



Prevalence of coccidiosis in poultry farms in District Chakwal Punjab Pakistan

Qualab Naveed¹, Rani Faryal^{2*}, Nargis Aasma Rani³, Farooq Ahmad⁴, Bilal Hussain⁵

¹Department of Microbiology Faculty of Biological Science Quaid-I-Azam University Islamabad, Pakistan

²Department of Microbiology Faculty of Biological Science Quaid-I-Azam University Islamabad, Pakistan

³Mphil, Department of Statistics, International Islamic University, Islamabad, Pakistan

⁴Assistant professor Higher Education Department, Punjab, Pakistan

⁵Department of Microbiology Faculty of Biological Science Quaid-I-Azam University Islamabad, Pakistan

Dr Rani Faryal, Professor, Department of Microbiology, Quaid-i-Azam University, Islamabad, Pakistan

Key words: Poultry, Season, Coccidiosis, *Eimeria*, Chakwal, Prevalence.

<http://dx.doi.org/10.12692/ijb/15.5.425-442>

Article published on November 28, 2019

Abstract

Coccidiosis is one of the most deadly and costly disease of poultry. In this study prevalence of coccidiosis and different species of *Eimeria* were studied in organized poultry industry (2013-2017), of District Chakwal, Punjab, Pakistan. From February 2013 to October 2017, 5700 gut and 5700 fecal samples of broiler and Layer chickens suspected for coccidiosis were collected. Overall prevalence of coccidiosis was 59.19% (Gut 55.93% and fecal 62.46%). Seven species of *Eimeria* were detected. *E. tenella* was dominant circulatory species and *E. brunette* was least one. Young flocks show significantly high susceptibility then older. Frequency of coccidiosis in broiler was 50.24% and 9.20% in Layer. There was significant difference in prevalence of coccidiosis in rainy season (August, 85.10%), then dry and hot season (Jun, 44.9%). The prevalence of clinical diseases was 27.97% and 32.36% were subclinical. Prevalence of coccidiosis in rise hull 32.42% and 27.90% in wood shaving. Prevalence of coccidiosis in difference good, normal, and poor management conditions are 14.47%, 17.17%, and 28.5% respectively. In gut and fecal samples similar trend was observed. Study shows coccidiosis was serious problem of local poultry industry. Better management of poultry farms, use of better management technique, bio-security, Stander operating procedure tend to lower outbreaks of disease.

* Corresponding Author: Rani Faryal ✉ ranifaryal@gmail.com

Introduction

Coccidiosis is caused by the genus *Eimeria* and nine species are known to occur in chicken, which are widely distributed throughout the world Soulsby (1982); Lillehoj and Trout (1993); McDougald and Reid (1997); Pant *et al.* (2018). *Coccidia* exhibit marked degree of host specificity (Becker (1948); Boles and Becker (1954); Hadipour *et al.* (2011); Amare *et al.* (2012); Bachaya *et al.* (2012); Dinka and Tolassa (2012); Kala *et al.* (2013) and Singh and Meitei (2015). *E. tenella* and *E. necatrix* are the most pathogenic species. *E. acervulina*, *E. maxima* and *E. mivati* are common and slightly moderate pathogenic; *E. brunetti* is uncommon but pathogenic when it does occur. *E. mitis*, *E. praecox* and *E. hageni* are relatively non-pathogenic species (Reid (1978); Soulsby (1982); Guale (1990); Lillehoj and Trout (1993); Lee *et al.* (2010); Jadhav *et al.* (2011). The occurrence of clinical coccidiosis is directly related to the number of sporulated oocysts ingested by a bird at one time, the pathogenicity of the *Eimeria* species, the age of the infected chicken and the management system (Reid (1990); Lillehoj and Trout (1993); Whitmarsh (1997); Awais *et al.* (2012).

Human beings are the main mechanical transmitters in disseminating oocysts, which could be carried over by manure clinging to shoes or by utensils carried about from one pen to another. Flies, beetles, cockroaches, rodents, pets and wild birds have also been incriminated as mechanical vectors (Reid (1978); Chapman (1997); Adib-Nishaboori *et al.* (2000); Gyorke *et al.* (2013); Zhang *et al.* (2013); Sharma *et al.* (2015). Oocysts may survive as long as 86 weeks in shaded soil. But sunlight assists in destruction of oocysts. Incubator temperature for several days will kill oocysts, so there is little danger of hatchery transmission to the baby chicks. Oocysts are so resistant to disinfectants that they survive stringent attempts to kill them (Patillo and Becker (1955); Lunden *et al.* (2000).

Coccidia infections in chicken cause greater financial losses than in other domesticated birds. It costs on yearly basis, for prophylaxis, as well as therapy exceed

two billion Euros (Graat *et al.* (1996); Dallouil and Lillehoj (2006); Ahad *et al.* (2015). In general, the losses caused by coccidiosis without including the sub clinical coccidiosis are estimated to be 2 billion USD throughout the world (O'Lorcain *et al.* (1996); Williams (1999; 2005); Gari *et al.* (2008). Losses due to sub clinical forms of the disease are heavy and can't be estimated (Lee *et al.* (2010); Awais *et al.* (2012). Methuselaha *et al.* (2002) has reported that coccidiosis contributes to 8.4% loss in profit in large scale farms and 11.86% loss in profit in small scale farms (Gari *et al.* (2008); Zaman *et al.* (2012).

In cage system the lowest rate of infection is 1% (Guale (1990); Reid (1990); Morris *et al.* (2007); Iacob and Duma (2009). In deep litter poultry houses, which offer optimal condition of temperature and humidity for oocyst sporulation, the risk of infection is further increased (Urquhart *et al.* (1996); Amin *et al.* (2014). Infection with single species of coccidium is rare in natural conditions, and mixed infections are common. Nevertheless, in many outbreaks the clinical entity can be ascribed principally to one species or occasionally a combination of two or three (Soulsby (1982); Ayaz *et al.* (2003); Gari *et al.* (2008); Gyorke *et al.* (2013). The clinical disease is dependent on the number of oocysts ingested by individual birds. If the environment hygiene is poor, this number may be very large which is particularly true for *E. tenella* that have high biotic potential. But in very light doses no clinically recognizable symptoms may occur and thus, the morbidity and mortality increase in proportion to the size of the dose ingested (Soulsby (1982); Kinung'hi *et al.* (2004); Williams (2005); Morris *et al.* (2007); Chapman (2009); Iacob and Duma (2009).

Preliminary studies on the prevalence of coccidiosis done in the past have shown that both clinical and sub clinical coccidiosis have been occurring with low prevalence rate in the local strain chicken kept under the backyard production system than in the commercially oriented production systems (Guale (1990); McDougald (1998, 2003b); Ashenafi, *et al.*

(2004); Williams (2005). The range of coccidial infection prevalence has been reported as low as less than 10 % to as high as more than 90 % in broilers globally (Morris and Gasser (2006); Haug *et al.* (2008); Karaer *et al.* (2012); Singh *et al.* (2015). The prevalence of the disease in broiler was 33.07% (Sultana *et al.*, (2009). Prevalence of coccidiosis in free-range chicken in Sidi Thabet, northeast Tunisia was 31.8% (Kaboudi *et al.* (2016). Infection rate was reported to be 54.3% in Turkey (Karaer *et al.* (2012), 20.6% and 70.9% in Ethiopia (Gari *et al.* (2008); Elmira *et al.* (2012) 31.7% and 39.6% in India (Nikam *et al.* (2012); Sharma *et al.* (2013), 36.7% and 52.9% in Nigeria (Muazu *et al.* (2008), 71.9% in Pakistan (Khan *et al.* (2006), 78% in Jordan (Al-Natour *et al.* (2002), 88.4% in Argentina (McDougald and Mattiello (1997) and 92% in Romania (Gyorke *et al.* (2013). In Iran, this rate was reported to be 75% in North, 64% in Southwest, 55.96% in Northwest, and 38% in Northeast regions (Razmi and Kalideri (2000); Nematollahi *et al.* (2009); Hadipour *et al.* (2011); Shirzad *et al.* (2011).

If we have knowledge of level of burden of species of *Eimeria* in local poultry industry, better model for control this menace will be formulated and can protect our poultry industry. Prevalence of coccidiosis will help us to formulated epidemiological database and frequencies of different species of *Eimeria* are necessary to control uncertain and sudden outbreak of disease and to design better preventive measure. It is useful with respect to geographical epidemiology of this disease.

Materials and methods

Study area

The study area comprised of Chakwal district which is located in salt range and Chail peaks at the height of 1128 meters or 3701 feet, above sea. District has an area of 6609 square kilometers. It has five Tehsils and 68 union councils. Chakwal located in Punjab, Pakistan is a semi-arid area. Average temperatures in summers, it remains between 15° C and 40° C and may go up to a maximum of 51° C. The winter temperatures remain between -4° C and 25° C.

Ethical approval

Research work was approved by ethical committee in Department of Microbiology, Quaid-i-Azam University Islamabad.

Sampling

From February 2013 to June October 2017, 5700 gut & 5700 faecal samples of broiler and Layer chickens suspected for coccidiosis were collected from poultry sale point and poultry farms in adjacent areas reported to have signs of coccidiosis. The samples i.e. intestines along with caeca were collected in 2.5% (w/v) potassium dichromate solution and stored in the dark at 4°C. At the time of sample collection, information regarding age, no. of birds per house, no. of houses in the poultry farm, house dimensions, season, use of anticoccidials, ventilation system, genotype of broilers, management practices like watering and feeding methods, condition of drinkers and feeders, nature and condition of litter, frequency of change of litter; were recorded from poultry farmers.

