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# **RESEARCH PAPER**

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# Etiological agents of Vulvovaginitis amongst women complaining of genital tract infection in Soran City, Kurdistan, Iraq

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# Abstract

Vulvovaginitis is one of the commonest reproductive tract infections in women worldwide. There are three types of infectious vaginitis including, bacterial vaginosis, valvovaginal candidiasis, and trichomoniasis. The present study aimed to characterize the causative agents of vulvovaginitis among women attending Soran Obstetrics and Gynecology hospital. Host -related and some behavioral risk factors which proposed as predisposing factors for this disease were also investigated. A pair of high vaginal swabs was collected from 97 vulvovaginitis women aged 18-52 years. Trichomoniasis was detected by wet and stained smears. Candidiasis was identified by direct examination and culturing on both Sabouraud dextrose agar and Chromagar medium. Bacterial infection was confirmed by Nugent scoring method after Gram staining. The present results showed that 85 (87.6%) cases were positive for etiologic agents and 12(12.3%) with nonpathogenic agents. Among the positive patients, the commonest organisms was bacteria 45%, followed by Candida species 15%, Trichomniasis as 7%, and mixed infection with these three agents in 33%. Moreover, infection with non-albicans Candida like C. galabrata, C. krusei, and C. dubliensis were also detected. Pregnant women showed high percentage of infection with trichomoniasis and candidiasis .Women that used IUD as contraceptive methods, as well as those of non- contraceptive users were more susceptible to bacterial infection than the other. High percentage of women was infected with the etiological agents that responsible for vulvovaginitis particularly with bacteria. Routine culturing of vaginal discharge must be performed for these patients; also antibiotics susceptibility of bacterial isolates should be determined. In addition, comprehensive healthcare education plan is needed to manage the disease.

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#### Introduction

Vulvovaginitis includes infectious and noninfectious conditions involving vulva and vagina (Mc Cormack and Augenbraun, 2015). It is a common disease that affects women and girls of all ages specifically the sexually active women .It considered as one of the most important public health issues in both developed and developing countries. Vaginal complaints are the most common reason for gynecological consultation and account for approximately 10 million office visits annually (Anderson *et al.*, 2004). Recently, Loveless and Myiant (2017) summarized that this disease is the most prevalent gynecological concern presenting in pediatric and adolescent girls.

Bacterial vaginosis (BV), vulvovaginal candidiasis (VC), and trichomoniasis are diagnosed in 70% of patients, including 40% to 50% with BV, 20% to 25% have VC, and 15% to 20% inflected with trichomoniasis. The remaining 30% are undiagnosed that can have physiologic discharge (leucorrhea), atrophic vaginitis, and vulvar dermatologic abnormalities, or vulvodynia. (Mulley, 2000; Vaginitis, 2006). The main characteristics of vaginitis are vaginal discharge, vulvar itching /irritation and malodor, which consider as important reason for women to seek help of a health care providers (Mills, 2017; Narayankhedar et al., 2015).

Normal vaginal ecosystem is a physiological important biomass, in which glycogen rich epithelial cells are always shed. As the cells autolysis, glycogen depolymerizes to glucose, which serves as an energy source for the commonly inhabit vagina Lactobacilli spp. These bacteria metabolize glucose to lactic acid, which lead to a normal vaginal pH of about 4.0 that inhibit the growth of pathogenic bacteria (Ravel et al., 2011). In addition to producing lactic acid, lactobacilli may also produce hydrogen peroxide, which is bactericidal alone and highly effective when integrate with physiologic amounts of myeloperoxidase and chloride. (Klebanoff et al .,1991) .Several researchers pointed that human vaginal microbiota seem to play an important role in preventing many urogenital diseases, like BV, VC, sexually transmitted infections,

urinary tract inflammations (Gupta et al.,1998; Martin et al., 1999; Wiesenfeld et al., 2003) and Human immunodeficiency virus type 1 infection. (Lai et al., 2009; Taha et al., 1998). Trichomonas vaginalis infection is the most prevalent non-viral sexually transmitted infection worldwide. It caused by unicellular, aerotolerant, flagellated protozoan parasite that restricted to human. Globally, an estimated of 170 million cases of trichomoniasis were reported .The majority of these cases were in developing countries (Gerbase et al., 1998). In United States, approximately 3.7 million people are infected with T. vaginalis, more than chlamydia and gonorrhea collectively (Meites, 2013; Satterwhite et al., 2013) .Although this infection is common worldwide, it has been considered a "neglected" parasitic infection, because of a lack of public awareness and due to restricted knowledge of its sequelae and associated costs. (Meites, 2013; Secor et al., 2014).

Furthermore, Alcaide and colleagues (2015)highlighted the high rates of reinfection or treatment failures in trichomoniasis women and emphasized the need for rescreening them after treatment by using sensitive and specific test. The center of disease control recommends the use of more sensitive method to detect T. vaginalis of both symptomatic and asymptomatic infections. Nucleic acid amplification tests (NAATs) are more sensitive than wet preparation in about 3-5 times, and both of vaginal secretions or urine can be tested by NAAT. (Workowski and Bolan, 2015).

Moreover, trichomoniasis increases the risk of complications in pregnancy. The percentage of preterm birth was increased by 42% among infected women. As well as the risks of delivery of a small for gestational age infant also increased. (Malone, 2017). There is strong evidence suggests the correlation between human immunodeficiency virus (HIV) and T. vaginalis. This parasitic infection in women infected with (HIV) enhances HIV transmission by increasing genital shedding of the virus (Hirt and Sherrard, 2015; Schwebke and Burgess, 2004). In addition. trichomoniasis has been interacted with incidence of cervical cancer (Boyle & Smith, 1999; Gram et al., 1992,).

VC is an infection caused by *Candida* species that affects millions of women yearly and considered as significant public health problem (Goncalves *et al.*, 2016). Some researchers have estimated that about 70-75% of adult women are inflected with at least one episode of VC during their lives .I t is considered as the second most common cause of vaginitis after BV (Sobel, 2007). Inhibition of normal microbial flora by antibiotics encourages the growth of yeasts, although VC sometimes happens after treatment of trichomoniasis or BV (Agnew and Hillier, 1995).

