

International Journal of Biosciences | IJB | ISSN: 2220-6655 (Print), 2222-5234 (Online) http://www.innspub.net Vol. 19, No. 5, p. 10-26, 2021

OPEN ACCESS

Anogeissus leiocarpus (DC.) Guill. & Perr. (Combretaceae), a medicinal plant traditionally used in small ruminant breeding in West and Central Africa: zootechnical performances, pharmacological activities and chemical compositions (bibliography synthesis)

Julienne Kuiseu^{1,3}, Tony Taofick Babalola Abiodoun Sounkere¹, Pascal Abiodoun Olounlade^{1,2*}, Claude Gbemeho Houssoukpe¹, Basile Saka Boni Konmy¹, Fréjus Tanguy Ablo Zinsou², Ibikounle Moudachirou⁴, Severin Babatounde⁵, Sylvie Mawule Hounzangbe-Adote², Patrick Aleodjrodo Edorh³

¹Zootechnics and Livestock Systems Research Unit (URZoSE), Animal and Fisheries Sciences Laboratory (LaSAH), Doctoral School of Agronomic and Water Sciences (EDSAE), National University of Agriculture (UNA), 01 BP 55, Porto-Novo, Republic of Benin

²Laboratory of Ethnopharmacology and Animal Health, Faculty of Agronomic Sciences, University of Abomey Calavi (UAC), 01 BP526 Cotonou, Republic of Benin

^sLaboratory of Toxicology and Environmental Health (LATSE), University of Abomey-Calavi (UAC), 01 BP 526 Cotonou, Republic of Benin

^{*}Zoology Laboratory of the Faculty of Science and Technology, University of Abomey-Calavi (UAC), 01 BP526, Cotonou, Republic of Benin

^sLaboratory of Zootechnics, Faculty of Agronomic Sciences, University of Abomey-Calavi (UAC), 01 BP 526, Cotonou, Republic of Benin

Key words: Ethno veterinarian, Anogeissus leiocarpus, Chemical composition, Pharmacological, Zootechnical utility and Small ruminant.

http://dx.doi.org/10.12692/ijb/19.5.10-26

Article published on November 30, 2021

Abstract

The genus *Anogeissus* has eight (8) species, of which 5 are found in Tropical Asia, 2 in Arabia and 1 in Tropical Africa. The only species of the genus *Anogeissus* found in Africa is *Anogeissus leiocarpus*. Also known as *Anogeissus leicarpa* or *Anogeissus schimperi*, Hochest. Ex Hutch & Dalz., *Anogeissus leiocarpus* is a multipurpose tree widely used in traditional medicine. However, it is less used in ethno-veterinary medicine and

its pharmacological evidence and especially in the treatment of parasitic diseases caused by *Haemonchus contortus* are poorly documented. This review is aimed at synthesizing current knowledge on the chemical composition, pharmacological and zootechnical utility of *A. leiocarpus in order* to explore gaps and propose research perspectives. Google Scholar was the main database used to compile most of the information available on *A. leiocarpus* in this review. Overall, 44 publications were analyzed, out of which 65.9% addressed the pharmacological properties of *A. leiocarpus*. The anthelmintic, antidiabetic, antioxidant, antimicrobial, antibacterial, antispasmodic, analgesic, anti-inflammatory, antihypertensive are the most pharmacological properties found in those publications. Similarly, many secondary metabolites including alkaloids, flavonoids, tannins, saponins, quinones, coumarins found in *A. leiocarpus* give it its pharmacological properties. The factorial correspondence analysis performed for the variable extracts and chemical compounds showed that the presence of a given chemical compound does not depend on the type of extract. This review allowed to gather essential information on *A. leiocarpus* and justifies its traditional use in the small ruminant breeding in West and Central Africa.

*Corresponding Author: Pascal Abiodoun Olounlade 🖂 abiodouno@yahoo.fr

Introduction

Plants are the basis of the traditional medical system (Dramane et al., 2010) and represent a primary resource for traditional medicine. Because of their importance in the prevention and treatment of human and animal diseases, several studies were conducted and aimed at identifying the chemical compounds responsible for their actions. In fact, in developing countries, more than 80% of the population use medicinal plants for the first intention because of their easy access compared to modern drugs (Muthu et al., 2006; OMS, 2014). In Benin, the accessibility and availability of medicinal plants throughout the country lead people, especially in the villages, to use them to treat diseases. Among these plants, there is Anogeissus leiocarpus belonging to the Combretaceae family and found in West Africa, from Senegal to Cameroon and extended to Ethiopia and East Africa (Arbab, 2014). Growing in dry forests and gallery forests (Arbab, 2014; Ouédraogo et al., 2013), its distribution goes from the borders of the Sahara to the outer layer of tropical rain forests. It is a 15 to 18 m tall tree with a trunk up to 1 m in diameter, found in woodlands and savannas of the Sudanese regional center of endemism, with drooping branches bearing alternate, opposite, pubescent, ovate and lanceolate leaves of 2 to 8 cm long and 1.3 to5 cm wide. The leaves are acute at the apex, attenuated at the base, pubescent below and the barks are greyish to beige in color with fine scales, a yellowish edge streaked with brown (Arbab, 2014). It is used in the manufacture of chewing sticks for oral hygiene (Arbab, 2014; Sereme et al., 2008) and in the treatment of some diseases (Olutayo et al., 2011). This literature review aims to gather information available on the zootechnical utility, pharmacological activities and chemical composition of A. leiocarpus in order to shed more light on this medicinal plant little documented in the literature.

Methodology

Study protocol

Information on the pharmacological properties and the zootechnical importance of *A. leiocarpus* in the breeding system was collected through publications selected using inclusion and exclusion criteria, data source, criteria for preliminary evaluation of papers. The retrieved papers were manually reviewed to identify and exclude any work that did not meet the above criteria.

Literature review strategy

The main electronic database used for the review is Google scholar. It was chosen after consulting other databases such as PubMed and SCORPUS which had almost the same publications with available information on A. leiocarpus. The review was carried out from March 2020 to April 2021 using keywords or expressions (see Table 1) to collect as much information as possible on the plant and its pharmacological properties. The strategy adopted is that used by (Li et al., 2013). The English correspondents for each of the keywords in Table 1 were used. This strategy allowed the inventory of 52 documents and the selection of 44 ones based on their close link with the research topic. Publications either or in French English addressing the in pharmacological properties of the plant via laboratory tests were identified and recorded. Data collected were inserted into the Excel spreadsheet (2016) for encoding, processed with the GraphPad Prism 8.4.3 software for graphs and histograms and analyzed using the R software 3.6.3 version for factorial correspondence analysis (CFA) to determine relations between the extracts and the chemical compounds obtained. The variables were linked to the pharmacological properties of the plant, the type of test (in vitro or in vivo) and of extract used, the field of application of the research (ethno-veterinary or ethnomedicine), the part of the plant used, and the country where the research was carried out. The animal model used was specified for in vivo tests and only the variables "extracted" and "chemical compounds" were considered for factorial analysis of the correspondences.

