



## RESEARCH PAPER

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## Effect of sunshine on the evolution of morphological and physicochemical parameters of preharvest mangoes (*Mangifera indica* L.)

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### Abstract

To reduce post-harvest losses due to an uncontrolled harvest date, the effects of sunshine on the ripening of mangoes on trees were studied in the Kent variety intended for export from Côte d'Ivoire. On trees, some mangoes exposed to direct daily sunlight (DDS) and others shaded by the leaves, therefore subject to indirect daily sunlight (IDS) were monitored. Mangoes growth and development were accessed through morphological (weight, length, circumference and volume) and physicochemical (firmness, pulp color, total sugars, reducing sugars, flavonoids and tannins) parameters. Furthermore, temperature induced by daily sunlight was recorded until harvest. The obtained results showed that, except for pulp firmness and coloration, indirect sun exposure (IDS) increased more rapidly morphological parameters than direct sun exposure (DDS). For example, 78 days after fruit set; IDS increased faster (0.46 kg/week) mangoes weight than DDS (0.37 kg/week). However, mangoes physicochemical parameters changed faster under DDS than under IDS. Investigation of daily temperature effect on mango parameters evolution revealed that low temperatures (28.72; 30.22; 30.53°C) promoted morphological growth while high temperatures (31.55; 31.56; 32.55°C) rapidly evolved mango internal physicochemical parameters. Consequently, sun direct exposition (DDS) reduce mangoes harvest time than indirect exposition i.e. shadiness under leaves (IDS). Sunshine can now be harnessed by dropping branches in orchards after the mango trees have fruited. This is so that the maximum number of mangoes are exposed to the sun on the tree. Thus, quality of the harvested mangoes is improved and mangoes will be competitive in the international market.

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## Introduction

Mango is known as an edible stone fruit produced by the tree *Mangifera indica*. Mangos trees grow in tropical and subtropical climates throughout the world (Dambreville, 2012). This edible fruit includes a large and varied range of nutrients. This makes it an exceptional nutritional quality (Djioua, 2010). Mango production is approximately 36 % of the world's tropical fruit production (FAOSTAT, 2001). Since its exportation, mango holds a prominent place in the Ivorian economy; making it the top mango exporter in West Africa (Mieu, 2017). The North is currently the main mango producing area for export in Côte d'Ivoire (Hala *et al.*, 2007). Mango generates substantial incomes for the local population. The Kent variety by itself contributes 60-70 % of these exports (Rey *et al.*, 2004). This is due to its very slow and gradual ripening process, which prevents rotting and improves storage (Touré, 2012). Moreover, the Kent mango variety offers more interesting sensory qualities than other commercialized varieties (Belem *et al.*, 2017). That is why it was considered as the reference for mangoes traded in the European Union. However, each year producers and exporters who are the mango sector main actors encounter numerous losses due to the uncontrolled harvest time. In fact, when harvested too early, mangoes shrivel up without really ripening, whereas harvested too late, their shelf life remains very limited. According to FIRCA (2011), because of their extremely high perishability, the mangoes post-harvest losses range between 30 and 40%. According to FIRCA (2011), the postharvest losses of mangoes are between 30 and 40 % thus showing their high perishability.

In addition, the difficulties encountered for better marketing include the optimum stage determining for harvesting mangoes and maintaining fruit during transport (Gomez, 1997; Johnson *et al.*, 1997). In mango tree, fruit exposure or not to the sun in preharvest influences their suitability for transport, storage and ripening (Ferguson *et al.*, 1999); i.r. the final quality of the mangoes offered to consumers. Therefore, obtaining and maintaining satisfactory quality levels, capable of meeting market

requirements, involves controlling the harvest date and the suitability of mangoes for storage. For this, methods based on physicochemical parameters such as the level of sugar, acidity, dry matter, starch, color of the pulp and firmness exist. They make it possible to assess the level of maturity and the harvest date of the fruit (Valente *et al.*, 2004).

Therefore, morphology-based methods were used to determine the level of maturity and predict the harvest date of mangoes. However, these knowledge leads to a high degree of uncertainty on these methods due to climate change. So take into account Factors such as sunshine are essential for long-lasting and high-quality mango production. However, this knowledge leads to a high degree of uncertainty about these methods, due to climate change. So take into account factors such as sunshine are essential for long-lasting and high-quality mango production. Moreover, Ganry (1978) reported that in bananas, the amount of heat received by the fruits during ripening could allow an estimate of their harvest date. In the mango tree, very few studies were carried out in these areas.

Thus, knowing the evolution of the morphological and physicochemical parameters of mangoes under sunshine effect could be an interesting tool to estimate the harvest date. This will allow the mango harvesting campaign for export to be organized with great flexibility and dexterity. This, in order to improve the quality of mangoes from Côte d'Ivoire. Therefore, the aim of the present study was to reduce postharvest losses by controlling the harvest date of mangoes of Kent variety, intended for export.

## Materials and methods

### *Plant material*

The trial was carried out in an experimental orchard of the National Agronomic Research Center (CNRA), in Lavononka in the north of Côte d'Ivoire (9 ° 22'56 "N; 5 ° 33'39" W). The trial was conducted on 10-year-old cultivar Kent mango trees. Kent was the most cultivated cultivar of mangoes in Côte d'Ivoire and was intended for export. The mangoes on the tree were constituted the plant material (Fig. 1).

## Methods

### *Selection of study plot, experimental mangoes and daily temperature measurements*

Experiments were carried out in a square orchard of 100 Kent mango trees. Orchard was chosen based on the careful respect of the mango technical itinerary: regulatory spacing of mango trees (10 m on the lines and 10 m between lines) and field maintenance (pruning, weeding, fertilizers, phytosanitary treatments). In view of minimizing a possible effect of soil fertility heterogeneity, the mango orchard was subdivided into four identical blocks of 25 mango trees each (Figure 2) for random sampling. Within each block, blooms of the same age and exposed to direct daily sunlight (DDS) and indirect daily sunlight (IDS) were identified and tagged on trees. After their development, 30 mangoes of the same age (resulting from fruit set on the same date) were randomly selected and tagged with string within each block, of which 15 were outside the foliage and hence directly exposed to the sun (DDS) and the other, 15 inside foliage and therefore indirectly exposed to the sun (IDS). Fifty days after fruit set, three mangoes were harvested per block (hence 12 mangoes in the orchard) each week for morphological and physicochemical analysis. From fruit set to harvest, daily-occurred temperatures were automatically monitored with a thermo-hygrometer (tinytag) installed in the orchard.

### *Determination of mangoes morphological parameters*

The morphological parameters concerned mangoes size (length, circumference and volume), weight and appearance. Mangoes length, from the point of peduncle insertion to fruit beak, was measured with a caliper. Each week, their weight was determined using an electronic balance (20 g sensitivity). Mango circumference was measured on the largest width using a tape measure. Fruit volume was determined by the difference in water displacement levels. Indeed, in a large graduated beaker of 2000 ml containing initial volume of water ( $V_i = 1000$  ml), mangoes were immersed until complete submersion. The final volume ( $V_f$ ) corresponded to the water ( $V_i$ ) + the mango ( $V_m$ ).

