

International Journal of Biosciences | IJB | ISSN: 2220-6655 (Print), 2222-5234 (Online) http://www.innspub.net Vol. 13, No. 1, p. 36-41, 2018

**RESEARCH PAPER** 

**OPEN ACCESS** 

Effect of chloride and sulfate salinity on in vitro regeneration

of rice

Md. Riyadh Arefin<sup>\*1</sup>, Mohammad Mehfuz Hasan Saikat<sup>2</sup>, Mr. Umakanta Sarker<sup>2</sup>, M. Abdul Karim<sup>3</sup>, Md. Sadiqur Rahman<sup>4</sup>

<sup>1</sup>Botany Division, Bangladesh Tea Research Institute, Srimangal, Moulvibazar, Bangladesh <sup>2</sup>Department of Genetics and Plant Breeding, Bangabandhu Sheikh Mujibur Rahman Agricultural University (BSMRAU), Gazipur, Bangladesh <sup>3</sup>Department of Agronomy, Bangabandhu Sheikh Mujibur Rahman Agricultural University (BSMRAU), Gazipur, Bangladesh <sup>4</sup>Seed Technology Discipline, Bangladesh Agricultural Research Institute (BARI), Akbarpur, Moulvibazar, Bangladesh

**Key words:** Rice, Chloride (Cl<sup>-</sup>), Sulphate (SO<sub>4</sub><sup>2-</sup>), *In vitro*, Regeneration

http://dx.doi.org/10.12692/ijb/13.1.36-41

Article published on July 15, 2018

# Abstract

Rice is the seed of the monocot plants *Oryza sativa* (Asian rice) or *Oryza glaberrima* (African rice). As a cereal grain, a great source of protein; it is the most widely consumed staple food for a large part of the world's human population, especially in Asia. Among the various factors limiting rice yield, salinity is one of the oldest and most serious environmental problems in the world. Rice yield can be reduced by up to 50% when grown under moderate (6dS/m) salinity level. Salinity in aquatic ecosystem is determined by the total amount of dissolved salts present in it. Four types of cations prevail in saline water such as Sodium (Na<sup>+</sup>), Potassium (K<sup>+</sup>), Calcium (Ca<sup>++</sup>) and Magnesium (Mg<sup>++</sup>) and 3 anions such as Chloride (Cl<sup>-</sup>), Carbonate (CO<sub>3</sub><sup>-</sup>) or Bi-Carbonate (HCO<sub>3</sub><sup>-</sup>) and Sulphate (SO<sub>4</sub><sup>2-</sup>). Even though Cl is an essential element for plant growth, but higher concentration may restrain plant growth or cause toxicity to some plants. Higher concentration of Chloride ions also causes leaf tip burn, disruption of membrane function, obstruction in internal solute balance that hampers nutrient uptake. On the other hand, Sulphur is an important factor in plant feeding but high sulphate concentrations may affect plant development and crop yield. In NaCl and Na<sub>2</sub>SO<sub>4</sub> salts, Sodium (Na<sup>+</sup>) is common but differs only by two anions: Chloride (Cl<sup>-</sup>) and Sulphate (SO<sub>4</sub><sup>2-</sup>). The present investigation was conducted through *in vitro* regeneration protocol of ten rice genotypes to observe the effect of these two salts and to identify which salt causes more inhibitor.

\* Corresponding Author: Md. Riyadh Arefin $\boxtimes$ riyadh.<br/>btri@gmail.com

#### Introduction

Rice is the seed of the monocot plants *Oryza sativa* (Asian rice) or *Oryza glaberrima* (African rice). As a cereal grain, a great source of protein; it is the most widely consumed staple food for a large part of the world's human population, especially in Asia (Datta *et al.*, 2003).

