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RESEARCH PAPER

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Influence of foliar application of moringa leaf extract and humic acid on growth, yield and chemical composition of cucumber

Asad Ullah¹, Ata Ullah², Fazle Amin¹, Barkat Ali², Waqas Ahmad¹, Imtiaz Khan³, Rahmatullah Khan^{1*}, Fayaz khan¹

Department of Horticulture Faculty of Crop Production Sciences, The University of Agriculture Peshawar-Pakistan

²Department of Plant Breeding and Genetics, Faculty of Crop Production Sciences, the University of Agriculture Peshawar-Pakistan

³Department of Plant Protection, Faculty of Crop Protection Sciences, the University of Agriculture Peshawar-Pakistan

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Abstract

An experiment to study the influence of moringa leaf extracts (MLE) and humic acid (HA) foliar application on growth, yield and physico-chemical components of cucumber was carried out at the University of Agriculture Peshawar-Pakistan during 2016.Randomized complete block design with split plot arrangement having three replicates was used. Different levels of MLE (control, 30, 40 and 50gL⁻¹) and HA concentrations (Control, 0.5, 1 and 1.5%) were applied for foliar application. Data were recorded on various yield and quality attributes. Statistical analysis showed that MLE and HA affected almost all the studied attributes, while the interaction was found non-significant. Results indicated that foliar application of MLE at the rate of 50 gL⁻¹ gave maximum vine length (164.75 cm), single fruit weight (172.25 g), fruit diameter (4.31 cm), total yield (40.93 tons ha⁻¹),TSS (2.91°Brix), titratable acidity (0.49%) and ascorbic acid content (5.79 mg 100g⁻¹) with minimum days to first harvest (54.08) and fruit juice pH (5.36). whereas HA concentration, 1.5 %showed best performance in all the studied attributes as compared to other treatments. Maximum vine length (166.08 cm), single fruit weight (171.50 g), fruit diameter (4.37 cm), total yield (41.84 tons ha⁻¹), TSS (2.83 °Brix), titratable acidity (0.48%), and ascorbic acid (6.00 mg 100g⁻¹) with minimum days to first harvest (54.00) and fruit juice pH (5.27) were recorded using HA at the rate of 1.5%. It was concluded that 50 gL⁻¹ MLE improved yield and chemical composition of cucumber, while 1.5% HA also showed best results.

^{*}Corresponding Author: Rahmatullah Khan ⊠ rahmatullah274@aup.edu.pk

Introduction

Cucumber (*Cucumis sativus* L.) is one of the most important and popular vegetable crops belonging to the family Cucurbitaceae. Cucumber is the main source of vitamins A, B6, C and minerals like Magnesium, Potassium and Copper (Vimala *et al.*, 1999). Throughout the world, China is the major producer of cucumber with estimated production of 54.34 million tons. About 73% of cucumber is produced by China. In Pakistan, the total area under cultivation is 23,268 hectare with production of 52,765 tones, with average production of 2.25 tones ha⁻¹ (MINNFSR, 2015). Generally, a balanced supply of nutrients is essential for optimum yield and fruit quality (Akhtar *et al.*, 2010).

Foliar spraying is a new method for crop feeding which micro and macro nutrients in form of liquid are used into leaves (Nasiri et al., 2010). Humic acid (HA) is one of the commercial products of an organic fertilizer which contains 44-58% C, 42-46% O, 6-8% H and 0.5-4% N as well as many other elements (Larcher, 2003). HA is a promising natural resource that can be used as an alternative to synthetic fertilizers to increase crop production. It exerts either a direct effect, such as on enzymatic activities and membrane permeability, or an indirect effect, mainly by changing the soil structure (Biondi et al., 1994). Furthermore, HA have the ability to increase the nitrogen use efficiency and which results to enhance the growth and development of the crop. Along with natural sources of PGRs and number of antioxidants, nowadays Moringa oleifera is receiving lot of interest (Foidle et al., 2001). Foliar spray of MLE accelerates plant growth and increase yield of crops (Marcu, 2005). Different substances like vitamin C as well as vitamin A, iron, beta-carotene, riboflavin, phenolic acid and calcium are present in the leaves of Moringa (Nambiaret al., 2005). The oil have antioxidant activity contained by the leaves (Njoku and Adikwu, 1997). Similarly zeatin, which is a purine adenine derivative from the group of plant hormone, cytokinin, found in abundant in MLE (MLE) which is recognized for stay green and stress tolerance capabilities. (Barciszweski et al., 2000).

