

International Journal of Biosciences | IJB | ISSN: 2220-6655 (Print) 2222-5234 (Online) http://www.innspub.net Vol. 19, No. 6, p. 150-157, 2021

RESEARCH PAPER

OPEN ACCESS

Characterization of *Colocasia esculenta* var. *esculenta* by cytogenetical analysis from Bangladesh

Ashma Ahmed Warasy*

Department of Botany, Jahangirnagar University, Savar, Dhaka, Bangladesh

Key words: Colocasia, Interphase, Prophase, Metaphase, Chromosome

http://dx.doi.org/10.12692/ijb/19.6.150-157

Article published on December 29, 2021

Abstract

The study was undertaken to characterize *Colocasia esculenta* var. *esculenta* by cytogenetical analysis after staining with orcein. Complex chromocentric type of interphase nuclei and Continuous type of prophase chromosomes was observed in this study. *Colocasia esculenta* var. *esculenta* was found to possesses 2n=28 chromosomes. Persistent nucleoli was observed in this plant material suggested the late transcription of rDNA to rRNA and late transportation of rRNA from the nucleus to the cytoplasm. This material possessed metacentric and submetacentric chromosome representing heterogeneous karyotype. Individual chromosome length ranged from $0.72\pm0.05-2.34\pm0.05\mu$ m. Distance between small and large chromosome was about 1.62μ m indicating the gradual decrease of chromosome length. The Total form percentage was 41.56%. Karyotype symmetry and asymmetry index was 71.11% and 58.44%, respectively. Moderately asymmetric karyotype was found in this material. Therefore, the interphase nuclei, prophase chromosome and metaphase chromosome of *Colocasia esculenta* var. *esculenta* can be characterized cytogenetically with the above features.

* Corresponding Author: Ashma Ahmed Warasy 🖂 aawarasy@yahoo.com

Introduction

Colocasia esculenta var. *esculenta* belongs to Araceae which is highly heterogenous family and consists of about 1400-1500 species under 150 genera (Lawrance, 1966). They are commonly known as taro, eddo, dasheen and kalo and one of the most ancient cultivated crops (*Denham, 2011*). It is a starchy root crop in tropical to temperate regions of Africa, the Mediterranean, Asia and the Pacific (Matthews, 2006). Many cultivars adapted to saline conditions, low water availability and/or seasonal flooded soils have been created and are also widely cultivated throughout tropical and subtropical regions (Onwueme, 1999; Acevedo-Rodríguez and Strong, 2005). In Bangladesh, different cultivated and wild taros are distributed in all over the country (Kreike *et al.,* 2004).

The different members of this widely cultivated crops have some economical and ethnobotanical importance. The tubers and roots are acrid stimulant at fresh stage and also an effective drug in case of snake bites when applied both externally and internally (Ghani, 1998; Rao and Henry, 1996). The young leaves and petioles are used as vegetables. This is used to cure piles also. The tubers are used as poultice. They also meet the iron deficiency. The rhizomes of Colocasia posses medicinal value in curing stomach ache, abdominal pain and cholera. The same is crushed into a paste and applied externally on the physical body to cure abscesses and bug bites (Heng, 1979). It is a very popular vegetable known as kochu or mukhi in Bangladesh. The stolons or stems are usually cooked with small prawns which are very favoured by Bangladeshis. It is used to cooked with the head of the ilish fish into a curry, but some dishes are cooked with dried fish. Green leaves and stem are also eaten as a favourite dish and usually ground to a paste or finely chopped to make shake. Food and Agriculture Organization (FAO) reported that taro production has doubled over the past decade (FAOSTAT, 2000) and is now the 5th most-consumed vegetable worldwide.

Colocasia esculenta is an extremely variable species and has many different varieties probably because its chromosomes are prone to unpredictable behaviour during cell division. Because of this natural variablity, many different varieties, cultivars have also arisen. Different regions produce many different types and even within one type there can be a marked variablity over the course of a season or two. Barrau (1957) recognized two species namely C. esculenta and C. antiquorum. The two prominent chromosome numbers in Colocasia esculenta are 2n=28 and 2n=42: the diploids with 2n=28 chromosomes being mainly Colocasia esculenta var. esculenta genotypes characterised by a large central tuber surrounded by smaller ones. In addition, different chromosome number such as 2n=36 (Huang et al., 1989), 2n=38 (Tanimoto and Matsumoto, 1986), 2n=42 (Zhang and Yang, 1984; Coates and Gaffey, 1988; Subramanian and Munian, 1988; Zhang, 1998), and 2n=84 (Sreekumari and Mathew, 1991) chromosomes number was also reported.

The Bangladesh National Herbarium (BNH) has been collecting different taro including *Colocasia esculenta* var. *esculenta* from all over the country and they maintained it in the garden of BNH. They are mainly collected and identified on the basis of plant morphology. Therefore, authentic identification problem is existing since specimen may show different morphology in different environment due to phenotypic plasticity.