Among the various factors limiting rice yield, salinity is one of the oldest and most serious environmental problems in the world (Islam et al., 2007). Rice yield can be reduced by upto 50% when grown under moderate (6dS/m) salinity level (Zhang *et al.*, 2002). Salinity in aquatic ecosystem is determined by the total amount of dissolved salts present in it. Four types of cations prevail in saline water such as Sodium (Na<sup>+</sup>), Potassium (K<sup>+</sup>), Calcium (Ca<sup>++</sup>) and Magnesium (Mg<sup>++</sup>) and 3 anions such as Chloride (Cl<sup>-</sup>), Carbonate (CO<sub>3</sub><sup>-</sup>) or Bi-Carbonate (HCO<sub>3</sub><sup>-</sup>) and Sulphate (SO<sub>4<sup>2-</sup></sub>) (Baten *et al.*, 2015). Even though Cl is an essential element for plant growth, but higher concentration may restrain plant growth or cause toxicity to some plants. Higher concentration of Chloride ions also causes leaf tip burn, disruption of membrane function, obstruction in internal solute balance that hampers nutrient uptake (Lantzke, 2004). On the other hand, Sulphur is an important factor in plant feeding but high sulphate concentrations may affect plant development and crop yield (Kowalska, 2005).

In NaCl and Na<sub>2</sub>SO<sub>4</sub> salts, Sodium (Na<sup>+</sup>) is common but differs only by two anions: Chloride (Cl<sup>-</sup>) and Sulphate (SO<sub>4</sub><sup>2–</sup>). The present investigation was conducted through *in vitro* regeneration protocol of ten rice genotypes to observe the effect of these two salts and to identify which salt causes more inhibitor.

## Materials and methods

#### Experimental location and materials

The experiment was conducted in the Advanced Plant Breeding Laboratory, Department of Genetics and Plant Breeding, Bangabandhu Sheikh Mujibur Rahman Agricultural University (BSMRAU), Salna, Gazipur-1706, Bangladesh. The following rice genotypes including a salt tolerant variety (BINA 8) were used as experimental materials in the present investigation.

### Salt concentrations and media preparation

The healthy, disease free and dehusked seeds were used as explants for the study. MS (Murashige and Skoog, 1962) medium supplemented with two salts (NaCl and Na<sub>2</sub>SO<sub>4</sub>) was used for callus induction and the callus pieces at appropriate size were transferred to MS regeneration medium.

For callus induction, the MS (Murashige and Skoog, 1962) medium supplemented with 2, 4-D (2mg/l), sucrose (30g/l), agar (10g/l) and six levels of salt concentrations was taken for each salt. For plant regeneration the MS (Murashige and Skoog, 1962) medium supplemented with sucrose (30g/l), agar (log/1), NAA (l mg/L), BAP (1 mg/l) and six levels of salt concentration was used for each salt. Six levels of NaCl and Na<sub>2</sub>So<sub>4</sub> salt solution (0, 0.1, 0.2, 0.4, 0.6 and 0.8%) were used in this experiment to observe the effect of salts.

#### Callus induction and regeneration

The dehusked seeds were sterilized and the seeds were air dried in the Laminar Air Flow cabinet for ten minutes and were transferred on callus induction media. This process was done for each genotype. The culture was inoculated with the treated seeds in the dark at  $25 \pm 2^{\circ}$ C for callus induction. After 3-6 weeks of inoculation, seeds of the responsive varieties started to produce callus. Callus induction frequency was calculated on the basis of the number of seeds producing callus. Some seeds produced more than one callus, but for this calculation, all calli originating from one were considered as one. Calli with a size of at least 2 mm were transferred to regeneration medium (MS basal semisolid medium), and were incubated in a temperature controlled growth room at  $25 \pm 2^{\circ}$ C under 16-hour light photoperiod with a light intensity of about 2000-3000 lux for plant regeneration. Day to day observations was carried out to note the response. Regenerated plants were counted on the basis of the number of callus producing plantlets.

# Int. J. Biosci.

## Statistical analysis of data

The data for the characters under study were statistically analyzed wherever applicable. The experiment was conducted in growth room and arranged in Factorial Completely Randomized Design (CRD) with three replications. The Analyses of Variance for different characters were preformatted and means were compared by the Duncan's Multiple Range Test (DMRT).

To observe the effect of different treatments of the experiment, data were collected on the following parameters:

Percent callus induction =	$\frac{\text{Number of seeds induced calli}}{\text{Number of seeds incubated}} X \ 100$

 $Percent plant regeneration = \frac{Number of calli with plantlets}{Number of incubated calli} X 100$ 

Table 1. Rice genotypes used in this experiment.

