

International Journal of Biosciences | IJB | ISSN: 2220-6655 (Print), 2222-5234 (Online) http://www.innspub.net Vol. 17, No. 4, p. 73-82, 2020

RESEARCH PAPER

OPEN ACCESS

A comparative analysis on mitotic interphase and prophase among twelve varieties of *Brassica* L. from Bangladesh: Brassicaceae

Susmita Saha, Kazi Nahida Begum*

Department of Botany, Faculty of Life and Earth Sciences, Jagannath University, Dhaka, Bangladesh

Key words: Brassica L., interphase, prophase, orcein-staining.

http://dx.doi.org/10.12692/ijb/17.4.73-82

Article published on October 10, 2020

Abstract

Brassica L. is an agronomical and economical important crop belonging to Brassicaceae family. The research was conducted to interrelate interphase nuclei and prophase chromosome of *Brassica* L. varieties. In the current analysis, the nature of interphase nuclei and prophase chromosome of twelve BARI varieties of *Brassica* L. was investigated based on orcein-staining properties. In the interphase nuclei only 'Diffuse type' and 'Simple Chromocenter type' were found in Tori-7, Dawlat, BS-11, BS-14 and SS-75, Rai-5, BS-6, BS-7, BS-8, BS-10, BS-12, BS-15, respectively. Considering prophase chromosome among these studied twelve varieties, 'Continuous type' showed in Dawlat, Rai-5, BS-7, Bs-8, BS-11, BS-14 while BS-6, BS-10, BS-12 and BS-15 observed 'Interstitial type' and 'Gradient type' showed in Tori-7, and SS-75. Therefore, the orcein staining property of interphase nuclei and prophase chromosome can provide essential information as cytological implement to discriminate the twelve analyzed *Brassica* L. varieties.

* Corresponding Author: Kazi Nahida Begum 🖂 kazinahida@yahoo.com

Introduction

Brassica L. is one of the most commercially important genus of Brassicaceae family which wildly distributed throughout the world. This genus mainly originated from close regions of Himalaya and then dispersed from Asia to European-Mediterranean territory (Downey and Robelen, 1989). About contrasting 37 species includes into the genus Brassica L. with divergent agronomic traits as vegetable and oilseed crops (Jahan et al., 2013). On the basis of divergent morphological characteristics and genetic diversity six interlinked species are found in the genus Brassica L., of which three amphidiploids species (B. carinata, B. juncea and B. napus) are evaluated from three diploid species, (Brassica campestris, B. nigra, and B. oleracea) (Nagahara, 1935). Among these six varieties, the four most widely cultivated species are Brassica campestris L., B. juncea (L.) Czern and Coss., B. napus L., and B. carinata Braun for oilseed and vegetables (Rakow, 2004).

Generally, the genus *Brassica* L. can be categorized into three groups: rapeseed, mustard and cole. Rapeseed-mustard is well known for edible oil and protein whereas cole is consumed as vegetables. As oil seed *Brassica* L. achieves second largest contributor role in global oil production after soybean (Raymer, 2002). Mustard oil is not only used for as cooking oil but also marinate food stuffs and salad dressings. Moreover, as an edible oil *Brassica* L. is worthier for human health due to the presence of linoleic acid (omega 3 fatty acid) and alpha linoleic acid (omega 3 fatty acid) (Mollika *et al.*, 2011).

Brassica L. is an excellent source of potassium, phenolics, dietary fiber, vitamins A, C and E as well as a renewable resource in the petro-chemical industry (Zhang *et al.*, 2006).

The oil meal of *Brassica* L. has a quality value in beef and poultry ration as a protein supplement. According to Luciano and Holley (2009), the mustard has antibacterial and antifungal properties considering the substance similar to glucosinolate. Brassinosteroids have a great influence to control both prostate and breast cancer which are highly distributed in pollen and seed of *B. campestris* and *B. napus* (Wachsman *et al.*, 2012).

