



Antimicrobial potential of two medicinal plants (*Psammogeton biternetum* and *Haloxylon griffithii*) of District Kalat Balochistan, Pakistan

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Abstract

The main objective of this study was to explore the medicinal potential of two important plant species including; *Haloxylon griffithii* and *Psammogeton biternetum*. The Leaf, stem, root and seed (methanolic and ethanolic) extracts of these plant were examined against some pathogenic microbes (fungal and bacterial strains) through agar well diffusion techniques. For anti-bacterial screening 4 gram positive (*Enterococcus faecalis*, *Staphylococcus aureus*, *Staphylococcus epidermidis* and *Bacillus subtilis*) and 14 gram negative (*Acinetobacter baumannii*, *Escherichia coli*, *Salmonella typhi*, *Proteus sp*, *Proteus mirabilis*, *Proteus vulgaris*, *Pseudomonas aeruginosa*, *Salmonella paratyphi A*, *Salmonella enterica*, *Shigella boydii*, *Shigella sonnei*, *Klebsiella oxytoca*, *Klebsiella pneumoniae* and *Serratia marcescens*) bacterial species were selected. For antifungal activities 5 fungal species (*Candida albicans*, *Aspergillus niger*, *Penicillin*, *Aspergillus flavus* and *Mucor*) were chosen for examination. Results revealed that *Haloxylon griffithii* leaves was effective against all test species except *Bacillus subtilis*. *Psammogeton biternetum* was not active against *Bacillus subtilis*, *Acinetobacter baumannii*, *Proteus mirabilis*, *Proteus vulgaris*, *Salmonella paratyphi A*, *Salmonella enterica*, *Shigella boydii*, and *Shigella sonnei*. *Psammogeton biternetum* exhibited highest antimicrobial activity for *Pseudomonas aeruginosa* with 37mm ZI (zone of inhibition). *Haloxylon griffithii* was highly effective against *Proteus* with 31mm ZI. Furthermore, ethanolic extracts were found to be more effective than methanolic extracts. The extracts were least active against fungal strains. Results revealed that *Candida albicans*, *Aspergillus flavus* and *Mucor* are completely resistant against the selected medicinal plants.

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Introduction

Use of medicinal herbs as ancient medication is one of the regularly used practices in subcontinent. Plants have always been a source of cure to different diseases due to presence of many phytochemical compounds. Drugs that are obtained from herbal sources have a remarkable influence in preventing and curing lethal diseases. People in most developed countries use traditional medicine in health care (Fransworth, 1993). Locally, various plants are used as medicines at the preliminary levels in different regions of the world. Majority of the people are dependent on herbal plants for health care (Bhalodia *et al.*, 2011).

Research by WHO on regular biological drugs like gentamycine showed such medicines are not good enough against the microbes any longer. There are many drugs that were effective once but do not show activity against harmful bacterial strains (Pushpa *et al.*, 2011). A few medications may likewise have symptoms that cause different abnormalities in the body as side effect. These circumstances desperately compelled researchers for seeking drugs which are affordable, protected and biologically degradable and with least side effects (Kumar *et al.*, 2010).

Traditional medication plays a pivotal role in the medicine of developed nations. Herbal plants have been used for a certain period as solutions for human diseases because these are medicinally important (Suneetha *et al.*, 2013). *Cassia alata* (L.) is an important medicinal plant native to Asian countries, especially India and Pakistan. Distinctive parts of the plant were observed to show medicinal properties such as antimicrobial and analgesic. The treatment of ring worm carried out with the help of leaves of this plant. It is traditionally very useful against skin diseases in man and cattle (Deepak *et al.*, 2014).

Population explosion increased the need of natural resources to meet the need of food, clothes medicine etc. The use of ethnomedicinal knowledge has gained great concerns over the past few years because of its effective and low or no side effects in comparison to allopathic medicine. Besides the plants are used to

make various lifesaving drugs. Rahman *et al.*, (2011) reported the use of medicinal plants through studying their medical application via latest methods. Long range of medicinal plant parts are used as raw material for medicines. However, numbers of plants have been explored for antibacterial activity but still some local plants of some areas remain under the shell and remain unexplored for the diverse antimicrobial potential that the local community is blissfully utilizing (Martins *et al.*, 2001). The growing problem of bacterial resistance to the available antibiotics has felt the need to search for new antibacterial agents.