Inclusion and exclusion criteria

The preliminary review consisted of a careful reading of the papers' title and abstract to ensure that they are linked with the research theme. Papers that passed

this stage were assessed on the quality and relevance of information. Those with supporting laboratory procedures (*in vitro* and or *in vivo* activities) were used as they ensure the accuracy of the results. Original research papers from 1983 to 2021 were included in the research. Equally, some references cited in publications were used to find other papers.

Results

Literature review

The literature review carried out over a one-year period allowed to select 44 documents published after a careful screening out of a total of 52 identified publications (research papers, conference reports and study reports). Among the 44 publications used, 29 dealt separately with the pharmacological properties of *A. leiocarpus,* representing 65.9% of the selected publications of interest.

The synthesis of data revealed 13 pharmacological properties of *A. leiocarpus* in West Africa, in Central Africa and in North Africa and also taking into account the unique publication dealing with several pharmacological properties in the same time as the case of Okpekon *et al.* (2004) in a single publication indicated three (3) different pharmacological properties which are: antimalarial, trypanocidal and anthelmintic properties (Table 3).

Table 1. Literature review strategy and terms used to identify data on the chemical composition, pharmacological and zootechnical uses of *Anogeissus leiocarpus* (Li *et al.*, 2013).

Searched terms	1. <i>Anogeissus leiocarpus</i> or <i>Anogeissus leiocarpa</i> , 2. Properties, 3. Activities, 4. Effect, 5. Distribution, 6. Ecology, 7. Use, 8. Pharmacology, 9. Compound, 10. Chemistry, 11. Animal				
	application, 12 Medicinal use, 13. Traditional use, 14. Local name, 15. Synonym, 16. Ethno				
	veterinarian, 17. Zootechnical utility, 18. Pharmacological utility, 19. Importance in traditional medicine, 20. Chemical compounds, 21. Bioactive compounds, 22. Chemical				
	composition, 23. Secondary metabolites, 24. Herbal medicine efficacy 25. Antibacterial				
	properties, 26. Antispasmodic properties, 27. Gastroenteritis properties, 28. Anthelmintic				
	properties, 29. Antimicrobial properties, 30. Antimicrobial properties, 31. Antidiabetic				
	properties, 32 Antiparasitic properties, 33. Antidiarrhoeal properties				
Strategy used	1 and (2 or 3 or 4 or 5 or 6 or 7 or 8 or 9 or 10 or 11 or 12 or 13 or 14 or 15 or 16 or 17 or 18 or				
	19 or 20 or 21 or 22 or 23 or 24 or 25 or 26 or 27 or 28 or 29 or 30 or 31 or 32 or 33).				

Country	Ethnic group Local name		References		
Benin	Fon	Abangnahi, hlihon	(Salifou <i>et al.</i> , 2017)		
	Hausa	Marke			
	Fulani	Kojoli	-		
	Kanuri	Annum	(Onoja <i>et al.</i> , 2018)		
	Yoruba	Ayinor orin-odan ainy	_		
	Igbo	Atara	_		
	Nupe	Kukunchi	_		
Burkina-Faso	Mooré	Siiga	(Zongo <i>et al.</i> , 2017)		
_	Sulphide	Kodjoli			
	Bornouam	Anum	-		
_	Doayo	Tarko			
_	Guziga	Duwui	_		
Cameroon — — — —	Mada	Euwe	(Gautier <i>et al.</i> , 2002)		
	Maha	Dawai			
	Mandara	Nawaya			
	Mundang	Kiere	_		
	Toupouri	Seou			
Côte d'Ivoire	Sinematiali population	Guenmin, N'galama	(Koné <i>et al.</i> , 2019)		
Niger	Niamey region	Maréké, N'gongo	(Souley Kallo et al., 2018)		

Table 2. Vernacular names of A. leiocarpus.

It is also important to note that in addition to these properties, the medicinal species is used against various ailments in traditional medicine in several African countries (Table 4).

Table 3. Pharmacological properties of *A. leiocarpus*, field of study carried out, part of the plant and extracts used.