Therefore, the mango corresponding volume ( $V_m$ ) was determined by the difference between the final volume ( $V_f$ ) and the initial volume ( $V_i$ ) through the following formula:  $V_m = V_f - V_i$ , where  $V_m$  was mango volume,  $V_i$  was water initial volume and  $V_f$  was water final volume.

### *Determination of mangoes aspects and physicochemical parameters*

The mango physicochemical parameters analyzed were ethano-soluble sugars, total sugars, phenolic compounds, tannins, fruit firmness and pulp coloration. Mango pulp ethano-soluble sugars were extracted using the method of Martinez-Herrera *et al.* (2006) and quantified using method of Bernfeld (1955). Total sugars were determined according to method of Dubois *et al.* (1956). Phenolic compounds were extracted and determined following Singleton *et al.* method (1999) and flavonoids, according to Meda and *al.* (2005). Tannins content was determined according to Bainbridge and *al.* (1996). Mangoes firmness was measured with a crossbow penetrometer equipped with a 4 mm diameter tip. Determining the necessary force required to penetrate the tip into the pulp, the firmness was estimated on mango's two faces. Therefore, the mean value of both measurements constituted the mango firmness value. Mango pulp coloration was determined using the scoring method by a panel of 11 persons. The scores were attributed according to the colorations indicated in Figure 3 by each panelist. The arithmetic means of the total scores of each panelist resulted in each mango pulp coloring score.

### *Determination of weekly variations of kent mangoes morphological, physicochemical and aspect parameters*

The weekly variations ( $\Delta V$ ) of mangoes morphological (weight, length, circumference and volume) and physicochemical (firmness, pulp color, total sugars, reducing sugars, flavonoids and tannins) parameters were determined using the following formula:  $\Delta V_n = V_{Wn} - V_{Wn-1}$ , where  $\Delta V_n$  is the weekly variation of the studied parameter at week n,  $V_{Wn}$  the parameter value at week n and  $V_{Wn-1}$ , the parameter value at the previous week n-1.

### Statistical analysis

The values of the morphological and physiological parameters of mangoes and the weekly temperature variations were compared with each other based on the two types of exposure (sun and shade) over time using a two-way analysis of variance (ANOVA 2). The comparison of the means was performed with Tukey's LSD test at 5% using Statistica Statistica 7.1 software.

### Results

#### Effect of sunshine on mangoes morphological parameters

Indirect sun exposed mangoes (IDS) grow faster in weight than direct exposed mangoes (DDS) (Figure 4). So direct sun exposure delays weight accumulation in mangoes of the kent variety.

Similarly, analysis of Figure 5 indicates that under shade (IDS) mangoes grow faster in length than under direct sun exposure (DDS). Therefore, mango length growth was delayed under direct sun exposure.

**Table 1.** Weekly variation of mango morphological parameters according to ambient temperatures.

Time	Temperature (°C)	<sup>3</sup> ΔWeight (Kg)	ΔCircumference (cm)	ΔLength (cm)	ΔVolume (cm <sup>3</sup> )
0WAB <sup>1</sup>	28.41 ± 0.32 <sup>e2</sup>	-	-	-	-
1 <sup>st</sup> WAB	31.56 ± 0.21 <sup>b</sup>	0.05 ± 0.17 <sup>c</sup>	0.97 ± 0.6 <sup>d</sup>	0.73 ± 0.32 <sup>d</sup>	33 ± 0.59 <sup>e</sup>
2 <sup>nd</sup> WAB	31.55 ± 0.01 <sup>b</sup>	0.05 ± 0.06 <sup>c</sup>	0.69 ± 0.33 <sup>e</sup>	0.92 ± 0.16 <sup>c</sup>	39.25 ± 1.1 <sup>d</sup>
3 <sup>rd</sup> WAB	32.55 ± 0.50 <sup>a</sup>	0.01 ± 0.04 <sup>d</sup>	0.53 ± 0.8 <sup>f</sup>	0.2 ± 0.27 <sup>e</sup>	31.75 ± 1.15 <sup>f</sup>
4 <sup>th</sup> WAB	30.53 ± 0.11 <sup>c</sup>	0.10 ± 0.08 <sup>a</sup>	2.53 ± 1.05 <sup>b</sup>	1.03 ± 1.04 <sup>bc</sup>	49.73 ± 1.26 <sup>c</sup>
5 <sup>th</sup> WAB	30.22 ± 0.19 <sup>d</sup>	0.07 ± 0.07 <sup>b</sup>	1.17 ± 1.12 <sup>c</sup>	1.13 ± 0.22 <sup>b</sup>	74.77 ± 1.11 <sup>a</sup>
6 <sup>th</sup> WAB	28.72 ± 0.41 <sup>f</sup>	0.11 ± 0.16 <sup>a</sup>	2.63 ± 0.75 <sup>a</sup>	1.78 ± 0.23 <sup>a</sup>	61.4 ± 1.53 <sup>b</sup>

<sup>1</sup>: WAB: weeks after bloom

<sup>2</sup>: In a column, values followed by the same letter aren't significantly different (Tukey HSD test at 5%)

<sup>3</sup>Δ: weekly variation of the considered morphological parameter.

Analysis of Figure 6 indicates that mango circumference increases greater under shade than under direct sun exposure. Direct daily sunlight therefore reduces mango length growth of the Kent variety. As for volume, it increases faster for mangoes

hidden under the foliage than for those exposed directly before harvest (Figure 7). In short, mangoes exposed to direct sunlight reduce their growth in size, weight and volume compared to those exposed to shade in the Kent variety.

**Table 2.** Weekly variation of mango physicochemical parameters and aspect depending on the ambient temperature.

Time	Mean temperature (°C)	<sup>3</sup> ΔFirmness (Kg/mm <sup>2</sup> )	Δ Pulp Coloration score	ΔTotal sugars (mg/100g)	ΔReducing sugars (mg/100g)	ΔFlavonoids (mg/100g)	ΔTannins (mg/100g)
0 WAB <sup>1</sup>	28.41±0.3 <sup>e2</sup>	-	-	-	-	-	-
1 <sup>st</sup> WAB	31.56±0.2 <sup>b</sup>	0.23±0.3 <sup>b</sup>	0 ± 0.0 <sup>e</sup>	2.04±1.0 <sup>b</sup>	0.61±0.6 <sup>b</sup>	1.66±0.5 <sup>a</sup>	-0.22±0.5 <sup>b</sup>
2 <sup>nd</sup> WAB	31.55±0.0 <sup>b</sup>	0.18±0.0 <sup>c</sup>	0.7 ± 0.0 <sup>b</sup>	1.57±0.6 <sup>c</sup>	0.61±0.7 <sup>b</sup>	1.33±1.0 <sup>b</sup>	-1.74±0.7 <sup>d</sup>
3 <sup>rd</sup> WAB	32.55±0.5 <sup>a</sup>	0.25±1 <sup>a</sup>	0.77 ± 0.3 <sup>a</sup>	2.44±1.2 <sup>a</sup>	0.68±0.3 <sup>a</sup>	0.77±1.1 <sup>c</sup>	-3.97±1 <sup>e</sup>
4 <sup>th</sup> WAB	30.53±0.1 <sup>c</sup>	0.11±0.6 <sup>d</sup>	0.16 ± 0.6 <sup>e</sup>	1.549±0.5 <sup>d</sup>	0.44±0.8 <sup>d</sup>	0.53±0.1 <sup>d</sup>	-5.62±0.7 <sup>f</sup>
5 <sup>th</sup> WAB	30.22±0.2 <sup>d</sup>	0.11±0.6 <sup>d</sup>	0.15 ± 0.6 <sup>e</sup>	0.927±0.5 <sup>e</sup>	0.46±0.5 <sup>c</sup>	0.44±0.3 <sup>e</sup>	-0.71±0.9 <sup>c</sup>
6 <sup>th</sup> WAB	28.72±0.4 <sup>f</sup>	0.05±0.0 <sup>e</sup>	0.06 ± 0.7 <sup>d</sup>	0.251±1.1 <sup>f</sup>	0.34±0.8 <sup>e</sup>	0.26±0.4 <sup>f</sup>	-0.17±0.1 <sup>a</sup>

