuokko H, Rinttila T, Paulin L, Corander J, Malinen E, Apajalahti J, Palva A.** 2007. The fecal microbiota of irritable bowel syndrome patients differs significantly from that of healthy subjects.

Gastroenterology **133**, 24–33.

Kato T, Park EY. 2012. Riboflavin production by *Ashbya gossypii*. Biotechnology Letters **4**, 611–18.

Kinoshita S, Udaka S, Shimono M. 1957a. Studies on the amino acid fermentation. Part 1. Production of L-glutamic acid by various microorganism. Journal of General and Applied Microbiology **3**, 193–205.

Kinoshita S, Nakayama K, Udaka S. 1957b. The fermentative production of L-ornithine (preliminary report). Journal of General and Applied Microbiology. **3**, 276–7.

Kim J, Park W. 2013. Indole inhibits bacterial quorum sensing signal transmission by interfering with quorum sensing regulator folding. Microbiology. **159**, 2616–25.

Kumagai H, Hatsui H, Yamada H. 1970. Formation of tyrosine phenol-lyase by bacteria. Agricultural and Biological Chemistry **34**, 1259–61.

Kumagai H, Kashima N, Torii H, Yamada H, Enei H, Okumura S. 1972. Purification, crystallization and properties of tyrosine phenol lyase from *Erwinia herbicola*. Agricultural and Biological Chemistry **36**, 472–82.

Kurbanoglu EB, 2001. Production of single-cell protein from ram horn hydrolyzate. Turkish Journal of Biology **25**, 371–377.

Lamberti C, Purrotti M, Mazzoli R, Fattori P, Barello C, Coisson JD. 2011. ADI pathway and histidine decarboxylation are reciprocally regulated in *Lactobacillus hilgardii* ISE 5211: proteomic evidence. Amino Acids **41**, 517–527.

Leathers TD. 2002. Dextran in biopolymers, Vol 5. Polysaccharides I: Polysaccharides from Prokaryotes, Vandamme, E.J., De Baets, S., Steinbuechel, A. (eds). Wiley-VCH, Weinheim, p 299–321.

Lipton M. 2001. Challenges to meet: Food and nutrition security in the new millennium. Proceedings of the Nutrition Society **60**, 203–214.

Mashaghi S, Jadidi T, Koenderink G, Mashaghi A. 2013. "Lipid nanotechnology". International Journal of Molecular Sciences **14**, 4242–82.

Misak A. 2004. Elsinan, an extracellular α -1,3 : 1,4 glucan produced by *Elsinoe leucospila*: Production, structure, properties and potential food utilization. Foods and food ingredients journal of Japan **209**, 286–97.

Miyata R, Yonehara T. 1999. Method for Producing Folic Acid. US Patent **5**, 968-788.

Mizuno T. 2002. Medicinal properties and clinical effects of culinary-medicinal mushroom *Agaricus blazei* Murill (*Agaricomycetidae*). International Journal of Medicinal Mushrooms **4**, 4.

Mizuno T, Hagiwara T, Nakamura T, Ito H, Shimura K, Sumiy T, Asakura A. 1990. Antitumor activity and some properties of water-soluble polysaccharides from 'Himematsutake', the fruiting body of *Agaricus blazei* Murrill. Agricultural and Biological Chemistry **54**, 2889–96.

Moriya T, Hikichi Y, Moriya Y, Yamaguchi T. 1999. Takeda Chemical Industries, Ltd., Process for producing D-pantoic acid and D-pantothenic acid or salts thereof. U.S. Patent **5**, 932.

Nagasawa T, Yamada H. 1989. Microbial transformations of nitriles. Trends in Biotechnology. **7**, 153–158.

Nakayama R, Kumagai H, Maruyama T, Tochikura T, Ueno T, Fukami H. 1981. Synthesis of γ -glutamylpeptides by γ -glutamylcysteine synthetase from *Proteus mirabilis*. Agricultural and Biological Chemistry **45**, 2839–2845.