Kalat is one of the big cities of Balochistan. It is located $28^{\circ} 57'$ & $29^{\circ} 20'$ north side and $66^{\circ} 35'$ & $67^{\circ} 31'$ east. It covers an area of 6621 sq. km with an elevation of 1525 to 1982 meters above sea level. Kalat is among the coldest cities of Balochistan. The average rainfall is recorded 15.23 mm in winter season and snowfall is also recorded in December and January.

The annual minimum temperature was -4.34°C for the years 1993-1998, which was recorded at the meteorological center Kalat, whereas maximum temperature was 31.71°C July (Tareen *et al.*, 2010). Kalat is blessed with diverse medicinal and ornamental plants due to its geographical location. The natives of Kalat basically depend on the use of herbal plants for treatment of various diseases.

Psammogeton consists of 6 species, majority is found in sandy dunes of deserts in different parts of Turkmenistan, Pakistan, Afghanistan and Iran (Nasrabadi *et al.*, 2009). *P. biternatum* Edgew commonly known as "izbotk" in Kalat and "sparki" in other regions. It is an annual herb that ranges from 15 to 22cm in length. It has small flowers that are white or pink in color. It flowers from March to April.

It is an annual plant that is located in several regions of District Kalat. *Psammogeton biternetum* is locally used for the cure of malaria, whooping cough, typhoid

and chest diseases by the people of region (Ikram *et al.*, 2015). *P. biternatum* Edgew seeds are very beneficial for postpartum infections (after delivery) if ground and mixed in water and taken between meals (Bibii *et al.*, 2017).

Haloxylon griffithii belongs to family Chenopodiaceae, it is also known by the name of cat family. It consists of hundred genera and approximately twelve hundred species (Ali & Qaiser., 2001). Some of its types are weed and grow in barren soils. Thirteen species of *Haloxylon* are located predominantly in the dry regions of the South-West Asia and Arabian deserts and North-African.

The species of this genus are shrubs or sometimes small trees (Ali *et al.*, 2001). Five species of *haloxylon* are found in Pakistan out of six species found in central Asia (Mhaskar *et al.*, 2000). *H. griffithii* is widely distributed in the forests of Balochistan. It is commonly found in the regions of Harboi, Kalat, Hanna Urak, Spinni road and Wali Tangi, Quetta. Hence, the present study was carried out to find out the antibacterial activity of *Haloxylon griffithii* and *Psammogeton biternetum* against some pathogenic microbes.

Material and methods

Procurement of plant samples

The plant samples were collected from the Kalat city and out skirts. These were recognized and authenticated by Dr. R. B. Tareen, Department of Botany, University of Balochistan, and Quetta.

Plant extract preparation

The collected plant parts were chopped into small pieces and shade dried at 32-35°C. About 50g of each plant specimen was powdered with the help of an electric blender. Each sample was extracted with methanol, and ethanol.

After 24 hours, the extracts were filtered via fine filtration with the help of Whatman filter paper. Then the filtrate was concentrated in rotary flask evaporator, until dried (Doughari, 2006).

Test species

Various microorganisms were used for the study that include fungi and number of bacteria. The test organisms were obtained from CASVAB (Center for Advance studies in Vaccinology and Biotechnology), Civil Hospital Quetta, Federal Urdu University Karachi and Microbiology Lab, Department of Microbiology, University of Balochistan, Quetta. All the bacterial strains were grown in nutrient broth and incubated at 37°C for at least two days (Doughari, 2006).

Agar well diffusion methods

Agar well diffusion method was used to explore antimicrobial potential of *Psammogeton biternetum* and *Haloxylon griiffthii*. It is a common method used to test the antimicrobial activity of plants extracts.

In this process the surface of agar plate was inoculated by dispersal of volume of the microbial inoculums on the total surface of agar. Then, a hole was punched with a diameter of 6 to 8 mm with a sterilized cork borer and an extract solution of volume (20–100 µL) at chosen concentration was poured into the well. Afterwards, agar plates were incubated under appropriate conditions depending upon the organism under investigation. The anti-microbial agent was diffused in the agar medium and inhibited the growth of the bacteria (Chaman *et al.*, 2013).