*Candida spp* perhaps become pathogenic and lead to candidiasis under some circumstances, such as immunesuppressed specifically due to steroid use and prolonged broad spectrum antibiotics treatment, pregnancy, used of oral contraceptive, malnutrition, diabetes, and obesity, (Nyirjesy 2014; Okungbowa *et al.*, 2003).

*C. albicans* is the most common cause (80%-90%), whereas other yeasts account for up to 20% of cases. *C. tropicalis* is isolated from 1% to 5% and may be associated with a higher rate of recurrence after standard treatments. (Horowitz *et al.*, 1985; Spinillo *et al.*, 1997). *C. glabrata* accounts for about 10% of vaginal yeast isolates (Geiger *et al.*, 1995; Spinillo *et al.*, 1997). The pathogenicity of *Candida* species is mediated by several virulence factors like, adhesion, hyphal and biofilm formation, extracellular hydrolytic enzyme production, and phenotypic switching (Goncalves *et al.*, 2016; Tamura *et al.*, 2007).

BV is considered as the main cause of symptomatic vaginitis with vaginal discharge and malodor with prevalence about 15% to 50%. It is the most common reason for lower genital tract disorder among women of child- bearing ages (Allsworth and Peipert, 2007). It reflects alteration in vaginal microbiota from H2O2-producing lactobacilli to many pathogenic bacteria (Hillier 1993). The disease is related to the disruption of vaginal environment rather than to the occurrence of these bacteria (Mills, 2017). Histologically, there is an absence of inflammation in biopsies of the vagina, therefor the term vaginosis rather than vaginitis was used. More recently, Yudin and Money (2017) recommended that diagnosis and treatment of BV is important for symptom resolution in pregnant women. In another current study ,they concluded that BV significantly associate with preterm delivery (Tellapragada *et al.*, 2017). Also this finding beside other adverse pregnancy outcomes were previously documented by several researchers (Carey and Klebanoff, 2005).

The goal of this study was to highlight the etiological agents of Vulvovaginitis among women complaining of genital tract infection in Soran city. Also the association between this disease and various behavioral risk factors was explained.

## Materials and methods

## Samples collection

High –vaginal swabs were collected from 97 vulvovaginitis women aged 18-52 years during October 2016 to March 2017. Pair of vaginal swab was obtained from each participant after speculum examination and clinical presentation which was confirmed by specialized gynecologist in Soran obstetrics and gynecology hospital and private clinics in Soran city/Erbil-Iraq. The first swab was used for culturing, and the second was used for direct wet smear examination, 10% KOH test, and stained smears.

## Trichomoniasis identification

*T. vaginalis* infections were identified by jerky motility of the organisms in wet direct mount. Dried smears stained with Giemsa and Leishman stains were also prepared. All slides were examined under 40 X and 100 X of compound microscopy (Garcia and Ash, 1979; Yazar *et al.*, 2002).

## Candidiasis identification

VC was confirmed by direct wet preparation with 10% KOH which clarify the appearance of yeast cells or pseudohyphae and killed the majority of bacteria, dry smear for Gram staining, and culturing on Sabouraud's dextrose agar supplemented with chloramphenicol. The plates were incubated at 37C for 48hr. Then smears were prepared from each growth and stained with lactophenol cotton blue for confirming the characteristic of *Candida* as blue budding yeast cells (AL-Attraqhchi *et al.*, 2013).

Moreover, Candida isolates were identified by germ tube formation for distinguishing C. albicans from other non- Candida albicans species (Cheesbrough, 2006) and by culturing on Chrom agar as differential and selective medium for classifying Candida spp. which differentiate Candida species depending upon the specific color that produced by each one( Yadav and Prakash, 2016). As well as the antifungal activity of Nystatin was assessed using agar disc diffusion 8.3, technique; three concentrations (16.6,4.15mg/ml) of the drug were tested against the clinical isolates of Candida spp. Diameters of the inhibition zone surrounding each disc was measured in comparison with zone Diameter Interpretive standards (Table. 2) (Maroszynska et al., 2013).

## Bacterial vaginosis identification

BV was assessed according to Nugent scoring system. Gram stained slides were prepared and examined under 100X; about 10-15 microscopic fields were checked for each samples, A Nugent score of 0-3 was considered as negative for BV and number of 4-6 account for intermediate, whereas a score of 7-10 was recorded as consistent with BV (Naravankhedkar *et al.*, 2015; Nugent *et al.*, 1991). In addition, the presence of Amsel *et al.* criteria for diagnosis of BV was observed in most of BV samples by presence of clue cells (the borders of epithelial cells intensively covered with bacilli after Gram staining), fishy amine odor or positive whiff test after adding 0%KOH solution, and presence of vaginal homogeneous white discharge (Amsel *et al.*,1983).

#### Statistical analysis

Chi-square test was used to analyze the data. If P value <0.05 so considered as statistically significant.

## Results

The results of the current study revealed that amongst the 97 enrolled vulvovaginitis women, 85 (87.6%) cases were positive for etiologic agents and 12(12.3) with nonpathogenic agents. Among the positive patients, BV was the most common type of vulvovaginitis (45%), followed by VC (15%) *,Trichomonas* vaginitis (TV) (7%), mixed infection of VC+BV (13%), TV+BV (8%) and (6%) for both of TV+VC and TV+VC +BV. The total mixed infections was about 33% Fig. 1. Regarding the age groups, the highest percentage of infection with TV was in 25-31 years age interval which was 14.28%. Candidiasis also distributed remarkably in the age group 25-31 years as 22.8% if we excluded the age group 46-52 years owing to few cases (just two). Moreover high percentage of BV was confirmed within 32-38 years age group 65.2%. As well as, maximum percentage of mixed infection with vaginitis pathogens was found in 25-31 age group. No significant differences was observed. Fig.2.



**Fig. 1.** Repartition of patients according to etiological agents Bacterial vaginosis (BV), Candidal vaginitis (CV), Trichomonas vaginitis (TV).



**Fig. 2.** Relationship between age groups and vulvovaginitis. Bacterial vaginosis (BV), Candidal vaginitis (CV), Trichomonas vaginitis (TV).