Processing

The samples were brought to Molecular Microbiology, Quaid-i-Azam University Islamabad for further analysis. For the isolation of oocysts, each sample was processed using the method described by Eckert *et al.* (1995).

Symptom of Clinical Coccidiosis

Depression, Blood in feces, something white water feces, Decrease in food consumption, discoloration, wing drooping, slow growth, stressed birds, high mortality, intestinal lesion diarrhea, emaciation, enteritis were used as clinical symptom of Coccidiosis. Clinical status of disease means that sign and symptom of disease are present poultry farms. There is great mortality of birds in flocks in clinical form of disease. In subclinical forms of this is no or less mortality.

Gut Examination

Intestine was freed from mesentery and surface of intestine was carefully observed for the presence of

blood clotting, or pitchy patchy or inflamed area. Different species of *Eimeria* infect different part of intestine and have characteristic lesion shape and pattern. Different parts of intestine were carefully open with the help of scissors and observed. Gross lesion score were taken along with other pathological problem ranging from petechi, reddening, thickening, ballooning, hemorrhage (bleeding), caecal core, whitish spot, ladder like appearance, from caecal destruction to swelling of whole intestine. Depending upon the amount of intestinal destruction, type of species and severity of diseases specific number were allotted ranging from 0-4.

Mucosal Scrapping Examination

When there is any clue from for the presence of Coccidiosis from fecal samples or from sign and symptom such as bloody diarrhea is due to *Eimeria tenella*, whitish diarrhea is indicator of *Eimeria acervulina*.

Fecal Examination

Fecal samples from each poultry farm were collected in plastic zipper bag. Fecal samples were collected in W shape path from each farm. Fresh feces and litter samples from each poultry farm were collected. Litter samples were also taken from places that are wet. Fecal sample were transported to laboratory and stored at 4°C till further processing (Maff, 1982; Conway and McKenzie, 2007).

Eimeria Species Identification

Combination of different methods was used for species specific identification of *Eimeria* in poultry. Pathological lesions, Oocysts index, and sporulation time help in differentiation of different species.

There is Specific infection site of each *Eimeria* species in the intestine of chicken, criteria for identification of present species was developed by Long and Reid (1982).

Data Analysis

Data was summarizing with the help of descriptive statistics by using Microsoft Excel worksheet. SPSS 16.0 version of SPSS statistical software package was used to analyze data. To calculate prevalence number of positive samples was divided by total number of samples and multiplied by hundred. Chi-square helps to draw relationship between prevalence of Coccidiosis and Risk factor of Coccidiosis. If resulting P-value is less than 0.05, association is statistical significant.

Results

Coccidiosis was widespread in prevalence was detected in each and every month, and Illustrate frequency of Coccidiosis in different months. Highest prevalence was in August (85.10%) and in Jun there was least (44.9%), burden of disease (Fig 1).

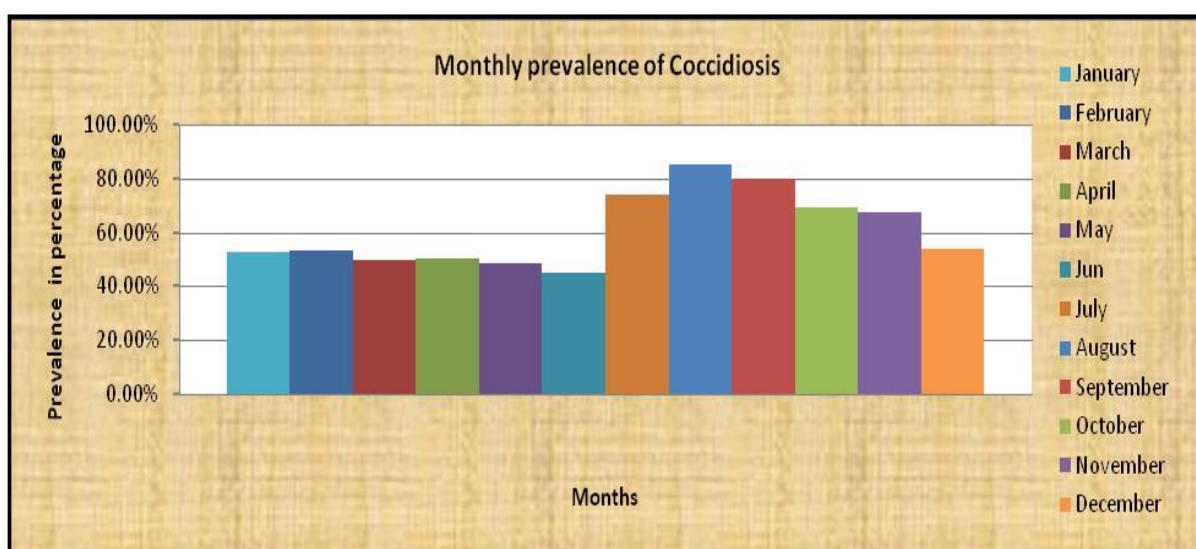


Fig. 1. Monthly variation in prevalence of coccidiosis.

Highest frequency of *Eimeria* was detected in Fecal and gut samples in August (81.2% in gut and 89% in fecal samples) during rainy season in summer and retreating Monsoon in October. In June dry and dry

season, lowest (39.2 % in gut and 50.6% in fecal samples) frequency was observed (Fig 2) in average highest prevalence of *Eimeria* was detected in summer and lowest in spring.

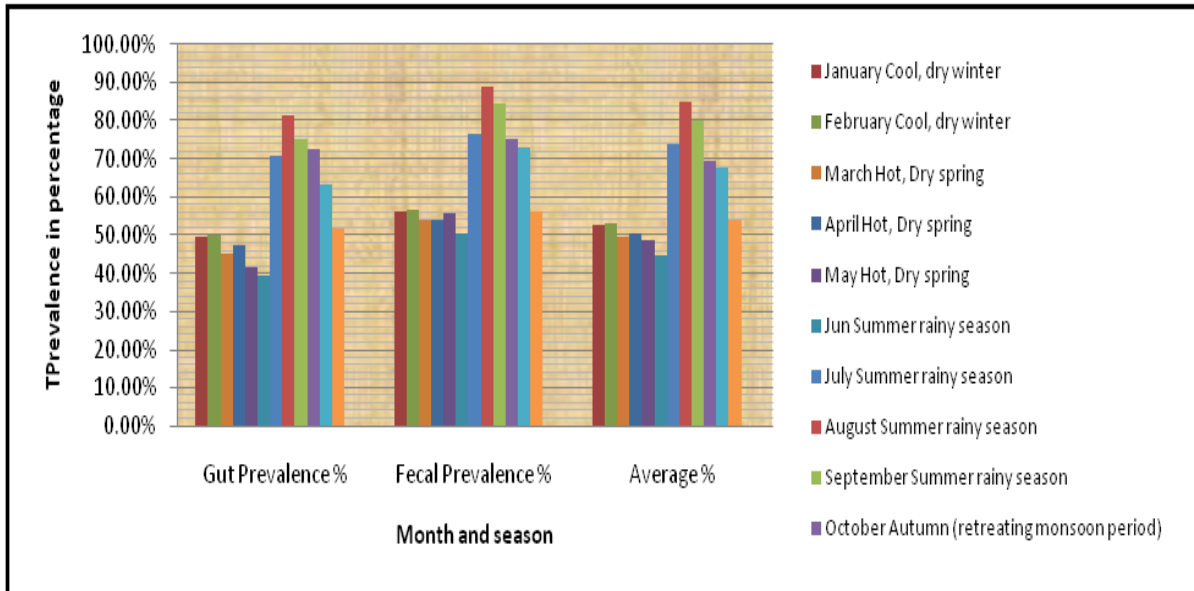


Fig. 2. Prevalence of coccidiosis in gut and intestinal samples during different month having particular weather (2013 -2017).

Similar trend was observed in gut and fecal samples (Fig 1). Gradual increase in frequency of diseases in commercial flock of poultry was observed.

pattern was observed in gut samples (Fig 4). Frequency of coccidiosis in different age groups in gut samples collected during study. Coccidiosis attack more in second age group and least outbreak of coccidiosis was observed in age group 4.