Due to the commercial values of Brassica L., a large number of tremendous investigations have been occurred for morphological, physiological and biochemical improvement in worldwide as well as Bangladesh (Zhang, 1996; Chen et al., 2001; Hu et al., 2001; Khan et al., 2002; Xiao et al., 2004; Liu, 2007; Mollika et al., 2011; Akbar and Begum, 2020; Paul et al., 2020). But still it is essential to know the cytological and cytogenetical information of a species because it plays a significant role to relevant its evolution and diversification (Ropiquet et al., 2008). Based on previous literature, researchers were concentrating to reveal the chromosomal information and molecular analysis of Brassica L. throughout the world (Takamine, 1916; Du et al., 1993; Olin-Fatih 1994; Cheng et al., 1995; Fukui et al., 1998; Kulak et al., 2002; Snowdon 2007; Fang et al., 2014; Sun et al., 2018). On account of small size chromosome, only chromosome number and classical cytogenetical information is insufficient for characterization of species or varieties (Begum and Alam, 2016). In such cases, the feature of interphase nuclei and prophase chromosome act as key cytological tool to characterization of any specimens. Therefore, the current investigation has been approached to reveal the nature of interphase nuclei and prophase chromosome of mitotic cell division using orcein staining that should provide convenient information for disguising twelve BARI varieties of Brassica L., because this information is not available in the contemporary scientific literatures.

Materials and methods

Plant materials

To conduct this current investigation, all the twelve varieties of *Brassica* L. (Table 1, Figs. 1A-W, 2A-W) were assembled from Oil Research Center (ORC) of Bangladesh Agricultural Research Institute (BARI), Joydebpur, Gazipur and then maintained in the Botanical garden of Jagannath University, Dhaka,

Bangladesh.

Sample preparation

At first the wet filter papers were placed in a Petri plate to germinate seeds for 36 h and 15 min at room temperature in the dark and Root tips (RTs) were gently taken from germinated seeds. Then, by using 8-hydroxyquinoline (0.002 M) for the RTs were pretreated 50 min at room temperature and fixed in Carnoy's fluid (Glacial acetic acid Ethanol 1: 3) at 4°C for 24 hours and then preserved in 70% alcohol for later use. Next, a mixture of 1% aceto-orcein and 1 N HCl was used to heat the pre-treated RTs through a spirit burner. Afterwards, the prepared aceto-orcein stained RTs were mildly squashed and finally prepared slides were examined under the Optika electric microscope and microscopic images were captured by the Euromax camera (CMEX 10, DC 10000C) with 100X magnification in Auto mode.

Analytic form

For considering the behavior of interphase and prophase of inspected twelve *Brassica* L. varieties, from 40 to 50 interphase nuclei and prophase chromosome were examined and then categorized on the basis of Tanaka's (1971) classification.

Results and discussion

Depending on orcein-staining properties, the conducting investigation disclosed that noticeable darkly stained heterochromatin blocks were formed by the interphase nuclei of eight BARI *Brassica* L. varieties among studied twelve varieties in which four varieties of *B. campestris* (SS-75, BS-6, BS-12 and BS-15), two varieties of *B. juncea* (Rai-5 and BS-10) and two varieties of *B. napus* (BS-7 and BS-8) (Table 2, Figs. 3B-D, 3F, 3G, 3I, 3L-K); Tanaka (1971) classified this nature of interphase nuclei as 'Simple Chromocenter type'.

Table 1. List of twelve BARI varieties of *Brassica* L. used in present cytological analysis.