- Nelis H, de Leenheer A.** 1991. Microbial sources of carotenoid pigments used in foods and feeds. *Journal of Applied Bacteriology* **70**, 181–191.
- Nagy K, Tiuca ID.** 2017. Importance of Fatty Acids in Physiopathology of Human Body. In *Fatty Acids*. InTech.
- Notararigo S, Nácher-vázquez M, Ibarburu I, Werning ML, De palencia PF, Duenasmt Aznar R, López P, Prieto A.** 2012. Comparative analysis of production and purification of homo- and hetero-polysaccharides produced by lactic acid bacteria. *Carbohydrates Polymers* **93**, 57–64.
- Osawa T.** 2007. Nephroprotective and hepatoprotective effects of curcuminoids. *Advances in Experimental Medicine and Biology* **595**, 407–423.
- Ohno N, Miura T, Miura NN, Adachi Y, Yadomae T.** 2001. Structure and biological activities of hypochlorite oxidized zymosan. *Carbohydrates Polymers* **44**, 339–49.
- Ohshiro T, Yamamoto M, Izumi Y, Bui BT, Florentin D, Marquet A.** 1994. Enzymatic conversion of dethiobiotin to biotin in cell-free extracts of a *Bacillus sphaericus* bioB transformant. *Bioscience, Biotechnology, and Biochemistry* **58**, 1738–41.
- Porto B, Mangiapane E, Pessione A, Neves MJ, Pessione E, Martins FS.** 2015. The effects of sodiums elenite on the probiotic *Saccharomyces cerevisiae* UFMG A-905:A physiological and proteomic analysis. *Journal of Functional Foods* **17**, 828–836.
- Pulz O, Gross W.** 2004. Valuable products from biotechnology of microalgae. *Applied Microbiology and Biotechnology* **65**, 635–48.
- Ratledge C.** 2005. Microbial production of gamma-linolenic acid. In *Handbook of Functional Lipids*. Akoh, A.A. (ed). Boca Raton, Taylor & Francis, p 19–45.
- Reichardt N, Duncan SH, Young P, Belenguer, A, McWilliam Leitch C, Scott K.** 2014. Phylogenetic distribution of three pathways for propionate production within the human gut microbiota. *ISME Journal* **8**, 1323–35.
- Ruas-Madiedo P, Hugenholtz J, Zoon P.** 2002. An overview of the functionality of exopolysaccharides produced by lactic acid bacteria. *International Dairy Journal* **12**, 163–71.
- Running JA, Huss RJ, Olson PT.** 1994. Heterotrophic production of ascorbic acid by microalgae. *Journal of Applied Phycology* **6**, 99–104.
- Sousa R, Halper J, Zhang J, Lewis SJ, Li W.** 2008. Effect of *Lactobacillus acidophilus* supernatants on body weight and leptin expression in rats. *BMC Complementary and Alternative Medicine*. **8**, 5–12.
- Sabra W, Zeng AP, Deckwer WD.** 2001. Bacterial alginate: physiology, product quality and process aspects. *Applied Microbiology and Biotechnology* **56**, 315–25.
- Sahm H, Eggeling L.** 1999. d-pantothenate synthesis in *Corynebacterium glutamicum* and use of panBC and genes encoding L-valine synthesis for d-pantothenate overproduction. *Applied and Environmental Microbiology* **55**, 1973–9.
- Sanyal I, Gibson KJ, Flint DH.** 1996. *Escherichia coli* biotin synthase: An investigation into the factors required for its activity and its sulfur donor. *Archives of Biochemistry and Biophysics* **326**, 48–56.
- Sato T, Tosa T.** 1993. Production of L-aspartic acid. *Bioprocess Technology* **16**, 15–24.
- Sato T, Yamada Y, Ohtani Y, Mitsui N, Murasawa H, Araki S.** 2001. Efficient production of menaquinone (vitamin K₂) by a menadione-