Due to high cost and skill labor in application of inorganic fertilizations and land pollution, harmful nature linked with use of inorganic fertilizer and contribution to climate change that has adverse effect on soil ecosystem, there is a need to examine for alternative sources for plant supplements. Such plants can be promoted as a multipurpose that may develop better, safer and cheaper way of increasing the yield of stable vegetable crops for healthy nourishment (Hussein and Abbaro, 1997) Application of MLE on to leaves of tomato, bell pepper, maize, soya bean, melon, chili and sorghum resulted in increase in their yields (Fuglie, 2000). Objectives of this study were to study the effects of MLE and HA on growth yield and physico-chemical components of cucumber.

Materials and methods

Experimental site

An experiment to study "Influence of foliar application of MLE and HA on growth, yield and chemical composition of cucumber" was conducted at Horticulture Research Farm and Post-harvest Laboratory, The University of Agriculture Peshawar during 2016.

Design of experiment

The research was worked out in Randomized Complete Block Design (RCBD) with two factorial split plot arrangement. HA concentrations along with control were allotted to main plot with four levels (0, 0.5, 1 and 1.5 %) while, MLE along with control were allotted to sub plot with four levels (0, 30, 40 and 50 %) having three replications were used. HA and MLEs was applied in liquid form as foliar spray to the cucumber plants.

Treatments

The total numbers of treatments were 48. The detail of the factors is given in table 1.

Preparation of moringa leaf extract (MLE)

MLE was prepared by collecting young and disease free leaves from moringa tree. These leaves were washed and then leaves were grinded in a grinder to extract the leaf juice. In order to remove all the green matters juice was filtered by passing through a muslin cloth. An amount of 30g, 40g and 50g of young moringa leaf juice were mixed in one liter of water to prepare solution of different concentrations like 30, 40 and 50 gL⁻¹ as experimental treatments.

Field preparation and sowing techniques

Before seed sowing, field was turned over, harrowed and leveled. Cultivar Poinsett 76 seeds were sown at 30 cm plant to plant and 1 meter row to row distance on ridges. Two to three seeds were sown at a space of 2.5 cm. When seedlings were germinated, thinning was carried out leaving only one healthy plant. Agronomic practices were applied such as weeding, watering and hoeing, while, insecticide and pesticides were sprayed.

Stage of foliar application

The crop was sprayed 30 days after seed sowing.

Studied attributes

Vine length (cm): From each treatment five plants were randomly selected and through measuring tape their height was measured from bottom to the top of the plant. Their mean was computed.

Days to 1st harvesting: From sowing to 1st appearance of flowering in all treatments of each replication, the days were counted and then average was computed. Fruit diameter (cm): The fruit diameter of the selected fruits were measured with the help of digital vernier caliper and their mean was taken.

Single Fruit weight (g): Mean was computed by weighing the randomly picked fruit from plants of each treatment of all replications.

Total yield (tons ha⁻¹): The total productivity was recorded by the following formulae:

$$Yield tons ha^{-1} = \frac{Yield per sub plot (kg)}{Area of sub plot (m^2) \times 1000} \times 10000 (m^2)$$

Total soluble solids (°Brix): Total soluble solids were analyzed by using hand held refractometer. Few drops of juice were taken on the prism and reading was recorded and the proposed method given in AOAC (2000) was applied. After every reading slide and prism of the instrument was cleaned.

Percent Titratable Acidity: The percent titratible acidity determined by standard method AOAC (2000) and calculated by following formula.

Percent titratable acidity (%) =
$$\frac{N \times T \times F \times 100}{D \times S} \times 100$$

Where

N = Normality of NaOH

T = ml of 0.1 N NaOH used

F = Constant acid factor 0.0067 (citric acid)

D = ml of sample taken of tomato juice.