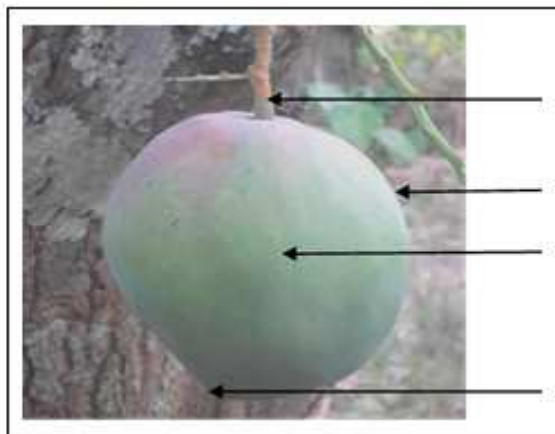
<sup>1</sup>: WAB: weeks after bloom

<sup>2</sup>: In a column, values followed by the same letter aren't significantly different (Tukey HSD test at 5%)

<sup>3</sup>Δ: weekly variation of the considered morphological parameter.

*Effect of sunlight on mangoes physicochemical parameters*

The analysis of figures 10, 11 and 12 indicates respectively that the total sugars, reducing sugars and flavonoids contents increased faster in mangoes exposed to direct sunlight than in those under shade during their development. Tannin content decreased more rapidly in mangoes exposed to direct sunlight than in those under shade (Figure 13). Therefore, direct exposure of mangoes to the sun favors the decrease of their tannin content.



**Fig. 1.** Fruit of Kent mango cultivar on the tree.

*Mangoes morphological evolution depending on pre-harvest ambient temperature*

The table 1 indicates that in Kent mangoes, the greatest weekly variations of their morphological parameters were obtained at the lowest ambient

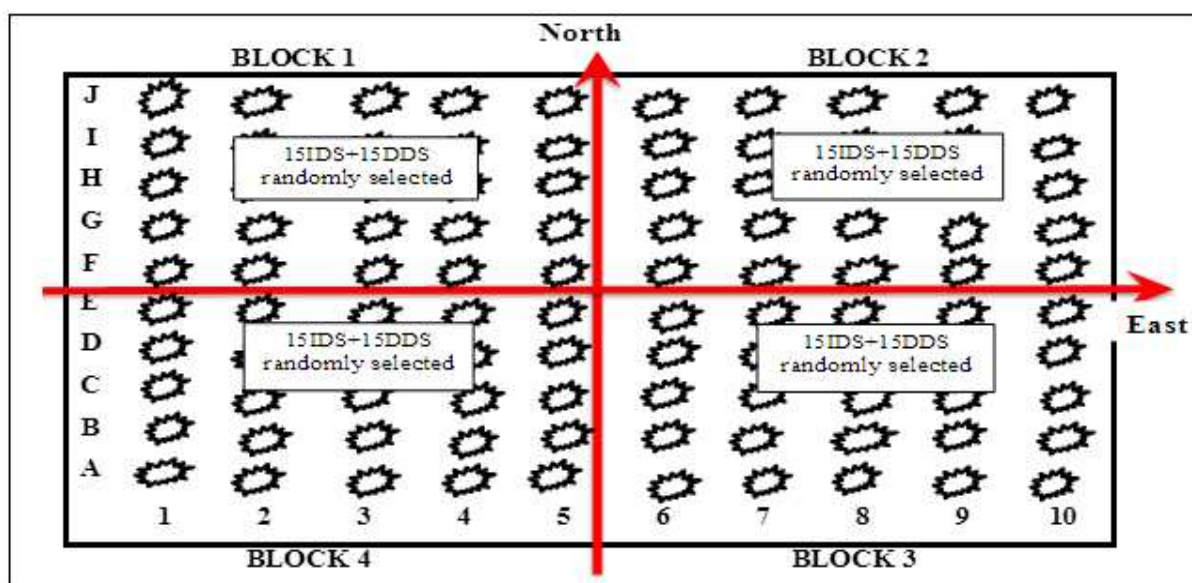
temperatures (28.72; 30.22 and 30.53 °C) while the least significant were achieved at relatively highest ambient temperatures (31.55; 31.56 and 32.55 °C).

*Effect of pre-harvest diurnal temperature on mango appearance and physicochemical parameters*

The relatively highest ambient temperatures (31.55; 31.56 and 32.55) were promoted the highest weekly variations in mango physicochemical parameters while the lowest ambient temperatures (28.72; 30.22 and 30.53) generated these parameters lowest variations (Table 2).

**Discussion**

Preharvest of mangoes exposure to sunlight significantly affects their morphological and physicochemical development. Indeed, morphological parameters of mangoes weight, length and volume not exposed to the sun (under foliage) showed a more rapid growth than those of mangoes exposed to direct sunshine. This indicated that mangoes morphological growth resulting from the accumulation of leaf photosynthesized reserve nutrients in fruit cells was more accentuated in under-shaded mangoes (Thibault, 2014). This finding could be explained by the fact that under shade, mangoes undergoing low direct evapotranspiration therefore have more available water for cell turgor than those exposed to direct sunlight (Gabrielle, 2001).



