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RESEARCH PAPER

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In-vitro assessment of anti-bacterial, anti-fungal and antioxidant potential of *Ficus carica* stems bark

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Abstract

Medicinal plants have been using for treatment of different diseases since ages. *Ficus carica* is a plant which is abundantly present and easily available in Pakistan. In this study methanolic extracts of *Ficus carica* stem bark was examined for anti-bacterial, anti-fungal and anti-oxidant potential. Anti-bacterial and anti-fungal potential was examined following disc diffusion method whereas anti-oxidant activity was determined using DPPH scavenging method. The maximum antibacterial activity was recorded 19 mm against *Klebsiella pneumonae* and *Escherichia coli*, and the highest anti-fungal potential was also recorded 19 mm against *Fusarium oxysporum* and *Mucor racemosus* each. The mean anti-oxidant activity was also recorded 63.45% in comparison to DPPH. The plant extracts was exhibited appropriate anti-oxidant potential and antimicrobial activity against various human pathogenic bacterial and fungal strains.

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Introduction

Even though a large number of anti-biotics have been produced synthetically in last few decades, but resistance to synthetic anti-microbial agents has also been increased (Aref et al., 2010; Nascimento, Locatelli, Freitas, & Silva, 2000). Anti-bacterial resistance has become a global concern; it has been increasing incidence of multiple resistance in human pathogenic microorganisms in recent years. Deaths have been reported worldwide, due to indiscriminate use of commercial antimicrobial drugs commonly employed in the treatment of infectious diseases (Parekh & Chanda, 2007). Commercially synthetic antibiotics development is highly expensive as compared to herbal medicines. The development of such drugs are declining as United States Food and Drug Administration (FDA) approval of new antibacterial agents have decreased by 56% over the past 20 years (Spellberg, Powers, Brass, Miller, & Edwards Jr, 2004). On the other hand the human as well as animals are using 250,000 to 500,000 plant species in daily life to resolve different microbial infections (Borris, 1996; Cowan, 1999). In this study Ficus carica is selected as one of medicinal plants, widely distributed from South Asia to Eastern Mediterranean and easily available in different areas of Pakistan. Traditionally it is used for healing various diseases. (Ahad et al., 2010; Dueñas, Pérez-Alonso, Santos-Buelga, & Escribano-Bailón, 2008). The stem bark of Ficus carica is beneficial as anti-oxidant, antifungal, anti-bacterial and anti-diabetic (Mopuri, Ganjayi, Meriga, Koorbanally, & Islam, 2017), antiinflammatory (Ramazani, Zakeri, Sardari, Khodakarim, & Djadidt, 2010) and anti-depressant (Badgujar, Patel, Bandivdekar, & Mahajan, 2014), anti-viral and anti-fungal effects (Lee & Cha, 2010). The objective of this study was to determine the of *F*. carica stem bark methanolic extracts and evaluate the anti-oxidant, anti-bacterial and anti-fungal potential.

Material and methods

Collection of plant materials

Ficus carica stem bark was collected from Canal rest house chichawatni district Sahiwal (Punjab). The plant was identified by Dr. Mansoor Hameed Associate prof. (Botanist). Shade dried plant materials were ground to fine powder and stored in air tight jar for further use.

Extraction of plant materials

400gm of sample was macerated in methanol for 7 days, filtered through whatman no.1 filter paper, the filtrates were then concentrated under reduced pressure using rotary evaporator at 45° C (Nebedum, Udeafor, & Okeke, 2010).

Anti-fungal activity

Sabouraud dextrose agar was sterilized in a flask and distributed into inoculated petri plates. Each fungal strain was inoculated in sabouraud dextrose plates. Sterile filter paper discs loaded with plant extracts and standard drug (clotrimazole) were placed on the top of sabouraud dextrose plates. The treated plates were kept at 4°C for 1-2 hour and then incubated for 24 hours at 28°C. Zone of inhibition was measured using vernier caliper in mm. Antifungal potential was founded as the mean diameter of zone of inhibition (mm) of plant extracts (Chaturvedi *et al.*, 2010; Shi *et al.*, 1996).