S = ml of diluted sample taken for titration

Ascorbic acid content (mg 100g⁻¹): "Dye method" was used to find ascorbic acid content. Fruit juice was extracted from the selected fruit samples at the rate of 10 ml and poured into the volumetric flask, then oxalic acid was supplemented to move up the volume to 100 ml. By this 10% solution was made. After that 10 ml solution was picked and titrated with Dye. The observance of pink color indicates the completion of reaction. Furthermore, the content of Vitamin C will be calculated through the given formula:

Ascorbic acid content (mg/ 100g) =
$$\frac{F \times T \times 10}{D \times S} \times 100$$

Where

T = Amount of Dye solution consumed from burette (ml)

F = Constant Factor of Dye

S = Fruit juice (g) taken for dilution

D = Amount of diluted sample used for titration (ml)

Fruit juice pH: Fruit juice pH was measured from the juice of the fruit with the help of pH meter by dipping the probe of the meter into the juice until the reading stop and then the reading was noted.

Statistical analysis

All the data collected was analyzed by using Randomized Complete Block Design. In case the data was found significant. Least significant difference was

applied for mean comparison. A statistical package (statistix8.1) was used for analyzing the data (Jan *et al.*, 2009).

Results

Plant growth and yield attributes

Table 1. Details of factors.

The foliar spray of MLE at the rate of 50 gL⁻¹ gave maximum vine length(164.75 cm), fruit weight (172.25 g), fruit diameter (4.31 cm) and highest fruit yield (40.93 tons ha⁻¹) with minimum days to first harvest (54.08) as shown in Table 2.

Factor A: (Moringa leaf extract gL ⁻¹)	Factor B: (Humic acid %)		
$C_0 = Control$	$H_0 = 0$		
$C_1 = 30 \text{ gL}^{-1}$	$H_1 = 0.5$		
$C_2 = 40 \text{ gL}^{-1}$	$H_2 = 1$		
C ₃ = 50 gL ⁻¹	H ₃ =1.5		

In case of HA, the maximum vine length(166.08 cm), fruit weight (171.50 g), fruit diameter (4.37cm) and total yield (41.84 tons ha⁻¹) with minimum days to first harvest (54.00) were recorded in treated plant with foliar spray of 1.5% HA (Table 2).

Physico-chemical components

The foliar spray of MLE@ 50 gL⁻¹gave maximum total soluble solids (2.91 °Brix), titratable acidity (0.49%) and ascorbic acid content (5.79 mg 100g⁻¹) with minimum fruit juice pH (5.36)as shown in Table 3.

Table 2. Influence of moringa leaf extract (MLE) and humic acid (HA) on plant growth and yield of cucumber.

Treatments	Characters							
Moringa leaf extracts (gL-1)	Vine length (cm)	Days to 1st	harvest	Fruit weight (g)	Fruit diameter (cm)	Total Yield (tons ha-1)		
Control	156.08C	62.67A		6.08C 62.67A		161.92C	3.25C	30.70C
30	160.17B	58.42B		165.33BC	3.75B	34.08BC		
40	40 160.42B		57.83BC		3.79B	36.07B		
50	164.75A	54.08C		172.25A	4.31A	40.93A		
LSD at 1%	3.33	3.99		4.65	0.49	4.42		
Humicacid (%)								
Control	Control 154.75c 62.42a		62.42a		3.18c	29.43c		
0.5	160.08b	58.50b		166.17bc	3.75b	35.14bc		
1	1 160.50b 58.08b		58.08b		3.79b	35.37b		
1.5	166.08a	54.00c		166.08a 54.00c		171.50a	4.37a	41.84a
LSD at 5%	5.20	3.82		4.42	0.54	5.75		

Mean values followed by different alphabets are statistically different using LSD test at 1 and 5% level of significance.

In case of HA, the increasing trend was recorded in total soluble solid (2.83 °Brix), titratable acidity(0.48%) and ascorbic acid(6.00 mg 100g⁻¹) with minimum fruit juice pH (5.27) in plants treated with 1.5% HA(Table 3).