According to pregnancy out of 85 infected women included in this study, 21 showed to be pregnant and 64 non –pregnant women. The distribution of TV was higher in non-pregnant women 7.8% compared in pregnant women 4.7% as single infection, but when take in account the total infection with both single and mixed, the result showed that pregnant women had higher percentage 38.09% (8/21) than non-pregnant women 23.4% (15/64) with no significant differences. VC was more prevalent in pregnant women 23.8% than in non-pregnant women 12.5%. Also the overall percentage of single and mixed infection of pregnant women with candidiasis was higher (14/21) 66.6% than that of non-pregnant women 31.2% (20/64).

Concerning bacteria, the present result showed that the non-pregnant women had high infection 51.5% in comparison with pregnant group23.8%. Finally the total mixed infections were detected in pregnant women 47.6% more than that recorded in nonpregnant 28.1%. Fig. 3.



**Fig. 3.** Prevalence of pathological agents among pregnant and non-pregnant women. Bacterial vaginosis (BV), Candidal vaginitis (CV), Trichomonas vaginitis (TV).

Moreover the current data showed just 19 women out of 85 women enrolled were used contraceptive methods. The most prevalent etiologic agent that detected in contraceptive user women was bacteria that reached 71.4% in Intra Uterine Device (IUD) user and 50% in women used oral pills. Beside one woman who used suppositories afflicted with BV.

No infection with trichomoniasis as single in contraceptives using women, but mixed infection with bacteria and Candida were detected in three cases. VC was recorded in low percentage as 7.1% in IUD user women for both of single and mixed infection of the three agents. Furthermore it was proved as a single infection in 1/4 (25%) and as mixed infection with bacteria 1/4 (25%) in pill user women Fig.4.

According to the residence of women, Within the 85 vulvovaginitis women, 69 and 16 of them lived in urban and rural area respectively. In women resident in urban area trichomoniasis was recorded in 8.6% as single infection and 7.24% in mixed infection with both bacteria and candida. Whereas rural women showed 12.5% as mixed infection with bacteria. VC was distributed as 15.9% and 12.5% in urban and rural area respectively; also mixed infections with the other two pathogens were detected. Infection with the three pathological agents was identified in urban women only as 7.2% Fig. 5. Regarding BV, 56.2% in women of rural area more than that of women in urban location 42%.

In addition to, mixed infections were identified in 18 and 5 cases related to urban and rural area respectively. Statistically no significant differences were confirmed. Fig.5.



**Fig. 4.** Distribution of pathological agents according to contraceptive methods Bacterial vaginosis (BV), Candidal vaginitis (CV), *Trichomonas* vaginitis (TV)



**Fig. 5.** Distribution of pathogens according to residence of women Bacterial vaginosis (BV), Candidal vaginitis (CV), Trichomonas vaginitis (TV).

## Identification of Trichomonas vaginalis

A total of 23 (27.05%) cases of trichomoniasis were recognized from 85 vulvovaginitis women by direct wet mounts (Fig.6.A) And dried smears stained with Giemsa and Leishman stains (Fig.6.B,C).



Fig.6. A. Lemon-shaped *T.vaginalis* in direct smear. B. stained with Giemsa stain C. Stained with Leishman stain.

## Identification of Candida spp

*Candida spp* were identified microscopically which appeared as unicellular, spherical shaped with bud in, Gram and Lactophenol cotton blue stained preparations. Fig.7.A &B. Also multicellular pueudohyphae was detected in direct specimen examination Fig.8.A. Moreover *C.albicans* was confirmed by germ tube formation Fig.8.B. and differentiation among *Candida* spp was achieved by culturing on Chrom Agar medium Fig.9.



**Fig. 7.A** Candida cells with buds, A. Stained with Lactophenol cotton blue.



Fig. 7.B. Candida cells in direct smear of vaginal exudate.

The prevalence of *C. albicans* was recorded as 26 (76.47%) and the non albicans included, *C. galabrata* 5(14.7%), *C.Krusei* 2 (5.88%) and *C.dubliensis* 1(2.94%). Also the present results proved the sensitivity of *Candida spp* to all Nystatin concentrations compared to interpretive chart of diameter zone of growth inhibition for Nystatin. Table 2.

	. Blanks :		
1. A	Candida pseudohyphae	6	ę
	1	7	
Epithelial cell Candida pseudo hypha 49	A	Germ tube	В

Fig. 8.A. Candida pseudohyphae. B. Germ tube of C.albicans.



Fig. 9. Candida spp on Chrom Agar Medium, A-C.albicans, B-C.dubliensis C-C.krusei, D-C.galabrata.

Species Number of		%	Germ tube	Colony on Chrom	Sensitivity to Nystatin		
	isolate			agar	Disc potency	Diameter	Interpretive
					(µg/ml)	(mm)	standard
Candida	26	76.47%	Positive	green	16.6	21 mm	Sensitive (S)
albicans					8.3	29 mm	S
					4.15	16 mm	S
Candida	5	14.70%	Negative	Dark pink or	16.6	20 mm	S
glabrata			purple	8.3	23 mm	S	
					4.15	17 mm	S
Candida	2	5.88%	Negative	Pink center with	16.6	24 mm	S
krusei		white edge	8.3	20 mm	S		
					4.15	16 mm	S
Candida	1	2.94%	Positive	Pale color	16.6	23 mm	S
dubliensis					8.3	20 mm	S
					4.15	17 mm	S

	Table 1.	Characterization	of Candida spp	beside sen	sitivity to Nystatin.
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**Table 2.** Interpretive chart with diameter zone of growth inhibition for Nystatin.

Antifungal agent	Zone di	Zone diameter interpretive standard (mm)			
	Resistant(R)	Intermediate(I)	Susceptible(S)		
Nystatin	No zone	10-14	≥15		
Nystatili	No zone	10-14	215		

## Identification of Bacterial vaginosis

According to Nugent- scoring method, smear preparations of vaginal discharges were stained with Gram stain for detection and counting of bacterial cells, as well as clue cells were identified as epithelial cells that heavily covered with bacilli. Fig,10.A and B.

The current data showed that, out of 85 positive vulvovaginitis women 60 (70.5%) of them were infected with bacteria depending upon Nugent scoring with milky-white, homogenous discharges and most of them with amine odor after addition of 10% potassium hydroxide to it, 15(17.6%) considered as intermediate bacterial flora, and 10 (11.7) classified as normal bacterial flora.Table 3.

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Table 9	Bacterial	Vaginosis	$hv N ll \sigma$	ent cooring
ranc 3.	Dacteriai	vasinosis	Dy mus	cint scoring.