## Results

### Effect of genotype and salt

Different rice cultivars responded differently to various levels of NaCl and Na<sub>2</sub>SO<sub>4</sub> salt. For both NaCl and Na<sub>2</sub>SO<sub>4</sub> salts, BINA 8 gave the highest response (Table 2).

The lowest callus induction response was observed in Binaphul (54.67%) with NaCl and BR11 (57.29) with Na<sub>2</sub>SO<sub>4</sub>. Plant regeneration were also effected by different rice genotypes and salts (Table 3).

In case of NaCl, the highest response was given by BINA 8 (38.27) while others gave statistically similar results. Again, BINA 8 (38.92) responded very well in Na<sub>2</sub>SO<sub>4</sub> salt, while BRRI dhan51 (26.51) and Gopalbhog (25.74) showed poorest response which were statistically similar.

Sl. No.	Rice genotypes	Sl. No.	Rice genotypes	
1.	BR10	6.	BINA 8	
2.	BR11	7.	Uknimadhu	
3.	BRRI dhan49	8.	Binaphul	
4.	BRRI dhan51	9.	Gopalbhog	
5.	BRRI dhan52	10.	Straw	

# Effect of salt and salt concentrations

Responses related to salt x salt concentration interaction are presented in Table 4. Percentage of callus induction was the highest (97.93%) at no salt concentration media. The lowest callus induction (1% & 2.7%) was found at 0.80% with both NaCl and Na<sub>2</sub>SO<sub>4</sub>.

Table 2. Effect of different rice genotypes and salts on callus induction (%).

Variety/genotype	NaCl	$Na_2SO_4$
BR10	59.75 ab	62.25 ab
BR11	54.79 b	57.29 c
BRRI dhan49	57 b	60.33 bc
BRRI dhan51	55.83 b	59.17 bc
BRRI dhan52	57 b	62 abc
BINA 8	65 a	66.17 a
Uknimadhu	58 b	62.17 ab
Gopalbhog	57.17 b	60.5 bc
Binaphul	54.67 b	62.17 ab
Straw	59.83 ab	59 bc
LSD at 5% level of significance	5.0987*	4.7918*

\*Significant at 5% level of significance

Table followed by the same letter in a column do not differ significantly by DMRT.

Again, highest plant regeneration percentage (77.05%) was recorded at no salt concentration in both NaCl and Na<sub>2</sub>SO<sub>4</sub> while the lowest was recorded (0.15%) at 0.80 % salt concentration in both NaCl and Na<sub>2</sub>SO<sub>4</sub> supplemented MS media.

# Effect of salt

The highest callus induction (61.10%) and plant regeneration (30.21%) were observed in the medium containing Na<sub>2</sub>So<sub>4</sub> while the lowest callus induction

(57.90%) and plant regeneration (29.17%) were observed in the NaCl containing media (Table 5).

## Discussion

### Effect of genotype and salt

The best performer variety in terms of plant regeneration and callus induction was found on  $Na_2So_4$  supplemented media. Ping *et al.* (2006) examined NaCl tolerance by observing callus growth state, germination rate and seedling height under solid medium in 13 genotypes of rice.

Table 3. Effect of different rice genotypes and salts on plant regeneration (%).

Variety/ genotype	NaCl	$Na_2SO_4$
BR10	30.28 b	31.68 abc
BR11	30.10 b	31.37 abc
BRRI dhan49	28.92 b	28.80 bc
BRRI dhan51	26.52 b	26.51 c
BRRI dhan52	30.85 b	34.13 ab
BINA 8	38.27 a	38.92 a
Uknimadhu	27.06 b	29.61 bc
Gopalbhog	25.78 b	25.74 c
Binaphul	27.31 b	27.87 bc
Straw	26.57 b	27.47 bc
LSD at 5% level of significance	6.8324*	7.4905*

\*Significant at 5% level of significance

Table followed by the same letter in a column do not differ significantly by DMRT.