Serial No	Species	Variety
1	Brassica campestris	Tori-7
2	B. campestris	Sonali Sarisha-75 (SS-75)
3	B. campestris	BARI Sarisha-6 (BS-6)
4	B. campestris	BARI Sarisha-12 (BS-12)
5	B. campestris	BARI Sarisha-14 (BS-14)
6	B. campestris	BARI Sarisha-15 (BS-15)
7	B. juncea	Dawlat
8	B. juncea	Rai-5
9	B. juncea	BARI Sarisha-10 (BS-10)
10	B. juncea	BARI Sarisha-11 (BS-11)
11	B. napus	BARI Sarisha-7 (BS-7)
12	B. napus	BARI Sarisha-8 (BS-8)

Table 2. Types of interphase nuclei and prophase chromosomes of the genus *Brassica* L. after staining with Orcein.

Variety	Type of orcein-stained	Type of orcein-stained prophase
	interphase nuclei	chromosomes
B. campestris var. Tori-7	Diffuse	Gradient
B. campestris var. Sonali sarisha-75	Simple chromocenter	Gradient
B. campestris var. BARI Sarisha-6	Simple chromocenter	Interstitial
B. campestris var. BARI Sarisha-12	Simple chromocenter	Interstitial
B. campestris var. BARI Sarisha-14	Diffuse	Continuous
B. campestris var. BARI Sarisha-15	Simple chromocenter	Interstitial
<i>B. juncea</i> var. Dawlat	Diffuse	Continuous
<i>B. juncea</i> var. Rai-5	Simple chromocenter	Continuous
B. juncea var. BARI Sarisha-10	Simple chromocenter	Interstitial
B. juncea var. BARI Sarisha-11	Diffuse	Continuous
B. napus var. BARI Sarisha-7	Simple chromocenter	Continuous
B. napus var. BARI Sarisha-8	Simple chromocenter	Continuous

On the other hand, the interphase nuclei of remaining four varieties were found to possess uniformly stained nucleus of which two varieties of *B. campestris* (Tori-7 and BS-14) and two varieties of *B. juncea* (Dawlat and BS-11) (Table 2, Figs. 3A, 3E, 3H, 3G, 3J), in other words heterochromatin blocks were not localized rather distributed homogenously all over the nucleus which regarded as 'Diffuse type' by Tanka (1971).

In interphase, two varieties of *B. campestris* (Tori-7 and BS-14) and two varieties of *B. juncea* (Dawlat and BS-11) occupied a prominent nucleolus (Figs. 3A, 3E, 3G, 3J, arrow) but not figure out in prophase.



Fig. 1. Habit of six varieties of *Brassica* L. A-D. *B. campestris* var. Tori-7; E-H. *B. campestris* var. SS-75; I-L. *B. campestris* var. BS-6; M-P. *B. campestris* var. BS-12; Q-T. *B. campestris* var. BS-14; U-X. *B. campestris* var. BS-15; A, E, I, M, Q, U: Plant morphology; B, J, N, R, V: Flowers; C, G, K, O, S, W: Pods; D, F, L, P, T, X: Seeds.

Although, the prophase chromosome of inquired varieties was categorized into three types found on orcein-staining properties.

The gradual staining nature of prophase chromosome categorized as 'Gradient type' in the classification of Tanaka (1971) that found in two varieties of *B. campestris* (Tori-7 and SS-75) and showed one darker

end to slightly stained end due to the appearance of comparative great amount of heterochromatins (Table 2, Figs. 4A-B). However, the interstitial area of prophase chromosome was observed to darkly stained meanwhile the other regions stained slightly in BS-6, BS-12, BS-15 and BS-10 (Table 2, Figs. 4C-D, 4F, 4I) and according to Tanaka (1971) theses were 'Interstitial type' of interphase nuclei.



Fig. 2. Habit of six varieties of *Brassica* L. A-D. *B. juncea* var. Dawlat; E-H. *B. juncea* var. Rai-5; I-L. *B. juncea* var. BS-10; M-P. *B. juncea* var. BS-11; Q-T. *B. napus* var. BS-7; U-X. *B. napus* var. BS-8; A, E, I, M, Q, U: Plant morphology; B, J, N, R, V: Flowers; C, G, K, O, S, W: Pods; D, F, L, P, T, X: Seeds.

