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**RESEARCH PAPER** 

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# Allelopathic potential of Diospyros kaki L. against Triticum aestivum L., Brassica compestris L. and Trifolium alexandrinum L.

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# Abstract

Allelopathic studies of Diospyros kaki L. were conducted by using aqueous extracts from leaves, bark, litter and mulching in various experiments. It was observed that the germination, radical growth, plumule growth, no. and growth of seminal roots, fresh weight and dry weight of Triticum aestivum L and Brassica campestris L. and Trifolium alexandrinum L. were significantly reduced in various bioassays. The aqueous extracts of leaves were inhibitorier than the bark extracts the Triticum for test species aestivum Aqueous extracts obtained at room temperature were inhibitorier than hot water extracts. Litter and mulching experiments also showed inhibitory effects. Further studies are required to see its allelopathic behavior under field condition against its associated species, to identify the toxic principle, and to evalvate as biocontrol agents for weeds, insect and disease control.

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#### Introduction

Allelopaty is direct or indirect effect of one plant to adjoining plants through release of chemical substances (Rice 1979). Allelopathy is "the release of phytotoxins by plants" (Bais et al., 2003). Allelopathy affects plant distribution, community formation, intercrop evolution and biodiversity conservation (Peneva 2007). Allelochemical can be present in root, stem, leaves, flowers and fruits, and inhibit root growth, shoot growth, germination percentage, nutrients uptake (Murawat and khan 2006).

Many plant species such as Terminalia bellirica, Terminalia chebula, Aegle marmelos and Sapindus mukorossi (Thapaliyal et al., 2008); Ficus subincisa , Bauhinia purpurea, and Toona hexandra (Sing et al., 2009); Cenchrus ciliaris and Bothriochloa pertusa (Hussain et al., 2010); Dodonaea viscose ( Barkatullah et al., 2010); Juniperus ashei (Young & bush 2009); Azadirachta indica ( Ashrafi et al., 2008); Cassia angustifolia (Hussain et al., 2007)and Cenchrus ciliaris and Bothriochloa pertusa (Hussain & Ilahi 2009) allelopathic effect on test species. The other workers, (Khan et al., 2005; Hussain et al., 2007; Maharjan et al., 2007; Uddin et al., 2007, Salih et al., 2009; Sun et al 2006; Kumar et al., 2006; Terzi 2008; khan et al., 2005; Thapaliyal et al., 2008). Also conducted Allelopathic studies of literature reveals that no such study was conducted on Diospyros kaki L.

Diospyros kaki Linn (Family Ebenaceae) is Small cultivated tree, up to 15 m tall, originating from Japan, Introduced and cultivated throughout Eastern Asia, Russia, Japan and China. The original habitat of the species is China, but it has been introduced and cultivated elsewhere. It also cultivated in Pakistan for its edible fruit, especially in Swat, Dir and Malakand. The aim of the study was to check out the Allelopathic potential of Diospyros kaki Linn against some selected valuable crops.

#### Material and method

Mature leaves and bark of Diospyros kaki L. were collected from Swat, dried at room temperature (25°C- 30°C) and powdered, Glass wares were washed with tap water and sterilized at 170°C for at least 4 hours. All the results were statistically analyzed through one way ANOVA.

#### Effect of aqueous extracts

Five and 10 gm of each part were separately soaked in 100 ml distilled water at 25°C for 24 and 48 hours respectively and filtered to get aqueous extracts. These extracts were used against Triticum aestivum Brassica campestris L. and Trifolium alexandrinum L. used as the test species on 2-fold filter paper in petri dishes following standard filter paper bioassay (Hussain & Ilahi 2009; Barkatullah et al., 2010, Hussain et al., 2010). The filter papers were moistened with the aqueous extracts, while distilled water was used as a control. For each treatment, five replicates, each with 10 seeds were made. The petri dishes were incubated at 25°C. After 72 hours, the percent germination, length of plumule and radical was noted. Twenty seedlings were randomly taken for fresh and dry weight determination and moisture contents. Seedlings were dried at 65°C for 72 hours for the determination of dry weight and moisture contents.