Nugent scoring	Total +ve cases	%
	(n=85)	
<4	10	11.76%
4-6	15	17.64%
7-10	60	70.58%



**Fig.10A.** Bacterial cells in smear preparation. Gram positive with clue cells and pus.



Fig. 10B. Clue cells and coccobacilli bacteria.

#### Discussion

Laboratory diagnostic results of the vulvovaginitis women enrolled in the present study showed that high percentage (45%) of them had BV as a single infection. This prevalence was approximately similar to the result reported in Ethiopia (48.6%) (Bitew et al., 2017). Whereas much higher than that documented in Canada (7.1%) (Pham et al., 2012) and in India (17.3%) (Narayankhedkar et al., 2015) . Beside this, Olowe et al., (2014) recorded it as (38%) in Nigeria. The present results suggested that large percentage of women of reproductive age is highly vulnerable to such infection. Lactobacilli are known to provide defence against infection by maintaining an acidic pH and presence of hydrogen peroxide in genital environment. In contrast, BV is a polymicrobial disease resulting in a decreased number of lactobacilli and an increase in pathogenic bacteria, mainly anaerobic or microaerophiles. These organisms include Gardnerella vaginalis, Mobiluncus spp., Bacteroides ,Prevotella spp and Mycoplasma spp. (Hillier, 1993).

Percentage of trichomoniasis in the present study (7%) was lower than the other vaginal infections. However, this prevalence was seemed to be higher than that recorded by (Kadir & Fattah, 2010; Al-Saeed, 2011; Nouraddin and Alsakee, 2015) in Sulaaimania, Dohok and Erbil respectively and consistent with( Khadir and Jerjis, 1999) in Kurkuk, whereas it was lower than that of (Al-Kaisi et al., 2008) in Baghdad as 19.16%, (Khadir et al., 1996) in Erbil as 10% and( Kharofa, 1999) in Mosul as14%. As well as an overall high prevalence of 17.5% was documented by (Iwueze et al., 2014) in Nigeria These differences may be owing to variation in the sample size, diagnostic techniques, environmental hygiene, personal hygiene, socioeconomic level, study population and sexual activity (Wondemagegn et al., 2015).

Regarding VC, the recent result showed 15% as single infection and 40% of both single and mixed infection which seemed to be higher than that recorded in Nigeria 14%; 20%; 36% (Emeribe *et al.*, 2015; Mbim *et al.*, 2017; Olowe *et al.*, 2014) respectively in Iran as 9.3% (Bonyadpour *et al.*, 2016) and in India as 30% (Narayankhedkar *et al.*, 2015). In contrast the present finding was lower than that in Baghdad which was 44.35% (Al-Attraqhchi *et al.*, 2013).

Progesterone and estrogens have direct effects on Candida cells, possibly contributing to VC. One direct effect of the hormones is the stimulation of estrogen and progesterone cytosolic receptors, which have been already identified in several Candida species. An estrogen-binding protein (EBP), that displays high affinity for estradiol and estrone was identified and characterized in C. albicans (Skowronski and Feldman, 1989; Wagner and Johnson, 2012) and an estrogen binding system was also detected in C. glabrata (Powell et al., 1984). In addition, a corticosteroid-binding protein (CBP) that exhibits high affinity for corticosterone and progesterone, but low affinity for estrogens was identified in C. albicans, C. guilliermondii, C. krusei, C. parapsilosis and C. tropicalis (Loose et al., 1983; Skowronski and Feldman, 1989).

Concerning the age group, the most effected one with VC and trichomoniasis was 25- 30 year. These result agreed with previous studies whom concluded that this age group was the most susceptible to infectious vaginitis (Al-akeel et al., 2013; Emeribe et al., 2015; Kadir et al., 2010 Nwadioha et al., 2010; Yadav and Prakash, 2016;). Lowest infection with VC was reported in the age group (46-52) years and this is in accordance with the study of (Al-akeel et al., 2013; Nwadioha et al., 2010) in Saudi Arabia and Nigeria respectively. Several studies explained the factors that responsible for the low percentage of candidiasis infection in old women like the reduced effect of estrogen hormone, reaching menopause, less or no sexual activity, no contraceptive, and also have good vaginal immunity because they have low estrogen and corticoids (Nelson et al., 2013). BV was higher in age group 32-38 years and this present result agree with the finding of (Nzomo et al., 2013) followed by (25-31) years. It is worthwhile to mention these age groups represent the peak of child bearing ages and also may attributed to high sexual activity. Several researchers explained the high frequency of infection of these groups by using of these women drugs indiscriminately and contraceptive to prevent pregnancy (Nelson et al., 2013). Moreover, Okungbowa et al. (2003) concluded that the women within this age group are vulnerable probably due to sexual promiscuity ,drug abuse and use of contraceptives.

Regarding pregnancy as a host related risk factor, the present result showed that trichomoniasis was most prevalent in pregnant women compared to non-pregnant. This finding agreed with the result of (Kadir *et al.*, 2010). In contrast, in Nigeria high percentage was detected in non-pregnant women compared to pregnant one but with non-significant differences (Iwueze *et al.*, 2014). The commensal dominant lactobacilli modulate the pathogenicity of this parasite (Phukan *et al.*, 2013). Recently some researchers proved that *T. vaginalis* is able to lyse T-cell and B-cells in vitro. Moreover they explained that this lysis of lymphocytes was mediated by contact – dependent and soluble factors (Mercer *et al.*, 2016).

Candidiasis in the current study was also documented in pregnant women (66.6%) more than that in nonpregnant group (31.2%). This result was consistent with that of (Al-akeel *et al.*, 2013). Also it was recorded that up to 40% of pregnant women may have vaginal candidiasis globally (Alli *et al.*, 2011; Alo *et al.*, 2012).

The high percentage of vaginal candidiasis in pregnant women has been attributed to the high level of the sex hormones which lead to increase Candida attachment, hyphae formation and decrease vaginal responses (Goncalves *et al.*, 2016). In addition, Olowe *et al.*, (2014) concluded that the incidence of candidiasis is highly increased in recent time among pregnant women particularly due to the widespread use of antibiotics and immunosuppressive drugs. Moreover, Babic and Hukic, (2010) reasoned this by the increasing amount of glycogen in the vagina and raising level of estrogen hormones. This condition provides good sources of carbon, which provoke the growth and germination of *Candida* spp.