In contrast, the prophase chromosomes of six varieties of inquired *Brassica* L. *viz. B. campestris* var. BS-14, three varieties of *B. juncea* (Dawlat, Rai-5 and BS-11) and two varieties of *B. napus* (BS-7 and BS-8), were considered as 'Continuous type' by

following Tanaka's classification (1971) where found to possess homogenous characteristics all over the inter-length after staining (Table 2, Figs. 4G-H, 4J, 4K-L) that means uniform distribution of heterochromatins.



Fig. 3. Orcein-stained mitotic interphase nuclei of twelve varieties of *Brassica* L. A. Tori-7; B. Sonali Sarisha-75; C. BARI Sarisha-6; D. BARI Sarisha-12; E. BARI Sarisha-14; F. BARI Sarisha-15; G. Dawlat; H. Rai-5; I. BARI Sarisha-10; J. BARI Sarisha-11; L. BARI Sarisha-7; K. BARI Sarisha-8; Arrows indicate the presence of nucleolus; Scale Bar= 10 µm.

Considering the nature of staining properties, three types of correlation could be remained within the orcein-stained interphase nuclei and prophase chromosomes (Tanaka, 1971). Firstly, the plant type which usually showed 'Diffuse type' of interphase nuclei had of staining in 'Continuous type' prophase chromosome and if 'Simple Chromocenter type' and 'Complex Chromocenter type' of interphase nuclei were formed by any organism, they could have 'Gradient type' and 'Interstitial type' of staining in prophase chromosome, respectively. *B. campestris* var. BS-14 and two varieties of *B. juncea* (Dawlat and BS-11) supported Tanaka's (1971) proposal as their 'Diffuse type' of interphase nuclei were found to possess 'Continuous type' of prophase chromosomes (Figs. 3E, 3G, 3J, 4E, 4G, 4J).



Fig. 4. Orcein-stained mitotic prophase chromosomes of twelve varieties of *Brassica* L. A. Tori-7; B. Sonali Sarisha-75; C. BARI Sarisha-6; D. BARI Sarisha-12; E. BARI Sarisha-14; F. BARI Sarisha-15; G. Dawlat; H. Rai-5; I. BARI Sarisha-10; J. BARI Sarisha-11; L. BARI Sarisha-7; K. BARI Sarisha-8; Scale Bar= 10 μm.

In case of *B. campestris* var. SS-75, the current findings assisted that abundance of localized heterochromatins in interphase nuclei and afterward occupied gradual and homogenous distribution in prophase chromosome which was as per assumption (Figs. 3B, 4B).

The homogenously distributed facultative heterochromatins of interphase nuclei further gradually distribution in the prophase chromosome of B. campestris var. Tori-7 (Figs. 3A, 4A). In the interphase nuclei of four varieties of Brassica L. viz. BS-6, BS-12, BS-15 and BS-10, vigorously arranged localized heterochromatins were found but in some way aggregated in interstitial area of the prophase chromosomes (Figs. 3C-D, 3F, 3I, 4C-D, 4F, 4I). Besides, the confined heterochromatins were showed in the interphase nuclei in B. juncea var. Rai-5 and two varieties of B. napus (BS-7 and BS-8) but the prophase chromosomes were homogeneously distributed (Figs. 3H, 3K-L, 4H, 4K-L). According to Sultana and Alam 2016, heterochromatin of interphase nuclei remained as condense status rather localized in a specific region that possibly diffused in the prophase chromosomes. So, the current detections did not assist the general directive concerning the distribution of heterochromatin in prophase chromosomes by the reason of facultative heterochromatins appearance.

Conclusion

It may be concluded that the present investigation correlated the nature of mitotic interphase nuclei and prophase chromosome of twelve BARI varieties of *Brassica* L. Moreover, this investigation might be a contribution to count the nature of mitotic interphase nuclei and prophase chromosome as a cytological tool and provided sufficient information to categorize as well as further investigation of investigated twelve varieties of *Brassica* L. from Bangladesh.