Results

The data clearly shows that *Haloxylon griffithii* leaves are comparatively more effective against all test plant species. *Psammogeton biternetum* was not effective against *Bacillus subtilis*, *Acinetobacter baumannii*, *Proteus mirabilis*, *P. vulgaris*, *Salmonella paratyphi A*, *S. enterica*, *Shigella boydii*, and *S. sonnei*. Both plants were found to be ineffective against *Bacillus subtilis*. *Psammogeton biternetum* shows the highest antimicrobial activity against *Pseudomonas aeruginosa* with 37mm ZI (zone of inhibition). *Haloxylon griffthii* was highly effective against *Proteus* with 31mm ZI. However, ethanolic extracts were found to be more effective than methanolic extracts (Table 1).

Table 1. Antibacterial activity of *Psammogeton biternetum* and *Haloxylon griffithii* leaf in methanolic and ethanolic extracts.

	Organisms	Inhibition Zone (mm)			
		<i>Psammogeton biternetum</i>		<i>Haloxylon griffithii</i>	
		Ethanol	Methanol	Ethanol	Methanol
Gram Positive					
1.	<i>Enterococcus faecalis</i>	15	4	17	15
2.	<i>Staphylococcus aureus</i>	27	12	27	20
3.	<i>Staphylococcus epidermidis</i>	15	6	19	11
4.	<i>Bacillus subtilis</i>	-	-	-	-
Gram Negative					
1.	<i>Acinetobacter baumannii</i>	-	-	17	14
2.	<i>Escherichia coli</i>	27	12	21	16
3.	<i>Salmonella typhi</i>	25	11	21	18
4.	<i>Proteus sp</i>	31	25	31	22
5.	<i>Proteus mirabilis</i>	-	-	11	10
6.	<i>Proteus vulgaris</i>	-	-	17	12
7.	<i>Pseudomonas aeruginosa</i>	37	8	19	13
8.	<i>Salmonella paratyphi A</i>	-	-	27	19
9.	<i>Salmonella enterica</i>	-	-	19	15
10.	<i>Shigella boydii</i>	-	-	25	19
11.	<i>Shigella sonnei</i>	-	-	17	16
12.	<i>Klebsiella oxytoca</i>	15	5	-	-
13.	<i>Klebsiella pneumoniae</i>	9	4	16	15
14.	<i>Serratia marcescens</i>	13	8	27	21

Considering, the stem extracts of both plants it was observed that the stem extracts were less active against various strains of bacteria. The maximum action was detected in methanolic extracts of *Haloxylon biternetum* against *Staphylococcus aureus* and *Pseudomonas aeruginosa*. The extracts showed

zero activity against *Staphylococcus aureus*, *Staphylococcus epidermidis*, *Bacillus subtilis*, *Acinetobacter baumannii*, *Salmonella paratyphi A*, *S. enterica*, *Shigella boydii*, *S. sonnei* and *Proteus mirabilis* (Table 2).

Table 2. Antibacterial activity of *Psammogeton biternetum* and *Haloxylon griffithii* stem in methanolic and ethanolic extracts.

	ORGANISMS	Inhibition Zone (mm)			
		<i>Psammogeton biternetum</i>		<i>Haloxylon griffithii</i>	
		Ethanol	Methanol	Ethanol	Methanol
Gram Positive					
1.	<i>Enterococcus faecalis</i>	13	11	-	-
2.	<i>Staphylococcus aureus</i>	-	-	12	19
3.	<i>Staphylococcus epidermidis</i>	-	-	-	-
4.	<i>Bacillus subtilis</i>	-	-	-	-
Gram Negative					
1.	<i>Acinetobacter baumannii</i>	-	-	-	-
2.	<i>Escherichia coli</i>	10	8	-	-
3.	<i>Salmonella typhi</i>	5	4	18	15
4.	<i>Proteus sp</i>	11	10	17	17
5.	<i>Proteus mirabilis</i>	-	-	-	-
6.	<i>Proteus vulgaris</i>	4	2	-	-
7.	<i>Pseudomonas aeruginosa</i>	8	6	11	19
8.	<i>Salmonella paratyphi A</i>	-	-	-	-
9.	<i>Salmonella enterica</i>	-	-	-	-
10.	<i>Shigella boydii</i>	-	-	-	-
11.	<i>Shigella sonnei</i>	-	-	-	-
12.	<i>Klebsiella oxytoca</i>	8	4	-	-
13.	<i>Klebsiella pneumoniae</i>	4	-	7	5
14.	<i>serratia marcescens</i>	10	4	-	-

The stem extracts of *Psammogeton biternetum* were found to be more effective against test strains as compared to *Haloxylon griffithii* (Table 2).