Pharmacological properties	Experimenta l model	Type of activity	Type of extract	Part of the plant used	Country	Field of study	References
Anthelmintic properties	Rats	In vivo	Methanolic extract	Bark	Nigeria	Ethno veterinarian and ethnomedicine	(Ibrahim <i>et al.</i> , 1983)
Antimalarial, trypanocidal and anthelmintic properties	-	In vitro	Methanolic extract	Leaves, bark, roots	Ivory Coast	Ethno veterinarian / Ethnomedicine	(Okpekon <i>et al.</i> , 2004)
Anthelmintic properties	-	In vitro	Ethanolic extract	Roots	Ivory Coast	Ethno veterinarian	(Koné <i>et al.</i> , 2005)
Anthelmintic properties	Sheep	In vivo	Raw aqueous extract	Leaves	Nigeria	Ethno veterinarian	(Agaie <i>et al.</i> , 2007a)
Anthelmintic properties	-	In vitro	Aqueous extract	Leaves	Burkina-Faso	Ethno veterinarian	(Kabore <i>et al.</i> , 2009)
Anthelmintic properties	Sheep	In vivo	Aqueous extract	Leaves	Burkina-Faso	Ethno veterinarian	(Kaboré, 2009)
Anthelmintic properties	-	In vitro	Acetone extract and	Leaves	Nigeria	Ethno veterinarian	(Ademola <i>et al.</i> , 2011)
			fractions (hexane,				
			chloroform, butanol				
			and 35% water in the				
			methanol fractions)				
Anthelmintic properties	Sheep	In vivo	Ethanolic extract	Roots	Ivory Coast	Ethno veterinarian	(Dramane <i>et al.</i> , 2013)
Anti trypanosomal properties	Rats	In vitro	Aqueous and mehanolic extracts	Leaves, bark and roots	Nigeria	Ethno veterinarian	(Wurochekke <i>et al.</i> , 2012)
Trypanocidal properties	Rats	In vivo	Methanolic extract	Stem bark	Nigeria	Ethno veterinarian	(Awobode <i>et al.</i> , 2015)
Antioxidant properties	-	In vitro	Methanolic extract	Sheets	Nigeria	Ethnomedicine	(Olutayo et al., 2011)
Antioxidant properties	-	In vitro	Ethanolic extract	Stem bark	Nigeria	Ethnomedicine	(Olugbami et al., 2014)
Antioxidant properties	-	In vitro	Aqueous extract	Root bark	Nigeria	Ethnomedicine	(Salau et al., 2015)
Antioxidant properties	Rats	In vivo	Aqueous extract	Root bark	Nigeria	Ethnomedicine	(Salau et al., 2015)
Anti-quorum and antioxidant properties	-	In vitro	Methanolic extract	Stem bark	Burkina-Faso	Ethnomedicine	(Ouedraogo <i>et al.</i> , 2016)
Antioxidant and antibacterial properties	-	In vitro	Methanolic extract	Leaves	Ghana	Ethnomedicine	(Barku <i>et al</i> ., 2013)
Antibacterial properties	-	In vitro	Methanolic extract	Stem bark	Niger	Ethnomedicine	(Mann <i>et al.</i> , 2010)
Antibacterial properties	-	In vitro	Aqueous and ethanolic extracts	Roots	Nigeria	Ethnomedicine	(Gbadamosi <i>et al.</i> , 2014)
Antibacterial properties	-	In vitro	Aqueous and methanolic extracts	Stem bark	Ivory Coast	Ethnomedicine	(Sanogo <i>et al.</i> , 2016)
Antibacterial properties	-	In vitro	Ethanolic extract and ethyl acetate fraction	Sheets	Nigeria	Ethnomedicine	(Ganfon <i>et al.</i> , 2019)
Antimicrobial and antifungal	-	In vitro	Methanolic extract and	Roots,	Sudan	Ethnomedicine	(Elsiddig <i>et al.</i> , 2015)
properties			fractions of petroleum ether, chloroform and	leaves and stems			
			ethyl acetate				
Antimicrobial, antibacterial and antifungal properties	-	In vitro	Aqueous extract	Rod	Senegal	Ethnomedicine	(Diatta <i>et al</i> ., 2019)
Antimicrobial properties	-	In vitro	Methanolic extract	Stem bark	Nigeria	Ethnomedicine	(Abdullahi, 2012)
Antimicrobial properties	-	In vitro	Aqueous, hexane, dichloromethane and ethanolic extracts	Stem bark	Nigeria	Ethnomedicine	(Usman <i>et al</i> ., 2020)
Antidiabetic and hypolipidemic properties	Rats	In vivo	Ethanolic extract	Leaves	Nigeria	Ethnomedicine	(Onoja <i>et al.</i> , 2018)
Antidiarrhoeal properties	Rats	In vivo	Aqueous extract	Leaves	Cameroon	Ethnomedicine	(Fokam Tagne <i>et al.</i> , 2019)
Antihypertensive properties	Rats	In vivo	Aqueous extract	trunk	Burkina-Faso	Ethnomedicine	(Ouédraogo <i>et al.</i> , 2008)
Antihypertensive properties	Pigs	In vivo	Dichloromethane fraction	Stem bark	Burkina-Faso	Ethnomedicine	(Belemnaba <i>et al.</i> , 2013)
Antimalarial properties	Mice	In vivo	Methanolic extract	Bark	Nigeria	Ethnomedicine	(Akanbi, 2017)
Anti-leishmanian properties	-	In vitro	Methanolic extract / Aqueous and butanolic	Stem bark	Nigeria	Ethnomedicine	(Shuaibu <i>et al</i> ., 2008)

Overview of Anogeissus leiocarpus

Anogeissus leiocarpus is a tree belonging to the phylum Spermaphytes, the family Combretaceae. It is from the Dicotyledonous class, Rosidae subclass and Myrtales order (Cronquist *et al.*, 1981). *A. leiocarpus*

has various vernacular names depending on the country of origin and the ethnic group. For example, it is called in English "African birch"(Arbab, 2014) and is known as " Hlihon " in Fon in Benin (Salifou *et al.*, 2017). Other names are given in Table 2.

Table 4. Use of A. leiocarpus in traditional medicine.

Part of the plant used	rt of the plant used Therapeutic indication		Medicinal preparation	References
Bark and seeds	Treatment or prevention of worm infestations in equine species, and in animal and human helminthoses.	Nigeria	-	(Ibrahim <i>et al.</i> , 1983)
Trunk bark powder with millet seeds	Jaundice, constipation, malaria, anorexia		Decoction	(Fane, 2003)
	Common cold	Mali	Infusion	_
Trunk bark	Cough	Ivory Coast	Decoction	(Koné <i>et al.</i> , 2019)
	Antioxidant virtues, blood pressure regulator	Burkina-Faso	Decoction	(Sereme et al., 2008)
	Diarrhea, dewormer, wounds, eczema, psoriasis, carbuncles, boils and ulcers	Burkina-Faso	Decoction	(Sereme <i>et al.</i> , 2008)
Bark	Febrifuge	Guinea (Upper Guinea)	Infusion	(Olutayo <i>et al.</i> , 2011)
	Cough	Sudan	Decoction	(Arbab, 2014)
	Trypanocidal effect on livestock	Burkina-Faso	Maceration	(Zongo et al., 2017)
Bark of Anogeissus leiocarpus and Khaya senegalensis	Increased milk production in cows	Benin	Maceration	(Salifou <i>et al.</i> , 2017)
Bark, leaves and roots	Antimicrobial and anthelmintic action, yellow fever, jaundice, various forms of hepatitis, common colds and headaches	Burkina-Faso	Decoction	(Sereme <i>et al.</i> , 2008)
Leaves, roots, bark of trunk	Helminthiasis, trypanosomiasis, malaria and dysenteric syndrome	Ivory Coast	-	(Okpekon <i>et al.</i> , 2004)
Leaves and leafy twigs	Jaundice, amoebic dysentery, headache.		Decoction	(Fane, 2003)
Leaves and gum	Dermatosis	Burkina-Faso	Decoction	(Sereme et al., 2008)
Leaves	Treatment of gastrointestinal parasitism in small ruminants (lyophilized product administered at a dose of 160 mg / kg)	Burkina-Faso	Decoction	(Kabore <i>et al.</i> , 2010)
Leaves	Changes in skin pigmentation, eye bath to fight against some ailments	Ivory Coast	Decoction	(Olutayo <i>et al.</i> , 2011)
	Stomach infections	Togo	Decoction	(Arbab, 2014)
Leaves, bark of stems and roots	Antifungal action against a number of pathogenic fungi	Burkina-Faso	Excerpts	(Sereme et al., 2008)
Leaves, stems, roots	Liver infection	Ivory Coast	Decoction	(Koné <i>et al.</i> , 2019)
Vegetative organs	Malaria	Burkina-Faso	-	(Sereme et al., 2008)
Sticks taken from Anogeissus leiocarpus	Strong activity against a broad spectrum of bacteria, including some that contribute to the deterioration of teeth.	Burkina-Faso	Chewing	(Sereme <i>et al.</i> , 2008)
Sticks	Oral hygiene	Nigeria	Chewing	(Arbab, 2014)
Roots	Intestinal worms, diarrhea, dysentery, anemia	Ivory Coast	-	(Koné et al., 2005)
Plant	Parasitic diseases, malaria, trypanosomiasis, helminthiasis and dysenteric syndrome	Ivory Coast	-	(Arbab, 2014)
	Fungal infections such as dermatitis and yeast infection	Togo	-	(Arbab, 2014)
	Treatment of general diabetic ulcer, body pain, blood clots, asthma, cough and tuberculosis	Ghana	-	(Arbab, 2014)

Table 5. Chemical compounds according to extracts and organs used.