Discussion

Vine length (cm)

The analysis of variance indicated that vine length of

cucumber plant was significantly affected by MLEs and HA concentrations. The increment in vine length might be due to the presence of macro and micro nutrients in MLE that positively increased vegetative and reproductive growth of plant. Similar results were also observed by Kato *et al.*, (2002). HA is a bio stimulant that stimulates the growth of the plant. It affects the morphological characteristics of plant particularly the vine length is directly related to the

concentration of HA. Sani (2013) who reported that when cucumber plants were sprayed at 15 days interval with HA then the morphological characters including vine length, have shown positive response.

Days to 1st harvest

The number of days to fruiting decreases with foliar application of MLEs as it increases nutrient absorption and nutrients supply that encourage more synthesis of assimilates and thus produce early flowering and fruiting. Our finding are supported by Nasir *et al.*, (2016) who claimed that kinnow mandarin treated with MLE produced early fruiting.

Plants utilize the foliar application of nutrients more efficiently as compared to soil application. Due to soil application leaching, non-availability of nutrient results in poor growth and ultimately the plant produce late flowers after the completion of its vegetative growth leads to delay harvesting.

Application of HA as foliar spray increases auxin concentration when plants absorb HA that results in early completion of its growth, flowering and hence early fruiting. Kazemi (2014) who reported that the application of HA and calcium chloride as foliar spray resulted in early fruiting.

Table 3. Influence of moringa leaf extract (MLE) and humic acid (MLE) on physico-chemical components of cucumber.

Treatments	Characters					
Moringaleaf extract (gL-1)	TSS (ºBrix)	Fruit juice pH	Titratable	Acidity (%)	Ascorbic acid (mg 100g ⁻¹)	
Control	2.22C	6.20A	0.39C		3.72C	
30	2.55B	5.77B	0.40BC		4.91B	
40	2.56B	5.71B	0.43B		4.92B	
50	2.91A	5.36C	0.49A		5.79A	
LSD at 1%	0.21	0.32	0.03		0.85	
Humic acid (%)						
Control	2.32c	6.29a	0.38c		3.63c	
0.5	2.53bc	5.76b	0.42bc		4.80b	
1	2.57b	5.72bc	0.43b		4.90b	
1.5	2.83a	5.27c	o.48a		6.00a	
LSD at 5%	0.24	0.46	0.04		1.08	

Mean values followed by different alphabets are statistically different using LSD test at 1 and 5% level of significance.

Fruit weight (g)

The increased in fruit weight might be due to the excessive level of potassium and zinc in MLE. Fruit quality is increased through potassium by promoting the translocation of carbohydrates from source to sink (Ramezani and Shekafandeh, 2011). Furthermore, Yasmeen (2011) found that spraying wheat, peas and tomato with MLE at 3.5% increased fruit weight. HA has been proven to promote the quantitative and qualitative characteristics of the fruit. Maximum single fruit weight was found at highest concentration of HA because it help in freeing up the nutrient in the soil due to which more nutrient are available for plant and help to convert the element into suitable plant

assimilates for their better fruit growth and yield. Similar result regarding availability of the nutrient for the fruit growth and yield was reported by Mahmoudi *et al.* (2013).

Fruit diameter (cm)

The increased in fruit diameter might be due to the high content of cytokinin in MLE which plays good role in enhancing cell division and expansion which results in increased fruit diameter (Sheren *et al.*, 2015). As MLE is a good source of ascorbic acid, the auxinic action of ascorbic acid along with its linkage in cell wall metabolism and cell expansion leads to big fruits (Samirnoff, 1996). The maximum fruit diameter

may be due to the direct involvement of the nutrient at growth and developmental stages of the fruit. Yildirim (2007) reported that foliar application of HA improved the fruit diameter of cucumber. Vasilenko (2002) reported that due to foliar application of HA the fruit diameter increased from 16-17% compared to control.

Total yield (tons ha-1)

Similar results were observed by Fuglie (2000) who stated that moringa leaves extract improve growth of young plants, increase number of roots, support plants, increase resistance to diseases and pests, produce more and superior fruits and generally increase yield by 20 to 30%. HA enable the plant to uptake more and more nutrients from the soil by making these nutrients in readily available form indirectly. HA improved the fruit weight and yield in term of quantity and quality as reported by Mahmoudi *et al.* (2013).