The seed extracts of *Psammogeton biternetum* showed great deal of activity against most of the test species. Ethanolic extracts were comparatively more

effective than the methanolic extracts. Minimum antimicrobial activity was observed against *Acinetobacter baumannii* and highest activity was observed against *Pseudomonas aeruginosa* *Shigella sonnei*, *Klebsiella oxytoca*, *Serratia marcescens* and *Salmonella paratyphi A* showed complete resistance against the extracts of *P. biternetum* seeds.

Table 3. Antibacterial activity of *Psammogeton biternetum* seeds in methanolic and ethanolic extracts.

	Organisms	Inhibition Zone (mm)	
		Ethanol	Methanol
Gram positive			
1.	<i>Enterococcus faecalis</i>	18	11
2.	<i>Staphylococcus aureus</i>	12	17
3.	<i>Staphylococcus epidermidis</i>	18	15
4.	<i>Bacillus subtilis</i>	8	-
Gram Negative			
1.	<i>Acinetobacter baumannii</i>	4	-
2.	<i>Escherichia coli</i>	7	12
3.	<i>Salmonella typhi</i>	18	17
4.	<i>Proteus sp</i>	26	27
5.	<i>Proteus mirabilis</i>	18	17
6.	<i>Proteus vulgaris</i>	15	11
7.	<i>Pseudomonas aeruginosa</i>	25	27
8.	<i>Salmonella paratyphi A</i>	-	-
9.	<i>Salmonella enterica</i>	19	27
10.	<i>Shigella boydii</i>	17	17
11.	<i>Shigella sonnei</i>	-	-
12.	<i>Klebsiella oxytoca</i>	-	-
13.	<i>Klebsiella pneumoniae</i>	5	11
14.	<i>Serratia marcescens</i>	-	-

Concluding, it was observed that the selected plants (*P. biternetum* and *H. griffithii*) are potentially effective against the selected bacterial strains. The seeds of izbotk and leaves of bundi are most effective with maximum zones of inhibition (Table 3). The

roots of *H. griffithii* showed zero activity in both ethanolic and methanolic extracts against all test species (Table 4). Both plants were highly effective against *Escherichia coli*, *Salmonella typhi* and *Pseudomonas aeruginosa*.

Table 4. Antibacterial activity of *Haloxylon griffithii* root in methanolic and ethanolic extracts.

	Organisms	Inhibition Zone (mm)	
		Ethanol	Methanol
Gram positive			
1.	<i>Enterococcus faecalis</i>	-	-
2.	<i>Staphylococcus aureus</i>	-	-
3.	<i>Staphylococcus epidermidis</i>	-	-
4.	<i>Bacillus subtilis</i>	-	-
Gram Negative			
1.	<i>Acinetobacter baumannii</i>	-	-
2.	<i>Escherichia coli</i>	-	-
3.	<i>Salmonella typhi</i>	-	-
4.	<i>Proteus sp</i>	-	-
5.	<i>Proteus mirabilis</i>	-	-
6.	<i>Proteus vulgaris</i>	-	-
7.	<i>Pseudomonas aeruginosa</i>	-	-
8.	<i>Salmonella paratyphi A</i>	-	-
9.	<i>Salmonella enterica</i>	-	-
10.	<i>Shigella boydii</i>	-	-
11.	<i>Shigella sonnei</i>	-	-
12.	<i>Klebsiella oxytoca</i>	-	-
13.	<i>Klebsiella pneumoniae</i>	-	-
14.	<i>Serratia marcescens</i>	-	-

The results show that the selected plants are least active against selected fungal species. The study carried out experiment on fungal strains i.e. *C. albicans*, *A. niger*, *penicillin sp*, *A. flavus* and *Mucor*. *P. biternetum* showed least activity against tested fungal strains. The seeds extract of *P. biternetum* are

only active against *A. niger* and *Penicillin sp* with 3mm and 2mm IZ (Table 5). *H. griffithii* showed activity against *A. niger* and *Penicillin sp*. Maximum activity was shown in the ethanolic extracts of leaf of *H. griffithii* (10mm IZ) (Table 6).