Extracts	Chemical compounds	References			
	Sheets	Stem bark Roots		•	
Methanolic		Hydrolyzable tannins, castalagine		(Shuaibu <i>et al.</i> , 2008)	
-	Tannins, saponosides		Tannins, saponosides and flavonoids	(Yemoa <i>et al.</i> , 2008)	
Methanolic		Alkaloids, flavonoids, saponins, tannins, phenolic compounds, cardiac glycosides and terpenoids		(Mann <i>et al.</i> , 2010)	
Aqueous	Flavonoids, tannins and polyphenols			Kabore <i>et al</i> . (2010)	
Methanolic	Tannins, glycosides, saponins, steroids and terpenoids			Olutayo <i>et al</i> . (2011)	
Aqueous and ethanolic			Alkaloids, saponins, tannins, phenols and glycosides	Gbadamosi <i>et al.</i> (2014)	
Aqueous		Saponosides, catechetical and gallic tannins, polyphenols, polyterpenes, anthocyanins, flavonoids		(Sanogo <i>et al.</i> , 2016)	
Methanolic		Saponosides, catechic and gallic tannins, polyphenols, polyterpenes, anthocyanins, flavonoids, alkaloids		(Sanogo <i>et al.</i> , 2016)	
Aqueous	Tannoids, catechetical tannins, flavonoids, coumarins and steroids			(Souley Kallo <i>et al.</i> , 2018)	
Ethanolic	Alkaloids, tannins (catechetical and gallic), flavonoids, leuco- anthocyanins, quinone derivatives, saponosides and reducing compounds			Ganfon <i>et al.</i> (2019)	
Aqueous		Alkaloids, saponins, flavonoids, tannins and reducing sugars and phenols		Usman <i>et al.</i> (2020)	
Hexanic		Alkaloids, steroids		Usman <i>et al</i> . (2020)	
Dichloromethane		Alkaloids, steroids and phenols		Usman <i>et al.</i> (2020)	
Ethanolic		Alkaloids, saponins, flavonoids, tannins and reducing sugars and phenols		Usman <i>et al.</i> (2020)	

Sometimes used as toothpicks for dental hygiene(Arbab, 2014; Sereme *et al.*, 2008), *A. leiocarpus* also has other therapeutic uses, especially

in traditional medicine for the treatment of various ailments in some African countries (Table 4).

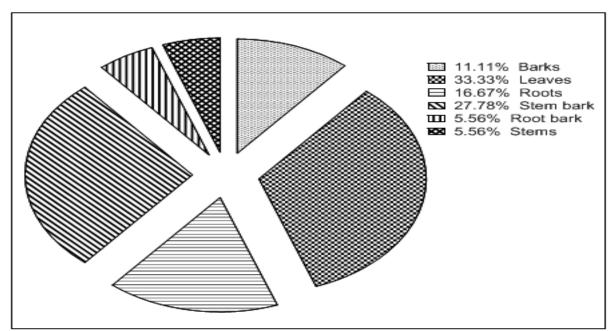


Fig. 1. Organs of A. leiocarpus used.

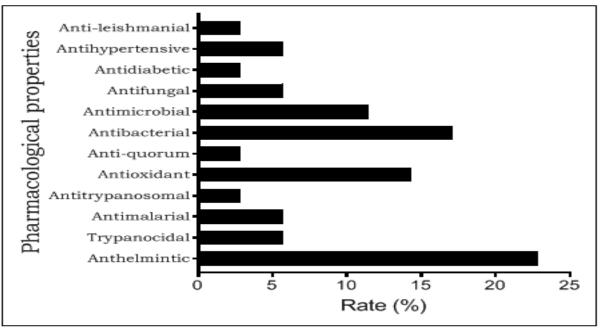


Fig. 2. Pharmacological properties of A. leiocarpus.

Pharmacological properties and zootechnical utilities of A. leiocarpus

A number of pharmacological studies have been carried out on the *A. leiocarpus* organs (Table 3). The literature has shown that the leaves are widely used (33.33%) against 25% for the bark of the stem (Fig. 1)

over the 13 pharmacological properties found. The review also revealed that the plant is used more for its anthelmintic properties. This was assessed in Burkina-Faso using aqueous extracts *in vitro* and *in vivo* on nematodes, parasites of small ruminants. In addition to the anthelmintic properties, *A. leocarpus*

is also known for its antimicrobial, antibacterial and antioxidant properties (Fig. 2) with most of the studies conducted in Nigeria (Fig. 3). This could be explained by the fact that Nigeria is one of the coastal countries in West Africa where the plant is widely distributed and where local people have incorporated it into their traditional health care practices.

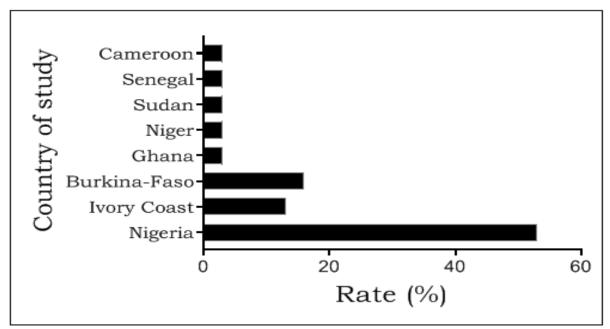


Fig. 3. Distribution of studies by country.

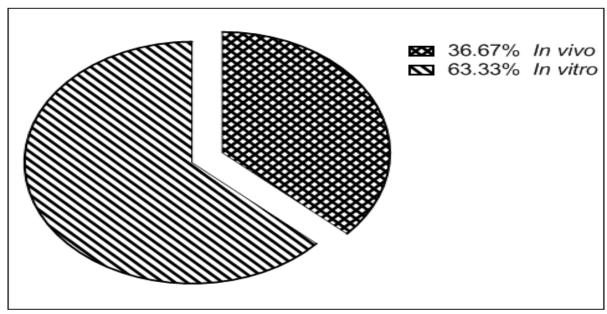


Fig. 4. Type of test carried out on the organs of A. leiocarpus.