Acknowledgment

The authors are thankful to Bangladesh Agricultural Research Institute (BARI) to supply the seeds of twelve varieties of *Brassica* L. in addition Ministry of Science and Technology, Government of the People's Republic of Bangladesh for funding to carry out the present investigation.

The authors also expressed their gratitude towards Faria Akbar, Research associate, Department of Botany, Jagannath University for her co-operation in the conducting research.

References

Akbar F, Begum KN. 2020. A comparative anatomical investigation of three taxa of *Brassica* L. from Bangladesh. Bangladesh Journal of Plant Taxonomy **27(1)**, 15–26.

https://doi.org/10.3329/bjpt.v27i1.47566

Begum KN, Alam Sk S. 2016. Karyomorphological analysis with differential staining of nine *Cicer Arietinum* L. Varieties. Bangladesh Journal of Botany **45(2)**, 327-334.

Chen XJ, Chen ZJ, Du GX, Zhang MF. 2001. Electrophoretic comparative study on polypeptides of chloroplast from cytoplasmic male sterile lines and their maintainer lines on tuber mustards. Journal of Zhejiang University **27**, 88-95.

Cheng BF, Heneen WK, Chen BY. 1995. Mitotic karyotypes of *Brassica campestris* and *Brassica alboglabra* and identification of the *B. alboglabra* in addition line. Genome **38**, 313-319.

https://doi.org/10.1139/g95-039

Downey RK, Robbelen G. 1989. *Brassica* species: oil crops of the world, their breeding and utilization. New York, USA. McGraw Hill Publishing Co., 339-374.

Du W, Chen QX, Mo H. 1993. Study on somatic chromosome karyotypes of Xinjiang wild rape, *B. campestris*, *B. nigra* and *B. juncea*. Journal of August 1st Agricultural College **16(4)**, 26-31.

Fang P, Chen FB, Yao QL. 2014. Karyotype analysis on three types of leaf mustard (*Brassica*

juncea). Journal of Changjiang Vegetables **12**, 13-17. <u>https://doi.org/10.1139/g95-039</u>

Fukui K, Nagayama S, Ohmido N, Yoshiaki H, Yamabi M. 1998. Quantitative karyotyping of three diploid *Brassica* species by imaging methods and localization of 45S rDNA loci on the identified chromosomes. Theoretical and Applied Genetics **96**, 325-330.

Hu XM, Chen ZJ, Wang BL. 2001.Comparison on leaf ultrastructure in cytoplasmic male sterile line for tuber mustard (*Brassica juncea* var. tumida). Journal of Zhejiang University **27**, 535-540.

Jahan N, Bhuiyan SR, Talukder MZA, Alam MA, Parvin M. 2013. Genetic diversity analysis in *Brassica rapa* using morphological characters. Bangladesh Journal of Agricultural Research **38(1)**, 11–18.

https://doi.org/10.3329/bjar.v38i1.15185

Khan MR, Rashid H, Quraishi A. 2002. Effect of various growth regulators on callus formation and regeneration in *Brassica napus* cv. Oscar. Pakistan Journal of Biological Science **5**, 693-695.

https://scialert.net/abstract/?doi=pjbs.2002.693.695

Kulak S, Hasterok R, Maluszynska J. 2002. Karyotyping of *Brassica* amphidiploids using 5S and 25S rDNA as chromosome markers. Hereditas **136**, 144-150.

https://doi.org/10.1034/j.1601-5223.2002.1360209.x

Liu YH, Leng R, Zhang ZR. 2007. Study on genetic divergence of quantitative characters in tunloros stem mustard (*Brassica juncea* var. tumida Tsen and Lee). Chinese Agricultural Science Bulletin 23, 328-331.