While screening fecal samples 59.72% of were positive in 2013 that increased to 69.30% in 2017. Same

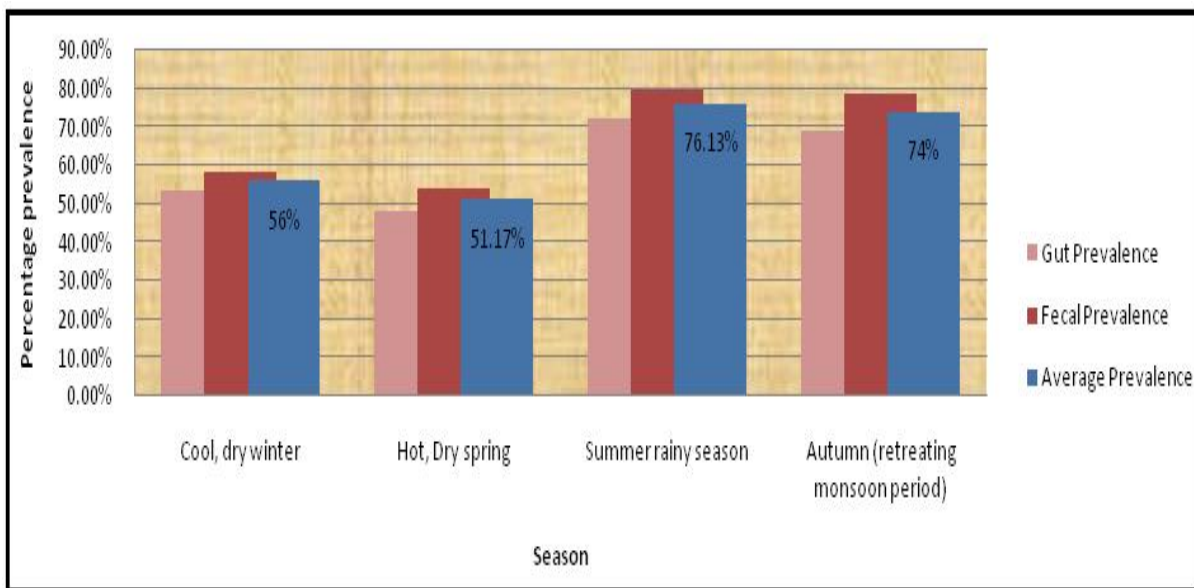


Fig. 3. Seasonal prevalence of coccidiosis.

The prevalence of disease was 55.93% out of which 25.69% in age group 1, 51.07% in group 2, 19.64% in group 3 and 3.64% in group 4. Highest burden of disease was observed during August and lowest in Jun (Fig 5). The highest infestation rate was observed in birds in age group 2 and infestation rate is lowest

in fecal samples of birds of age 43 days or more. August and September are months of heaviest infestation and Jun show lowest infestation of parasites. Prevalence of coccidiosis in age group 1, age group 2, age group 3, and age group 4 was 24.07%, 52.25%, 26.07%, 3.82% respectively (Fig 6).

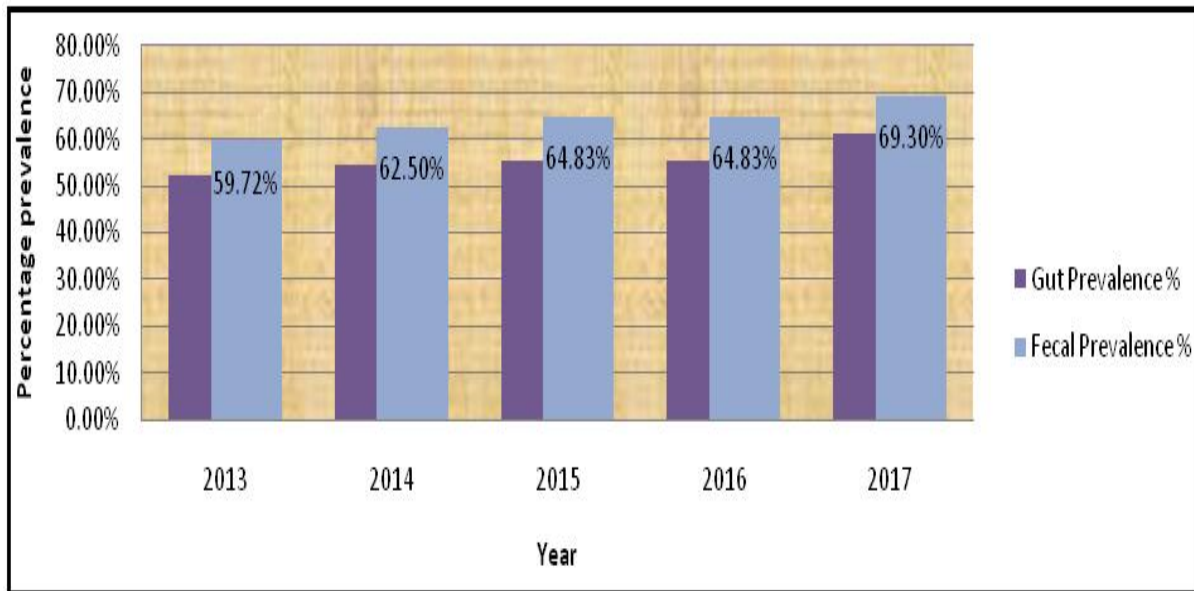


Fig. 4. Trend of coccidiosis in successive years (2013-2017).

Relative prevalence of seven species of *Eimeria*, in local poultry industry, revealed that there was little fluctuation in prevalence in circulatory species of *Eimeria* in each year. *E. tenella* is the most prevalent species and *E. brunette* is least one (Fig 7).

There is gradual increase in frequency of *E. tenella* and *E. maxima* on the other hand *E. acervulina* frequency decrease on the years, other species follow zigzag pattern. Different patterns of prevalence indicated by each species.

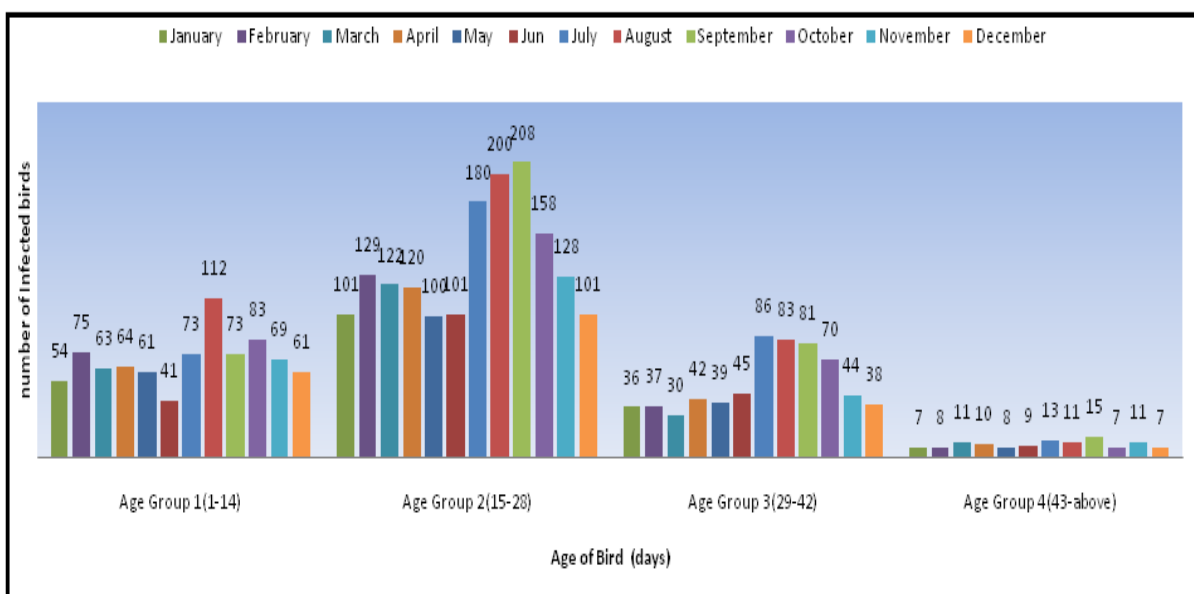


Fig. 5. Intestinal Prevalence of coccidiosis in different Age group in different months.

Accumulative frequency of different species of *Eimeria* was *E. tenella*, *E. acervulina*, *E. neatrix*, *E. mitis*, *E. maxima*, *E. brunette* and *E. praecox* was 33.86%, 26.33%, 25.01%, 5.60%, 22.08%, 4.87%, 14.33% respectively (Fig 8). Frequency of coccidiosis outbreaks in different poultry farms using rice hull and wood shaving as bedding material during flocks. Prevalence of coccidiosis in rise hull 32.42% and in

wood shaving 27.90% was observed (Fig 9).

There is more outbreak of coccidiosis in broiler than in Layers. Frequency of outbreak in broiler was 50.24% and in 9.20% in Layer flocks.

Highest incidence was reported in August and lowest incidence was observed in Jun (Fig 10).

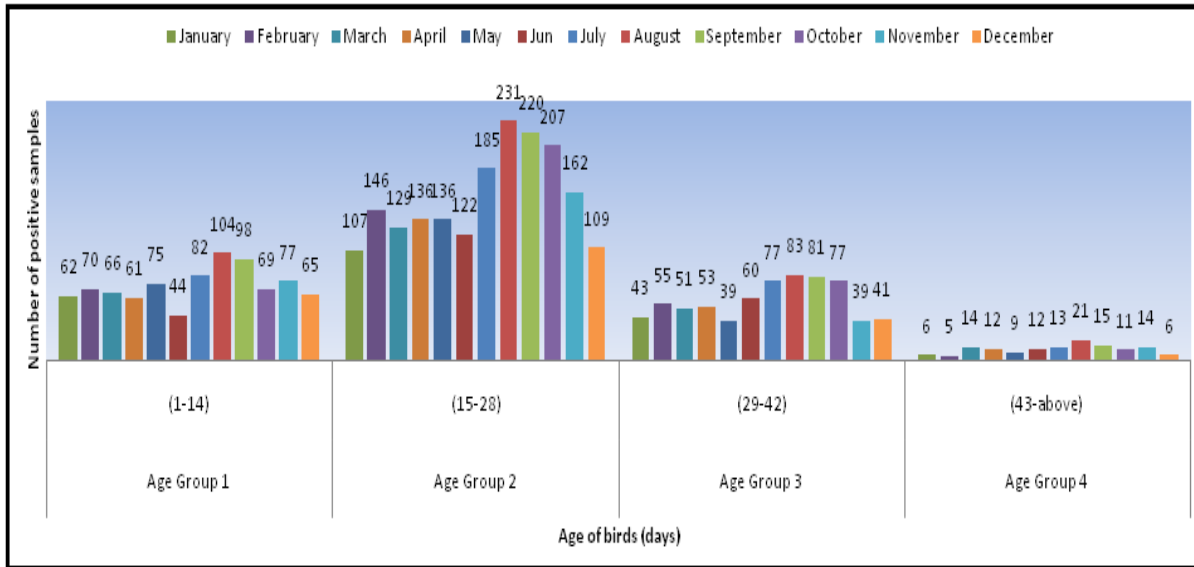


Fig. 6. Relative age wise susceptibility of Coccidiosis in fecal samples.