More than 60% of the studies exploited in this review are *in vitro* studies (Fig. 4) using mainly rats as animal models. This is the rodent model typically used in ethnomedicine to assess the pharmacological properties or toxicity of a plant. This justifies the high rate of ethnomedical data found in this review (Fig. 5). Moreover, seven (7) types of solvents are used to extract chemical compounds and assess the pharmacological properties of *A. leiocarpus* (Fig. 6). The methanolic extract is widely used, followed by the ethanolic and n-hexane extracts, while the aqueous decoction is less used (Fig. 7).

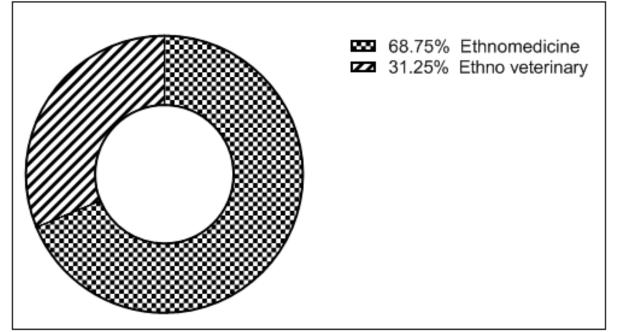


Fig. 5. Field of studies carried out on the organs of *A. leiocarpus*.

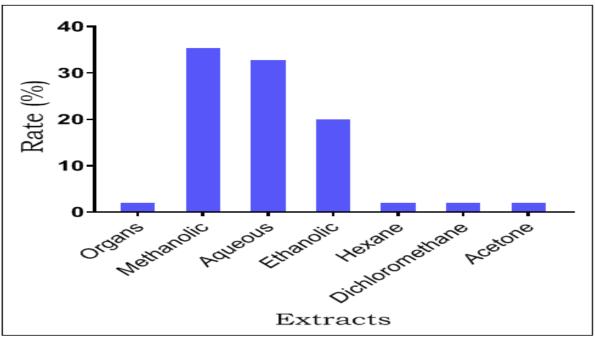


Fig. 6. Type of extract used in the studies.

$Chemical\ composition\ of\ A.\ leiocarpus$

The analysis of Table 5 shows that few chemical composition studies are conducted on the roots of *A. leiocarpus*. The few data available show that the roots contain alkaloids, flavonoids, saponins, tannins, phenols and glycosides (Fig. 9).

Some of these compounds, mainly tannins, saponins, alkaloids and flavonoids, are also present in the leaves

and stem bark of the plant (Figs 7 and 8) and are responsible for some pharmacological activities in the plant. According to Akpona et al. (2009), alkaloids have a local anesthetic action, tannins (healing, antibacterial, antiseptic, antioxidant, enzymatic inhibition: 5-lipooxygenase), flavonoids (antiinflammatory, antibacterial, antiviral in vitro), saponins (antibacterial, antiseptic, antiviral, antianti-edematous inflammatory, and analgesic),

terpenoids and steroids (antiviral, analgesic, antiinflammatory and antiseptic). Regardless of the extract used, the leaves, roots and/or bark of the stem of *A. leiocarpus* contain flavonoids, saponins, tannins and alkaloids. In addition to these compounds, the leaves and bark of the stem contain terpenoids and steroids. The presence of chemical compounds does not depend on the extract.

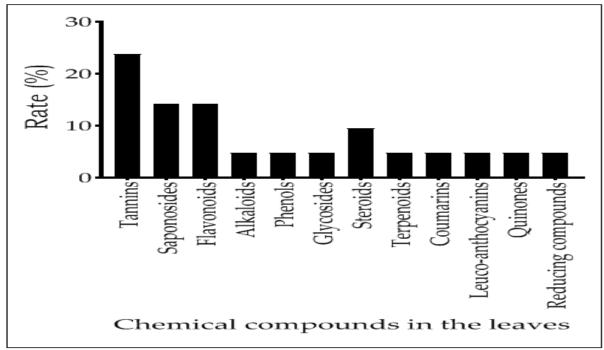


Fig. 7. Chemical compounds in the leaves of *A. leiocarpus*.

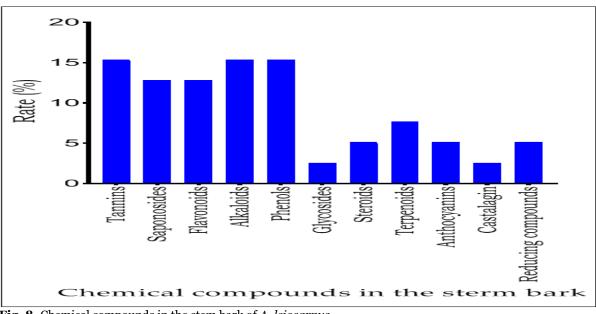


Fig. 8. Chemical compounds in the stem bark of A. leiocarpus.

Use of A. leiocarpus in animal husbandry

Anogeissus leiocarpus plays a key role in the management of pathologies. In Benin, it is used in animal production as a lactogenic plant in order to increase milk productivity in cows with a frequency of appearance in recipes amounting to 2.11(Salifou *et al.*, 2017). In addition, it is used in traditional medicine as an alicament for various diseases including parasitic diseases of small ruminants.(Agaie *et al.*, 2007a; Ibrahim *et al.*, 1983; Koné *et al.*, 2005; Wurochekke

et al., 2012).

Toxicity of A. leiocarpus

The use of *A. leiocarpus* as an alicament in African countries is of paramount necessity. Yet the reports of Ouédraogo *et al.* (2008) on the toxicity of the plant in

Burkina Faso showed that the acute general toxicity of the aqueous extract of *A. leiocarpus* revealed a lethal dose $50 (LD_{50})$ of 290.81 mg/kg body weight.

According to the WHO and Hodge and Sterner scales, such a drug would be classified as moderately toxic.

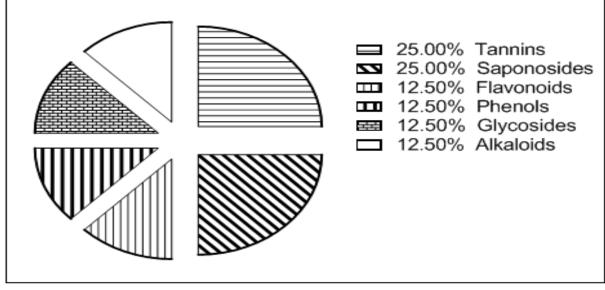


Fig. 9. Chemical compounds in the root of *A. leiocarpus*.