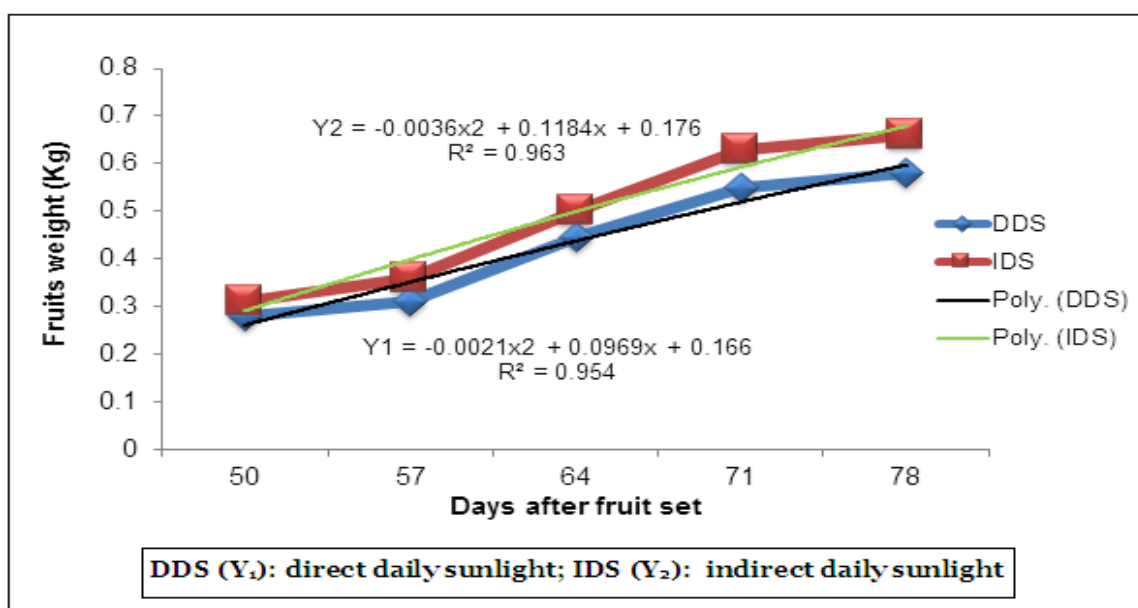
**Fig. 2.** Experimental design showing the four blocks where mangoes were collected.

In addition, compared to direct sunlight, exposition of mangoes under-shade improved their physicochemical parameters and appearance. So, mangoes remained more firm under direct sunlight

than under shade. The relatively high temperatures induced by direct pre-harvest sunlight enhance fruit's ability to bear heat by favoring their progressive hardening (Woolf *et al.*, 1999).



**Fig. 3.** Mango pulp staining scores after longitudinal sectioning.



**Fig. 4.** Evolution of mango Kent variety weight during ripening.

The ripening characterized by a progressive change in pulp color could be explained because of chloroplast conversion into chromoplasts and an increased carotenoid synthesis, specifically  $\beta$ -carotene, the mango flesh main carotenoid (Godoy and Rodriguez-Amaya, 1989).

However, this pulp coloration change differed following direct or indirect sunlight exposure. Besides, sunshine is one of the most important factors involved in carotenoids biosynthesis (Gautier *et al.* (2005).

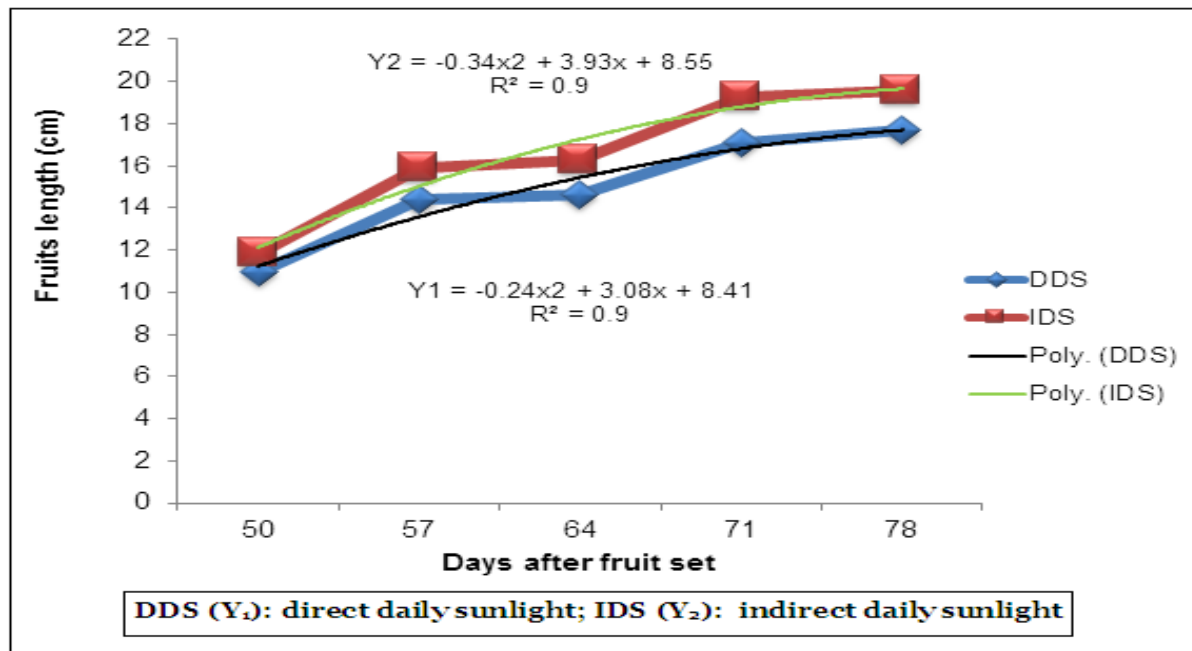


Fig. 5. Evolution of mango Kent variety length during ripening.

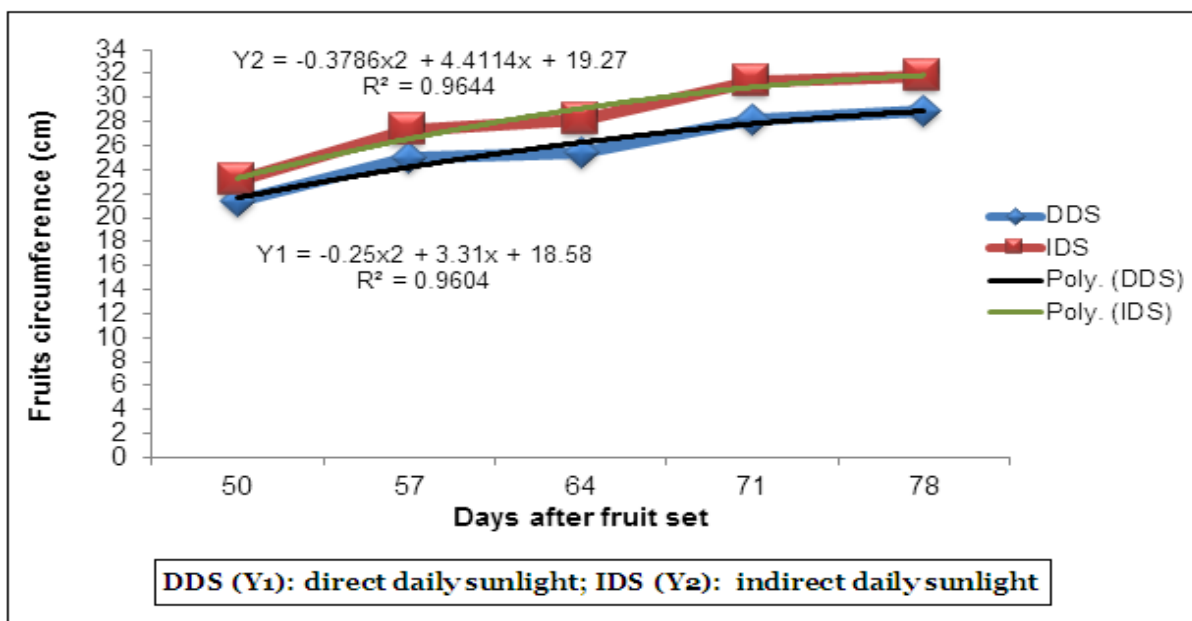


Fig. 6. Evolution of mangoes Kent variety circumference during ripening.