Represent 59.19% of observed cases are identified positive for coccidiosis. Prevalence of coccidiosis in difference good, normal, and poor management conditions are 14.47%, 17.17%, and 28.5%

respectively. Highest incidences of disease were observed in August and September, most humid month in year and few incidences were come to light in June, dry and hot environment (Fig 11).

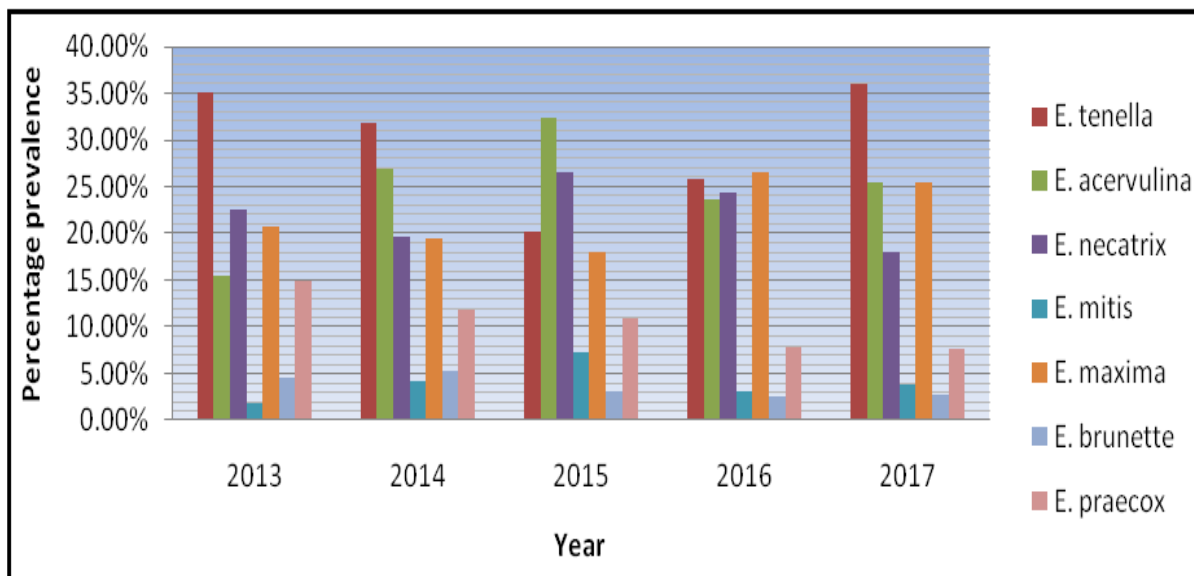


Fig. 7. Year wise relative prevalence of different species of *Eimeria* in Gut samples.

Eimeria is omnipresent parasite, present in each and every farm and in every part of world. The prevalence of clinical diseases was 27.97% and subclinical 32.36% were subclinical (Fig 12).

Discussion

In our study overall prevalence of coccidiosis was 59.19%. In disagreement to our results in northern Jordan Half (50%) of the farms surveyed clinical coccidiosis but overall prevalence of coccidiosis was 77% (Al-Natour *et al.* (2002). In Romania prevalence and distribution of *Eimeria* species, was

21 (91%) out of 23 flocks, and in 11 (92%) out of 12 farms (Gyorke *et al.* (2013). 33.33 % coccidian infection found in studies conducted by Sood *et al.* (2009) from Jammu. 46.04 % prevalence of coccidiosis was reported by Jadhav *et al.* (2011) in Aurangabad district of Maharashtra. 31.25 % coccidian infection found by Bandyopadhyay *et al.* (2006) in West Bengal.

Through microscopic examination, it was determined that 78.7% of the tested farms were positive in *Eimeria*-infection (Lee *et al.* (2010).

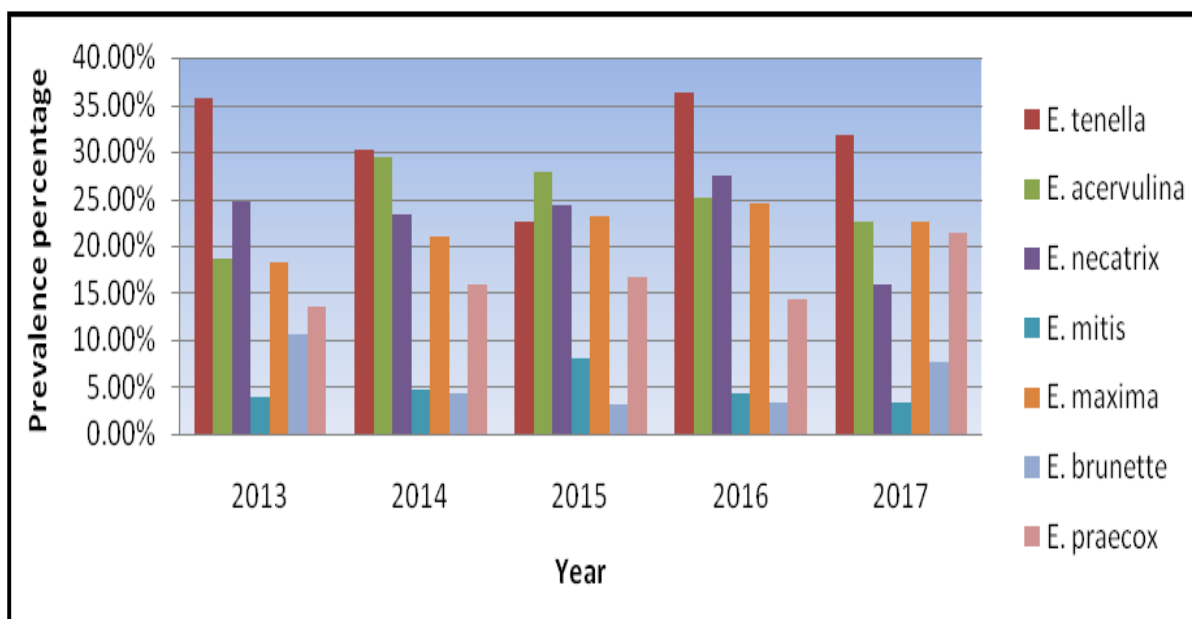


Fig. 8. Percentage prevalence of different species of *Eimeria* in feces samples during study period.

Out of total 5700 gut samples collected 3188 (55.93%) gut samples were positive. 3560 (62.46%) of total fecal 5700 isolates were positive. In disagree to this research 43.89% of chicken guts were positive in study by Awais *et al.*, (2012). Less prevalence 39.58 % of coccidiosis on examination of fecal samples was reported by Sharma *et al.*, (2015). Higher prevalence of coccidiosis may be due to more humid weather in district Chakwal and due to variation of geographical region, different type of management system and drug are used to control coccidiosis.

Frequency of outbreak in broiler was 50.24% and in 9.20% in Layer flocks. Much higher to our study Bachaya *et al.*, (2015) reported 65% rate of coccidiosis

in broiler chickens in Muzaffargarh District. Khan *et al.*, (2006) reported (71.8%) prevalence of coccidiosis in broiler in District Rawalpindi. Ayaz *et al.*, (2003) reported the (37.9%) prevalence of coccidiosis in research conducted in 2000-2001 in District Faisalabad-Punjab-Pakistan. Haug *et al.* (2008) reported 30% prevalence of disease were lower than our result. Number of other studies reported different prevalence of coccidiosis than our results, in broiler farms in Hamedan province, western Iran was 31.8%. Chicks and fecal samples were collected from 220 broiler farms in this region were positive for coccidiosis (Gharekhani *et al.* (2014). An overall prevalence rate of 31.8%) was obtained. Higher prevalence rates were recorded in growing

birds 58.9% and broiler birds 68.7%. Similarly, higher infection rates were also observed among birds sampled from Mairi ward 66.7% (Lawal *et al.* (2016). Yunus *et al.* (2008) reported 19.6% broiler flocks

were positive for coccidiosis. Sultana *et al.* (2009) reported the prevalence of coccidiosis was slightly different from our finding in broiler flocks was 42.85% and in layer flocks was 33.07%.