# Effect of hot water extracts

Five gm and 10gm of dried plant parts were separately boiled in 100 ml water for 5 minutes and filtered. The room cooled extracts were applied against the same test species as before.

# Effect of litter

Five gm of litter each from leaves and bark were crushed and placed over one fold of filter paper in a petri dish. The filter papers were moistened with 5 ml water. In control treatment fine pieces of filter paper were used instead of plant material.

Table 1. Effect of cold aqueous extract on germination, plumule and radical growth of test species. Each value is a mean of 5 replicates each with 10 seedlings.

Test species		Triticu a	estivum L.		В	Brassica compestris L.			Trif	Trifolium alexandrinum <u>L.</u>			
Soaking duration and concentration	5g/ 24h	5g/ 48h	10g/24h	10g/48h	5g/ 24h	5g/ 48h	10g/24h	10g/48h	5g/ 24h	5g/ 48h	10g/24h	10g/48h	
					Ger	mination '	%						
Control	82	82	82	82	92	92	92	92	94	94	94	94	
Bark	78	84	68*	80	82	72	88	86	100	96	90	100	
% of control	95.12	102.43	82.92	97.56	89.13	78.26	95.65	93.47	106.38	102.12	95.74	106.38	
Leaves	74	70	94	88	58***	40***	22***	14***	92	98	96	94	
% of control	90.24	85.36	114.63	107.31	63.04	43.47	23.91	15.21	97.87	104.25	10212	100	
Control	8.18	8.18	8.18	8.18		e growth		6.0	- 60	- 60	- 60	- 60	
Bark	5.76**	3.78***	5.56***	3.7***	6.9 4.26***	6.9 1.18***	6.9 2.82***	6.9	5.68 8.26	5.68	5.68 6	5.68	
% of control		3./6 46.21		- ,			40.86	1.54***		4.56 80.28		5.76	
% of control Leaves	70.14 3.78***	1.86***	67.97 3.8***	45.23 1.62***	61.73	17.10 0.58***	0.32***	23.04 0.20***	145.42 2.22***	2.8***	105.63 1.76***	101.40 1.96***	
% of control	3./6 46.21	22.73	3.6 46.45	1,02	1.54*** 22.31	8.40	3.91	2.89	39.08	49.29	30.98	_	
70 OI COILLOI	40.21	22./3	40.45	19.00	22.31	0.40	3.91	2.09	39.00	49.29	30.90	34.50	
					Radical	growth (	mm)						
Control	14.84	14.84	14.84	14.84	9.52	9.52	9.52	9.52	8.16	8.16	8.16	8.16	
Bark	12.94	7.74***	13.7	7.82***	5.96***	2.98***	5.02***	4.16***	7.6	7.18	5.72***	9.34	
% of control	87.19	52.15	92.31	52.69	62.60	31.30	52.73	43.69	93.13	87.99	70.09	114.46	
Leaves	12.62	7.94***	14.54	6.92***	4.04***	1.06***	0.92***	0.36***	3.94***	6.12***	4.0***	6.22***	
% of control	85.04	53.50	97.97	46.63	42.43 Seminal r	11.13 oot growt	9.6 h (mm)	3.78	48.28	75.0	49.01	76.22	
Control	11.56	11.56	11.56	11.56	0	0	0	0	0	0	0	0	
Bark	8.65*	3.94***	8.64*	4.14***	0	0	0	0	0	0	0	0	
% of control	74.82	34.08	74.74	35.81	0	0	0	О	O	0	0	О	
Leaves	8.86*	2.32***	9.16*	2.92***	0	0	0	О	O	0	0	О	
% of control	76.64	20.06	79.23	25.25	0	0	0	О	О	0	0	О	
			,,		No of	Seminal r	oot						
Control	1.52	1.52	1.52	1.52	0	0	0	0	0	0	0	0	
Bark	1.46	1.64	1.32	1.30	0	0	0	О	О	0	0	О	
% of control	96.05	107.89	86.84	85.52	0	0	0	0	0	0	0	0	
Leaves	1.38	1.46	1.92	1.72	0	0	0	0	0	0	0	0	
% of control	90.78	96.05	126.31	113.15	0	0	0	0	0	0	0	0	