In addition, the reports of Agaie et al. (2007b) on the acute oral toxicity of the aqueous extract of the leaves of A. leiocarpus at a dose of 3200 mg/kg in Nigeria revealed that the animals showed signs of depression and inappetence, but no mortality was observed in the test groups. However, the intraperitoneal toxicity testing revealed signs of dose-related toxicity and deaths occurred in the test groups at doses of 1200, 1600, 2000 and 2400 mg/kg within 14 hours of administration of the extract, while the animals treated at the dose of 2800 mg/kg of extract died within 8 hours (Agaie et al., 2007b). Likewise, the study of Kabore et al. (2010) on the acute toxicity of the aqueous extract of the leaves of A. leiocarpus in Burkina-Faso revealed a change in the behavior of mice at the dose of 2000 mg/kg resulting in fatigue and the loss of appetite. The work of Onoja et al. (2018) in Nigeria revealed that the ethanolic foliar extract of A. leiocarpus administered to rats orally in acute toxicity at a dose of 5000 mg/kg did not cause death. However, acute toxicity studies of the ethanolic fraction of the root bark of A. leiocarpus in Côte d'Ivoire by the oral route carried out in rats are nontoxic and are well tolerated by the animals at an LD50 of up to 5000 mg/kg body weight. As for subacute toxicity, doses up to 500 mg/kg body weight of the ethanolic fraction of the root bark of *A. leiocarpus* did not affect the behavior and body weight development of the rats in any way. No mortality was observed. Ultimately, the ethanolic fraction of *A. leiocarpus* has no negative effect on the normal development of the body and it is safe orally (Kouangbé *et al.*, 2019). Figures 10 and 11 show some organs of the species *A. leiocarpus*.

Discussion

About thirty studies dealing with the pharmacological properties of *A. leiocarpus* were included in this study, with most of them carried out in Nigeria. Most of the studies of interest were on human medicine (68.75%) against 31.25% on veterinary medicine, yet this plant has shown efficiency in traditional veterinary medicine (Ibrahim *et al.*, 1983; Zongo *et al.*, 2017).



Fig. 10. A leiocarpus tree and inflorescences (Photo Kuiseu, 2019).

With the exception of Nigeria, few scientific studies focused on *Anogeissus leiocarpus* in West Africa despite its wide distribution. However, *A. leiocarpus* was a species with multiple therapeutic potentials which can be very useful in traditional veterinary medicine as an alternative to conventional drugs which their ineffectiveness, high cost, counterfeiting and resistance were reported in some cases.



Fig. 11. Leaves and trunk bark of A. leiocarpus (Koné, 2009).

The promotion of *A. leiocarpus* in traditional veterinary medicine after having its scientific effectiveness evidence would relieve several farmers, especially smallholders who face many animal health problems in their herds, including parasitic diseases caused by gastrointestinal parasites that negatively affect the expression of the zootechnical performance

of livestock and cause serious loss of income for herders. In the papers reviewed, leaves (33.33%) and stem bark (25%) were the organs of the plant more used as they contain several secondary metabolites. To assess the pharmacological properties, different types of extracts were made from the powders of the leaves and/or bark of the stem, root and trunk of *A*.

leiocarpus. Methanol was the most widely used extraction solvent, followed by water. It was the most polar solvent after water and its high polarity would be favorable to the elimination of metabolites such as flavonoids, phenols, saponins, tannins, phlobatanine, cardiac glycosides, alkaloids, terpéniodes, steroids, resins, proteins, carbohydrates, fats and oils, much more than ethanol, butanol, chloroform, petroleum ether, n-hexane or water. Although methanol has many metabolites, its use in medicine can have adverse effects on human and animal health. It would therefore be more difficult, if not impossible, for people with low income to have access to this solvent for the preparation of traditional herbal remedies. It would therefore be advisable to promote natural solvents such as water, which were available, nontoxic and easily accessible to rural populations.

Conclusion

The current review on the pharmacological knowledge of A. leiocarpus in traditional medicine shows that the plant has anthelmintic, antimalarial, antimicrobial, antibacterial, antihypertensive, antidiabetic and antidiarrhoeal properties. It is rich in alkaloids, flavonoids, saponosides, tannins, glycosides, steroids, terpenoids which give it its pharmacological properties. The documentation on this medicinal plant also revealed that it has many therapeutic properties but is underused in veterinary medicine. Therefore, the prospect of in-depth studies on this species in the application field of ethnoveterinary arises in order to support the promotion of A. leiocarpus as an anthelmintic in domestic ruminants for a better improvement of animal productivity.

Acknowledgement

The authors warmly thank the Doctoral School of Agricultural and Water Sciences of the National University of Agriculture (UNA) of Benin for the training on writing methodologies.

Conflicts of interest

The authors declare no conflict of interest regarding the publication of this paper.

References

Abdullahi M. 2012. Evaluation of Antimicrobial Activity of *Anogeissus leiocarpus* and *Terminalia avicennioides* against Infectious Diseases Prevalent in Hospital Environments in Nigeria. Journal of Microbiology Research **2(1)**, 6-10.

http://dx.doi.org/10.5923/j.microbiology.20120201. 02

Ademola IO, Eloff JN. 2011. *In vitro* anthelmintic effect of *Anogeissus leiocarpus* (DC.) Guill. & Perr. leaf extracts and fractions on developmental stages of *Haemonchus contortus*. African Journal of Traditional Complementary Alternative Medecine **8(2)**, 134-139.

Agaie BM, Onyeyili PA. 2007a. Anthelmintic activity of the crude aqueous leaf extracts of *Anogeissus leiocarpus* in sheep. African Journal of Biotechnology **6(13)**, 1511-1515.

Agaie BM, Onyeyili PA, Muhammad BY, Ladan MJ. 2007b. Acute toxicity effects of the aqueous leaf extract of *Anogeissus leiocarpus* in rats. African Journal of Biotechnology **6(7)**, 886-889.

Akanbi OM. 2017. Antiplasmodial Potential of Combination Therapy of Methanolic Bark Extracts of *Terminalia avicennioides* and *Anogeissus leiocarpus* and Its Effect on Haematological Parameters on Mice Infected with *Plasmodium berghei*. Journal of Advances in Medical and Pharmaceutical Sciences **15(2)**, 1-9.

http://dx.doi.org/10.9734/JAMPS/2017/35105

Akpona HA, Akpona JDT, Awokou SK, Yemoa A, Dossa LOSN. 2009. Inventory, folk classification and pharmacological properties of plant species used as chewing stick in Benin Republic. Journal of Medicinal Plants Research **3(5)**, 382-389.

Arbab AH. 2014. Review on *Anogeissus leiocarpus* a potent african traditional drug. International Journal of Research in Pharmacy and Chemistry **4(3)**, 496-500.