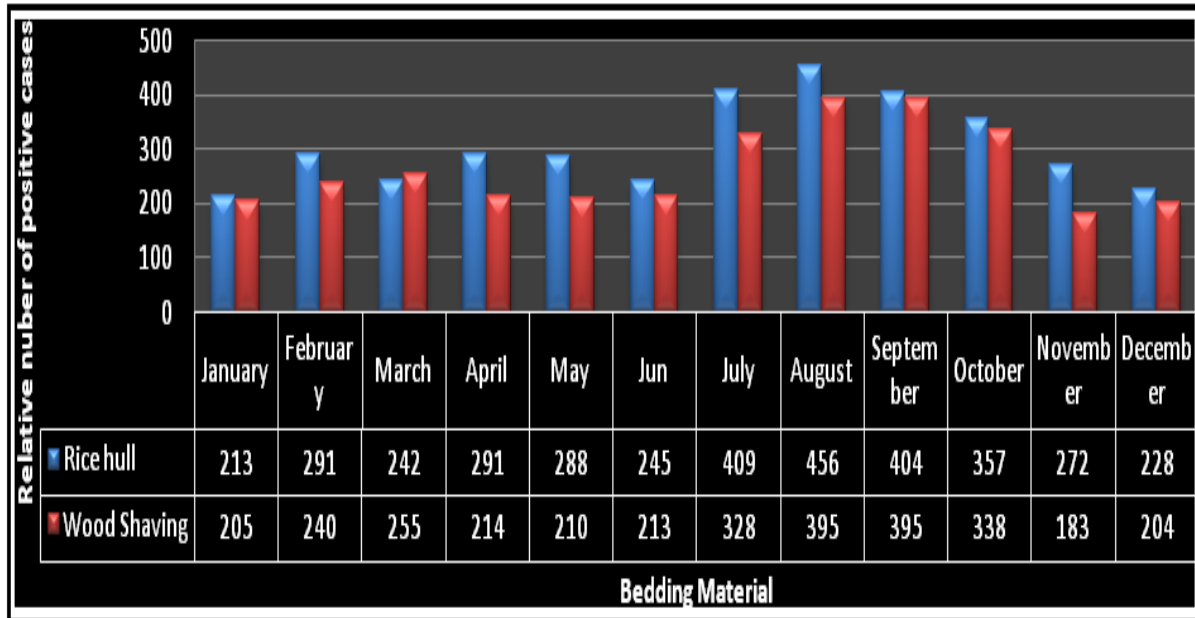


Fig. 9. Coccidiosis in different Bedding materials.

In our study mix species infection was 93.72% and 6.28% of samples reveal infection of single species. Prevalence of coccidiosis in free-range chicken in Sidi Thabet, northeast Tunisia was 31.8%. Mixed *Eimeria* species infection was 26.5% (Kaboudi *et al.* (2016).

Highest prevalence of coccidiosis was in August (85.10%) and in Jun it was least (44.9%), June outbreak of disease. Highest frequency of *Eimeria* was detected in Fecal and gut samples in August (81.2% in gut and 89% in fecal samples) during rainy season in summer and retreating Monsoon in October. In June lowest frequency was observed (39.2% in gut and 50.6% in fecal samples), the outbreak is least due to less humidity and hot season suppress the germination of *Eimeria* Oocysts was in line with Khan *et al.* (2006) and Bachaya *et al.* (2015).

Environmental conditions are different in District Muzaffargarh from District Chakwal. Different in prevalence of coccidiosis may be due to more rain fall

and humid weather. It is well known fact that humidity plays a vital role for the sporulation of the coccidial oocysts (Haug *et al.*, (2008) and Bachaya *et al.* (2012). In Faisalabad, Punjab, Pakistan, Awais *et al.* (2012) reported strong co-relation of different environmental condition with prevalence of coccidiosis in broiler flocks. Temperature, rainfall and humidity are main factor contributing to increase or decrease parasitic burden of different species of *Eimeria*, in intensive broiler industry (Awais *et al.* (2012). Age group, sampling sites, management conditions, litter management, breeds and exotic breeds are factor that leads to significant difference in prevalence of coccidiosis (Lawal *et al.* (2016).

Pattern of disease prevalence that fluctuates with changing temperature was in line with our results, rain fall and humidity as these parameters are key elements associated with outbreak of coccidiosis and favours the development or sporulation of coccidial oocysts in any particular area (Sharma *et al.* (2013). Presence of *Eimeria* in poultry farms in Chakwal region and prevalence of coccidiosis reported by Amin

et al., (2014) were different due to difference in geographical and weather condition. Difference rate of infestation of poultry farms in different area could be due to variation in, study design, methods and weather condition in different geographical regions (Al-Natour *et al.* (2002); Haug *et al.* (2008); Bachaya *et al.* (2012); Nikam *et al.* (2012).

Similar to this study high prevalence of coccidiosis during the moon soon due to warmth and moisture climatic condition favors transmission and

contamination of the oocytes was reported by number of researchers (Alawa *et al.* (2001); Jithendran (2001); Renaudeau *et al.* (2013). During hot and humid climatic conditions of monsoon season its prevalence was significantly high, during the spring it was lowest (Amin *et al.*, 2014). Humid weather results more outbreaks, humidity plays a vital role for the sporulation of the coccidial oocysts (Al-Natour *et al.* (2002); Haug *et al.* (2008); Bachaya *et al.* (2012); Nikam *et al.* (2012).

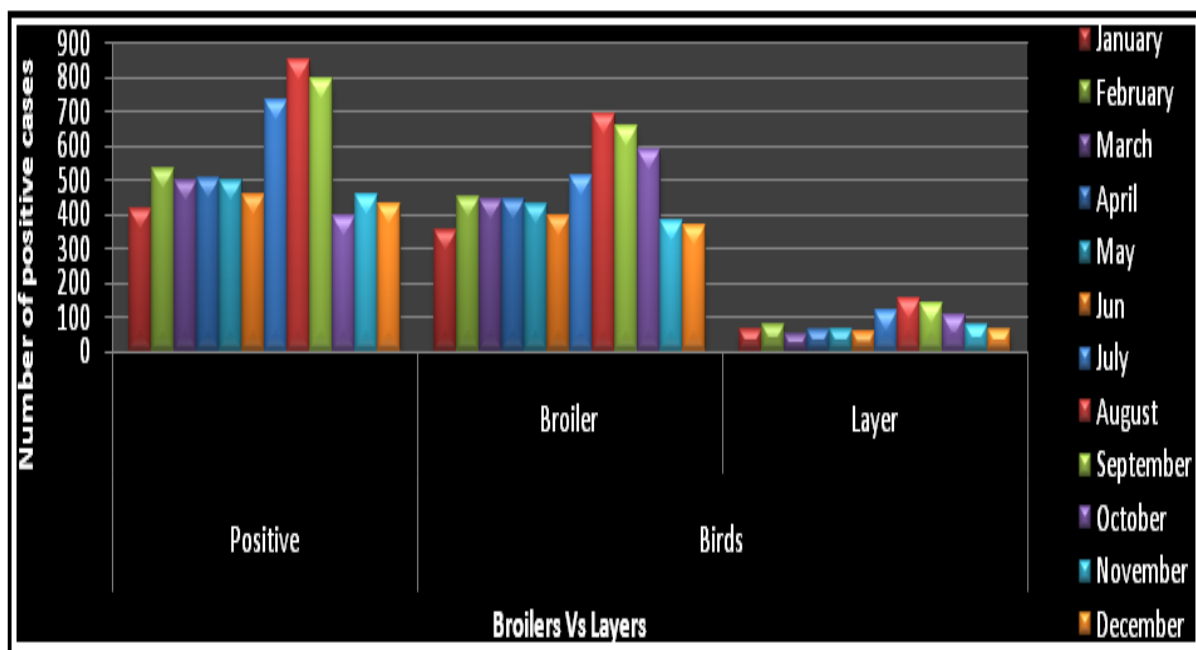


Fig. 10. Prevalence of coccidiosis in Broilers and Layers.

The highest prevalence of the disease during late summer and lowest during spring season was reported by Sultana *et al.* (2009) was in agreement to our results. Higher prevalence of infection during the rainy season of monsoon observed indicating that warmth temperature and humid condition favors the oocytes sporulation and subsequent transmission (Amin *et al.* (2014). In deviation from our study Graat *et al.* (1998) found coccidial infection to occur more often in autumn and winter in the Netherland.

Different type of material used as bedding material have different properties. Different rate of outbreak in different material is due to unique properties of each used as bedding material and different management

practice used in different area. Prevalence of coccidiosis in rise hull 32.42% and in wood shaving 27.90% was observed. Wetting of litter increased outbreak and prevalence of coccidiosis in intensive poultry raring systems. Younger birds are growing at rapid pace, at the age of 4-6 weeks, birds produce large volume of feces resulting dampness in litter. Damp litter serves as breeding sanctuaries of protozoan parasite that cause coccidiosis (Sultana *et al.* (2009).

When emergence of coccidiosis was studied in poultry house having soiled or cemented floor. Type of floor and bedding material observed as critical factor in prevalence of coccidiosis was in confirmation with

Sharma *et al.*, (2013) and Amin *et al.*, (2014). The prevalence of disease in gut was 55.93% out of which 25.69% in age group 1, 51.07% in group 2, 19.64% in group 3 and 3.64% in group 4. Same pattern results by screening of fecal samples. Our study was in confirmation with Sultana *et al.*, (2009), that young birds are most affected by coccidiosis than old birds.