Identification of *Candida* species is essential due to the emergence of new pathogenic species and owing to the various antifungal susceptibility profiles. Regarding this aspect the present result showed that *C. albicans* represent 76.4% and the others as 23.5% of the vaginal isolates. This finding agree with the results of other researchers whom concluded that *C. albicans* is the most common cause (80%-90%), whereas other yeasts account for up to 20% of cases. C. galbrata was also identified in the current study in 14.7% which is higher than that mentioned by (Geiger et al., 1995; Ray et al., 2007; Spinillo et al., 1997;) which accounts as 10% of vaginal yeast isolates. Symptomatic vaginitis caused by this organism is associated with less intense itching and dyspareunia (Geiger et al., 1995) than that caused by other Candida species, but the organism may be more difficult to eliminate with standard medications. (Ray et al., 2007). In other study, Nyirjesy (2008) recorded that 70% of complicated cases of recurrent vulvovaginitis are caused by C. albicans. Whereas C. glabrata and C. parapsilosis represent 30%. (Nyirjesy et al., 1995). Al-akeel et al., (2013) found that C. albicans and C. glabrata constitute 80.3 and 12.7% respectively in vaginal specimens. Recently, Goncalves et al., (2016) concluded that in spite of C. albicans is the main causative agent for VC, the identification of non-candida albicans Candida species, particularly C. glabrata, as responsible for this disease, appears to be high.

In addition, the present results identified *C. krusei* and *C. dubliensis* in 5.8% and 2.9% respectively. Whereas other recorded C. *tropicalis* in 1% to 5% and concluded it may be associated with a higher rate of recurrence after standard treatments. (Horowitz *et al.*, 1985; Spinillo *et al.*, 1997).

According to the sensitivity of *Candida* spp to Nystatin, the present data showed that all species seemed to be sensitive for the three concentrations of Nystatin. These results agreed with the studies of (Nelson *et al.*, 2013; Toua *et al.*, 2013) in Kenya and Cameron respectively.

On the other hand, the present result of single BV showed that non- pregnant women were more infected than pregnant group. This result disagree with the result of (Afolabi *et al.*, 2016; Bahram *et al.*, 2009) in Nigeria and Iran respectively. The present findings may be related to the population size which included (21) pregnant women, while non-pregnant women were (64).

In general the recent total mixed infections were detected in pregnant women more than that recorded in non-pregnant ones with significant differences. This finding may be related to the abundant of reproductive hormones in pregnancy. Estrogen is responsible for increasing the vulnerability of pregnant women to vaginitis by reducing the ability of epithelial cells to inhibit the growth of candida beside reduction of the immunoglobulins in the vaginal secretion (Anorlu et al., 2004). Moreover, Progesterone has suppressive effects on the activity of neutrophils (Adad et al., 2001). It is well known that the alteration of the vaginal environment PH toward alkalinity during pregnancy lead to overgrowth of the pathogens responsible for vaginitis due to reducing level of Doderlein bacilli which convert glycogen of the epithelial cells into lactic acid that keep the vagina acidic.

Bacteria were the most prevalent etiological agent that detected in contraceptive user women which reached 71.4% in intrauterine device users. Several researchers pointed out that IUD increase the likelihood of BV in women (Calzolari *et al.*, 2000; Madden *et al.*, 2012), whereas another in recent study found no correlation between the IUD contraception and the alteration of the vaginal microbiota composition. (Bassis *et al.*, 2017). Because of the few number of contraceptive users among the vulvovaginitis women included in the current study, no clear coloration was recognized between the disease and the type of contraceptive.

## Conclusions

Relatively high percentage of women whom enrolled in the current study was infected with the three etiological agents that responsible for vulvovaginitis particularly with bacteria. Routine culture of vaginal discharge must be performed for these patients; also drug susceptibility of bacterial isolates should be determined. Comprehensive healthcare education is needed to reduce this disease among women particularly those of child bearing age. Further studies with large number of infected women will be needed to highlight this important disease, which can be managed successfully.

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#### References

Adad SJ, de Lima RV, Sawan ZT, Silva ML, de Souza MA, Saldanha JC, Falco VA, da Cunha AH, Murta EF. 2001. Frequency of Trichomonas vaginalis, *Candida* spp. and Gardnerella vaginalis in cervical-vaginal smears in four different decades. Sao Paulo Med J. 1;119(6), 200-205.

Afolabi BB, Olusanjo Moses, Oyinlola E, Oduyebo O. 2016. Bacterial Vaginosis and Pregnancy Outcome in Lagos, Nigeria Infect Dis. 2016 Jan (3), 1-17. DOI: 10.1093/ofid/ofw030,

Agnew KJ, Hillier SL. 1995. The effect of treatment regimens for vaginitis and cervicitis on vaginal colonization with lactobacilli. Sex Transm Dis **22**, 269-273.

**Al- Kaisi AAR.** 2008 Trichomoniasis among females with vaginal discharge in Baghdad Medical City. J. Fac. Med. Baghdad, 2008, **50**, 37-41.

**Al-akeel RA, El-kersh TA, Al-Sheikh YA, Al-Ahmadey ZZ.** 2013. Prevalence and comparison for detection methods of Candida species in vaginal specimens from pregnant and non-pregnant Saudi women. Afr J Microbiol Res **7(1)**, 56-65. DOI: 10.5897/AJMR12.1979.

**Al-Attraqhchi AAF, Abbas N, Al-jeboori M.** 2013. Detection of *Candida* spp. responsible for vulvovaginitis in women with contraceptives. Journal of genetic and environmental resources conservation **1(3)**, 254-261.

Alcaide ML, Feaster DJ, Duan R, Cohen S, Diaz C, Castro JG, Golden MR, Henn S, Colfax GN, Metsch LR. 2015. The incidence of *Trichomonas vaginalis* infection in women attending nine sexually transmitted diseases clinics in the USA. Sex Transm Infect.

DOI: 10.1136/sextrans-2015-052010

Alli JAO, Okonko IO, Odu NN, Kolade AF, Nwanze JC. 2011. Detection and prevalence of Candida isolates among patients in Ibadan, Southwestern Nigeria. J. Microb. Biotechnol. Res 1, 176-184.