Awobode HO, Fagbemi FT, Afolayan FID. 2015. Antitrypanosomal activity of *Khaya senegalensis* and *Anogeissus leiocarpus* stem bark on *Trypanosoma brucei brucei* infected rats. African Journal of Biotechnology **14(6)**, 525-529. <u>http://dx.doi.org/10.5897/AJB2014.14136</u>

Barku VYA, Opoku-Boahen Y, Owusu-Ansah E, Dayie NTKD, Mensah FE. 2013. *In vitro* assessment of antioxidant and antimicrobial activities of methanol extracts of six wound healing medicinal plants. Journal of Natural Sciences Research **3(1)**, 74-80.

Belemnaba L, Ouedraogo S, Auger C, Chataigneau T, Traore A, Guissou IP, Lugnier C, Schini-Kerth VB, Bucher В. 2013. Endothelium-independent and endotheliumdependent vasorelaxation by a dichloromethane fraction from Anogeissus leiocarpus (DC) Guill. Et Perr. (Combretaceae): possible involvement of cyclic nucleotide phosphodiesterase inhibition. African of Traditional, Complementary Journal and Alternative Medecine 10(2), 173-179.

Cronquist A, Takhtadzhian AL. 1981. An integrated system of classification of flowering plants: Columbia university press.

Diatta BD, Houël E, Gueye M, Niass O, Boetsch G. 2019. Activités antimicrobiennes des plantes utilisées comme bâtonnets frotte-dents (curedents) par les Peul de la commune de Tessékéré (Ferlo Nord, Sénégal). International Journal of Biological and Chemical Sciences **13(3)**, 1444-1457.

Dramane S, Mamidou K, Kamanzi K. 2010. Evaluation of antimicrobial and free radical scavenging activities of some bioactive taxons from Côte d'Ivoire. European Journal of Scientific Research **40(2)**, 307-317.

Dramane S, Witabouna MK, Bassirou B, Bernadin D, Kassédo BT, Kagoyire K. 2013. *In vivo* anthelmintic activity of *Anogeissus leiocarpus* Guill & Perr (Combretaceae) against nematodes in naturally infected sheep. Parasitology Research **112(7)**, 2681-2688.

http://dx.doi.org/10.1007/s00436-013-3435-y

Elsiddig IME, Muddather AK, Ali HAR, Ayoub SMH. 2015. A comparative study of antimicrobial activity of the extracts from root, leaf and stem of *Anogeissus leiocarpus* growing in Sudan. Journal of Pharmacognosy and Phytochemistry **4(4)**, 107-113.

Fane S. 2003. Etude de la toxicité de certaines plantes vendues sur les marchés du district de Bamako. (Thèse), Université de Bamako, Mali.

Fokam Tagne MA, Tagne MAF, Rékabi Y, Noubissi PA, Fankem GO, Akaou H, Wambe H, Kamgang R. 2019. Evaluation of Antidiarrheal Activity of Aqueous Leaf Extract of *Anogeissus leiocarpus* on Castor Oil-Induced Diarrhea in Rats Mice. American Journal of Biomedical Science and Research **3(1)**, 27-34.

http://dx.doi.org/10.34297/ajbsr.2019.03.000629

Ganfon H, Houvohessou J, Assanhou AG, Bankole HS, Gbenou J. 2019. Activité antibactérienne de l'extrait éthanolique et des fractions de *Anogeissus leiocarpa* (DC) Guill. Et Perr. (Combretaceae). International Journal of Biological and Chemical Sciences **13(2)**, 643-651. http://dx.doi.org/10.4314/ijbcs.v13i2.6

Gautier D, Hautdidier B, Ntoupka M, Onana J, Perrot N, Tapsou T. 2002. Fiches techniques des arbres utiles aux paysans du Nord Cameroun. Caractéristiques de l'arbre, ce qu'en font les paysans et ce qu'ils pourraient en faire. (Thèse), Cameroun.

Gbadamosi IT, Ogunsuyi AO. 2014. An appraisal of the potency of roots of *Anogeissus leiocarpus* (DC.) Guill. & Perr. and *Terminalia glaucescens* Benth. in the management of *E. coli* related infections. Journal of Applied Biosciences **78(1)**, 6646-6653. http://dx.doi.org/10.4314/jab.v78i1.10

Ibrahim MA, Nwude N, Ogunsusi RA, Aliu YO. 1983. Screening of West African plants for anthelmintic activity. International Symposium on Medicinal Plants **5**, 1-34.

Kaboré A. 2009. Activité anthelminthique de deux plantes tropicales testées *in vitro* et *in vivo* sur les strongles gastro-intestinaux des ovins de race Mossi du Burkina-Faso. (Thèse), Université Polytechnique de Bobo-Dioulasso, Burkina-Faso.

Kabore A, Belem AMG, Tamboura HH, Traore A, Sawadogo L. 2009. *In vitro* anthelmintic effect of two medicinal plants (*Anogeissus leiocarpus* and *Daniellia oliveri*) on *Haemonchus contortus*, an abosomal nematode of sheep in Burkina Faso. African Journal of Biotechnology **8(18)**, 4690-4695.

Kabore A, Tamboura HH, Traore A, Traore A, Meda R, Kiendrebeogo M, Belem AMG, Sawadogo L. 2010. Phytochemical analysis and acute toxicity of two medicinal plants (*Anogeissus leiocarpus* and *Daniellia oliveri*) used in traditional veterinary medicine in Burkina Faso. Archives of Applied Science Research **2(6)**, 47-52.

Koné D. 2009. Enquête ethnobotanique de six plantes médicinales maliennes : extraction, identification d'alcaloïdes - caractérisation, quantification de polyphénols : étude de leur activité antioxydante. (Thèse), Université Paul Verlaine -Metz (France).

Koné KHC, Coulibaly K, Konan KS. 2019. Plantes à usage médicinale en élevage d'ovins à Sinématiali (Nord de la Côte d'Ivoire). Journal of Animal and Plant Sciences **41(1)**, 6828-6839. <u>http://dx.doi.org/10.35759/JAnmPlSci.v41-1.9</u>

Koné WM, Atindehou KK, Dossahoua T, Betschart B. 2005. Anthelmintic activity of medicinal plants used in Northern Côte d'Ivoire against intestinal helminthiasis. Pharmaceutical biology **43(1)**, 72-78.

http://dx.doi.org/10.1080/13880200590903408

Kouangbé MA, Sonan KHG, Ouattara K, Bahi C, Djaman AJ, N' Guessan JD. 2019. Assessment of acute and subacute oral toxicity of Ethanolic fraction of root barks of *Anogeissus leiocarpa* (DC) Guill & Perr in albinos wistar rats. The Pharma Innovation Journal **8(4)**, 211-217.