Identification problems were also found in different members of Araceae. Taxonomist faced problem to identify them authentically (Ara, 2000). Latter extensive cytological investigation had been carried out for proper identification of these taxa. The cytological data indicated sharp difference among the various sort of Colocasia esculanta, Xanthosoma violaceum, Typhonium trilobatum etc. (Alam and Deen, 2002; Deen and Alam, 2002; Warasy and Alam 2009). For authentic identification of a specimen, genomic information is very essential. Karvotype analysis is one among the traditional methods that provide a preliminary idea about the genome of a specimen. Generally karyotype analysis plays important role in determining the taxonomic status. Staining properties of interphase nuclei and prophase chromosomes is another approach for distinguishing different individual.

Int. J. Biosci.

This is usually done by differential staining (Fawzia and Alam, 2011; Bonna *et al.*, 2018). Tanaka (1971) classified the different types of interphase nuclei and prophase chromosomes on the basis of orcien staining property. The outcome of this study showed that various taxa including sort of many plant species might be distinguished by their staining properties.

Therefore, an extensive research should be undertaken to characterize *Colocasia esculenta* var. *esculenta* through cytogenetical analysis.

The aim of the present study was-

i. to compare the staining property of the interphase nuclei and prophase chromosomes after staining with orcein.

ii. to find out the chromosome number of *Colocasia* esculenta var. esculenta.

iii. to prepare and analysis the convantional karyotype from mitotic metaphase plate.

Material and methods

Material

For Cytogenetical analysis, *Colocasia esculenta* var. *esculenta* was used in this study as material.

Study area

The plant materials was collected from Bangladesh National Herbarium (BNH) and maintained in the botanic garden of the Department of Botany, Jahangirnagar University, Bangladesh.

Cytogenetical methods

Healthy roots were collected from the Botanical garden, Department of Botany, Jahangirnagar University. The optimum time of root collection for obtaining maximum number of dividing cells was 9.30-10.00 am. The collected root were pretreated with 0.002 M 8-hydroxyquinoline for 1.30 h at room temperature (28-30° C) followed by 15 min fixation in 45% acetic acid at 4°C. The pretreated roots were then hydrolyzed in a mixture of 1 N HCl and 45% acetic acid (2:1) at 60° C for 7-8 sec. Then the hydrolyzed root were soaked on a filter paper and taken in a clean slide. The meristematic region was cut with fine blade. A drop of 1% aceto-orcein was added to the material.

The materials were tapped gently by a tooth pick and squashed by placing thumbs. Finally the slides were observed under microscope (Olympus-DP72, Japan) with a digital camera. Final measurement was calculated by dividing the magnification of the objective 100x.

Results and discussion

The conducting investigation disclosed the karyotypic features of *Colocasia esculenta* var. *esculenta*. The nature of staining property of interphase nuclei and prophase chromosomes were accomplished to get additional information towards characterization of this germplasm.

Orcein-staining properties of interphase nuclei and prophase chromosomes

The staining properties of interphase nuclei and prophase chromosomes usually provide karyomorphological features that help to characterize different germplasm. Tanaka (1971) was the pioneer of proposing these criteria for karvomorphological investigation. He found that the nature of staining of heterochromatins present in the interphase nuclei and prophase chromosomes were different in different species. On the basis of the staining property he classified interphase nuclei and prophase chromosomes in five different categories. Later different investigators applied these criteria in characterizing different plant materials of diverge nature (Alam et al., 1993; Begum and Alam, 2004; Hossain et al., 2016; Sultana and Alam, 2016; Bonna et al., 2018).

In the present study, few darkly stained large heterochromatic regions were found in interphase nuclei of *Colocasia esculenta* var. *esculenta*. A clear nuclear boundary was visible. A big and prominent nucleolus was found in the interphase nuclei (Fig 1). The presence of prominent nucleoli indicated the active transcription of rDNA for the synthesis of rRNA. According to Tanaka (1971) it may be regarded as complex chromocentric type.

The prophase chromosomes stained uniformly from one end to another with orcein. A prominent nucleolus was also found in this stage (Fig 2). According to Tanaka (1971) it may be regarded as continuous type. Usually, germplasm with "Complex chromocenter type" of interphase nuclei showed "Gradient type" of prophase chromosomes. In this investigation, the studied material did not followed the usual regulation of heterochromatin. Presence of facultative heterochromatin might be one of the reasons for this type of observation. Whatever the reason is, this germplasm could be characterized on the basis of these characters of interphase nuclei and prophase chromosomes.