### Effect of mulching

Five gm each of the crushed dried leaves and bark were placed in plastic cups which were half filled with sterilized moist sand. For each treatment five replicate, each with 10 seeds were made. Control consisted of fine pieces of filter paper. The plastic cups were incubated at 25°C and observed for germination. After 7 days growth of plumule and radical were measured. Twenty seedlings were randomly taken for fresh and dry weight and moisture contents.

#### Results and discussion

### Effect of aqueous extracts

Percent germination was not affected by the application of aqueous extracts of both bark and leaves in all test species (Siddiqui et al., 2009), but radical and plumule length, fresh weight, dry weight and moisture contents are declined (Table 2). (Barkatullah et al., 2010; Hussain et al 2010: Samreen et al., 2009, hussain & ilahi 2009) have also reported similar results for allelopathic effects of various plants. Except 5g/24h aqueous extract percent germination in Triticum aestivum L. and the aqueous extracts of leaves reduce germination in Brassica compestris L. (Alagesaboopathi. 2010; Uniyal & Chhetri 2010). Plumule, radical and percent germination are not effected by application of aqueous extracts of bark in Trifolium alexandrinum L. Seminal roots were also inhibited in Triticum aestivum L. More inhibition occured by increasing concentration of the aqueous extracts. (Samreen et al., 2009; Hussain et al., 2010 and Barkatullah et al., 2010).

**Table 2**. Effect of aqueous extract on fresh weight, dry weight and moisture content of seedlings. Each value is a mean of 20 randomly selected seedlings.

				0								
	Fresh weight (mg)											
Control	2322	2322	2322	2322	408.1	408.1	408.1	408.1	319.1	319.1	319.1	319.1
Bark	2003.6	1587.2	2122	1592	175.6	128.3	211.4	161.4	299.6	193.9	299.8	185.4
% of	86.28	68.35	91.38	68.56	43.02	31.43	51.80	39.54	93.88	60.76	93.95	58.10
control												
Leaves	1798.3	1364.8	1869.4	1498.8	156.3	0	0	0	205.5	184.1	211.8	136
% of	77.44	58.77	80.50	64.54	38.29	О	0	0	64.39	57.69	66.37	42.61
control												
					Dry	weight (	mg)					
Control	1249.6	1249.6	1249.6	1249.6	42.1	42.1	42.1	42.1	31.5	31.5	31.5	31.5
Bark	1058.9	935.8	1152.4	974.4	18.6	23.1	31.2	29.2	22.4	29.1	37.1	23.6
% of	84.73	74.88	92.22	77.97	44.18	54.86	74.10	69.35	71.11	92.38	117.7	74.92
control												
Leaves	1109.2	905.2	1155.3	932.9	31.8	0	0	0	25.1	28.1	34.6	27.8
% of	88.76	72.43	92.45	74.65	75.53	0	0	0	79.68	89.20	109.84	88.25
control												
					Moistur	re conten	ts (mg)					
Control	85.86	85.86	85.86	85.86	869.35	869.35	869.35	869.35	913.01	913.01	913.01	913.01
Bark	89.21	69.60	84.08	63.38	844.08	455.4	577.56	455.73	1237.5	566.32	708.08	685.59
% of	96.24	81.06	97.92	73.81	97.09	52.38	66.43	52.42	135.54	62.02	77.55	75.09
control												
Leaves	62.12	50.77	100.3	60.66	391.5	О	О	0	718.72	555.16	512.13	389.2
% of	72.35	59.13	116.81	70.64	45.03	0	0	0	78.71	60.80	56.09	42.62
control												

<sup>\*</sup>Significantly different from control at alpha 0.050 according to one way ANOVA

The inhibition of early growth of seedling is a critical step by the aqueous extracts. In many cases germination may be stimulated but seedling growth is strongly reduced. In present case the plumule and radical growth of both test species was significantly retarded in all the test conditions (Table 1). These findings agree with those of Khan *et al.*, (2008); Barkatullah *et al.*, (2010); Hussain *et al.*, (2010), who observed inhibited growth of seedlings by inhibitors from other plants. The present results are also in line with those of Uniyal & Sachin (2010), Abugre & Sam (2010) and Hussain & Ilahi (2009) who also observed allelopathic inhibition of radical and plumule growth of test species.