This is probably why mangoes exposed to direct sunlight ripened faster through a strong pulp coloration, contrary to those indirectly exposed to the sunlight (under shade).

Likewise, the mangoes exposure to pre-harvest sunlight also influenced the evolution of other physicochemical parameters like firmness, pulp coloring, and total sugars, reducing sugars, flavonoids and tannins. In fact, mangoes exposed to the direct

sunlight favored their rapid accumulation of total sugars compared to unexposed mangoes left under shade. Sugars, on the other hand, represent a large proportion of chemical compounds in the ripen mango (Subramanyam *et al.*, 1975). Starch accumulates during fruit growth phase and then hydrolyzes to the reducing sugars (maltose and glucose) during ripening (Delroise, 2003). Therefore, this hydrolysis results in an increase of reducing sugars and a decrease of starch content in mangoes,

which become sweeter (Kalra and Tandon, 1983). Indeed, this hydrolysis intensity was a function of temperature; it was intense at high temperatures and less intense at low temperatures (Paull *et al.*, 1999). Moreover, studies in pineapple showed that fruit

quality depends mainly on climatic conditions (Yapo, 2013). In fact, sun-exposed mangoes had better grow under light and temperature conditions than those not exposed to sunlight (Larkindale and Knight, 2002).

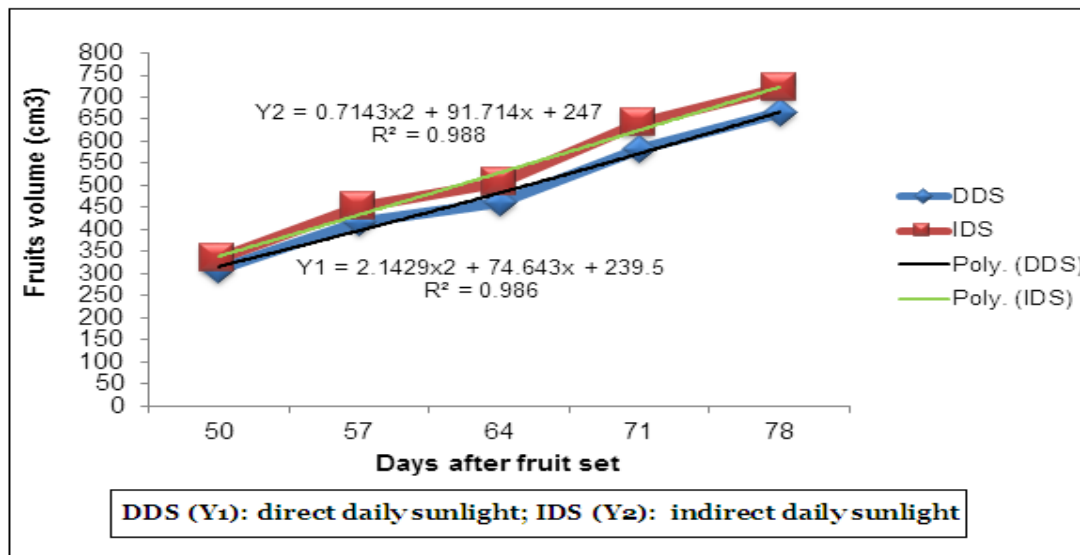


Fig. 7. Evolution of mangoes Kent variety volume during ripening.

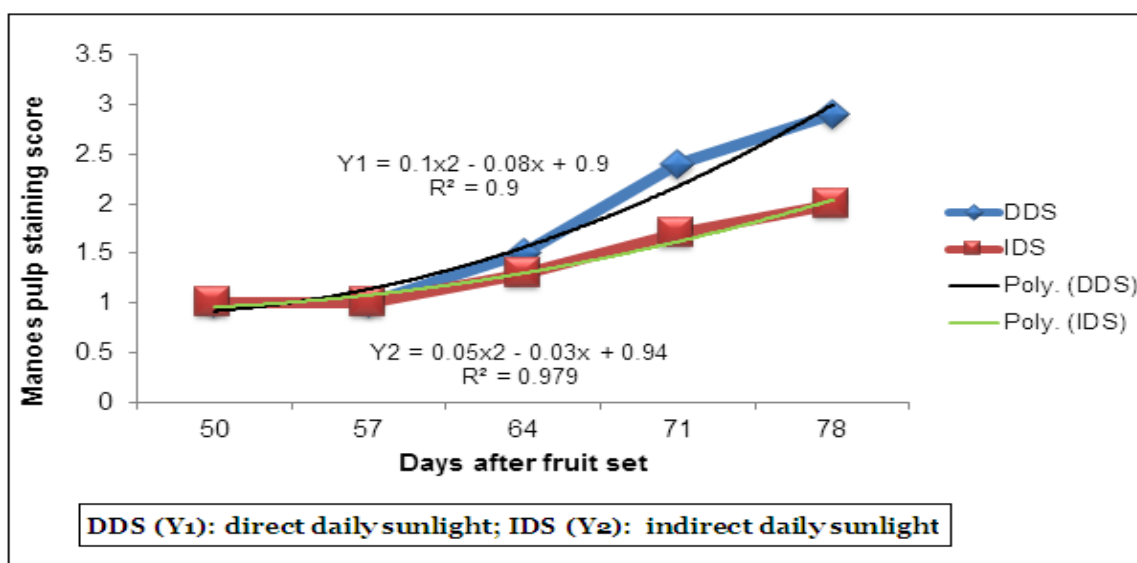


Fig. 8. Evolution of mangoes pulp coloring in the Kent variety during ripening.

Sunshine seems to affect mangoes flavonoid contents. Therefore, direct exposure of fruits to sunshine induces a greater synthesis of flavonoids comparatively to those hidden by the leaves (Bergqvist *et al.*, 2001). Sunshine during mangoes growth are beneficial effect on secondary metabolism, so on the fruit quality. Moreover, many studies on

others speculations have clearly show that flavonoid synthesis induction and their associated enzymes in cuticle or epidermal cells were strongly influenced by sunshine exposure and especially to ultraviolet (UV) radiation (Beggs and Wellman, 1985; Mazza *et al.*, 2000; Wang *et al.*, 2000; Solovchenko and Schimitz-Eiberger, 2003).



Before harvesting, daily sunlight induced temperatures which also influenced mango morphology.

Indeed, the highest morphological growths were obtained at relatively high ambient temperatures (28.72, 30.22 and 30.53°C).