Li XX, Zhou XN. 2013. Co-infection of tuberculosis and parasitic diseases in humans: a systematic review. Parasites & Vectors **6(1)**, 1-12. https://doi.org/10.1186/1756-3305-6-79

Mann A, Barnabas BB, Daniel II. 2010. The Effect of Methanolic Extracts of *Anogeissus Leiocarpus* and *Terminalia Avicennioides* on the Growth of Some Food-borne Microorganisms. Australian Journal of Basic and Applied Sciences **4(12)**, 6041-6045.

Muthu C, Ayyanar M, Raja N, Ignacimuthu S. 2006. Medicinal plants used by traditional healers in Kancheepuram District of Tamil Nadu, India. Journal of Ethnobiology ethnomedicine **2(1)**, 1-10.

Okpekon T, Yolou S, Gleye C, Roblot F, Loiseau P, Bories C, Grellier P, Frappier F, Laurens A, Hocquemiller R. 2004. Antiparasitic activities of medicinal plants used in Ivory Coast. J Ethnopharmacol **90(1)**, 91-97.

http://dx.doi.org/10.1016/j.jep.2003.09.029

Olugbami JO, Gbadegesin MA, Odunola OA. 2014. *In vitro* evaluation of the antioxidant potential, phenolic and flavonoid contents of the stem bark ethanol extract of *Anogeissus leiocarpus*. African Journal of Medecine and Medical Sciences **43(1)**, 101-109.

Olutayo O, Doyinsola I, Simon O, Abayomi O, Thomas S. 2011. Phytochemical and antioxidant properties of some Nigerian medicinal plants. American Journal of Scientific and Industrial Research **4(3)**, 328-332.

http://dx.doi.org/10.5251/ajsir.2013.4.3.328.332

OMS. 2014. Stratégie de l'OMS pour la médecine traditionnelle pour 2014-2023. Genève.

Onoja US, Ugwu CC, Uzor PF, Nweze IE, Omeje EO, Nnamani PO, Nwachukwu E, Njoku I, Effiong EJ. 2018. Effect of *Anogeissus leiocarpus* Guill and Perr Leaf on Hyperglycaemia and Associated Dyslipidaemia in Alloxan-induced Diabetic Rats. Journal of Pharmaceutical Sciences 17(1), 65-72.

Ouédraogo A, Kakaï RG, Thiombiano A. 2013. Population structure of the widespread species, *Anogeissus leiocarpa* (DC.) Guill. & Perr. across the climatic gradient in West Africa semi-arid area. South African Journal of Botany **88**, 286-295. <u>http://dx.doi.org/10.1016/j.sajb.2013.07.029</u>

Ouédraogo S, Belemnaba L, Traoré A, Lompo M, Bucher B, Guissou IP. 2008. Etude de la toxicité et des propriétés pharmacologiques de l'extrait aqueux de *Anogeissus leiocarpus* (DC) Guill. et Perr (Combretaceae). Pharmacopée et Médecine traditionnelle Africaines **15**, 18-22.

Ouedraogo V, Kiendrebeogo M. 2016. Methanol Extract from *Anogeissus leiocarpus* (DC) Guill. et Perr. (Combretaceae) Stem Bark Quenches the Quorum Sensing of *Pseudomonas aeruginosa* PAO1. Medicines **3(4)**, 1-10.

http://dx.doi.org/10.3390/medicines3040026

Salau AK, Yakubu MT, Oladiji AT. 2015. *In vitro* and *In vivo* Antioxidant Activity of Aqueous Extracts of *Anogeissus leiocarpus* (DC) Guill & Perr and *Terminalia avicennioides* Guill & Perr Root Barks. Cameroon Journal of Biological and Biochemical Sciences 23, 9-16.

Salifou CFA, Kassa KS, Ahounou SG, Moussa H, Dotché IO, Agbozo JM, T IM, Youssao IAK. 2017. Plantes lactogènes des bovins et leurs modes de préparation dans les élevages traditionnels au Bénin. Livestock Research for Rural Development **29(2)**, 1-5. Sanogo Y, Guessennd NK, Tra Bi HF, Kouadio NJ, Konan FK, Bamba M, Danho N, Bakayoko A, Yao K, Dosso M. 2016. Evaluation *in vitro* de l'activité des écorces de tige de *Anogeissus leiocarpus* (DC) Guill. et Perr. (Combretaceae) sur des bactéries responsables de maladies courantes en Afrique et criblage phytochimique. International Journal of Biological and Chemical Sciences **10(3)**, 1139-1152. http://dx.doi.org/10.4314/ijbcs.v10i3.19

Sereme A, Millogo-Rasolodimby J, Guinko S, Nacro M. 2008. Propriétés thérapeutiques des plantes à tanins du Burkina-Faso. Pharmacopée et Médecine traditionnelle Africaines **15**, 41-49.

Shuaibu M, Pandey K, Wuyep P, Yanagi T, Hirayama K, Ichinose A, Tanaka T, Kouno I. 2008. Castalagin from *Anogeissus leiocarpus* mediates the killing of Leishmania *in vitro*. Parasitology Research **103(6)**, 1333-1338.

Souley Kallo M, Adamou R, Sawadogo J, Ayouba Mahamane A, Maman Maarouhi I, Ikhiri K. 2018. Enquête ethnobotanique et criblage phytochimique de quelques plantes tinctoriales du Niger en vue d'une valorisation en énergie solaire. International Journal of Biological and Chemical Sciences 12(2), 867-883.

http://dx.doi.org/10.4314/ijbcs.v12i2.20

Usman AH, Ali T, Danjani AG. 2020. Phytochemical and antimicrobial studies of stem-bark extracts of *Anogeissus leiocarpus* found in Dutsin-Ma, Katsina - Nigeria. FUDMA Journal of Sciences **4(2)**, 156 - 167.

http://dx.doi.org/10.33003/fjs-2020-0402-209

Wurochekke AU, Anyanwu GO. 2012. Antitrypanosomal activity of *Anogeissus leiocarpus* in rats infected with *Trypanosoma brucei brucei*. International Research Journal of Biotechnology **3(1)**, 5-9.

Yemoa AL, Gbenou JD, Johnson RC, Djego JG, Zinsou C, Moudachirou M, Quetin-Leclercq J,

Bigot A, Portaels F. 2008. Identification et étude phytochimique de plantes utilisées dans le traitement traditionnel de l'ulcère de Buruli au Bénin. Ethnopharmacologia **42**, 48-55.

Zongo A, Kaboré A, Traoré A, Zabré G, Traoré A, Tamboura H, Belem AMG. 2017. Constraints of Ruminant Rearing and Ethno-veterinary Practice

Against African Animal Trypanosomosis in the Pastoral Area of Gaongho in Burkina Faso. Animal and Veterinary Sciences **5(1)**, 1-7.

http://dx.doi.org/10.11648/j.avs.20170501.11