The results showed that NaCl tolerance was different among different genotypes of rice. Muhammad *et al.* (2013) selected salt-tolerant soma clone from 3 local rice varieties (Rajashail, Katicota, and BRRI-22). The best responded regarding the callus induction frequency, the regeneration frequency and the number of shoot per callus were obtained from Rajashail and Katicota among three varieties. Qamaruz *et al.* (2011) found two genotypes MR219 and MR219 consistently performed the best in both callus culture (93.51 and 92.22%) and plant regeneration capacity (27.03 and 26.34%), respectively.

Treatment		Callus induction (%)	Plant regeneration (%)
Salt	Salt concentrations (%)		
NaCl	0	97.93 a	77.05 a
	0.10	89.00 b	42.18 b
	0.20	73.00 c	36.76 c
	0.40	51.50 d	17.91 d
	0.60	35.00 e	1.00 e
	0.80	1.00 f	0.15 e
	LSD at 5% level of significance	3.9494*	5.2924*
$Na_2SO_4$	0	97.93 a	77.05 a
	0.10	92.50 b	45.70 b
	0.20	78.00 c	36.76 c
	0.40	55.00 d	19.35 d
	0.60	40.50 e	2.25 e
	0.80	2.70 f	0.15 f
	LSD at 5% level of significance	3.7117*	5.8026*

\*Significant at 5% level of significance

Table followed by the same letter in a column do not differ significantly by DMRT

Similarly, in this experiment, BINA 8 performed very well in both NaCl and  $Na_2SO_4$  containing media because it is a salt tolerant variety.

### Effect of salt and salt concentrations

Adil *et al.* (2009) found a reverse relationship between salt and salt concentration. Nahar *et al.* (2013) studied to evaluate the effect of salt stress on two rice genotypes (BRRI dhan38 and Chini Kanai). Plant regeneration of BRRI Dhan38 was 80% at 0 mM NaCl, but decreased to 20% at 100 mM NaCl. There was no plant regeneration at 150 mM NaCl for BRRI dhan38 and Chini Kanai respectively. Puspasree (2013) reported that calluses were subjected to NaCl stress at 5 levels (0.2, 0.4, 0.6, 0.8 and 1.0%) for 3 successive passages. Among the cultivars, IR 50 showed the maximum percentage of callus induction (92.0) and Dasal showed the minimum (78.5).

Table 5. Mean effect of different salts on c	allus in	duction (%)	and pl	lant regeneration	(%)	) in rice genotypes.
--	----------	-------------	--------	-------------------	-----	----------------------

Treat	ment	Callus induction (%)	Plant regeneration (%)
	NaCl	57.9 b	29.17 b
	Na <sub>2</sub> SO <sub>4</sub>	61.1 a	30.21 a
Salt	LSD at 5% level of significance	3.0368*	1.0268*

\*Significant at 5% level of significance

Table followed by the same letter in a column do not differ significantly by DMRT.

There was a decrease in callus score with increase in salt concentration and number of passages. Forkan *et al.* (2013) reported that callus was grown on agar solidified media containing 0.2, 0.4, 0.6, 0.8 and 1.0% Na<sub>2</sub>SO<sub>4</sub> salt. The callus growth decreased with increasing Na<sub>2</sub>SO<sub>4</sub> concentration in the medium. Generally, saline condition decreases the growth rate of rice plant. Again, amongst different doses of salt concentration, the highest dose most adversely affects the growth. It was clearly observed from the Table 4 that, increasing level of salt concentration decrease the response of callus induction and plant regeneration both.

### Effect of salt

Na<sub>2</sub>So<sub>4</sub> gave better response for *in vitro* plant regeneration of rice than NaCl salinity. Two salts (NaCl & Na<sub>2</sub>So<sub>4</sub>) were used to see their inhibitory differences in this experiment. Both callus induction and plant regeneration were affected by NaCl and Na<sub>2</sub>So<sub>4</sub>. Among the two salts, Na<sup>+</sup> is common in NaCl and Na<sub>2</sub>So<sub>4</sub> but differ only by two anions: Chloride (Cl<sup>-</sup>) and Sulphate (SO<sub>4</sub><sup>2-</sup>). So, Cl<sup>-</sup> containing salts are more inhibitory in *in vitro* response of rice while better response can be expected from Na<sub>2</sub>So<sub>4</sub> salt.