Allsworth JE, Peipert JF. 2007. Prevalence of bacterial vaginosis: 2001-2004 National Health and Nutrition Examination Survey data. Obstet Gynecol **109(1)**, 114-120.

Alo MN, Anyim C, Onyebuchi AK, Okonkwo EC. 2012. Prevalence of asymptomatic Co Infection of Candidiasis and Vaginal Trichomoniasis among Pregnant Women in Abakaliki, South-Eastern Nigeria. J. Nat. Sci. Res (2), 87-91.

**Al-Saeed WM.** 2011. Detection of *Trichomonas vaginalis* by different methods in women from Dohok province, Eastern Mediterranean Health Journal **17(9)**, 706-709.

**Amsel R, Totten PA, Spiegel CA, Chen KC, Eschenbach D, Holmes KK.** 1983. Nonspecific vaginitis. Diagnostic criteria and microbial and epidemiologic associations. Am J Med **74(1),** 14-22.

Anderson MR, Klink K, Cohrssen A. 2004. Evaluation of vaginal complaints. JAMA **291(11)**, 1368-79.

Anorlu R, Imosemi D, Odunukwe N, Abudu O, Otuonye M. 2004. Prevalence of HIV among women with vaginal discharge in a gynecological clinic. J Natl Med Assoc. 2004 Mar **96(3)**, 367-71.

**Babic M, Hukic M.** 2010. *Candida albicans* and non-*albicans* species as etiological agent of vaginitis in pregnant and non-pregnant women. Bosn J Basic Med Sci **10(1)**, 89-97.

**Bahram A, Hamid B, ZohreT.** 2009. Prevalence of bacterial vaginosis and impact of genital hygiene practices in non-pregnant women in Zanjan, Iran, Oman Medical Journal **24(4)**, 288-293. Bassisa CM, Allsworth JE, Wahl HN, Sack DE, Young VB, Bell JD. 2017. Effects of intrauterine contraception on the vaginal microbiota. Contraception **96**, 189-195.

**Bitew A, Abebaw Y, Bekele D, Mihret A.** 2017. Prevalence of bacterial vaginosis and associated risk factors among women complaining of genital tract infection. Int. J. Microbial 1-8. DOI: ORG/10.1155/ 2017/4919404

**Bonyadpour B, Akbarzadeh M, Mohagheghzadeh A.** 2016. A Descriptive study on the prevalence of vulvovaginal infections and speciesspecific distribution of vulvovaginal candidiasis in married women of the south of Iran. J. Midwifery & Reproductive health **4(4)**, 741-747.

**Boyle DC, Smith JR.** 1999. Infection and cervical intraepithelial neoplasia. Int J Gynecol Cancer **9**, 177-186. pmid:11240764e

Calzolari E, Masciangelo R, Milite V, Verteramo R. 2000. Bacterial vaginosis and contraceptive methods. Int J Gynecol Obstet **70**, 341-6.

**Carey JC, Klebanoff MA.** 2005. Is a change in the vaginal flora associated with an increased risk of preterm birth? Am J Obstetr Gynecol **192(4)**, 1341-1346.

**Cheesbrough M.** 2006. District laboratory practice in tropical countries. 2<sup>nd</sup> edition, part **2**, 243. Cambridge university press.

**Emeribe AU, Nasir IA, Onyia J, Ifunanya AL.** 2015. Prevalence of Vulvovaginal candidiasis among nonpregnant women attending a tertiary health care facility in Abuja, Nigeria. Research and Reports in Tropical Medicine **6**, 37-42.

Gallo MF, Macaluso M, Warner L, Fleenor ME, Hook III EW, Brill I, *et al.* 2012. Bacterial vaginosis, gonorrhea, and chlamydial infection among women attending a sexually transmitted disease clinic: a longitudinal analysis of possible causal links. Ann Epidemiol **22(3)**, 213-20.

**Garcia LS, Ash LR.** 1979. Diagnostic parasitology. 2<sup>nd</sup> edition, Mosby Company.

**Geiger AM, Foxman B, Sobel JD.** 1995. Chronic vulvovaginal candidiasis: characteristics of women with *Candida albicans*, *C glabrata* and no Candida. Genitourin Med **71**, 304-307.

Gerbase AC, Rowley JT, Heyman HL, Berkley SFB, Piot P. 1998. Global prevalence and incidence estimates of selected curable STDs. Sex Trans Inf 74, S12-S16.

Goncalves B, Ferreira C, Alves CT, Henriques M, Azeredo J, Silva S. 2016. Vulvovaginal candidiasis: Epidemiology, microbiology and risk factors. Crit Rev Microbiol **42(6)**, 905-927. DOI: 10.3109/1040841X.2015.1091805.

**Gram IT, Macaluso M, Churchill J, Stalsberg H.** 1992. *Trichomonas vaginalis* (TV) and human papillomavirus (HPV) infection and the incidence of cervical intraepithelial neoplasia (CIN) grade III. Cancer Causes Control **3**, 231-236.

**Gupta K, Stapleton AE, Hooton TM, Roberts PL, Fennell CL, Stamm WE.** 1998. Inverse association of H2O2-producing lactobacilli and vaginal Escherichia coli colonization in women with recurrent urinary tract infections. J Infect Dis **178**, 446-450.

**Hillier SL.** 1993. Diagnostic microbiology of bacterial vaginosis. Am J Obstet Gynecol **169**, 455e9.

**Hirt RP, Sherrard J.** 2015. *Trichomonas vaginalis* origins, molecular pathobiology and clinical considerations. Curr Opin Infect Dis **28**, 72-79.

Horowitz BJ, Edelstein SW, Lippman L. 1985. *Candida tropicalis* vulvovaginitis .Obstet Gynecol **66**, 229-232.

Iwueze MO, Ezeanyanwu LN, Okafor FC, Nwaorgu OC, Ukibe SC. 2014. Prevalence of *Trichomonas vaginalis* infection among women attending hospitals/health centres in Onitsha community, Onitsha North Local Government Area of Anambra State. Bioscientist **2**, 54-64. **Kadir MA, Fattah COD.** 2010. Trichomonas vaginalis among women in Sulaimania Governorate-Iraq. Tikrit J pharmaceutical .science **6**, 1-9.