Prevalence of coccidiosis in difference good, normal, and poor management conditions are 14.47%, 17.17%, and 28.5% respectively. Prevalence of coccidiosis was 66.7%, intensive management system 46.5% and constructed local cages 54.0% was in line with Lawal

et al., (2016). Poor management conditions have important role in development of clinical coccidiosis. Contaminated and leaking drinkers, feeders, poor ventilation, overcrowding, wet litter exacerbates mortality and morbidly cause economic losses (Ruff (1993); Ashenafi *et al.* (2004). Poor management such as high humidity in litter favor sporulation of oocysts, sporulation of oocysts directly increases circulation of parasite among birds (Bachayha *et al.*, 2015). *Eimeria* was detected in backyard and intensive rearing system, poor management leads to build up of oocysts of *Eimeria* (Lawal *et al.* (2016).

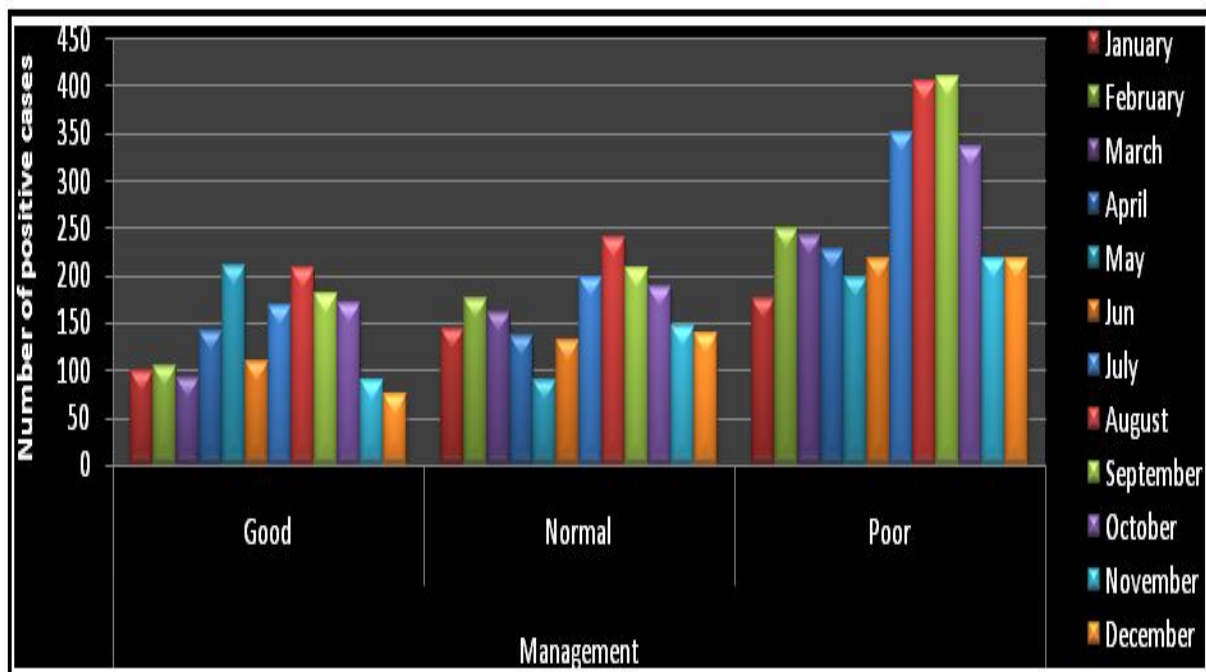


Fig. 11. Prevalence of coccidiosis in Management System.

Good management helps to reduce losses due to outbreaks and parasitic burden in poultry farm can be minimizing by improving management. Good ventilation, cleaning of drinker and feeder, use of disinfectant in poultry house between successive flocks, dry litter use of lime, timely vaccination and stander stocking densities in poultry flocks can minimize outbreak of disease (Abbas *et al.* (2011). Through cleanout of poultry house during successive flocks cannot eliminate oocysts completely but reduction in outbreaks of coccidiosis (Bachayha *et al.* (2015); Abbas *et al.* (2011); Razmi and Kalideri (2000); Sharma *et al.* (2013). Poor management

conditions are major impediments to prevent outbreak. By improving management we can prevent emergence of disease in poultry flocks. If strict management protocols are followed there is great reduction in coccidiosis incidence in flocks.

The prevalence of clinical diseases was 27.97% and subclinical 32.36% were subclinical. Shirzad *et al.* (2011) study contribution of different factor in sub-clinical coccidiosis. Age of bird, size of flock show direct relation while season, strain, drugs, and poultry farms don't reveals direct relation in sub clinical form of disease.

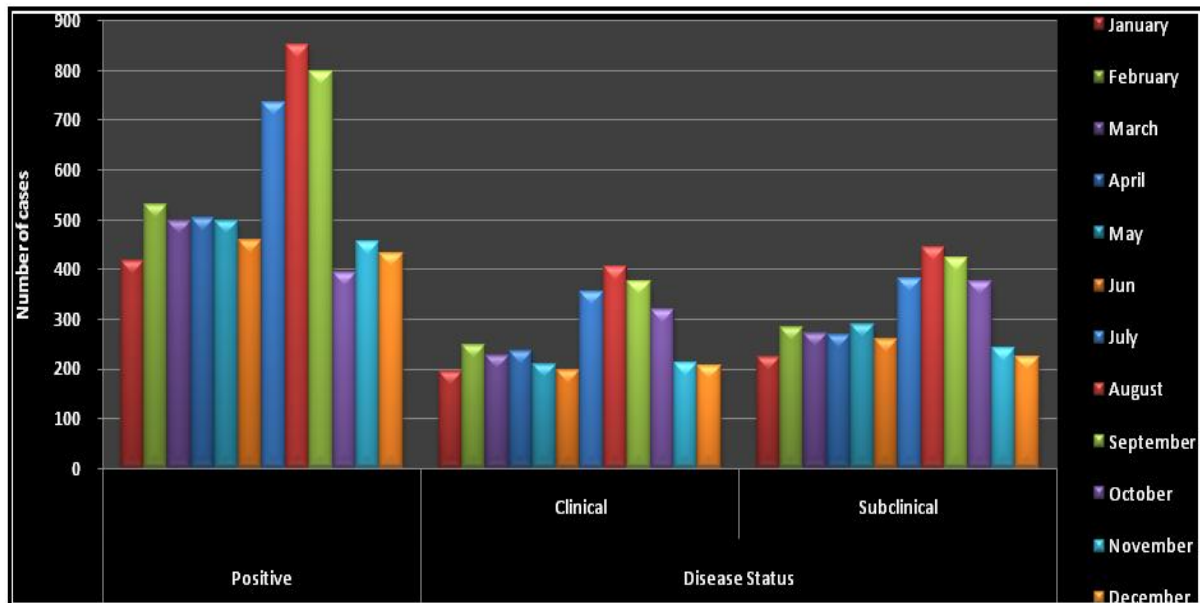


Fig. 12. Clinical vs. Subclinical Coccidiosis.

Effective control of subclinical coccidiosis depends on the knowledge of types and number of *Eimeria* species present in local poultry farms. Epidemiology of *Eimeria* helps to design efficient system to prevent and control coccidiosis. In Shandong province in Eastern China, five species of *Eimeria* were circulating in subclinical form in broiler flocks and to protect flocks from disease and to reduce economic loss to farmer at least five species of *Eimeria* should present (Sun *et al.* (2009).

Conclusion

Coccidiosis is widespread disease of poultry; it was detected in all type and management system. Seven species of *Eimeria* was detected. *E. tenella* was most dominant and *E. mitis* was observed in least frequency. Better management of poultry farms, use of better management technique, bio-security Stander operating procedure tend to lower outbreaks of disease.

Overcrowding, poor management system, rainy weather, less time period between successive flocks, use of same drugs for long time in same area, or in successive flocks, age of birds, bedding material and farmer level of education are critical factors that leads to emergence of disease and effect population dynamics of *Eimeria* in local poultry farms. More

humidity in litter system due to rainy season, poor management or production of more fecal material in young birds, litter material of poor quality serve as sanctuaries for *Eimeria* oocyst germination and outbreaks. Better management and study of local population of parasite will help in development of epidemiological database. Knowledge of parasite infestation, parasite history, emergence of resistance in local population, effectiveness of vaccine is useful designing quick, economic, long term, sustainable control program all are based on epidemiological study of coccidiosis.

Recommendation

It is recommended there should be more research on prevalence of coccidiosis in local poultry industry. There may be discovery of more species of *Eimeria*. It will help us to develop database on population dynamics of *Eimeria*. It has been observed seven species of *Eimeria* are present in local poultry; there should be routine monitoring by veterinary authorities. We recommend epidemiological study of coccidiosis using more advance and molecular techniques. It will help to develop better and quick response from veterinary practitioners to control outbreak. Rational use of coccidiosats and shuttle program will be helpful to avoid development of resistance in local population of *Eimeria* and to

control coccidiosis outbreaks. For uplift of poultry industry long term and sustainable strategy is required on urgent basis. We recommend that vaccination combined with good management practices will provide a feasible and sustainable strategy to control coccidiosis and improve the overall health of poultry. It is recommended educate farmer about coccidiosis to avert possible loss to flocks. Infected birds should be separated from healthy birds. In order to avoid buildup of Oocyst, litter should be changed as soon as possible. Early monitoring will be helpful for intervention and reduced mortality.