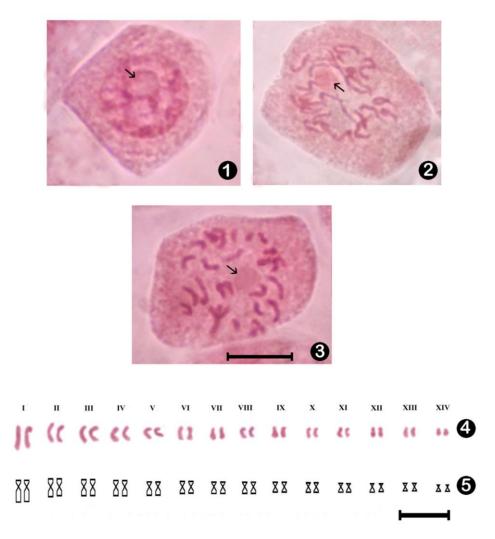
Somatic chromosome Number

Colocasia esculenta var. *esculenta* was found to possess 2n=28 chromosomes (Figs. 3,4,5 Table 1,2). Same chromosome number was reported earlier by different scientist (Zhang and Yang, 1984; Coates and Gaffey, 1988; Sreekumari and Mathe, 1989; Kuruvilla *et al.*, 1989; Ivancic and Lebot, 1999). In addition, different chromosome number such as 2n=36 (Huang et al., 1989), 2n=38 (Tanimoto and Matsumoto, 1986), 2n=42 (Zhang and Yang, 1984; Coates and Gaffey, 1988; Subramanian and Munian, 1988; Zhang, 1998), and 2n=84 (Sreekumari and Mathew, 1991) chromosomes number was also reported. Several scientist considered the basic chromosome number as x=14 for this species, then specimen with 2n=28 could be regarded as deploid, 2n=42 could be the triploid and 2n=84 is considered as hexaploid, respectively. So the present findings correlate with 2n=28 of the earlier reports and agree with the basic number of x=14. However, 2n=36 and 38 report previously did not agree with the basic number of x=14. In this case, they might be evolved from nearer ploidy level by chromosomal aberration.

Table 1. Length (in μ m), arm ratio, centromeric index, relative length and centromeric type of metaphase chromosomes of *Collocasia esculenta* var. *esculenta*.

Pair	Long arm (l)µm (x̄ ± SD)	Short arm (s)µm (x̄ ± SD)	Total length (Τ)μm (x̄ ± SD)	Arm ratio (l/s)	Relative length (RL)	Centro-meric Index (CI)	Centro- meric type (CT)
Ι	1.62 ± 0.03	0.72 ± 0.02	2.34 ± 0.05	2.25	0.06	30.77	sm
	1.53 ± 0.01	0.72 ± 0.02	2.25 ± 0.03	2.13	0.06	32.00	sm
II	1.17 ± 0.03	0.90 ± 0.01	2.07 ± 0.04	1.30	0.05	43.48	m
	0.99 ± 0.02	0.90 ± 0.01	1.89 ± 0.03	1.10	0.05	47.62	m
III	0.99 ± 0.01	0.81 ± 0.01	1.80 ± 0.01	1.22	0.05	45.00	m
	0.99 ± 0.01	0.81 ± 0.00	1.80 ± 0.01	1.22	0.05	45.00	m
IV	0.99 ± 0.03	0.72 ± 0.01	1.71 ± 0.04	1.38	0.05	42.11	m
	0.99 ± 0.03	0.72 ± 0.02	1.71± 0.04	1.38	0.05	42.11	m
V	0.90 ± 0.02	0.54 ± 0.02	1.44 ± 0.04	1.67	0.04	37.50	sm
	0.90 ± 0.03	0.54 ± 0.02	1.44 ± 0.04	1.67	0.04	37.50	sm
VI	0.72 ± 0.02	0.63 ± 0.03	1.35 ± 0.05	1.14	0.04	46.67	m
	0.72 ± 0.04	0.63 ± 0.02	1.35 ± 0.05	1.14	0.04	46.67	m
VII	0.72 ± 0.01	0.54 ± 0.01	1.26 ± 0.02	1.33	0.03	42.86	m
	0.72 ± 0.02	0.54 ± 0.01	1.26 ± 0.03	1.33	0.03	42.86	m
VIII	0.72 ± 0.02	0.54 ± 0.01	1.26 ± 0.03	1.33	0.03	42.86	m
	0.72 ± 0.02	0.54 ± 0.01	1.26 ± 0.03	1.33	0.03	42.86	m
IX	0.63 ± 0.03	0.54 ± 0.02	1.17 ± 0.05	1.17	0.03	46.15	m
	0.63 ± 0.03	0.54 ± 0.02	1.17 ± 0.05	1.17	0.03	46.15	m
Х	0.63 ± 0.03	0.45 ± 0.01	1.08 ± 0.04	1.40	0.03	41.67	m
	0.63 ± 0.03	0.45 ± 0.02	1.08 ± 0.04	1.40	0.03	41.67	m
XI	0.63 ± 0.01	0.45 ± 0.03	1.04 ± 0.03	1.54	0.03	41.67	sm
	0.63 ± 0.02	0.36 ± 0.01	0.99 ± 0.03	1.75	0.03	36.36	sm
XII	0.63 ± 0.02	0.36 ± 0.01	0.99 ± 0.03	1.75	0.03	36.36	sm
	0.55 ± 0.01	0.36 ± 0.02	0.91 ± 0.03	1.53	0.02	39.56	sm
XIII	0.45 ± 0.02	0.45 ± 0.01	0.90 ± 0.03	1.00	0.02	50.00	m
	0.45 ± 0.03	0.45 ± 0.01	0.90 ± 0.04	1.00	0.02	50.00	m
XIV	0.45 ± 0.04	0.27 ± 0.01	0.72 ± 0.05	1.67	0.02	37.50	sm
	0.45 ± 0.04	0.27 ± 0.01	0.72 ± 0.05	1.67	0.02	37.50	sm
			GT=37.83± 0.91				