A weak seedling is disadvantage to a growing plant as it cannot properly take up water and minerals from the habitat. The reduction in growth could be due to water loss that leads to poor biomass. It has been seen that inhibited seedlings also have poor fresh and dry weight (Table 2), which means poor accumulation of food and growth. Similar reduction in fresh and dry weight of seedlings has been reported by many workers (Uniyal & Sachin 2010; Barkatullah et al., 2010; Hussain et al., 2010: Samreen et al., 2009, hussain & ilahi 2009) and our findings agree with them. The reduction of moisture contents of seedlings mean failure of seedlings to absorb sufficient soil moisture. A drought like condition prevailed imparity in the functioning of root, radical mean poor growth performance as was evident in the present study. Such a reduction in moisture contents as been reported by earlier workers (Hussain et al 2010; Barkatullah et al., 2010; Hussain & Ilahi 2009; Uniyal & Sachin 2010; Abugre & Sam 2010).

<sup>(\*</sup>less significant, \*\*moderately significant, \*\*\*highly significant)

Table 3. Effect of hot water extract on the germination, plumule and radical growth of test species. Each value is a mean of 5 replicates, each with 10 seedlings.

Test species	Triticu a	estivum L.	Brassica co	mpestris L.	Trifolium ale	Trifolium alexandrinum <u>L</u>		
Hot water extract	5g	10g	5g	10g	5g	10g		
and concentration								
			Germination %					
Control	82	82	92	92	94	94		
Bark	84	80	92	94	100	98		
% of control	102.43	97.56	100	102.17	106.38	104.25		
Leaves	74	84	64	68	96	98		
% of control	90.24	102.43	69.56	73.91	102.12	104.25		
a . 1			umule growth (m					
Control	8.18	8.18	6.9	6.9	5.68	5.68		
Bark	5.84	7.28	2.62***	2.7***	1.3***	16.16***		
% of control	71.39	88.99	37.97	39.13	22.88	284.50		
Leaves	2.76***	3.26***	0.82***	0.92***	4.64	3.26***		
% of control	33.74	39.85	11.88	13.33	81.69	57.39		
Control	0 .		adical growth (m		8.16	8.16		
Bark	14.84	14.84	9.52	9.52				
	16.36	16.42	7.76	10.04	16.3	14.06		
% of control	110.24	110.64	81.51	105.46	199.75	172.30		
Leaves	9.28*	8.9*	2.62***	2.42***	3.28***	2.72***		
% of control	62.53	59.97	27.52	25.42	40.19	33.33		
		Semi	inal root growth	(mm)				
Control	11.56	11.56	0	0	0	О		
Bark	11.26	10.98	0	0	0	0		
% of control	97.40	94.98	0	0	0	О		
Leaves	6.38*	6.94	0	0	0	О		
% of control	55.19	60.03	0	0	0	О		
		1	No of Seminal roo	ot				
Control	1.52	1.52	0	0	0	0		
Bark	1.64	1.48	0	О	0	0		
% of control	107.89	97.36	O	0	0	0		
Leaves	1.26	1.42	O	О	0	0		
% of control	82.89	93.42	0	0	0	0		

# *Effect of hot water extracts*

Use of hot water extracts is unusual in natural environment, but to facilitate and save time, similar studies have been made ( Hussain et al., 2010; Barkatullah et al., 2010; Samreen et al., 2009; Hussain & Ilahi 2009). In the present study hot water extract significantly caused inhibition (Table 3), Seminal root were also inhibited in wheat. But the hot water extracts of bark has not affected test species Triticum aestivum L. while inhibited plumule growth in Brassica and Trifolium. The leaf extract inhibited plumule, radical and seminal root in Triticum aestivum L., plumule, radical and percent germination in Brassica. Only 5g hot water

leaves extracts inhibited radical growth in Trifolium alexandrinum L.