Anti-bacterial activity

Nutrient agar was sterilized in flask and distributed into petri plates. Each bacterial strain was inoculated in nutrient agar broth plates. Sterile filter paper discs loaded with methanolic extracts of fig stem bark and standard drug (gentamycin) were placed on the top of nutrient agar plates. The treated plates were kept at 4°C for 1-2 hour and then incubate for 24 hours at 37°C. Zone of inhibition were measured using vernier caliper in mm. Anti-bacterial potential was founded as the mean diameter of zone of inhibition (mm) of plant extracts (Shi *et al.*, 1996).

DPPH scavenging assay

The antioxidant activity of *Ficus carica* stem bark extracts was assessed by measuring their scavenging ability to 1, 1-diphenyl-2-picrylhydrazyl stable radicals (DPPH). The assay was performed as described by (Neves, Matos, Moutinho, Queiroz, & Gomes, 2009) with slight modifications. Methanolic extracts (10,20 and 30µl were added an equal volume in ethanolic solution of DPPH (0.1mM). After 30 minutes it was incubated at room temperature. Absorbance was recorded at 517nm. The experiment was repeated thrice. Butylated Hydroxytoluene (BHT) was used as standard control. Inhibition of free radical by DPPH was calculated using following way: I (%) = 100 x (A $_{blank}$ —A $_{sample}$ / A $_{blank}$)

 A_{blank} is the absorbance of the control reaction mixture excluding the test compounds, and A_{sample} is the absorbance of the test compounds.

Results

Anti-fungal

Results obtained, shows *Ficus carica* stem bark methanolic extracts have the maximum zone of inhibition (ZOI) against *Fusarium oxysporum*, *Mucor racemosus* 19 mm each, 18mm against *Mucor mucedo*, 17 mm against *Aspergillus flavus* and 16 mm against *Candida albicans* as indicated in Table 1. The lowest zone of inhibition was 14 mm recorded each against *Rhizopus stolonifer* and *Aspergillus nigar*.

Table 1. Anti-bacterial potential of methanolicextracts of *Ficus carica* stem bark.

Bacteria strain	Zone of Inhibition (mm)	
	Sample	Standard
Staphylococcus	16	25
aureus		
Klebsiella	19	23
pneumonae		
Pseudomonas	14	20
aeruginosa		
Salmonella typhi	18	25
Escherichia coli	19	22
Bacillus subtilis	17	25
Pseudomonas	14	25
fluorescens		
Staphylococcus	16	24
epidermidis		
Staph. Haemolyticus	13	20
MRSA	15	22

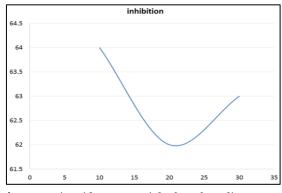


Fig. 1. Anti-Oxidant potential of methanolic extracts of *Ficus carica* stem bark.

whereas 18mm was recorded against *Staphylococcus typhi*, 17mm against *Bacillus subtilis*, 16mm against *Staphylococcus aureus*, shown in Table 2. The lowest zone of inhibition was recorded 13mm against *Staph*. *haemolyticus*.

Anti-bacterial

Table 2. Anti-fungal potential of methanolic extracts

 of *Ficus carica* stem bark.

Ficus carica stem bark methanolic extracts exhibit

maximum ZOI against Klebsiella pneumonae and

Escherichia coli i.e. 19mm each respectively

	Zone of Inhibition (mm)	
Fungi strain	Sample	Standard
Candida albicans	16	21
Fusarium	19	22
oxysporum		
Aspergillus nigar	14	20
Mucor mucedo	18	22
Mucor racemosus	19	22
Aspergillus flavus	17	21
Rhizopus stolonifer	14	20

Anti-oxidant activity

Ficus carica stem bark methanolic extracts were exhibited 63.45% anti-oxidant potential using DPPH scavenging assay method.