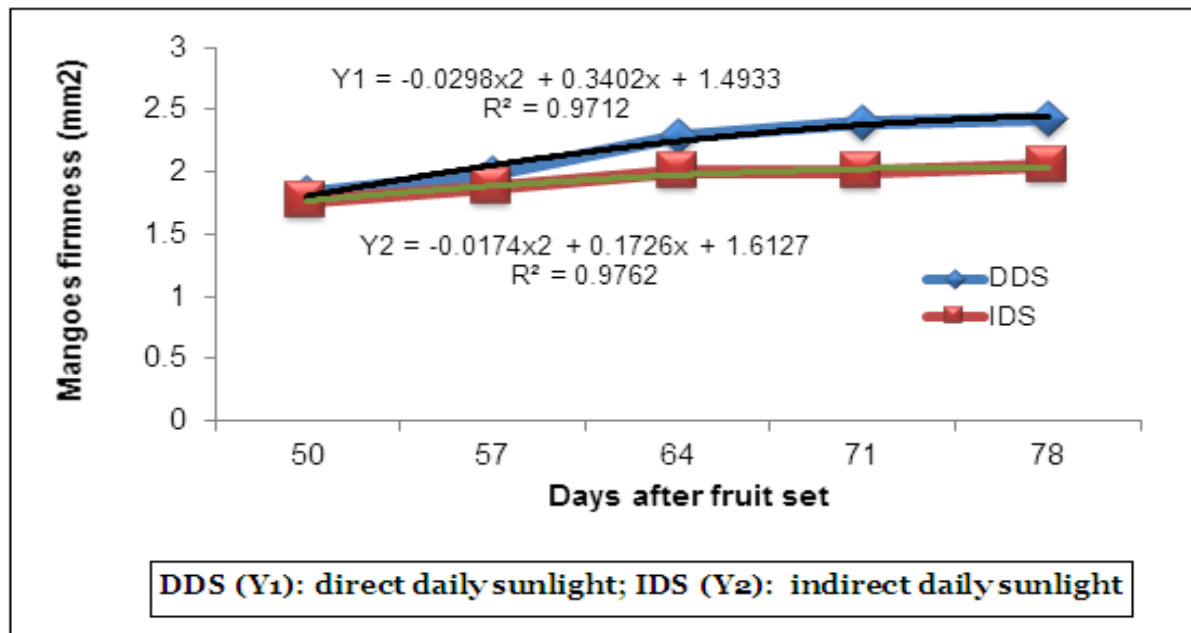


Fig. 9. Evolution of mango firmness in the Kent variety during ripening.

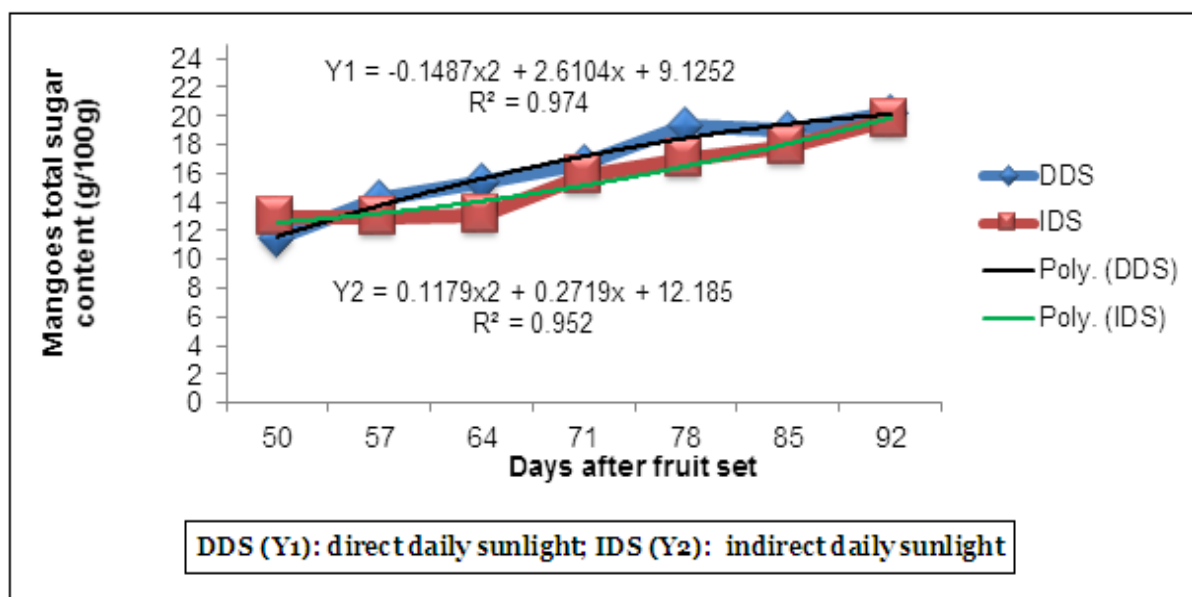


Fig. 10. Evolution of mangoes total sugar content following the type of sun exposure during ripening.

This seems to mean that temperatures influence fruit development by determining its growth rate (Monselise and Goren, 1987; Pantin *et al.*, 2011).

Furthermore, temperature influences enzymatic activities responsible for changes in physicochemical parameters and fruit appearance as mentioned Yao

(2013) in papaya. Ultimately, sunshine and duration of exposure to high temperatures promote important metabolic changes in mangoes before harvest (Ferguson *et al.*, 1998). This has the advantage of predicting the harvest date, improving the quality of mangoes and facilitating their marketing on the international market where competition is tough.

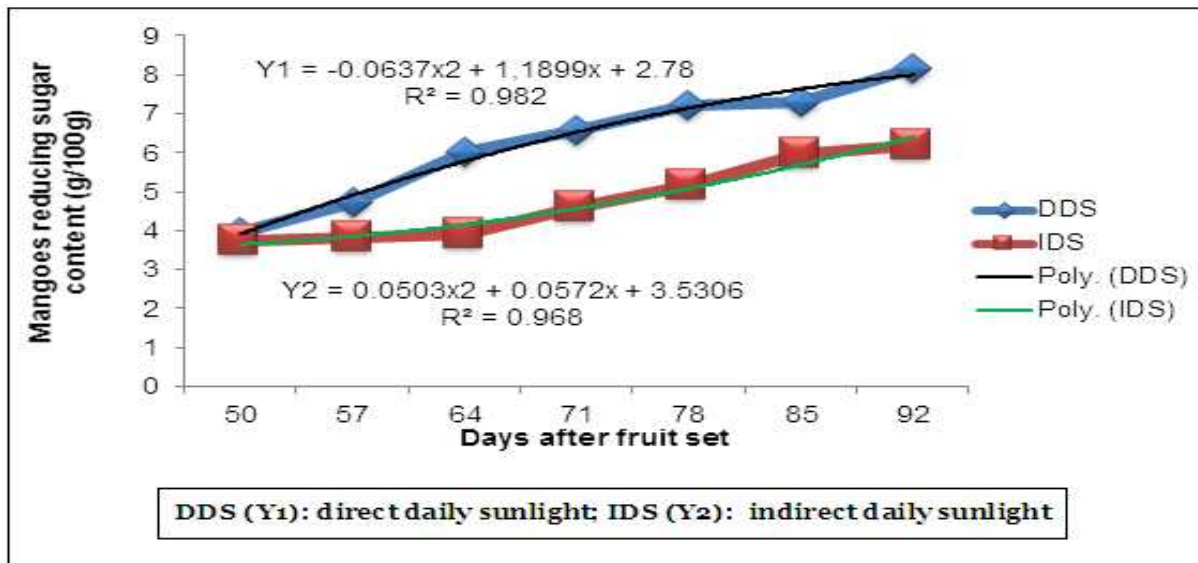


Fig. 11. Evolution of mangoes reducing sugars content following the type of sun exposure during ripening.