Proper Bio-security and hygienic of poultry house, churning and use of disinfectant after disposal of litter material, using time gap between successive flocks and use of vaccine against coccidiosis develop from local strain of *Eimeria* will help to save our industry and early vaccination against other poultry disease is required. Moderate stocking density to avoid overcrowding and congestion special in rainy season, fecal material of young birds leads to more humidity in litter. Better litter management will reduce outbreak of disease in successive flocks. Research is need to know the effect of local herbs, antibiotics, probiotics, cheap and effective litter material available in local area that will be more useful in local management system.

Acknowledgement

Authors acknowledge the service of Mr. Khalid Mahmood and Mr Aftab for facilitation in samples collection from poultry flocks and sale point of poultry. We will like to appreciate Mr Nasir Madni for transportation of samples. We are thankful to Dr Aslam for his serves in diagnosis of coccidiosis and preservation of gut samples.

Authors acknowledge effort of Mr Bilal Hussain in laboratory works (Postmortem and species specific diagnosis of coccidiosis by lesion scoring). We are thankful to Livestock center Chakwal Punjab and other poultry disease diagnosis laboratories for help in sample collections.

Conflict of Interest

In this research project there is no conflict of interest among authors. There is no conflict between poultry farmer, society and authors.

Reference

Abbas RZ, Iqbal Z, Blake D, Khan MN, Saleemi MK. 2011. Anticoccidial drug resistance in fowl coccidia: the state of play revisited. *World's Poultry Science Journal* **67(2)**, 337–350.

Adib-Nishaboori M, Razmi G, Kalidari G. 2000. A study of coccidiosis in the pullets of laying hens in Mashhad area, *Journal of Pajohesh and Sazandegi* **71**, p 31–35.

Ahad S, Tanveer S, Malik TA. 2015. Seasonal impact on the prevalence of coccidian infection in broiler chicks across poultry farms in the Kashmir valley. *Journal of Parasitic Diseases* **39**, 736-740.

Alawa CBI, Mohammed AK, Oni OO, Adeyinka IA, Lamidi OS, Adamu AM. 2001. Prevalence and seasonality of common health problems in Sokoto Gudali cattle at a beef research station in the Sudan ecological zone of Nigeria. *Nigerian Journal of Animal Production* **28**, 224-228.

Al-Natour MQ, Suleiman M. 2002. Flock-level prevalence of *Eimeria* sp. Among broiler chicks in northern Jordan. *Preventive Veterinary Medicine* **53**, 305-310.

Amare A, Mengistu A, Nazir SH. 2012. Prevalence and Aetiology of Poultry Coccidiosis and Associated Risk Factors in White Leghorn Grower Chickens at Kombolcha Poultry Farm Ethiopia. *Journal of World's Poultry Research* **2(3)**, 54–9.

Amin Y, Aslam A, Anwar K, Pervez Ali Z. 2014. Seasonal prevalence of eimeriosis in broiler chicken. *Advances in Life Sciences* **1(3)**, p 160-164.

Ashenafi H, Tedessa S, Medhin G, Tibbo M. 2004. Study on coccidiosis of scavenging indigenous

chickens in central Ethiopia. *Tropical Animal Health and Production* **36** (7), 693-701.

Awais MM, Akhtar M, Iqbal Z, Muhammad F, Anwar MI. 2012. Seasonal prevalence of coccidiosis in industrial broiler chickens in Faisalabad, Punjab, Pakistan. *Tropical Animal Health and Production*, **44**, 323-328.

Ayaz M, Akhtar M, Hayat CS, Hafeez MA, Haq A. 2003. Prevalence of coccidiosis in broiler chickens in Faisalabad, Pakistan. *Pakistan Veterinary Journal*. **23**, 51-52.

Bachaya HA, Raza MA, Khan MN, Iqbal Z, Abbas RZ, Murtaza S, Badar N. 2012. Predominance and detection of different Eimeria species causing coccidiosis in layer chickens. *Journal of Animal and Plant Sciences* **22**(3), 597-600.

Bachaya HA, Abbas RZ, Raza MA, Iqbal Z, Rehman TU, Baber W, Hussain R. 2015. Existence of coccidiosis and associated risk factors in broiler chickens in Southern Punjab, Pakistan. *Pakistan Veterinary Journal* **35**, 81-84.

Bandyopadhyay PK, Bhatka JN, Shukla R. 2006. A new Eimeria species (Protozoa: Apicomplexa: Sporozoea) from the Blue Rock Pigeon *Columba livia* (Aves: Columbidae). *Zoos' Print Journal* **21**, 2386-2387.

Becker ER. 1948. Coccidiosis of the chicken. In: *Diseases of poultry*, 2nd ed. H. E. Biester and L. H. Schwarte, eds. Iowa State College Press, Ames, IA., p 863-895.

Boles JI, Becker ER. 1954. The development of Eimeria brunetti Levine in the digestive tract of chickens. *Iowa State College Journal of Science* **29**, 1-26.

Chapman HD. 1997. Biochemical, genetic and applied aspects of drug resistance in Eimeria parasite of the fowl. *Avian Pathology* **26**, 221-224.

Chapman HD. 2009. A landmark contribution to poultry science prophylactic control of coccidiosis in poultry. *Poult Science* **88**, 813-815.

Conway DP, McKenzie ME. 2007. *Poultry coccidiosis: diagnostic and testing procedures*. Ames Iowa 50014, USA: Blacwell Publishing Professional p 32-33.

Dalloul RA, Lillehoj HS. 2006. Poultry coccidiosis: recent advancements in control measures and vaccine development. *Expert Review of Vaccines* **5**, 143-163.

Dinka A, Tolossa YH. 2012. Coccidiosis in Fayoumi Chickens at Debre Zeit Agricultural Research Center Poultry Farm, Ethiopia. *European Journal of Scientific Research*. **4**(5), 191-195.

Eckert J, Taylor M, Catchpole J, Licois D, Coudert P, Bucklar H. 1995. Morphological characteristics of oocysts. In: J. Eckert, R. Braun, M.W. Shirley and P. Coudert (Eds.), *Guidelines on techniques in coccidiosis research*. European Commission, Directorate-General XII, Science, Research and Development Environment Research Programme, p 103-119.

Elmira D, Melaku A, Bogale B. 2012. Prevalence and risk factors of coccidiosis in poultry farms in and around Ambo Town, western Ethiopia. *The American-Eurasian Journal of Science Research*, **7**(4), p 146-149.

Gari G, Tilahun G, Dorchies P. 2008. Study on Poultry Coccidiosis in Tiyo District, Arsi Zone, Ethiopia. *International Journal of Poultry Science*. **7**(3), 251-256.

Gharekhani J, Sadeghi-Dehkordi Z, Bahrami M. 2014. Prevalence of coccidiosis in broiler chicken farms in Western Iran. *Journal of Veterinary Medicine* **4**, 800-804.

Graat EA, Ploeger HW, Henken AM, De Vries,

- RG, Noordhuizen JP, Van Beek PN.** 1996. Effects of initial litter contamination level with *Eimeria acervulina* on population dynamics and production characteristics in broilers. *Veterinary Parasitology* **65**, 223-232.
- Graat EAM, Van der Kooij E, Frankena K, Henken AM, Smeets JFM, Hekerman MTJ.** 1998. Quantifying risk factors of coccidiosis in broilers using on-farm data based on a veterinary practice. *Preventive Veterinary Medicine* **33**, 297-308.
- Guale F.** 1990. Poultry Coccidiosis and Effect of Management System. An Assessment Trial in Debre Zeit and Its Surroundings. Faculty Veterinary Medicine, Addis Ababa University, Debre Zeit, DVM thesis.
- Gyorke L, Pop V, Cozma.** 2013. Prevalence and distribution of *Eimeria* species in broiler chicken farms of different capacities. *Parasite* **20**, article 52.
- Hadipour MM, Olyaie A, Naderi M, Azad F, Nekouie O.** 2011. Prevalence of *Eimeria* species in scavenging native chickens of Shiraz (Iran). *African Journal of Microbiology Research*. **5(20)**, 3296-3299.
- Haug AG, Gjevne Skjerve E, Kaldhusdal M.** 2008. A survey of the economic impact of subclinical *Eimeria* infections in broiler chickens in Norway, *Avian Pathology* **37(3)**, p 333-341.
- Iacob OC, Duma V.** 2009. Clinical paraclinical and morphopathological aspects in cecal eimeriosis of broilers. *Scientia Parasitologica* **10**, 43-50.
- Jadhav N, Nikam SV, Bhamre SN, Jaid EL.** 2011. Study of *Eimeria necatrix* in broiler chicken from Aurangabad District of Maharashtra state India. *International Multidisciplinary Research Journal* **1**, p 11-12.
- Jithendran K.** 2001. Coccidiosis-an important disease among poultry in Himachal Pradesh. *ENVIS Bulletin* **9(2)**, 35.
- Kaboudi K, Umar S, Munir TM.** 2016. Prevalence of Coccidiosis in Free-Range Chicken in Sidi Thabet, Tunisia. *Scientifica (Cairo)* **70(75)**, 195-600.
- Kala S, Gattani A, Kumar A, Samantaray S.** 2013. Infectious dynamics of different species of *Eimeria* in chicken. *Animal Science Reporter* **7**, 139-145.
- Karaer Z, Guven E, Akcay A, Kar S, Nalbantoglu S, Cakmak A.** 2012. Prevalence of subclinical coccidiosis in broiler farms in Turkey. *Tropical Animal Health and Production* **44(3)**, p 589-594.
- Khan GA, Siddque M, Sheereen N, Javed T,** 1990. Studies on the prevalence and pathogenicity of natural coccidiosis. *Archieve- Veterinaria Bucuresti*. **20**, 89-96.
- Khan MQ, Irshad H, Anjum R, Jahangir M, Nasir U.** 2006. Eimeriosis in poultry of Rawalpindi/Islamabad area. *Pakistan Veterinary Journal* **26**, 85-87.
- Kinung'hi SM, Getachew T, Hafez MH, Moges W, Moses K, Mathias G, Maximillian PO.** 2004. Assessment of economic impact caused by poultry coccidiosis in small and large poultry farms in Debre Zeit, Ethiopia. *International Journal of Poultry Science* **3**, 715-718.
- Lawal JR, Jajere SM, Ibrahim.** 2016. Prevalence of coccidiosis among village and exotic breed of chickens in Maiduguri, Nigeria. *Veterinary World*. **9(6)**, p 653-659.
- Lee BH, Kim WH, Jeong J, Yoo J, Kwon YK, Jung BY, Kwon JH, Lillehoj HS, Min W.** 2010. Prevalence and cross-immunity of *Eimeria* species on Korean chicken farms. *Journal of Veterinary Medical Science* **72(8)**, 985-989.

