

SHORT COMMUNICATION

OPEN ACCESS

Evaluation of allelopathic effects of *Calotropis procera* against wheat (*Triticum aestivum*) using seedling vigor and germination indices

Syed Amir Manzoor^{1*}, Muhammad Zubair², Syed Bilal Hussain², Sarwat N. Mirza¹, Rizwan Akhtar², Wasif Nouman²

¹Department of Forestry and Range Management, PMAS-Arid Agriculture University Rawalpindi, Pakistan

²Faculty of Agriculture Science and Technology, Bahauddin Zakariya University Multan, Pakistan

Article published on May 16, 2013

Key words: Calotropis procera, Triticum aestivum, Wheat, Germination Behavior, allelochemicals.

Abstract

Many plants are known for their phytotoxic potential. Such plants tend to suppress the germination and normal growth processes in other plants under their effect. *Calotropis procera* is one of the most dominant weed plants in Pakistan and is largely known for its insecticidal, pharmacological and phytotoxic properties. It is found abundantly around and inside the agricultural field crops. This study was aimed at investigating the allelepathic effects of *C. procera* on *Triticum aestivm* (wheat) which is one of the most important agricultural crops in most of the Asian countries. The allelopathic effects were gauged using different indices for germination behavior and seedling growth. Results indicated that *C. procera* leaf extracts has such allelopathic compounds which can significantly reduce the germination of and seedling vigor of *Triticum aestivum*.

*Corresponding Author: Syed Amir Manzoor 🖂 amir.kzd@gmail.com

J. Bio. & Env. Sci. 2013

Introduction

Apart from the fact that weeds compete with agricultural crops for light, nutrients, moisture and space, these unwanted plants also pose threat to the productivity and growth of the agricultural crops by release of certain allelochemicals in the environment (Kadioglue et al., 2005). There are a number of water soluble compounds in leaves, roots, stems and fruit of weed plants which when released into the soil, inhibit the germination and subsequent growth of crops plants (Batish et al., 2007). The phytotoxic nature of these allelochemicals is featured as selective (varies against different crops) and concentration dependant (more obvious at higher concentrations and vice versa) (Cheema et al., 2004). Within the subject these crops, allelochemicals can disturb a number of metabolic processes like photosynthesis, respiration and water uptake (Einhelling, 2002).

Calotropis procera, locally known as "Akk" in Pakistan (Yasin et al., 2012) is a member of family Asclepiadaceae (Parihar et al., 2011). C. procera has been reported as one of the dominant weeds in agricultural crops of Pakistan (Qureshi et al., 2009). Being abundant in tropical regions across the world, it is generally found along roads, sand dunes, fallow lands and on agricultural farmlands (Sastry and Kavathekar, 1990). C. procera has been studied by many a researchers for its anti-phytopathogenic and insecticidal properties (Begum et al., 2010 and Kareem et al., 2008). Studies conducted by different researchers (Yasin et al., 2012; Al-Zohrani and Al-Robai, 2007; Samreen et al., 2009) showed that leaf extract of C. procera has a number of water-soluble organic compounds which tend to inhibit the normal germination and growth performance of agricultural crop plants by suppressing the production of growth regulating hormones, checking the uptake of water and nutrients, decreasing the photosynthetic efficiency and affecting the seedling length of the subject plants.

Since wheat is one of the most important crops grown in most parts of the world including Pakistan, numerous studies have been carried out to assess the allelopathic potential of different weeds against wheat (Kadioglue *et al.*, 2005; Singh *et al.*, 2005; Mishra *et al.*, 2004). However, no significant study has yet been brought to light regarding the allelopathic effects of *C. procera* specifically on wheat crop.

Considering the fact that *C. procera* plants are found quite frequently in and around wheat field in Pakistan, it gives rise to serious question regarding the possible phytotoxic effects of this weed on wheat crop. Thus, keeping in view the susceptibility of wheat to different weeds, the current study was planned and carried out to evaluate the phytotoxic effects of *C. procera* on the germination behavior and seedling vigor of wheat plant (*Triticum aestivum*).

Material and methods

The experiment was conducted during December 2012 at room temperature (25°C±2) in the laboratory of Department of Forestry Wildlife and Range Management, Faculty of Agricultural Sciences and Technology, Bahauddin Zakariya University Multan, Pakistan.

Calotropis procera leaves were collected from *C. procera* plants growing in the cropped area of Agriculture Research Farm of Bahauddin Zakariya University Multan. The leaves were chopped down and boiled in tap water for 60 minutes in following ratios to make respective treatments.