These results intimated that phytotoxins were easily extracted within short time and the phytotoxins retained phytotoxicity after boiling. The present study also showed that hot water extracts effectively exhibited allelopathy. Inspite of all favours, it is an unusual process that hardly can be possible in nature. The results of cold and hot water extracts almost agree with each other and strengthen the view that extracts of Diospyrus kaki L. are inhibitory to the test species.

**Table 4.** Fresh weight, dry weight and moisture content of test species in hot water extract bioassay.

				·	
2322	2322	408.1	408.1	319.1	319.1
1904	1763.6	115.6	99.4	95.3	100.6
81.99	75.95	28.32	24.35	29.86	31.52
1581.2	1616	225	61.3	124.6	39.6
68.09	69.59	55.13	15.02	39.04	12.40
1249.6	1249.6	42.1	42.1	31.5	31.5
1015.3	943	37.8	40.2	25.7	21.3
81.25	75.46	89.78	95.05	81.58	67.61
1009.8	992.3	39.9	44.1	26.9	26.4
80.80	79.40	94.77	104.75	85.39	83.80
85.86	85.86	869.35	869.35	913.01	
87.56	87.02	205.8	147.2	326.76	
101.97	101.35	23.67	16.93	35.78	
56.58	62.85	463.9	39.0	363.19	
65.89	73.20	53.36	4.48	39.77	
	1904 81.99 1581.2 68.09 1249.6 1015.3 81.25 1009.8 80.80 85.86 87.56 101.97 56.58	1904 1763.6   81.99 75.95   1581.2 1616   68.09 69.59   1249.6 1249.6   1015.3 943   81.25 75.46   1009.8 992.3   80.80 79.40   85.86 85.86   87.56 87.02   101.97 101.35   56.58 62.85	1904 1763.6 115.6   81.99 75.95 28.32   1581.2 1616 225   68.09 69.59 55.13   1249.6 1249.6 42.1   1015.3 943 37.8   81.25 75.46 89.78   1009.8 992.3 39.9   80.80 79.40 94.77   85.86 85.86 869.35   87.56 87.02 205.8   101.97 101.35 23.67   56.58 62.85 463.9	1904 1763.6 115.6 99.4   81.99 75.95 28.32 24.35   1581.2 1616 225 61.3   68.09 69.59 55.13 15.02   1249.6 1249.6 42.1 42.1   1015.3 943 37.8 40.2   81.25 75.46 89.78 95.05   1009.8 992.3 39.9 44.1   80.80 79.40 94.77 104.75   85.86 85.86 869.35 869.35   87.56 87.02 205.8 147.2   101.97 101.35 23.67 16.93   56.58 62.85 463.9 39.0	1904 1763.6 115.6 99.4 95.3   81.99 75.95 28.32 24.35 29.86   1581.2 1616 225 61.3 124.6   68.09 69.59 55.13 15.02 39.04   1249.6 1249.6 42.1 42.1 31.5   1015.3 943 37.8 40.2 25.7   81.25 75.46 89.78 95.05 81.58   1009.8 992.3 39.9 44.1 26.9   80.80 79.40 94.77 104.75 85.39   85.86 85.86 869.35 869.35 913.01   87.56 87.02 205.8 147.2 326.76   101.97 101.35 23.67 16.93 35.78   56.58 62.85 463.9 39.0 363.19

<sup>\*</sup>Significantly different from control at alpha 0.050 according to one way ANOVA

**Table 5.** Effect of litter and mulching on germination, plumule and radical growth of test seedlings. Each value is a mean of five replicates, each with 10 seedlings.