Table 5. Antifungal activity of *Psammogeton biternetum* (stem, leaf and seed) in methanolic and ethanolic extracts.

Fungal strains	Zone of inhibition(mm)					
	Stem		Leaf		Seed	
	Ethanol	Methanol	Ethanol	Methanol	Ethanol	Methanol
1. <i>Candida albicans</i>	-	-	-	-	-	-
2. <i>Aspergillus niger</i>	-	-	-	-	3	-
3. <i>Penicillin</i>	-	-	-	-	2	-
4. <i>Aspergillus flavus</i>	-	-	-	-	-	-
5. <i>Mucor</i>	-	-	-	-	-	-

Discussion

From the present research, it was evident that *P. biternetum* and *H. griffithii* possess a promising antibacterial action against the investigated organisms. The results obtained from Agar diffusion assay method supports that gram-negative bacteria are sensitive to the extracts than gram-positive bacteria. Similar observations were observed Cimanga *et al.*, (2002) that *P. aeruginosa* was found to be most sensitive against tested concentrations. Similar results were also reported by Salama *et al.*,

(2010). Ethanolic extracts had high antimicrobial potential than methanolic extracts, the difference in antibacterial and antifungal actions for different plant parts was also stated Ajaib *et al.*, (2014).

The results of present research clearly show that the antibacterial and antifungal activity fluctuate with the plants and plant part used. Thus, the study establishes the value of plants to be used in ayurveda, which could be of substantial interest to the growth of new medicinal drugs.

Table 6. Antifungal activity of *Haloxylon griffithii* (stem, leaf and root) in methanolic and ethanolic extracts.

Fungal strains	Zone of inhibition(mm)					
	Stem		Leaf		Root	
	Ethanol	Methanol	Ethanol	Methanol	Ethanol	Methanol
1. <i>Candida albicans</i>	-	-	-	-	-	-
2. <i>Aspergillus niger</i>	2	-	10	-	-	-
3. <i>Penicillin</i>	3	-	2	-	6	-
4. <i>Aspergillus flavus</i>	-	-	-	-	-	-
5. <i>Mucor</i>	-	-	-	-	-	-

Furthermore, the selected plants show high inhibition zone against *Pseudomonas aeruginosa*, *Candida albicans* and *Aspergillus niger* ZI of 13, 14 and 18mm respectively. Chaman *et al.* (2013) reported similar results. The disappearance of bacterial colonies may be due to the presence of chemicals like

alkaloids. Ramya *et al.*, (2008) stated several positive outcomes of ethanolic leaf extracts of *Sadbahar* against many bacteria (gram positive and gram negative) i.e. *E. coli*, *P. aeruginosa*, *S. marcescens*, *S. typhi*, *S. aureus*, *S. pyrogens*, *B. cereus* and *B. subtilis*. Methanolic extracts of both plants (*P.*

biternetum and *H. griffithii*) were least active against bacterial and fungal strains. However, Ethanolic extracts have shown good antimicrobial activity. From the previous research it has been found that extracts prepared in specific solvents enhances the antimicrobial potential of plant. Thongson *et al.*, (2004) findings strongly support these findings.

The appearance and spread of multidrug-resistant (MDR) bacterial pathogens have significantly threatened the present antibacterial treatment (Boucher *et al.*, 2009). MDR bacterial infections are responsible for increasing mortality, staying longer in hospitals, and increased cost of treatment and attention (Giamarellou, 2010). The addition of traditionally used drugs including phytomedicine, if they prove harmless and operative, into national health care system is recommended by World Health Organization (Marasini *et al.*, 2015). Although many medicinal plants (>500 plants) have been reported to be used by people since ancient times for primary health care. However, there has been a paucity in data regarding their in vitro or in vivo efficacy (Kunwar and Bussmann, 2008).

Various reported literature like Cheruiyot *et al.*, (2009) and Ramzi *et al.*, (2005) were in accordance to our findings, claiming the antimicrobial activities of methanolic plant extracts of *Psidium guajava* and *Boswellia elongate* respectively against test species were least effective. Ajaib *et al.*, (2013) found alike results during the investigation of antioxidant and antimicrobial activities of *Rivina humilis* L., *Echinochloa colona* (Linn.) and *Sporobolus coromandelianus* (Retz.) Kunth. Thus, it can be concluded that test plant species has great antibacterial potential against diverse bacterial strains that could help the pharmaceutical industries to efficiently use these plants that were remained unexplored yet.