Discussion

Anti-fungal Activity

F. carica stem bark was used in traditional medicine for management of different fungal infections (Debib, Tir-Touil, Mothana, Meddah & Sonnet, 2014). *Candida albicans, Fusarium oxysporum, Aspergillus nigar, Mucor mucedo, Mucor racemosus, Aspergillus flavus* and *Rhizopus stolonifer* were used in my study. *Ficus carica* stem bark has an effective potential against.

These strains as compared to previous research work because it has no activity previously while in my study it has significant results. Anti-fungal potential of this species was also justified by the research work of (Oyelana *et al.*, 2011). A specie of same genus, *Ficus bubu* methanolic extracts of stem bark and leaves extracts were examined for anti-microbial and antifungal activity (Oyelana *et al.*, 2011).

Anti-bacterial activity

Plants having anti-bacterial activity are used against management of different disorders because of the

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presence of active phytochemicals (Joshi & Boyd, 2009). According to this study, *F. carica* stem bark has an effective potential against bacterial strains (*Staphylococcus aureus*, *Klebsiella pneumonae*, *Pseudomonas aeruginosa*, *Salmonella typhi*, *Escherichia coli*, *Bacillus subtilis*, *Pseudomonas fluorescens*, *Staphylococcus epidermidis* and *Staph. haemolyticus*). Previously by the research work of (Jeong, Kim & Cha, 2009), anti-bacterial activity of fig leaves methanolic extracts were evaluated with significant results using MIC assay (broth dilution method).

Anti-oxidant activity

Previous research work on *Ficus carica* stem bark was exhibited less anti-oxidant potential as compared to this analysis (Ao, Li, Elzaawely, Xuan & Tawata, 2008). Results were also justified by comparison of other species of same genus i.e. *Ficus racemosa* stem bark aqueous extracts being evaluated for antioxidant activity (Ahmed, Siddesha, Urooj & Vishwanath, 2010).

Conclusion:

Ficus carica is one of the oldest and nutritious known medicinal plant species in the world. Its history goes back to ancient times and people from different fields of life have been using its (leaves, fruits and bark) for both nutritious and medicinal purposes. Keeping in view the current review, fig bark can be useful in management of various disorders but recent scientific approaches are required for bio-assay guided isolation of different phytochemicals present in bark.

Conflict of Interest:

The authors has declared that there is no conflict of interest

References

Ahad H, Sreeramulu J, Kumar C, Anuradha C, Reddy K, Sushma K, Savithri R. 2010. Fabrication and in-vitro permeation studies of indomethacin-Ficus carica fruit mucilage patches. International Journal of Applied Biology and Pharmaceutical Technology **1(3)**, 786-792. Ahmed F, Siddesha JM, Urooj A, Vishwanath BS. 2010. Radical scavenging and angiotensin converting enzyme inhibitory activities of standardized extracts of *Ficus racemosa* stem bark. Phytotherapy research **24(12)**, 1839-1843.

Ao C, Li A, Elzaawely AA, Xuan TD, Tawata S. 2008. Evaluation of antioxidant and antibacterial activities of *Ficus microcarpa* L. fil. extract. Food control **19(10)**, 940-948.

Aref HL, Salah K, Chaumont JP, Fekih A, Aouni M, Said K. 2010. In vitro antimicrobial activity of four *Ficus carica* latex fractions against resistant human pathogens (antimicrobial activity of Ficus carica latex). Pak Journal Pharm Science **23(1)**, 53-58.

Badgujar SB, Patel VV, Bandivdekar AH, Mahajan RT. 2014. Traditional uses, phytochemistry and pharmacology of *Ficus carica:* A review. Pharmaceutical Biology **52(11)**, 1487-1503.