- resistant mutant of *Bacillus subtilis*. *Journal of Industrial Microbiology and Biotechnology* **26**, 115–20.
- Shirasug N, Misaki A.** 1992. Isolation, characterization, and antitumor activities of the wall polysaccharides from *Elsinoe leucospila*. *Bioscience, Biotechnology, and Biochemistry* **56**, 29–33.
- Shibasaki T, Mori H, Ozaki A.** 2000. Enzymatic production of trans-4-hydroxyl-proline by regio- and stereospecific hydroxylation of L-proline. *Bioscience, Biotechnology, and Biochemistry* **64**, 746–50.
- Shimizu S.** 2008. Vitamins and related compounds: microbial production. *Biotechnology: Special Processes*, Volume 10, 2nd edition, Rehm, H.J. and Reed, G. (eds). Wiley-VCH, Weinheim, Germany.
- Stahmann KP, Revuelta JL, Seulberger H.** 2000. Three biotechnical processes using *Ashbya gossypii*, *Candida famata* or *Bacillus subtilis* compete with chemical riboflavin production. *Applied Microbiology and Biotechnology* **53**, 509–516.
- Suzuki H, Kato K, Kumagai H.** 2004. Development of an efficient enzymatic production of γ -d-glutamyl-L-tryptophan (SCV-07), a prospective medicine for tuberculosis, with bacterial γ -glutamyltranspeptidase. *Journal of Biotechnology* **111**, 291–5.
- Tani Y.** 1989. Microbial production of vitamin B6 derivatives. *Biotechnology of Vitamins, Pigments and Growth Factors*, Vandamme, E.J. (ed.). Elsevier Applied Science London, p 221–30.
- Thauer RK, Moller-Zinkhan D, Spormann AM.** 1989. Biochemistry of acetate catabolism in anaerobic chemotrophic bacteria. *Annual Review of Microbiology* **43**, 43–67.
- UNICEF.** 2016. Global nutrition report. data.unicef.org/resources/2016-global-nutrition-report.
- Valentin HE, Qi Q.** 2005. Biotechnological production and application of vitamin E: Current state and prospects. *Applied Microbiology and Biotechnology* **68**, 436–44.
- Visick KL, Fuqua C.** 2005. Decoding microbial chatter: cell-cell communication in bacteria. *Journal of Bacteriology* **187**, 5507–5519.
- Watanabe F.** 2007. Vitamin B12 sources and bioavailability. *Experimental Biology and Medicine* **232**:1266–74.
- Wojcikowski K, Johnson DW, Gobe GC.** 2006. Herbs or natural substances as complementary therapies for chronic kidney disease: ideas for future studies. *Journal of Laboratory and Clinical Medicine* **147**, 160–166.
- Wen, Z.Y, Chen F.** 2003. Heterotrophic production of eicosapentaenoic acid by microalgae. *Biotechnology Advances* **21**, 273–94.
- Yamada H, Takahashi S, Kii Y, Kumagai H.** 1978. Distribution of hydantoin hydrolyzing activity in microorganisms. *Journal of Fermentation Technology* **56**, 484–91.
- Yamamoto S, Wakayama M, Tachiki T.** 2005. Theanine production by coupled fermentation with energy transfer employing *Pseudomonas taetrolens* Y-30 glutamine synthetase and baker's yeast cells. *Bioscience, Biotechnology, and Biochemistry* **69**, 784–9.
- Yang B, Chen H, Gu Z, Tian F, Ross RP, Stanton.** 2014. Synthesis of conjugated linoleic acid by the linoleate isomerase complex in food-derived lactobacilli. *Journal of Applied Microbiology* **117**, 430–9.
- Yasuda T, Ueda J, Ohsawa K.** 2001. Urinary metabolites of genistein administered orally to rats. *Chemical and Pharmaceutical Bulletin* **49**, 1495–7.

Ye VM, Bhatia SK. 2012. Pathway engineering strategies for production of beneficial carotenoids in microbial hosts. *Biotechnology Letters* **34**, 1405–14.

Zeevi D, Korem T, Zmora N, Israeli D, Rothschild D, Weinberger. 2015. Personalized nutrition by prediction of glycemic responses. *Cell*. **163**, 1079–94.

Zhan XB, Lin CC, Zhang HT. 2012. Recent advances in curdlan biosynthesis, biotechnological production, and applications. *Applied Microbiology and Biotechnology* **93**, 525–31.

Zhu T, Pan Z, Domagalski N, Koepsel R, Ataii, MM, Domach MM. 2005. Engineering of *Bacillus subtilis* for enhanced total synthesis of folic acid. *Applied and Environmental Microbiology* **71**, 7122–9.