- $T_2 = 50$ g *C. procera* leaves boiled in 1 L tap water for one hour,
- $T_3 = 100 \text{ g } C$.procera leaves boiled in 1 L tap water for one hour,
- $T_4 = 150$ g *C. procera* leaves boiled in 1 L tap water for one hour,
- T_5 = 200 g *C. procera* leaves boiled in 1 L tap water for one hour.

 $T_1 = Tap$ water,

After filtering through filtering through muslin cloth, the leaf extracts were collected in bottles and were tagged.

The experiment was carried out in Completely Randomized Design with five replications. Each replication had two soil filled plastic pots. The pots were placed on the laboratory table according to the procedure laid down by Steel and Torrie (1997). Ten seeds of wheat were sown in each pot. Different concentrations of *C. procera* leaf extracts were applied to respective pots to keep the soil moistened. The study was carried out for 21 days.

Germination Parameters

Following the standard procedure (AOSA, 1990), germinated seeds were counted and recorded after every 24 hours interval. A seed was considered as germinated when its radical become at least 2 mm in size. Ten days after sowing, the final germination percentage was calculated.

Following indices were used to evaluate the allelopathic effects of C. procera on germination behavior of wheat: Germination Energy (GE), Mean Germination Time (MGT), Final Germination Percentage (FGP), Relative Germination Rate (RGR) and Germination Index (GI) were determined using the formula as mentioned by Li (2008). Moreover, Days Required For 50% Germination of Total Seeds (T₅₀'), Days Required For 50% of Total Germinated Seeds (T₅₀), Speed of Germination (S), Speed of Accumulated Germination (AS), Coefficient of The Rate of Germination (CRG) were calculated using formulas as mentioned by Anjum and Rukhsana (2005). Germination Value (GV), Peak Value (PV), Percent Inhibition (PI) and Delay Index (DI) were determined using formulas as used by Malaviya and Anuradha (2011). Corrected Germination Rate Index was calculated using formula as used by Dewir et al., (2011) and Germination Distribution (GD) was determined following the mentioned by Schrader and Williams (2000). Besides seedling vigor index

was determined using the formula mentioned by Li *et al.,* (2008).

Statistical Analysis

The data was subjected to ANOVA technique and Least Significance difference test was applied to separate the treatment means as explained by Steel and Torrie (1997).

Results and discussion

Table 1 shows the allopathic effects of different concentrations of C. procera leaf extracts on Triticum aestivum (wheat) in terms of various germination and seedling vigor indices. Taking into consideration all the mentioned indices (in Table 1) depicting germination behavior it becomes clear that higher concentrations (T5) of C. procera leaf extract have significantly negative effect on the germination behavior of wheat crop indifferent of the index used. Interpreting the germination data through all of the germination indices, no concentration of treatment seemed to have enhanced the germination of wheat seeds while 100% concentration of leaf extracts appears most suppressing concentration in all cases. In fact, there are a number of compounds in C. procera leaf extracts which can significantly inhibit the germination of many agricultural crops. Moradshahi et al., (2003) concluded that higher concentration of allelochemicals in C. procera tend to inhibit the seed germination by checking the production of indole acetic acid and gibberellins in seeds. Al-Zahrani and Al-Robai (2007) also reported similar results in their stdudy carried out on C. procera leaves which suggested that C. procera had inhibitory effects on seed germination of agricultural crops. (Samreen et al., 2009, Hussain & Ilahi, 2009) have also reported similar results for allelopathic effects of various plants.

Seedling length and seedling vigor also decreased with increasing concentrations of leaf extract. Highest suppression of seedling length was recorded in treatment T_5 which was followed by T_4 and T_3 while there was no significant different between control and T_2 . In case of seedling vigor, value of index stood inversely proportional to the increase in leaf extract concentration. Root length seemed to have been more adversely affected by the allelopathic compounds present in *C. procera* leaf extract. This is possibly due to the fact that roots are first to encounter the allelochemicals in the soil. *C. procera* leaf extract has been widely reported to be a suppressing agent for seedling vigor of many crops (Oudhia and Tripathi, 1999). Allelopathic compounds like those of p-Menthane-3 and 8-transdiol are present in *C. procera* leaves and these compounds can inhibit the cell division and cell elongation process in the root and shoot apical, leading to a decrease in seedling length (Moradshahi *et al.*, 2003). Many other scientists (Samreen *et al.*, 2009, Hussain & Ilahi, 2009) have also reported similar results for studies carried out on allelopathic effects of *C. procera*.

Table 1. Allelopathic Effects of *Calotropis procera* on seed germination behavior and seedling vigor of *Triitcum* aestivum.