Borris RP. 1996. Natural products research: perspectives from a major pharmaceutical company. Journal of Ethnopharmacology **51(1-3)**, 29-38.

Chaturvedi V, Springer DJ, Behr MJ, Ramani R, Li X, Peck MK, Samsonoff WA. 2010. Morphological and molecular characterizations of psychrophilic fungus Geomyces destructans from New York bats with white nose syndrome (WNS). PLoS One **5(5)**, e10783.

Cowan MM. 1999. Plant products as antimicrobial agents. Clinical microbiology reviews **12(4)**, 564-582.

Debib A, Tir-Touil A, Mothana R, Meddah B, Sonnet P. 2014. Phenolic content, antioxidant and antimicrobial activities of two fruit varieties of Algerian *Ficus carica* L. Journal of food biochemistry **38(2)**, 207-215.

Dueñas M, Pérez-Alonso JJ, Santos-Buelga C, Escribano-Bailón T. 2008. Anthocyanin composition in fig (*Ficus carica* L.). Journal of Food Composition and Analysis **21(2)**, 107-115.

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Jeong MR, Kim HY, Cha JD. 2009. Antimicrobial activity of methanol extract from *Ficus carica* leaves against oral bacteria. Journal of Bacteriology and Virology **39(2)**, 97-102.

Joshi S, Boyd S. 2009. Sensor selection via convex optimization. Institute of Electrical and Electronics Engineers Transactions on Signal Processing 57(2), 451-462.

Lee YS, Cha JD. 2010. Synergistic antibacterial activity of Fig (*Ficus carica*) leaves extract against clinical isolates of methicillin-resistant Staphylococcus aureus. korean journal of microbiol biotechnology **38(4)**, 405-413.

Mopuri R, Ganjayi M, Meriga B, Koorbanally NA, Islam MS. 2017. The effects of *Ficus carica* on the activity of enzymes related to metabolic syndrome. Journal of Food and Drug Analysis.

Nascimento GG, Locatelli J, Freitas PC, Silva GL. 2000. Antibacterial activity of plant extracts and phytochemicals on antibiotic-resistant bacteria. Brazilian journal of microbiology 31(4), **247-256**.

Nebedum J, Udeafor P, Okeke C. 2010. Comparative effects of ethanolic extracts of Ficus carica and *Mucuna pruriens* leaves on haematological parameters in albino rats. Biokemistri **22(2)**. **Neves JM, Matos C, Moutinho C, Queiroz G, Gomes LR.** 2009. Ethnopharmacological notes about ancient uses of medicinal plants in Trás-os-Montes (northern of Portugal). Journal of Ethnopharmacology **124(2)**, 270-283.

Oyelana O, Durugbo E, Olukanni O, Ayodele E, Aikulola Z, Adewole A. 2011. Antimicrobial activity of *Ficus* leaf extracts on some fungal and bacterial pathogens of *Dioscorea rotundata* from Southwest Nigeria. Journal of Biological sciences **11(5)**, 359-366.

Parekh J, Chanda S. 2007. Antibacterial and phytochemical studies on twelve species of Indian medicinal plants. African Journal of Biomedical Research **10(2)**.

Ramazani A, Zakeri S, Sardari S, Khodakarim N, Djadidt ND. 2010. *In vitro* and *in vivo* antimalarial activity of *Boerhavia elegans* and *Solanum surattense*. Malaria Journal **9(1)**, 124.

Shi J, Ross CR, Chengappa M, Sylte MJ, McVey DS, Blecha F. 1996. Antibacterial activity of a synthetic peptide (PR-26) derived from PR-39, a proline-arginine-rich neutrophil antimicrobial peptide. Antimicrobial agents and chemotherapy **40(1)**, 115-121.

Spellberg B, Powers JH, Brass EP, Miller LG, Edwards Jr JE. 2004. Trends in antimicrobial drug development: implications for the future. Clinical Infectious Diseases **38(9)**, 1279-1286.