Kadir MA, Jerjis KJ. 1999. Incidence of trichomoniasis in Kirkuk city. Journal of the Faculty of Medicine, Baghdad **28(2)**, 75-79. 15.

Kadir MA, Salehy A, Hamed EE. 1996. Studies on Trichomonas vaginalis in Erbil teaching hospital. Journal of the Faculty of Medicine, Baghdad **23(1)**, 83-88. 16.

Kadir MA, Sulyman MA, Dawood IS, Shams-Eldin S. 2014. *Trichomonas vaginalis* and associated microorganisms in women with vaginal discharge in Kerkuk-Iraq. Ankara Medical journal 14(3), 91-99. DOI: 10.17098/amj.47284.

**Kharofa WA.** 1999. An epidemiological study and cultivation of Trichomonas vaginalis in Mosul city MSc thesis. Department of Microbiology, College of Medicine, University of Mosul, Mosul, Iraq.

**Klebanoff SJ, Hillier SL, Eschenbach DA, Waltersdorph DA.** 1991. Control of the microbial flora of the vagina by H202-Generating Lactobacilli. J Infect Dis **164**, 94-100.

Lai SK, Hida K, Shukair S, Wang Y-Y, Figueiredo A, Cone R, Hope TJ, Hanes J. 2009. Human immunodeficiency virus type 1 is trapped by acidic but not by neutralized human cervicovaginal mucus. J Virol **83(21)**, 11196-11200. DOI: 10.1128/JVI.01899-08

Loose DS, Stevens D, Schurman DJ, Feldman D. 1983. Distribution of a corticosteroid-binding protein inCandida and other fungal genera.J Gen Microbiol **129**, 2379-85.

**Loveless M, Myiant O.** 2018. Vulvovaginitispresentation of more common problems in pediatric and adolescent gynecology. 48,14-27. DOI: 10.1016/j.bpobgyn. Madden T, Grentzer JM, Secura GM, Allsworth JE, Peipert JF. 2012. Risk of bacterial vaginosis in users of the intrauterine device: a longitudinal study. Sexually Transmitted Diseases 39,217–222

**Malone MA.** 2017. Vulvovaginitis in: Bope ET, Kellerman RD, (Eds.), Conn's Current Therapy. Elsevier, USA pp.1092.

Maroszyńska M, Kunicka-Styczyńska A, Rajkowska K, Maroszyńska I. 2013. Antibiotics sensitivity of Candida clinical and food-borne isolates. Acta Biochim Pol **60**, 719-724.

Martin HL, Richardson BA, Nyange PM, Lavreys L, Hillier SL, Chohan B, Mandaliya K, Ndinya-Achola JO, Bwayo J, Kreiss J. 1999 .Vaginal lactobacilli, microbial flora, and risk of human immunodeficiency virus type 1 and sexually transmitted disease acquisition. J Infect Dis **180**, 1863-1868.

**Mbim EN, Mboto CI, George UE, Umego CF, Edet UO, Orajiaka NA.** 2017. Prevalence of Vaginal Candidiasis among Female Students of a Hostel in the University of Calabar, Calabar. J Applied Life Sciences International **13(3)**, 1-7.

**Mc Cormack WM, Augenbraun MH.** 2015. Vulvovaginitis and Cervicitis. Mandell, Douglas, and Bennett's Principles and Practice of Infectious Diseases, Updated Edition **110**, 1358-1371.

Meites E. 2013. Trichomoniasis .The "Neglected" Sexually Transmitted Disease. Infect Dis Clin North Am **27(4)**, 755-764. DOI: 10.1016/j.idc.2013.06.003

Mercer F, Diala FGI, Chen YP, Molgora BM, Ng SH, Johnson PJ. 2016. Leukocyte Lysis and Cytokine Induction by the Human Sexually Transmitted Parasite *Trichomonas vaginalis*. PloS Negl Trop Dis **10**, 1-19. DOI: 10.1371/journal.pntd.0004913

Mills BB. 2017. Vaginitis: Beyond the Basics. Obstet Gynecol Clin N Am 44, 159-177. DOI: doi.org/10.1016/ j.ogc.2017.02.010.

**Mulley AG.** 2000. Approach to the patient with a vaginal discharge. In: Goroll AH, Mulley AG, editors. Primary Care medicine: office evaluation and management of the adult patient. Philadelphia: Lippincott Williams & Wilkins p. 702-7.

Narayankhedar A, Hodiwala A, Mane A. 2015. Clinicoetiological Characterization of Infectious Vaginitis amongst Women of Reproductive Age Group from Navi Mumbai, India. J Sex Transm Dis 817092.

DOI: 10.1155/2015/817092

**Nelson M, Wanjiru W, Margaret MW.** 2013. Prevalence of vaginal candidiasis and determination of the occurrence of Candida species in pregnant women attending the antenatal clinic of Thika District Hospital, Kenya. Open J Med Microbiol **3**, 264-72.

**Nouraddin AS, Alsakee HM.** 2015 . Prevalence Of *Trichomonas Vaginalis* Infection Among Women In Erbil Governorate, Northern Iraq: An Epidemiological Approach. European Scientific Journal August 2015 edition vol. **11**, No. 24.243-254.

**Nugent RP, Krohn MA, Hillier SI.** 1991. Reliability of diagnosing bacterial vaginosis is improved by a standardized method of Gram stain interpretation. J Clin Microbiol **29**, pp. 297-301.

**Nwadioha SI, Egah DZ, Alao OO, Iheanacho E.** 2010. Risk factors for vaginal candidiasis among women attending primary health care centers of Jos, Nigeria. J Clin Med Res **2**, 110-13.

Nyirjesy P, Seeney SM, Grody MH, Jordan CA, Buckley HR. 1995 .Chronic fungal vaginitis: the value of cultures. Am J Obstet Gynecol **173**, 820-3. Nyirjesy P. 2008. Vulvovaginal candidiasis and bacterial vaginosis. Infect Dis Clin North Am **22**, 637-52, vi.

Nyirjesy P. 2014. Management of persistent vaginitis. Obstet Gynecol **124(6)**,1135-46.