Treatment		Litter			Mulching	
Test species	Triticum	Brassica	Trifolium	Triticum	Brassica	Trifolium
	aestivumL.	compestris l.	alexandrinum L.	aestivum L.	$compestris\ L.$	alexandrinum L.
			Germination %			
Control	82	92	94	76	68	84
Test	86	76	96	42	40**	80
% of control	104.87	82.60	102.12	55.26	58.82	95.23
			Plumule growth (mn	n)		
Control	8.18	6.9	5.68	74.98	7.7	22.42
Test	5.52	3.86***	5.46	10.9***	3.38***	21.28
% of control	67.48	55.94	96.12	14.53	43.89	94.91
			Radical growth (mm	1)		
Control	14.84	9.52	8.16	31.24	30.56	10.38
Test	15.26	8.46	9.02	5.12***	11.44***	12.5
% of control	102.83	88.86	110.53	16.38	37.43	120.42
		S	eminal root growth (n	nm)		
Control	11.56	0	0	20.48	0	0
Test	10.62	О	0	3.14***	O	О
% of control	91.86	0	0	15.33	0	0
			No. of Seminal root			
Control	1.52	0	0	4.3	0	0
Test	1.68	0	0	1.84	0	0
% of control	110.52	0	0	42.79	0	0

<sup>(\*</sup>less significant, \*\*moderately significant, \*\*\*highly significant)

Effect of mulching and litter

It is commonly understood that litter improves soil nutrients and physiochemical features of soil. However, it is also agreed (Inderjit & Duke 2003; Sasikumar et al., 2001; Hussain & Ilahi 2009; Barkatullah et al., 2010; Samreen et al., 2009; Hussain et al., 2010) that litter prior to decay might release phytotoxins in the soil. This possibility was envisaged by performing litter and mulching experiments. In both these experiments it was seen

that only plumule growth are inhibited by mulch in Triticum aestivum L. Brassica compestris L. while there was no effect on Trifolium alexandrinum L. (Table 5). Litter inhibited percent germination, plumule and radicle growth of Triticum aestivum L. Brassica compestris L. while Trifolium alexandrinum L. was not affected. The fresh and dry weight and moisture contents of test species, in both the experiments decreased significantly (Table 6).

**Table 6.** Effect of added litter and mulch on the fresh weight, dry weight and moisture contents of seedlings.

	Fresh weight (mg)									
Control	2322	408.1	319.1	2859.9	533	292.9				
Test	1904.6	152.3	254.9	1722.6	310	209				
% of control	82.02	37.31	79.88	60.23	58.16	71.35				
		]	Dry weight (mg)	)						
Control	1249.6	42.1	31.5	1042	98.9	54.8				
Test	1129.8	25.7	25.6	896.3	39.1	54.4				
% of control	90.41	61.04	81.26	86.01	39.53	99.27				
		Moi	isture content (1	mg)						
Control	85.86	869.35	913.01	174.46	438.9	434.48				
Test	68.57	492.6	418.43	92.19	692.8	284.1				
% of control	79.86	56.66	45.82	52.84	157.84	65.38				

<sup>\*</sup>Significantly different from control at alpha 0.050 according to one way ANOVA

(\*less significant, \*\*moderately significant, \*\*\*highly significant)

It was evident that added litter and mulch proved inhibitory just like the aqueous extracts. The litter and mulch reduced germination, seedling growth and physiological aspects of tested plants. It can be visualized that addition of litter from Diospyros kaki L. might intoxicate the soil as observed in the present case.The litter of Dodonaea viscosa (Barkatullah et al., 2010), Cenchrus ciliaris and Bothriochloa pertusa (Hussain et al., 2010) have been reported to exhibit similar inhibition and our findings agree with these. The present investigation also indicated that leaves and bark were differentially toxic to the Triticum, Brassica and trifolium. These test species had their own differential response towards the same extract. Similar differential behavior of Cenchrus ciliaris and Bothriochloa pertusa (Hussain & Ilahi 2009), Calotropis procera (Samreen et al., 2009), Eucalyptus camadulensis (Mohammad & Rajaie, 2009) has been reported, which support the present

findings. Furthermore, phytotoxicity is depended upon the concentration, soaking duration and the physiological responses of test species.

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