	MGT	T50	GI	T50'	FGP	CGRI
T1	7.42±.04(a)	4.79±.19(a)	12.6±.25(a)	5.15±.18(a)	91±1.87(a)	13.97±.39(a)
T2	7.7±.048(b)	5.48±.17(b)	10.9±.483(b)	5.78±.16(ab)	87±21(a)	12.66±.64(a)
T3	7.93±.031(b)	5.82±.05(b)	9.8±.234(c)	7.83±1.71(abc)	77±1.52(b)	12.83±.32(ab)
T4	8.07±.08(c)	5.591±.41(b	8.1±.25(d)	6.89±.7(bc)	73±1.5(b)	10.98±.35(ab)
T5	8.64±.12(d)	6.408±.12(c)	6.07±.13(e)	9.04±.15(c)	50±41(c)	12.387±11(b)
	RGR	DI	CRG	S	AS	GV
T1	-	-	13.50±.11(a)	9.12±.16(a)	14.0±.44(a)	4.1
T2	0.95±.021(a)	0.39±.07(a)	12.75±.07(ab)	7.51±.25(b)	10.4±.37(b)	2.66
T3	0.846±.02(b)	0.34±.08(b)	13.14±.13(bc)	7.74±.32(bc)	11.1±.61(bc)	2.63
T4	0.802±.05(b)	0.53±.09(b)	13.02±.19(cd)	6.57±.61(c)	9.2±11(c)	2.12
T5	0.549±.04(c)	0.86±.21(b)	12.61±.21(d)	$4.69 \pm .35(d)$	6.3±.534(d)	1.54
	PV	GE	RL	SL	SdL	SV
T1	2.2	65	9.99±.10(a)	18.37±.32(a)	28.3±.42(a)	2578.8±31.1(a)
T2	2.79	65	9.66±.11(a)	17.91±.38(a)	27.5±.41(a)	2395.6±27.1(b)
T3	1.75	55	8.30±.2(b)	15.22±.3(b)	23.5±.35(b)	1811.5±16(c)
T4	1.51	55	6.61±.3(c)	10.71±.5(c)	17.3±.7(c)	1265.5±60(d)
T5	1.31	45	3.68±.1(d)	7.584±.4(d)	10.2±.4(d)	500±58(e)

 $^{*}T_{1}$ = Control (Tap Water), $^{*}T_{2}$ = 25% *C. procera* leaf extract, $^{*}T_{3}$ = 50% *C. procera* leaf extract, $^{*}T_{4}$ = 75% *C. procera* leaf extract, $^{*}T_{5}$ = 100% *C. procera* leaf extract.

*T50'= Days Required for 50% Germination of total germinated seeds ,*T50= Days Required for 50% Germination of total seeds ,*DI= Delay Index ,*GI= Germination Index,*FGP= Final Germination Percentage ,*MGT= Mean Germination Time,*PV=Peak Value ,*S= Speed of Germination,*AS= Accumulated Speed of Germination ,*GE= Germination Energy ,*GV= Germination Value ,*CGRI= Corrected Germination Rate Index ,*RGR= Relative Germination Rate, CRG=Coefficient of Rate of Germination. *RL=Root Length, *SL= Shoot Length, *SdL= Seedling Length, *SV= Seedling Vigor.

Conclusion

Keeping in view the fact that *C. procera* can significantly suppress the germination activity in wheat seeds, it would be recommended to make sure that wheat fields should be kept free of *C. procera* plants. Presence of this weed plant in or around wheat fields can result in release of allelocheicals in the microenvironment of wheat field, especially with

the rain water, which in turn can reduce the crop yield to a great extent.

Acknowledgement

Authors are thankful to the administration and laboratory staff of department of forestry, wildlife and management and department of plant breeding and genetics, Faculty of Agricultural Sciences, Bahauddin Zakariya University Multan for their cooperation and kind support during this study.

References

Al-Zahrani HS, Al-Robai SA. 2007. Allelopathiceffect of Calotropis procera leaves extract on seedgermination of some plants. Journal of KingAbdulazizUniversity19,115-126.http://dx.doi.org/10.4197/Sci.19-1.9

Anjum T, Rukhsana B. 2005. Importance of germination indices in interpretation of allelochemical effect on seed germination. International Journal of Agriculture & Biology **7**, 417-419.

Association of Official Seed Analysts (AOSA). 1990. Rules for testing seeds. Journal of Seed Technology **12**, 1-112.