#### Conclusion

The response of callus induction and plant regeneration were decreased with the increasing level of salt concentration. Being a salt tolerant variety, BINA 8 gave best response in both NaCl and Na<sub>2</sub>SO<sub>4</sub>. But individual effect of NaCl and Na<sub>2</sub>SO<sub>4</sub> on callus induction and plant regeneration was different. The responses were least affected in the medium containing Na<sub>2</sub>SO<sub>4</sub>. It thus appears that Cl<sup>-</sup> containing salts are more inhibitory in *in vitro* response of rice.

#### References

DattaK,Baisakh N, Oliva N, Torrizo L, Abrigo E, Tan L, Rai M,Rehana S, Beyer P, Potrykus I, Datta SK. 2003. Bioengineered golden indicarice cultivars with  $\beta$  carotene metabolism in the endosperm with hygromycin and mannose selection systems. Journal of Plant Biotechnology 1(2), 81-90.

https://doi.org/10.1046/j.1467-7652.2003.00015.x

**Islam M. Z, Mia MAB, Akter A, Rahman MH.** 2007. Biochemical attributes of mutant rice under different saline levels. International Journal of Sustainable Crop Production **2(3)**, 17-21.

# Int. J. Biosci.

Zhang L, Xie CH, Li WF. 2002. Studies on tissue of mature embryo in rice. Hybrid- Rice 17(2), 44-46.

**Baten MA, Seal L, Lisa KS.** 2015. Salinity intrusion in interior coast of Bangladesh: Challenges to agriculture in south-central coastal zone. American Journal of Climate Change **4(3)**, 248.

http://dx.doi.org/10.4236/ajcc.2015.43020

Lantzke N, Calder T. 2004. Water salinity and crop irrigation. Farmnote **34**, 2004.

Kowalska I. 2005. Effects of sulphate level in the nutrient solution on plant growth and sulphur content in tomato plants. Folia Horticulturae 17(1), 91-100.

**Murashige T, Skoog F.** 1962. A revised medium for rapid growth and bioassays with tobacco tissue culture. Physiologia Plantarum **15(3)**, 473-497.

**Ping W, Gang W, Jing J.** 2006. NaCl tolerance of different genotypes in rice under tissue culture. Journal of Jilin Agricultural University **28(6)**, 601-605.

**Muhammad HM, Azdi ZA, Prodhan SH.** 2013. Development of callus initiation and regeneration system of different indigenous *Indica* rice varieties. Journal of Biology **1(2)**, 46-51. **Qamaruz A, Muhammad FZ, Zain AM, Maziah M, Ling HC, Hatwe NN.** 2011. Responses of some selected Malaysian rice genotypes to callus induction under *in vitro* salt stress. African Journal of Biotechnology **10(3)**, 350-362. https://doi.org/10.5897/AJB10.097

Adil M, Hasan M, Kamal MM, Zakaria M, Mian MAK. 2009. In vitro plant regeneration of aromatic rice under salt stress. International Journal of Biological Research **6(6)**, 48-52.

Nahar AM, Krishnan SR, Ramesh M. 2013. Rapid regeneration and ploidy stability of 'cv IR36' indica rice (*Oryza Sativa*. L) confers efficient protocol for in vitro callus organogenesis and *Agrobacterium tumefaciens* mediated transformation. Botanical Studies **54(1)**, 47-48.

https://doi.org/10.1186/1999-3110-54-47

**Puspasree P, Ebrahimali AK, Binyamin A.** 2013. Protocol optimization and evaluation of rice varieties response to *in vitro* regeneration. Advances in Bioscience and Biotechnology **4(5)**, 7-9. http://dx.doi.org/10.4236/abb.2013.45085

**Forkan MA, Rudra B, Khaleda L.** 2013. Screening of salt tolerant potentiality and development of *in vitro* tissue culture system for some local rice varieties. The International Journal of Biotechnology **2(12)**, 193-195.