Total soluble solid (oBrix)

This enhancement in the amount of soluble solid contents might be due to the high sugar and starch contents in aqueous solution of MLE. Furthermore, moringa leaf aqueous extract also contain vital amount of cytokinins. Cytokinins helps in the metabolism of carbohydrate and form new source to sink relationship which in term increased the fruit total soluble solids (Dyer et al., 1990). These results are similar with the findings of Sheren et al. (2015), who reported that foliar application of MLE increases the total soluble solids in 'Le Conte' pear. This favourable nutritional status, induced by foliar applications of HA could be the indirect cause of the accumulation of sugar in fruit. These findings are in line with those findings of Neri et al. (2002) who reported that increased of TSS in strawberry was due to the foliar application of HA. The increase in total sugar in response to HA might be due to formation of maximum amount of carbohydrate within the leaf and fruit tissues, which than converted to the specific sugar like glucose and sucrose (Abbas et al., 2013). Balibrea et al., have reported that an increase of TSS in tomato fruits may depend on a higher sugar import

and accumulation.

Percent Titratable Acidity

The increased in the total acidity of cucumber fruit juice might be due to the fact that MLE is a good source of certain acids like ascorbic acid, which is directly involved in the rising of total acidity at final harvest, moreover certain nutrients such potassium also directly involved in the acidity of fruit juice. Higher the potassium, higher will be the juice acidity and lower the potassium, lower will be the juice acidity (Zekri and Obreza, 2009). Plant keep their C:N ratio constant, but due to application of humic the cell utilized extra carbon to make organic acid such as malic acid and citric acid that result in maximum acidity of the fruit (Toor et al.,2006). Similar result also reported by Dogan and Demir (2004) in tomato increased titratable acidity found by the addition of HA aggregate culture in green house condition.

Ascorbic acid content (mg 100g-1)

The increased amount of ascorbic acid in cucumber fruit might be due to the high amount of ascorbate in MLE. The exogenous application of ascorbate in the solution form of MLE trigger the production of internal ascorbic acid, which leads to the high quantity of ascorbic acid in the fruits of treated plants (Mengel, 1997). Nasira *et al.* (2016) also found that foliar application of moringa leaf aqueous extract increased vitamin C of 'Kinnow' mandarin. The application of HA increased the ascorbic acid content in cucumber fruit. It might be due to HA increase the permeability of bio membranes for electrolytes accounted for increased uptake of phosphorus and potassium which increase the ascorbic acid percentage of the fruit (Yildrim, 2007).

Fruit juice pH

The decreased in the juice pH of cucumber fruit might be due to the high content of potassium. Higher the potassium application, higher will be the juice acidity and lower the application of potassium, the lower will be the juice acidity (Zekri and Obreza, 2009). Ashraf *et al.* (2012), who reported that foliar application of

MLE on 'Kinnow' mandarine decreased the acidity of fruit juice. Increase the concentration of HA the pH will decrease due to the acidic nature of the chemical also the fruit produce by that plant will be acidic and the juice extracted from that fruit will be also acidic in nature. Kazemi (2014) reported that significant decrease in pH of tomato crop treated with HA as compare to control.

Conclusion

The results showed that the application of MLE (50 gL⁻¹) with HA concentrations significantly effected all the growth, yield and quality attributes. The MLE (50 gL⁻¹) enhanced vine length, fruit weight, fruit diameter, Total yield (tons ha⁻¹), total soluble solids, titratable acidity and ascorbic acid content. Application of HA at 1.5 % improved the growth, yield and quality attributes of cucumber. The interactive effect of MLE and HA application for all the studied attributes were found non-significant.

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References

Abbas T, Ahmad S, Ashraf M, Adnan M, Yasin M, Balal RM. 2013. Effect of humic and application at different growth stages of Kinnow mandarin (citrus reticulateblanco) on the basis of physio-biochemical and reproductive responses. Academia Journal of Biotechnology **1(1)**, 014-020.

http://dx.doi.org/10.15413/ajb.2012.0106

Abdel FH, Boshra G, Sayed AE, Shahin SM. 2008. The role of humic acid in reducing the harmful effect of irrigation with saline water on tifway turf. Journal of Biological Chemistry and Environmental Sciences **3(1)**, 75-89.