Luciano FB, Holley RA. 2009. Enzymatic inhibition by Allyl-isothiocyanate and factors affecting its antimicrobial action against *Escherichia coli* O157:H7. International Journal of Food Microbiology **131(2-3)**, 240-245.

https://doi.org/10.1016/j.ijfoodmicro.2009.03.005

Mollika SR, Sarkar RH, Hoque MI. 2011. In *vitro* plant regeneration in *Brassica* spp. Plant Tissue Culture and Biotechology **21(2)**, 127-134. https://doi.org/10.3329/ptcb.v21i2.10235

Nagahara U. 1935. Genome analysis in *Brassica* with special reference to the experimental formation of *B. napus* and peculiar mode of fertilization. Japanese Journal of Botany 7, 389-452.

Olin-Fatih M. 1994. A new method for differential staining of *Brassica* Metaphase chromosomes and karyotypes of *B. campestris*, *B. oleracea* and *B. napus*. Hereditas **120**, 253-259. https://doi.org/10.1111/j.1601-5223.1994.00253.x

Paul M, Islam T, Hoque MI, Sarker RH. 2020. Analysis of genetic diversity in oilseed *Brassica* germplasm through ISSR markers and isozyme profiling. Bangladesh Journal of Botany **49(1)**, 147-158.

Rakow G. 2004. Species origin and economic importance of *Brassica*. In: Biotechnology in Agriculture and Forestry, Ed. E. C. Pua and C. J. Douglas, Berlin: Springer, 3–11.

Raymer PL. 2002. Canola: an emerging oilseed crop, in trends in new crops and new uses. In: Janick J & Whipkey A (eds). American Society for Horticultural Science, Vol. I. Alexandria, USA, ASHS Press, 122–126.

Ropiquet A, Gerbault-Seureau M, Deuve JL, Gilbert C, Pagacova E, Chai N, Rubes J, Hassanin A. 2008. Chromosomal evolution in the subtribe Bovina (Mammalia, Bovidae): The karyotype Cambodian of the banteng (Bos javanicus suggested birmanicus) that Robertsonian related translocation are to interspecific hybridization. Chromosome Research 16, 1107-1118. https://doi.org/10.1007/s10577-008-1262-2

Snowdon RJ. 2007. Cytogenetics and genome analysis in *Brassica* crops. Chromosome Research **15**, 85-95.

https://doi.org/10.1007/s10577-006-1105-y

Sultana SS, Alam Sk S. 2016. Karyomorphology of eleven varieties of *Gossypium hirsutum* L. Bangladesh Journal of Botany **45(1)**, 151-159.

Sun B, Xia X, Tian Y, Zhang F, Tang H. 2018.Karyotype analysis of *Brassica napus* cv. huayou no.Advances in Biological Sciences Research 5, 78-81.

Takamine N. 1916. Über die ruhenden und die präsynaptischen. Phasen der Reduktionsteilung. Botanical Magazine **30**, 293–303.

Tanaka R. 1971. Types of resting nuclei in Orchidaceae. Botanical Magazine Tokyo **84**, 118-1220.

https://doi.org/10.15281/jplantres1887.84.118

Wachsman MB, Ramirez JA, Galagovsky LR. 2012. Antiviral activity of Brassionsteroids derivatives against measles virus in cell cultures. Antiviral Chemical and Chemotherapy **13(1)**, 61–66. https://doi.org/10.1177/095632020201300105

Xiao CG, Guo XH, Han HB, Peng HJ. 2004. Identification of the pathogen for club root on

diseased stem mustard in Fuling, Chongqing and its major characteristics. Journal of Southwest Agricultural University **24**, 539-541.

Zhang MF. 1996. Tsatsai vegetable growth and flowering response to Paclobutrazol. Acta Agriculturae Zhejiangensis **8**, 110-112.

Zhang S, Chiang CY, Xie YF, Park SJ, Lu Y, Hu JW, Dostrovsky JO, Sessle BJ. 2006. Central sensitization in thalamic nociceptive neurons included by mustard oil application to rat molar tooth pulp. Neuroscience **142(3)**, 833-842.