Conclusion

The present study concludes that the selected plants (*P. biternetum* and *H. griffithii*) extracts have prominent effects against propagation of investigated

bacterial species but did not show efficient results against fungal strains. Hence, the results exhibit the possibility of using these extracts for the ailment of harmful infections. The study further concludes that the seeds and leaves of *P. biternetum* are better active for microbial activities. Moreover, ethanolic extracts were better than the methanolic extracts and proved to be highly effective against vast range of microbial strains. So, it can be concluded that both plants possess medicinal properties against wide range of bacterial and fungal strains and can be used as better substitute to expensive drugs available in market.

References

- Ajaib M, Ali S, Khan Z.** 2014. Antioxidant and antimicrobial activities of an ethnobotanically important plant *Notholirion thomsonianum* from district Kotli, Azad Jammu & Kashmir. Journal of Animal and Plant Sciences **24**, 774-780.
- Ajaib M, Khan KM, Perveen S, Shah S.** 2013. Antimicrobial and antioxidant activities of *Echinochloa colona* (Linn.) Link and *Sporobolus coromandelianus* (Retz.) Kunth. Journal of the Chemical Society of Pakistan **35**, 960-965.
- Ajaib M, Zikrea A, Khan KM, Perveen S, Shah S, Karim A.** 2013. *Rivina humilis* L: A potential antimicrobial and antioxidant source. Journal of the Chemical Society of Pakistan **35**, 1384-1398.
- Ali SI, Qaiser M, Freitag H, Hedge IC, Jafri SMH, Kothe-Heinrich G, Omer S, Uotila P.** 2001. Flora of Pakistan, Published by Department of Botany, University of Karachi, Karachi, Pakistan **204**, 188-190.
- Ali SI, Qaiser M.** 2001. Flora of Pakistan Chenopodiaceae Jointly published by Department of Botany, University of Karachi, Karachi, Pakistan & Missouri Botanical Press: Missouri Botanical Garden, St Louis, Missouri USA **204**, 185.
- Aziz-UL-Ikram NBZ, Shinwari ZK, Mohammad Q.** 2015. Ethnomedicinal review of

folklore medicinal plants belonging to family Apiaceae of Pakistan. *Pakistan Journal of Botany* **47**, 1007-1014.

Bhalodia NR, Shukla VJ. 2011. Antibacterial and antifungal activities from leaf extracts of *Cassia fistula* L: An ethnomedicinal plant. *Journal of Advanced Pharmaceutical Technology & Research* **2**, 104-109.

Bibii T, Ahmad A, Baloch AI, Muhammad S, Manzoor R. 2017. Ethnomedicinal uses of plants for child birth and postpartum recovery in district Pishin, Northern Balochistan, and Pakistan. *International Journal Biological Pharmacy and Allied Sciences* **6**, 1730-1760.

Boucher HW, Talbot GH, Bradley JS, Edwards JE, Gilbert D. 2009. Bad bugs, no drugs: no ESKAPE! An update from the Infectious Diseases Society of America. *Clinical infectious diseases* **48**, 1-12.

Chaman S, Sharma G, Shalini, Reshi AK. 2013. Study of Antimicrobial properties of *Catharanthus roseus* by Agar Well Diffusion method. *International Research Journal of Pharmaceutical and Applied Sciences* **5**, 65-68.

Cheruiyot K, Olila R, Kateregga D. 2009. In vitro antibacterial activity of selected medicinal plants from Longisa region of Bomet district, Kenya. *African Health Sciences* **9**, 42-46.

Cimanga K, Kambu K, Tona L, Apers S, De Bruyne T, Hermans N, Totte J, Pieters L, Vlietinck AJ, 2002. Correlation between chemical composition and antibacterial activity of essential oils of some aromatic medicinal plants growing in the Democratic Republic of Congo. *Journal of ethnopharmacology* **79**, 213-220.

Deepak S, Pawar A, Shinde P. 2014. Study of antioxidant and antimicrobial activities of *Andrographis paniculate*, *Asian Journal of Plant Science and Research* **4**, 31-41.