Akhtar ME, Khan MZ, Rashis MT, Ahsan Z, Ahmad S. 2010. Effect of potash application on yield and quality of tomato (Lycopersicon esculentum

Mill.). Pakistan Journal of Botany 42, 1695-1702.

Alloway BJ. 2004. Zinc in Soils and Crop Nutrition. International Zinc Association Brussels, Belgium (accessed 11.04.14.). Website, http://www.zinc-crops.org/

Ashraf MY, Yaqub M, Akhtar J, Khan MA, Ebert G. 2012. Control of excessive fruit drop and improvement in yield and juice quality of Kinnow (Citrus deliciosa × Citrus nobilis) through nutrient management. Pakistan Journal of Botany 44, 259–265.

Azra Y. 2011. Exploring the Potential of moringa (Moringaoleifera) Leaf Extract as Natural Plant Growth Enhancer. Ph.D. Faculty of Agriculture, University of Agriculture, Faisalabad, Pakistan.

Balibrea MA, Martínez-Andújar C, Cuartero J, Bolarín MC, Pérez-Alfocea F. 2006. The high fruit soluble sugar content in wild Lycopersicon species and their hybrids with cultivars depends on sucrose import during ripening rather than on sucrose metabolism. Functional Plant Biology 33, 279-288.

http://dx.doi.org/10.1071/FP05134

Barciszweski J, Siboska G, Rattan SIS, Clark BFC. 2000. Occurrence, biosynthesis and properties of kinetin (N6-furfuryladenine). Plant Growth Regulation **32**, 257-265.

Biondi FA, Figholia A, Indiati R, Izza C. 1994. Effects of fertilization with humic acids on soil and plant metabolism: a multidisciplinary approach. NoteIII: phosphorus dynamics and behaviour of some plant enzymatic activities. In humic Substances in the Global Environment and Implications on Human Health, ed. Senesi N and Miano TM. Elsevier, New York, p 239-244.

Carvajal M, Martinez-Sanchez F, Alcaraz CF. 1995. Improvement of fruit colour quality of paprika

combined treatments of Ti (IV) and humic acids. ActaAlimentaria **24**, 321-329.

Dogan E, Demir K. 2004. Determination of yield and fruit characteristics of tomato crop grown in humic acids-added aggregate culture in greenhouse condition. VI. National Vegetable Symposium 21-24 September, Canakkale, Turkey, 218-224.

Dyer D, Cotterman D, Kerr PS, Carlson DR. 1990. Cytokinins as metabolic stimulants which induce pod set. Plant Growth Substances Springer Verlag, p 67-70.

El-Hamied SAA, El-Amary EI. 2015. Improving growth and productivity of pear trees using some natural plants extracts under north Sinai conditions. IOSR Journal of Agriculture and Veterinary Science **8**, 01–09.

El-Razek EA, Abd-Allah ASE, Saleh MMS. 2012. Yield and fruit quality of 'Florida Prince' peach trees as affected by foliar and soil application of humic acid. Journal of Applied Sciences Research 8(12), 5724-5729.

Foidle N, Makkar HPS, Becker K. 2001. The potential of Moringa oleifera for agricultural and industrial uses. In: L.J. Fugile (ED.), the miracle tree: the multipurpose attributes of Moringa. CTA publications Wageningen, The Netherlands, p 45–76.

Fuglie LJ. 2000. New uses of moringa studied in Nicaragua: ECHO's Techincal Network Sitenetworking global hunger solution ECHO, Nicaragua. Gaborone. Botswana.

Hafez OM, El-Metwally IM. 2007. Efficiency of zinc and potassium spray alone or in combination with some weed control treatments on weed growth, yield and fruit quality of Washington Navel oranges. Journal of Applied Sciences Research. Egypt. **3,** 613-621.

Helgi O, Rolfe SA. 2005. The Physiology of

Flowering Plants. 4thEdn. Cambridge University Press, Cambridge UK., p 392.