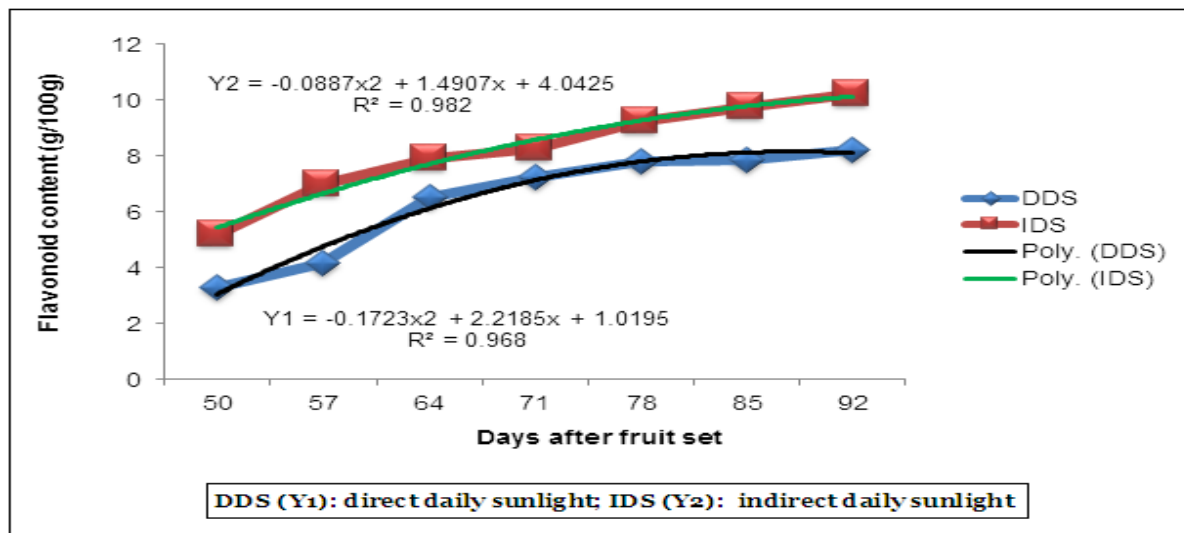


Fig. 12. Variation of mangoes flavonoid content depending on daylight exposure type during ripening.

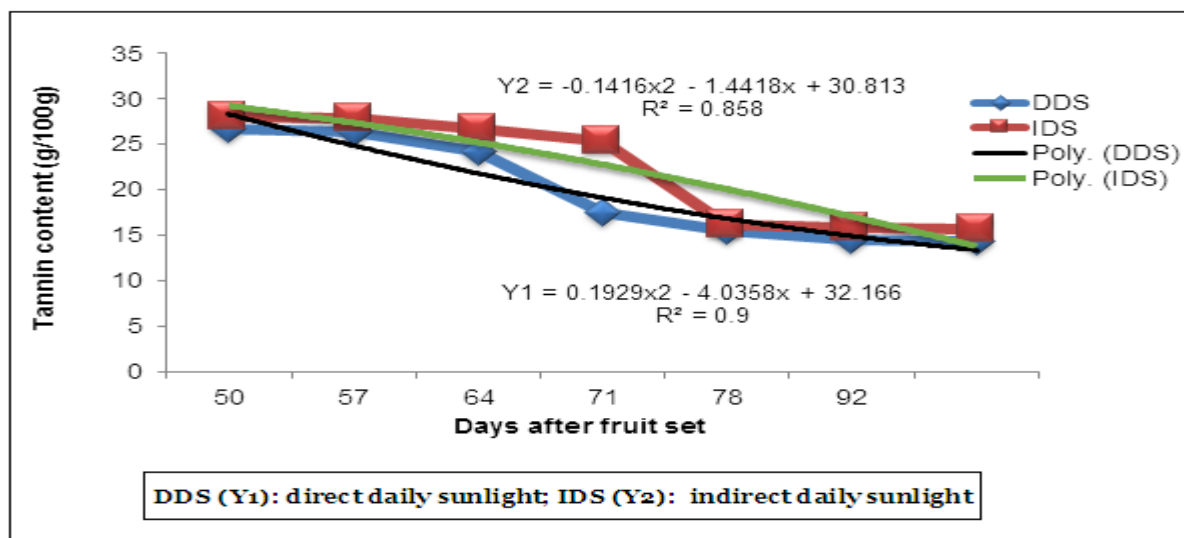


Fig. 13. Variation of mangoes the tannin content following daylight exposure type during ripening.

## Conclusion

Direct exposure of mangoes in tree to sunlight retards their morphological evolution. On the other hand, it improves the physicochemical parameters (internal part) and the appearance of mangoes. The variation of these parameters was therefore closely linked to the sunshine. Indeed, the weak temperatures promoted morphological growth and while the high temperatures were beneficial to the rapid evolution of physicochemical parameters. Thus, mangoes directly exposed to sunshine (DDS) can be harvested earlier than those indirectly exposed (IDS) i.e. hidden under the foliage. Therefore, the mangoes position on the tree with respect to the sunshine was an important factor to take into account when harvesting mangoes of the Kent variety

## References

- Bainbridge Z, Tomlins K, Willings K, Westby A.** 1996. Methods for assessing quality characteristics of non-grain starch staple. Part 4 advanced methods. National Resources Institute. University of Greenwich **1**, 43-79.
- Beggs C, Wellman E.** 1985. Analysis of light controlled anthocyanins formation in coleoptile of *Zea maize* L.: the role of UV-B, blue, red and far-red light. *Photochemistry and Photobiology* **41**, 481- 486.
- Belem A, Tapsoba F, Ouattara ITS, Zongo C, Savadogbo A.** 2017. Étude de la qualité organoleptique de trois variétés de mangues Amélie, Lippens, Brooks séchées au cours du stockage par technique de brunissement enzymatique des peroxydases (POD) et des polyphénoloxydases (PPO). Université d'Ouagadougou 1, Burkina-Faso, p 39.
- Bergqvist J, Dokoozlian N, Ebisuda N.** 2001. Sunlight exposure and temperature effects on berry growth and composition of Cabernet Sauvignon and Grenache in the central San Joaquin Valley of California. *American Journal of Enology and Viticulture* **52**, 1-7.
- Bernfeld P.** 1955. Amylase  $\beta$  and  $\alpha$  (assay method), in methods in enzymology I. Colowick and Kaplan. Edition. Academy press. New York, USA. **1**, 149-158.
- Dambreville A.** 2012. Croissance et développement du manguier (*Mangifera indica* L.) in natura : approche expérimentale et modélisation de l'influence d'un facteur exogène, la température, et de facteurs endogènes architecturaux. Thèse. Université Montpellier 2, France, p 188.
- Delroise A.** 2003. Caractérisation de la qualité et étude du potentiel de maturation de la mangue (*Mangifera indica* L.) en fonction de son stade de récolte, p 37.
- Djioua T.** 2010. Amélioration de la conservation des mangues 4<sup>ème</sup> gamme par application de traitements thermiques et utilisation d'une conservation sous atmosphère modifiée. Thèse de Doctorat. Université d'Avignon et des Pays de Vaucluse, France, p 169.
- Dubois M, Gilles K, Hamilton J, Rebers P, Smith F.** 1956. Colorimetric method for determination of sugars and related substances. *Analytical Chemistry* **28(3)**, 350-356.
- FAOSTAT.** 2001. Market situation Commodity Committee second session. Available: <http://www.fao.org/docrep/Meeting/004/Y1982F.htm> Accessed 5/6/2021
- Ferguson, IB, Snelgar WP, Lay YM, Watkins CB, Bowen JH.** 1998. Expression of heat shock protein genes in apple fruit in the field. *Australian Journal of Plant Physiology* **25**, 155-163.
- FIRCA.** 2011. Répertoire de technologies et de procédés de transformation de la mangue et de l'ananas, p 120.
- Gabrielle F.** 2001. Étude de la composition biochimique de la mangue (*Mangifera Indica* L.) en fonction de son stade de maturité, p 30.
- Ganry J.** 1978. Calcul des sommes de températures moyennes journalières à partir du minimum et du maximum journalier de températures sous climat tropical et équatorial. *Fruit* **3(4)**, 221-236.