Doughari JH. 2006. Antimicrobial Activity of *Tamarindus indica* Linn. *Tropical Journal of Pharmaceutical Research* **5**, 597-603.

Farnsworth NR, Soejarto DD. 1991. Global importance of medicinal plants. In: Akerele, O., Heywood, V., Synge, H. (Eds.). *The Conservation of Medicinal Plants*. Cambridge University Press, Cambridge, 25-52.

Giamarellou H. 2010. Multidrug-resistant Gram-negative bacteria: how to treat and for how long. *International Journal of Antimicrobial Agents* **36**, 50-54.

Hediat MH, Salama, Najat Marraiki. 2010. Antimicrobial activity and phytochemical analyses of *Polygonum aviculare* L. (Polygonaceae), naturally growing in Egypt. *Saudi Journal of Biological Sciences* **17**, 57-63.

Kumar AO, Mutyala NL, Raja Rao KG. 2010. Antibacterial Evaluation of Snake Weed (*Euphorbia Hirta* L.), *International Journal of Pharm Tech Research* **2**, 1383-1385.

Kunwar RM, Bussmann RW. 2008. Ethnobotany in the Nepal Himalaya. *Journal of Ethnobiology and Ethnomedicine* **4**, 24.

Marasini BP, Baral P, Aryal P, Ghimire KR, Neupane S. 2015. Evaluation of antibacterial activity of some traditionally used medicinal plants against human pathogenic bacteria. *BioMed Research International* 2015, 6 pages, Article ID 265425, <http://dx.doi.org/10.1155/2015/265.425>

Martins AP, Salgueiro L, Goncalves MJ, Proencacunha V, Vila R, Canigueral S, Mazzoni V. 2001. Essential oil composition and antimicrobial activity of three Zingiberaceae from S.Tomeeprincipe, *Journal of Planta Medica* **67**, 580-584.

Mhaskar KS, Blatter E, Caius JF. 2000. Indian

Medicinal plants published by Indian Books centre, Delhi, India IV, 1212-1214.

Rahimi-Nasrabadi M, Gholivand MB, Batooli H, Vatanara A. 2009. Chemical Composition of the Essential Oil from Leaves and Flowering Aerial Parts of *Psammogeton canescens* (DC.) Vake from Iran. *Journal of Medicinal Plants* **8**, 82-86.

Rahman SA, Abd-Ellatif SA, Deraz SF, Khalil AA, 2011. Antibacterial activity of some wild medicinal plants collected from western Mediterranean coast, Egypt: Natural alternatives for infectious disease treatment. *African Journal of Biotechnology* **10**, 10733-10743.

Ramya S, Govindaraji V, Kannan KN, Jayakumararaj R. 2008. In Vitro-Evaluation of Antibacterial Activity Using Crude Extracts of *Catharanthus roseus* L. (G.) Don, *Ethno botanical Leaflets* **1**, 1067-72.

Ramzi A, Mothana A, Lindequist U. 2005. Antimicrobial activity of some plants of the Island soqotra. *Journal of Ethnopharmacology* **96**, 177-181.

Samy RP, Ignacimuthu S. 2000. Antibacterial activity of some folklore medicinal plants used by tribals in Western Ghats in India. *Journal of Ethnopharmacology* **69**, 63-71.

Suneetha G, Ravi V. 2013. Antimicrobial Activity of *Andrographis paniculata* Flower Extracts, *International Journal of Research and Reviews in Pharmacy and Applied science* **2**, 604-610.

Tareen RB, Bibi T, Khan MA, Ahmad M, Zafar M. 2010. Indigenous knowledge of Folk medicine by the women of Kalat and Khuzdar regions of Balochistan, Pakistan. *Pakistan Journal of Botany* **42**, 1465-1485.

Thongson C, Davidson PM, Mahakarnchanakul W, Weiss J. 2004. Antimicrobial activity of ultrasound assisted solvent-extracted spices. *Letters in Applied Microbiology* **39**, 401-6.

Tona L, Kambu K, Ngimbi N, Cimanga K, Vlietinck AJ. 1998. Antiamoebic and phytochemical screening of some Congolese medicinal plants. *Journal of Ethnopharmacol* **61**, 57-65.