Hussein SG, Abbaro MB. 1997. The influence of prunings of Leucaena leucocephala and Eucalyptus microtheca used as green manure mulch on growth and yield of fodder sorghum (Sorghum bicolor L.) University of Khartoum Journal of Agricultural Science **5**, 106-114.

Jan MT, Shah, Hoolinton PA, Khan MJ, Sohail Q. 2009. Agriculture research: Design and analysis. Dept. Agronomy, KPK Agriculture University Peshawar, Pakistan.

Jason P. 2013. Pepe's fruit trees, response of sweet bell pepper to moringa leaf extract and organo-bio degradable fertilizer. Asian Journal of Agriculture and Biology **3(4)**, 117-123.

Jyotsna V, Srivastava AK. 1998, Physiological basis of salt stress resistance in pigeon pea (Cajanuscajan L.)-II, Pre-sowing seed soaking treatment in regulating early seedling metabolism during seed germination. Plant Physiology and Biochemistry **25**, 89-94.

Kato C, Kato H, Asami T, Yoshida TH, Noda H, Kamada Satoh S. 2002. Involvement of xylem sap zeatin-O-glucoside in cucumber shoot greening plant physiology and Biochemistry **40(11)**, 949-995.

KazemiM. 2014. Effect of foliar application of humic acid and calcium chloride on tomato growth. Bulletin of Environment, Pharmacology Life Sciences **3(3)**, 41-46.

Khan AS, Nasir M, Malik AU, Basra SMA, Jaskani MJ. 2015. Combined application of boron and zinc influence the leaf mineral status, growth, productivity and fruit quality of 'Kinnow' mandarin (Citrus nobilisLour × Citrus deliciosa Tenora). Journal of Plant Nutrition 38, 821–838.

Larcher W. 2003. Physiological plant ecology:

Ecophysiology and stress physiology functional groups, 4th Edition, Springer, New York.

Mahmoudi M, Samavat S, Mostafavi M, Khalighi A, Cherati A. 2013. The effects of proline and humic acid on quantitative properties of kiwi fruit. International Research Journal of Applied and Basic Sciences 6(8), 1117-1119.

Marcu MG. 2005. Miracle tree. K.O.S. Health publication, Canada. p. 108-115.

Mengel K. 1997. Impact of potassium on crop yield and quality with regard to economic and ecological aspects. International Potash Institute, Bornova, Turkey, p 157-174.

MINNFSR. 2015. Fruits, vegetables, and condiments statistics of Pakistan, Ministry of National Food Security and Research. Economic wing, Islamabad, p 21-22.

Nagar PK, Leyer RI, Sircar PK. 2006. Cytokinins in developing of pear Gaertn. Physiologia Plantarum **55**, 45-50.

Nambiar VS, Mehta R, Daniel M. 2005. Polyphenol content of three Indian green leafy vegetables. Journal of Food Science and Technology **42(6)**, 312-315.

Nasira M, Khan AS, Basra SMA, Malik AU. 2016. Foliar application of moringa leaf extract, potassium and zinc influence on yield and fruit quality of 'kinnow' mandarin. Scientia Horticulturae. 210, 227-235.

https://doi.org/10.1016/j.scienta.2016.07.032

Nasiri Y, Zehtab-Salmasi S, Nasrullahzadeh S, Najafi N, Ghassemi-Golezani K. 2010. Effects of foliar application of micronutrients (Fe and Zn) on flower yield and essential oil of chamomile (Matricaria chamomilla L.). Journal of Medicinal Plants Research. 4(17), 1733-1737.

http://dx.doi.org/10.5897/JMPR10.083

Neri EM, Lodolini G, Savini P, Sabbatini G, Zucconi F. 2002. Foliar application of humic acids on Strawberry cv. Onda. Acta Horticulturae **594**, 297-302.

http://dx.doi.org/10.17660/ActaHortic.2002.594.35

Njoku OU, Adikwu MU. 1997. Investigation on some physico-chemical antioxidant and toxicological properties of Moringa oleifera seed oil. Acta Pharmaceutica **47(4)**, 287-290.