Nzomo J, Waiyaki P, Waihenya R. 2013. Bacterial vaginosis and correlates in women of reproductive age in Thika, Kenya. Advances in Microbiol, 2013, **3**, 249-254. **Okungbowa FI, Isikhuemhen OS, Dede APO.** 2003. The distribution frequency of Candida species in the genitourinary tract among symptomatic individuals in Nigerian cities. Rev Iberoam Micol **20**, 60-3.

**Olowe O, Makanjuola O, Olowe R, Adekanle D.** 2014. Prevalence of vulvovaginal candidiasis, trichomoniasis and bacterial vaginosis among pregnant women receiving antenatal care in Southwestern Nigeria. European Journal of Microbiology and immunology **4(4)**, 193-197. DOI: 10.1556/EUJMI-D-14-00027.

Pham AT, Kives S, Merovitz L, Nitsch R, Tessler K, Yudin MH, *et al.* 2012. Screening for bacterial vaginosis at the time of intrauterine contraceptive device insertion. J Obstet Gynaecol Can **34**, 179-185.

Phukan N, Parsamand T, Brooks AE, Nguyen TN, Simoes-Barbosa A. 2013. The adherence of Trichomonas vaginalis to host ectocervical cells is influenced by lactobacilli. Sex Transm Infect **89**, 455-459. DOI: 10.1136/sextrans-2013-051039.

**Powell BL, Frey CL, Drutz DJ.** 1984. Identification of a 17b-estradiol binding protein in Candida albicans and Candida (Torulopsis) glabrata. Exp Mycol **8**, 304-13.

Ravel J, Gajer P, Abdo Z, Schneider GM, Koenig SSK, McCulle SL *et al.* 2011. Vaginal microbiome of reproductive-age women. Proc Natl Acad Sci US A **108**, 4680-4687.

Ray D, Goswami R, Banerjee U, *et al.* 2007. Prevalence of Diabetes Care **30**, pp. 312-317.

SatterwhiteCL, TorroneE, MeitesE, DunneEF, MahajanR, OcfemiaMC, SuJ, XuF, Weinstock H. 2013.Sexually transmitted infectionsamong US women and men:prevalence and incidenceestimates.Sex Transm Dis 40(3), 187-93.DOI:10.1097/OLQ.ob013e318286bb53

Schwebke JR, Burgess D. 2004. Trichomoniasis. Clin Microbiol Rev 17, 794-803.

Secor WE, Meites E, Star MC, Workowski KA. 2014. Neglected Parasitic Infections: Trichomoniasis. Am J Trop Med Hyg **90**, 800-804. Skowronski R, Feldman D. 1989. Characterization of an estrogen-binding protein in the yeast Candida albicans. Endocrinology **124**, 1965-1972.

**Sobel JD.** 2007. Vulvovaginal candidosis. Lancet **369**, 1961-1971.

**Spinillo A, Capuzzo E, Gulminetti R, Marone P, Colonna L, Piazzi G.** 1997Prevalence of and risk factors for fungal vaginitis caused by non-albicans species. Am J Obstet Gynecol **176**, 138-141.

Taha TE, Hoover DR, Dallabetta GA, KumwendaNI, MtimavalyeLA, YangLP, LiombaGN, BroadheadRL, Chiphangwi JD, MiottiPG.1998. Bacterial vaginosis and disturbances of vaginalflora: association with increased acquisition of HIV.AIDS 12(13), 1699-1706.

TamuraNK, NegriMF, BonassoliLA,Svidzinski TI. 2007.Virulence factors for Candidaspprecovered from intravascular catheters andhospital workers hands.Rev Soc Bras Med Trop.Jan-Feb 40(1), 91-93.

**Tellapragada C, Eshwara VK, Bhat P, Kamath A, Aletty S, Mukhopadhyay C.** 2017. Screening of vulvovaginal infections during pregnancy in resource constrained settings: Implications on preterm delivery. J Infect Public Health. 2017; **10(4)**, 431-437. DOI: 10.1016/j.jiph.2016.06.003.

Toua V, Djaouda M, Gaké B, Menye DE, Christie EA, Tambe E, Akindoh VV, Njiné T. 2013. Prevalence of Vulvovaginal candidiasis amongst pregnant women in Maroua (Cameroon) and the sensitivity of Candida albicans to extracts of six locally used antifungal plants. International Research Journal of Microbiology (IRJM) **4**, 89-97.

**Vaginitis.** 2006. ACOG Practice Bulletin No. 72. American College of Obstetricians and Gynecologists. Obstet Gynecol **107**, pp. 1195-1206. Wagner RD, Johnson SJ. 2012. Probiotic Lactobacillus and estrogen effects on vaginal epithelial gene expression responses to *Candida albicans*. J Biomed Sci **19**, 58.

DOI: 10.1186/1423-0127-19-58.

**Wei Y, Feng J, Luo Z.** 2010. Isolation and genotyping of vaginal non-albicans *Candida* spp. in women from two different ethnic groups in Lanzhou China. Int J Gynecol Obstetrics **110**, 227-230.

Wiesenfeld HC, Hillier SL, Krohn MA, Landers DV, Sweet RL. 2003. Bacterial vaginosis is a strong predictor of Neisseria gonorrhoeae and Chlamydia trachomatis infection. Clinical Infect Dis **36**, 663-668.

**Wondemagegn M, Yimer M, Zenebe Y, Abera B.** 2015. Common causes of vaginal infections and antibiotic susceptibility of aerobic bacterial isolates in women of reproductive age attending at Felegehiwot referral Hospital, Ethiopia: a cross sectional study, BMC Women's Health 15(42), 1-9.

Workowski KA, Bolan GA. 2015. Centers for Disease Control and Prevention: Sexually transmitted diseases treatment guidelines, MMWR Recomm Rep **64**, 1-137.

Yadav K, Prakash S. 2016. Prevalence of vulvovaginal candidiasis in pregnancy. Glob. J. Med. Med. Sci **4(1)**, 108-116.

Yazar S, Dagci H,Aksoy U, Ustun S, Akisu C, Mucide AK, Daldal N. 2002. Frequency of *Trichomonas vaginalis* among women having vaginal discharge in Izmir, Turkey. Inonu. Univ. Tip. Fakut. Derg **9**, 159-161.

Yudin MH, Money DM. 2017. Screening and Management Bacterial Vaginosis in Pregnancy. J Obstet Gynaecol Can Aug **39(8)**, e184-e191. DOI: 10.1016/j.jogc.2017.04.018