Batish DR, Lavanya K, Singh HP, Kohli RK. 2007. Root mediated allelopathic interference of Nettle-leaved Goosefoot *(Chenopodium murale)* on wheat *(Triticum aestivum)*. Journal of Agronomy and Cop Science **193**, 37-44. http://dx.doi.org/10.1111/j.1439-

<u>037X.2006.00243.x</u>

Begum N, Sharma B, Pandey RV. 2010. Evaluation of insecticidal efficacy of *Calotropis procera* and *Annona squamosa* ethanol extracts against *Musca domestica*. Journal of Biofertilizer and Biopesticide **1**, 1-6.

Cheema ZA, Khaliq A, Saeed S. 2004. Weed control in maize (*Zea mays L.*) through sorghum allelopathy Journal of Sustainable Agriculture **23**, 73-86.

Dewir YH, El-Mahrouk ME, Naido Y. 2011 Effect of some mechanical and chemical treatments on seed germination of Sabal palmetto and Thrinax morrisii palms. Australian Journal of Crop Science 5, 248-253. **Einhelling FA.** 2002. The physiology of allelochemical action: clues and views. In Reigosa MJ & Pedrol N (eds.) Allelopathy from Molecules to Ecosystem. Science Publishers, Enfield, New Hampshire, USA.

Kadioglue I, Yanar Y, Asav U. 2005. Allelopathic effects of weed leachates against seed germination of some plants. Journal of Environemtnal Biology **26**, 169-173.

Kareem SO, Akpan I, Ojo OP. 2008. Antimicrobial activities of *Calotropis procera* on selected pathogenic microorganisms. African Journal of Biomedical Research **11**, 105-110. http://dx.doi.org/10.4314/ajbr.v11i1.50674

Kayode J. 2004. Allelopathic effect of aqueous extracts of *Calotropis procera* on germination and seedling growth of maize. Pakistan Journal of Scientific and Industrial Research **47**, 69-72.

Li Y. 2008. Effect of salt stress on seed germination and seedling growth of three salinity plants. Pakistan Journal of Biological Sciences **11**, 1268-1272.

Malaviya P, Sharma A. 2011. Impact of distillery effluent on germination behaviour of *Brassica napus L.* Journal of Environmental Biology **32**, 91-94.

Mishra BP, Tripathi OP, Tripathi RS, Pandey HN. 2004. Effects of anthropogenic disturbance on plant diversity and community structure of a sacred grove in Meghalaya, northeast India. Biodiversity and Conservation **13**, 421-43.

Moradshahi A, Ghadiri H, Ebrahimikia F. 2003. Allelopathic effects of crude volatile oil and aqueous extracts of *Eucalyptus camaldulensis* Dehnh. Leaves on crops and weeds. Allelopathy Journal **12**, 189-195.

Oudhia P, Tripathi RS. 1999, Allelopathic research on rice seeds in chhattisgarh (India) region:

An overview. In: Abstract. National Seminar on Institute Industry Co-operation Programme for Developing Skills in Students of Seed Technology, Govt. Motilal Vigyan Mahavidyalaya, Bhopal(india)20-21 November 1999, 88-89.

Parihar G, Sharma A, Ghule S, Sharma P, Deshmukh P, Srivastava DN. 2011. Antiinflammatory effect of *Calotropis procera* root bark extract. Asian Journal of Pharmacy and Life Science 1, 29-44.

Qureshi R, Khan WA, Khan B. 2009. Study of vegetation and smooth coated otter in chotiari wetlands complex, sanghar, sindh, pakistan. Pak. J. Bot **41(5)**, 2507-2516.

Samreen U, Hussain F, Sher Z. 2009. Allelopathic potential of *Calotropis procera* (AIT.) AIT. Pakistan Journal of Plant Science **15**, 7-14.

Sastry CST, Kavathekar KY. 1990. Plants for reclamation of wasteland. Publication and Information Directorate, CSIR, New Delhi. 175-179 .p. Schrader JA, Willian RG. 2000. Seed germination and seedling growth of *Alnus maritima* from its three disjunct populations. Journal of the American Society For Horticultural Science **125**, 128–134.

Singh HP, Batish DR, Shalinder K, Kohli RK, Dogra KS. 2005. Allelopathic interference of *Ageratum conyzoides L.* against some crop plants. Weed management: balancing people, planet, profit 14th Australian Weeds Conference, Wagga Wagga, New South Wales, Australia, 6-9 September-2004. Papers and proceedings 558-561

Steel RGD, Torrie JH, Dicky D. 1997. Principles and Procedures of Statistics. Multiple comparison. 3rd Ed. McGraw Hill Book Co., New York, USA, 178-198.

Yasin M, Safdar ME, Iqbal Z, Ali A, Jabran K, Tanveer A. 2012. Pakistan Journal of Weed Science Research 18, 379-392.