Prabhu M, Kumar AR, Rajamani K. 2010. Influence of different organic substances on growth and herb yield of Orange (Washington Novel). Indian Journal of Agricultural Research **44**, 48–52.

Prince ML. 2000. The moringa tree. ECHO, 17391 Durrance Rd., North Ft. Myers FL 33917, USA., p 78-92.

Ramezani S, Shekafandeh A. 2011. Influence of Zn and K sprays on fruitand pulp growth in olive (Olea europaea L. Cv. 'Amyg dalifolia'). Iran Agricultural Research 30, 1-10.

http://dx.doi.org/10.22099/iar.2012.489

Reuther W. 1973. The citrus industry. University of California, Division of Agricultural Science., U. S. A.

Sani B. 2013. Foliar applicator of humic acid on plant height in canola. APCBEE. **8(1)**, 82-86.

Sheren A, El-Hamied A, El-Amary I. 2015. Improving growth and productivity of "Pear" trees using some natural plants extracts under North Sinai conditions. Journal of Agriculture and Veterinary Science ISSN: 2319-2380, p-ISSN: 2319-2372.

Smirnoff N. 1996. The function and metabolism of ascorbic acid in plants. Annals of Botany **78**, 661–669.

http://dx.doi.org/10.1093/aob/mcg079

Taiz L, Zeiger E. 2006. Auxin: The Growth Hormone. In: Plant Physiology. 4th edition, Sinauer

Associates, Inc. Sunderland p 467-504.

Thanaa SHM, Kassim NE, Abou Rayya MS, Abdalla AM. 2017. Influence of Foliar Application with moringa (Moringa oleifera L.) Leaf Extract on Yield and Fruit Quality of Hollywood Plum Cultivar Journal of Horticulture 4, 193.

http://dx.doi.org/10.4172/2376-0354.1000193

Toor RK, Geoffrey PS, Annuschka H. 2006. Influence of different types of fertilizers on the major antioxidant component of tomatoes. Journal of Food Composition and Analysis 19, 20-27.

Vasilenko V. 2002. Hydroponcs and humates: Ancient acid for modern agriculture, the best of the growing edge international select cream of the crop articles for soilless growers. New moon publishing 375-375.

Vaughan D, McDonald IR. 1976. Some effects of humic acid on cation uptake by parenchyma tissue. Soil Biology and Biochemistry 8(5), 415-421.

http://dx.doi.org/10.1016/0038-0717(76)90043-2

Vimala P, Ting CC, Salbiah H, Ibrahim B, Ismail L. 1999. Biomass production and nutrient yields of four green manures and their effect on the yield of cucumber. Journal of Tropical Agriculture and food Science 27, 47-56.

Wang YH, Thomas CE, Dean RA. 1997. A genetic map of melon (Cucumis melo L.) based on amplified fragment length polymorphism (AFLP) markers. Theoretical and Applied Genetics 95, 791-798. http://dx.doi.org/10.1007/s001220050627

Yaseen M, Arshad M, Khalid A. 2006. Effect of acetylene and ethylene gases released from encapsulated calcium carbide on growth and yield of wheat and cotton. Pedobiologia 50, 405-411.

http://dx.doi.org/10.1016/j.pedobi.2006.08.002

Yasmeen A. 2011. Exploring the Potential of moringa (Moringa oleifera) Leaf Extract as Natural Plant Growth Enhancer PhD Thesis. University of Agriculture Faisalabad, Pakistan.

Yildirim E. 2007. Foliar and soil fertilization of humic acid affect productivity and quality of tomato. ActaAgriculturae Scandinavica, Section B - Soil and Plant Science 57, 182-186.

http://dx.doi.org/10.1080/09064710600813107

Zachariakis A, Tzorakakis E, Kritsotakis I, Siminis CI, Manios V. 2001. Humic substance stimulate plant growth and nutrient accumulation in grapevine rootstocks. Acta Horticulturae 549, 131-136.

http://dx.doi.org/10.17660/ActaHortic.2001.549.14

Zekri M, Obreza TA. 2009. Plant Nutrients for Citrus Trees. UF/IFAS extension service, University of Florida, IFAS, Florida, USA.