- Lillehoj HS, Trout JM.** 1993. Coccidia: A Review of Recent Advances on Immunity and Vaccine Development. *Avian Pathology* **22(1)**, 3-31.
- Long PI, Reid W.** 1982. A Guide for the Diagnosis of Coccidiosis in Chickens. Research reports 404 University of Georgia, College of Agriculture, and Athens.
- Lunden A, Thebo P.** 2000. Eimeria infection in litter-based, high stocking density systems for loose-housed laying hens in Sweden. *British Poultry Science* **41**, 440-447.
- MAFF.** 1986. Parasitological Laboratory Techniques. Technical Bulletin no.18, Manual of Veterinary, Her Majesty's Stationary Office, Ministry of Agriculture, Fisheries and Food, London.
- Mc-Dougald LF, Mattiello RA.** 1997. Survey of coccidia on 43 poultry farms in Argentina. *Avian Disease* **41(3)**, 923-929.
- Mcdougald LR.** 2003. Coccidiosis. Disease of poultry (11th edn.) Iowa State University Press, Ames, FA, USA.
- McDougald LR.** 1998. Intestinal Protozoa Important to Poultry. *Poultry Science* **77(8)**, 1156-1158.
- McDougald LR, Reid WM.** 1997. Coccidiosis In: B.W.Calnek Disease of Poultry 10th edition by Mosby - Wolfe, p 780-797.
- Methusela SK, Tilahun G, Hafez HM, Wolemesked M.** 2002. Studies on poultry coccidiosis in different production systems in Debre Zeit and surrounding areas, Ethiopia. *Bull. Animal. Health Production. Africa* **50**, 41-52.
- Morris GM, Woods WG, Richards DG, Gasser RB.** 2007. The application a polymerase chain reaction (PCR)-based capillary electrophoretic technique provides detailed insights into Eimeria populations in intensive poultry establishments. *Molecular and Cell Probes* **21(4)**, 288-294.
- Morris GM, Gasser RB.** 2006. Biotechnological advances in the diagnosis of avian coccidiosis and the analysis of genetic variation in Eimeria. *Biotechnology Advances* **24(6)**, 590-603.
- Muazu AA, Masdooq J, Ngbede.** 2008. Prevalence and identification of species of Eimeria causing coccidiosis in poultry within Vom, Plateau state, Nigeria. *International. Journal of Poultry. Science* **7(9)**, p 917-918.
- Nematollahi A, Gholamali M, Reze FP.** 2009. Prevalence of Eimeria species among broiler chicks in Tabriz, Iran. *Munis Entomology and Zoology Journal* **4**, 53-58.
- Nikam VS, Kanse SV, Jadab N, Jaid EL.** 2012. Comparative study of seasonal incidence of chicken coccidiosis in eight districts of Marathwada region, Maharashtra state," *Trends in Parasitology Research* **1(1)**, p 7-9.
- O'Lorcain P, Talebi A, Mulcahy G.** 1996. Mapping for B-Cell Epitopes in the 6x3262 Antigenic Sequence Derived from Eimeria tenella Sporulated Oocysts. *Vetinary. Parasitology* **66**, 159-169.
- Patillo WH, Becker ER.** 1955. Cytochemistry of Eimeria brunetti and Eimeria acervulina of the chicken, *Journal of Morphology* **69**, 372-377.
- Reid WM.** 1990. History of Avian Medicine in the United States. X Control of Coccidiosis. *Avian Disease* **34**, 509-525.
- Reid WM.** 1978. Coccidiosis. In: Hofstad, M. S., Calnek, B. W., Helmboldt, C. F., Reid, W. M. and Yoder, Jr, H. W. (ed.), *Diseases of Poultry*, 7th Edition. USA, Iowa State University Press. Ames, Iowa. P, 784-805.
- Renaudeau D, Collin A, Yahav S, De Basilio V,**

- Gourdine J.** 2013. Adaptation to hot climate and strategies to alleviate heat stress in livestock production. *Animal* **6(05)**, 707-728.
- Reza Razmi G, Ali Kalideri.** 2000. Prevalence of subclinical coccidiosis in broiler-chicken farms in the municipality of Mashhad, Khorasan, Iran," *Preventive Veterinary Medicine* **44(4)**, p 247-253.
- Ruff MD.** 1993. External and internal factors affecting the severity of avian coccidiosis. In: Proc 6th Int Coccidiosis Conf, June 21-25, p 73-79.
- Sachin Pant, Prakash Bhatt, Shekhar S, Gopal Krishna.** 2018. Epidemiological Investigation of Poultry Coccidiosis in and around Tarai Region of Uttarakhand. *International Journal of Current Microbiology and Applied Sciences* **7(7)**, 374-380.
- Sharma S, Iqbal A, Azmi S, Shah HA.** 2013 Study of poultry coccidiosis in organized and backyard farms of Jammu region," *Veterinary World*, **6(8)**, p 467-469.
- Sharma S, Iqbal A, Azmi S, Mushtaq I, Wani ZA, Ahmad S.** 2015. Prevalence of poultry coccidiosis in Jammu region of Jammu & Kashmir State," *Journal of Parasitology Disease* **39(1)**, p 85-89.
- Shirzad MR, Seifi S, Gheisari HR, Hachesoo BA, Habibi H, Bujmehrani H.** 2011. Prevalence and risk factors for subclinical coccidiosis in broiler chicken farms in Mazandaran province, Iran," *Tropical Animal Health and Production* **43(8)**, p 1601-1604.
- Singh LJ, Meitei NM.** 2015. Prevalence and intensity of infection of different Eimeria species in Broiler chicken, *Gallus gallus domesticus* from Imphal, Manipur, India. *Indian Journal of Applied Research* **5(4)**, 817-819.
- Sood S, Yadav A, Vohra S, Katoch R, Ahmad BD, Borkatari.** 2009. Prevalence of coccidiosis in poultry birds in R.S. Pura region, Jammu. *Veterinary Practice* **10(1)**, 69-70.
- Soulsby E.J.L.** 1982. Helminths, Arthropods and Protozoan's of Domesticated Animals, 7th edition. London: Bailliere Tindall p, 167-169.
- Sultana R, Hussain SA, Maqbool A, Shabnum, IC, Hussain S.** 2009. Epidemiology of Emeriosis in broiler and layer flocks in and around Lahore, Pakistan. *Punjab University Journal of Zoology* **24**, 1-2, p 81-86.
- Sun XM, Pang M, Jia T, Yan WC, He G, Hao LL, Bentue M, Sue X.** 2009. Prevalence of Eimeria species in broilers with subclinical signs from fifty farms. *Avian Diseases* **53**, 301-305.
- Urquhart GM, Amour J, Duncan JL, Dunn JN, Jennings FW.** 1996. Coccidiosis of poultry. Blackwell Publishers, London p 228-230.
- Whitmarsh SH.** 1997. Protozoan Poultry Diseases. Poultry Science Home page, College of Agricultural and Life Sciences, Mississippi State University. <http://www.misstate.edu/dept/poultry/disproto.Htm>
- William RB.** 1996. A survey of Eimeria species in commercially-reared chickens in France during. *Avian Pathology* **25**, 113-130.
- Williams R.** 1999. A compartmentalised model for the estimation of the cost of coccidiosis to the world's chicken production industry. *International Journal of Parasitology* **29**, 1209-1229.
- Williams R.** 2005. Intercurrent coccidiosis and necrotic enteritis of chickens: rational integrated disease management by maintenance of gut integrity. *Avian Pathology* **34(3)**, 159-180.
- Williams RB.** 1995. Epidemiological studies of coccidiosis in the domestic fowl (*Gallus gallus*). II. Physical condition and survival of Eimeria acervulina oocysts in poultry house litter. *Applied parasitology*

36, 90-96.

Zaman MA, Z Iqbal, RZ Abbas, Khan MN. 2012. Anticoccidial activity of herbal complex in broiler chickens challenged with *Eimeria tenella*. *Parasitology* **139**, 237-243.

Zhang Y, Jia Q, Zhang X. 2013 Screening of multiple drug resistant genes of *Eimeria tenella* infesting chicken in China. *Pakistan Veterinary Journal* **33(2)**, 150– 154.