- Gautier H, Rocci A, Buret M, Grasselly D, Dumas Y, Causse M.** 2005. Effect of photoselective filters on the physical and chemical traits of vine-ripened tomato fruits. *Canadian Journal of Plant Science* **85**, 439-446.
- Godoy HR, Rodriguez-Amaya DB.** 1989. Carotenoid composition of commercial mangoes from Brazil. *Lebensmittel-Wissenschaft und Technologie* **22**, 100-103.
- Gomez L.** 1997. *Postharvest Physiology. The mango: botany, production and uses.* (Litz RE) Homestead, CAB International: 425-445.
- Hala N, Coulibaly F.** 2007. Étude diagnostique de l'état sanitaire du verger mangoier et acquis de la recherche agronomique sur la lutte intégrée contre les mouches des fruits et la cochenille farineuse du mangoier en Côte d'Ivoire. Rapport Final, Convention CNRA / FIRCA Mangue, p 68.
- Johnson G, Sharp J, Milne D, Oosthuysen S.** 1997. Postharvest technology and quarantine treatments, in: Litz R.E. (Ed.). *The mango botany production and uses.* CAB International. Wallingford, UK. 447-507.
- Kalra SK, Tandon DK.** 1983. Ripening behaviour of « Dashehari » mango in relation to harvest period. *Scientia Horticulturae* **19**, 263-269.
- Larkindale J, Knight MR.** 2002. Protection against heat stress-induced oxidative damage in *Arabidopsis* involves calcium, abscisic acid, ethylene and salicylic acid. *Plant Physiology* **128**, 682-695.
- Martinez HJ, Siddhuraju P, Francis G, Davila OG, Becker K.** 2006. Chemical composition, toxic/anti-metabolic constituents, and effect of different treatments on their levels, in four provenances of *Jatropha curcas* L. From Mexico. *Food Chemistry* **96(1)**, 80-89.
- Mazza CA, Boccalandro HE, Giordano CV, Battista D, Scopel A, Ballaré CL.** 2000. Functional significance and induction by solar radiation of ultraviolet-absorbing sunscreens in field-grown soybean crops. *Plant Physiology* **122**, 117-125.
- Meda A, Lamien CE, Romito M, Millogo J, Nacoulma OG.** 2005. Determination of the total phenolic, flavonoid and proline contents in Burkina Faso honey, as well as their radical scavenging activity. *Food Chemistry* **91**, 571-577.
- Mieu B.** 2017. Côte d'Ivoire: The government wants to restructure the strategic sector of mango. Available: <http://www.Jeuneafrique.com/mag/386974/economie/cotedivoire-gouvernement-veut-restructurerfilierestrategique-de-mangue> Accessed 2/5/2021
- Monselesse SP, Goren R.** 1987. Preharvest growing conditions and postharvest behavior of subtropical and temperate-zone fruits. *HortScience* **22**, 1185-1189.
- Pantin F, Simonneau T, Rolland G, Daudat M, Muller B.** 2011. Control of leaf expansion: A developmental switch from metabolics to hydraulics. *Plant Physiology* **156**, 803-815.
- Paul RE, Gross K, Qiu YX.** 1999. Changes in papaya cell walls during fruit ripening. *Postharvest Biology and Technology* **16**, 79-89.
- Rey JY, Diallo TM, Vannière H, Didier C, Kéita S, Sangaré M.** 2004. La mangue en Afrique de l'Ouest francophone, Synthèse historique. *Fruits* **59**, 121-129.
- Singleton VL, Orthofer R, Lamuela-Raventos RM.** 1999. Analysis of total phenols and other oxidation substrates and antioxidants by means of Folin-Ciocalteu reagent. *Methods Enzymology* **299**, 152-178.
- Solovchenko A, Schmitz-Eiberger M.** 2003. Significance of skin flavonoids for UV-B protection in apple fruit. *Journal of Experimental Botany* **54(389)**, 1977-1984.

- Subramanyam H, Krishnamurthy S, Parpia HAB.** 1975. Physiology and biochemistry of mango fruit. *Advances in Food Research* **21**, 223-305.
- Thibault N.** 2014. Analyse expérimentale et modélisation de l'hétérogénéité de la qualité et de la maturité des mangues. Thèse de Doctorat de l'Université d'Avignon et des pays de Vaucluse, France, p 163-170.
- Touré S.** 2012. Étude nationale mangue. La Côte d'Ivoire et le centre du commerce International, Ed. ECOWAS, p 27.
- Valente M, Dornier M, Piombo G, Grotte M.** 2004. Relation entre la fermeté de la mangue fraîche et la teneur en amidon de la pulpe. *Fruit* **59**, 399-410.
- Wang H, Arakawa O, Motomura Y.** 2000. Influence of maturity and bagging on the relationship between anthocyanin accumulation and phenylalanine ammonia-lyase (PAL) activity in Jonathan apples. *Postharvest Biology and Technology* **19**, 123-128.
- Woolf AB, Bowen JH, Ferguson IB.** 1999. Preharvest exposure to the sun influences postharvest responses on 'Hass' avocado fruit. *Postharvest Biology and Technology* **15**, 143-153.
- Yao NB.** 2013. Conservation du fruit du papayer (*Carica papaya* L. var. solo) par le contrôle du stade de maturité à la récolte et quelques activités biochimiques. Thèse de l'Université Nangui Abrogoua, Côte d'Ivoire, p 120.
- Yapo SE.** 2013. Propagation et régénération in vitro de l'ananas [*Ananas comosus* var. comosus (L. Merrill) Coppens et Leal] cultivé en Côte d'Ivoire et étude physicochimique des fruits issus des vitrocultures. Thèse de de l'Université Nangui Abrogoua, Côte d'Ivoire 18-19.