m = metacentric, sm = sub-metacentric chromosome.



Figs. 1-5. Orcein-stained mitotic interphase, prophase, metaphase, karyotype and Idiogram of *Colocasia esculenta* var. *esculenta*. 1. Interphase nuclei, 2. Prophase chromosome, 3. Mitotic metaphase chromosome, 4. Karyotype prepared from mitotic metaphase chromosomes, 5. Idiogram. In figure, arrow (\rightarrow) indicates nucleolus. Bar=5 μ .

A big and prominent nucleolus was observed in metaphase stage. Usually the nucleolus disappears at late prophase of mitosis. There is considerable evidence that it is not unusual for plant nucleoli to continue in mitotic metaphase or later. Persistent nucleolar materials were of frequent occurrence at prometaphase, metaphase, anaphase and even sometimes in telophase. In the majority of cases, nucleoli appeared as clear entities, usually almost round in shape. They varied in size from small, hardly detectable structures to large conspicuous ones. These observations suggested the late transcription of rDNA to rRNA and late transportation of rRNA from the nucleus to the cytoplasm. Persistent nucleolus was observed in few species such as Spartocera fusca (Cattani and Papeschi, 2004), Zea mays (Zirkle,

1928), *Ceiba pentandra* (Tijo, 1948), 11 species of *Gossypium* (Sultana and Alam, 2016) and 45 species of the family Gramineae (Walter and Emery, 1957). However, no available report on persistent nucleolus was found in the studied species. So the persistent nature of nucleolus is considered as a salient feature of *Colocasia esculenta* var. *esculenta*.

Chromosomal length

In the present study, the total length of 2n chromosome complement was recorded as $37.83\pm$ 0.91µm (Table 1,2). The range of chromosomal length was $0.72\pm0.05 - 2.34\pm0.05$ µm. Distance between small and large chromosomes was about 1.62µm indicating the gradual decrease of chromosome length (Table 1, 2). According to Stebbins (1971) the

asymmetric karyotype showed gradual decrease of chromosome length in their karyotype representing advance nature of germplasm. Average chromosomal length was about 1.48µm. The range of relative length was 0.02-0.06. Difference between relative lengths was 0.04 (Table 2). These features have been limitedly considered as cytological parameters in this species. Therefore, this is very important approaches by which this species could be characterized.

Table 2. Orcein stained karyotype analysis of*Collocasia esculenta* var. *esculenta*.

Cytologgenetical Parameters	Collocasia esculenta var. esculenta.		
2n	28		
Total length of chromosome complement (μm)	37.83 ± 0.91		
Range of individual chromosome length (μm)	0.72 ± 0.05 -2.34 ± 0.05		
Average chromosomal length (µm)	1.48		
Range of relative length	0.02-0.06		
Difference of relative length (DRL)	0.04		
Centromeric index range (CI)	31.77-50.00		
Centromeric formula	18m+10 sm		
Total form percentage (TF%)	41.56		
Karyotype symmetry index (Syi%)	71.11		
Karyotype Asymmetry index (AsK%)	58.44		

m = metacentric, sm = sub-metacentric chromosome.

Centromeric feature

Centromeric index range of Colocasia esculenta var. esculenta was 31.77-50.00. In case of centromeric formula, studied material was found to possess 18 metacentric and 10 sub-metacentric chromosomes. Combination of meta and sub-metacentric chromosomes were also found in previous reports (Sreekumari and Mathew, 1991; Parvin et al., 2008; Rattanavalee et al., 2018). Beside meta and submeta, acrocentric chromosome were reported earlier by different scientist (Sreekumari and Mathew, 1991; Parvin et al., 2008). All metacentric chromosome also reported (Parvin et al., 2008). The result of present study representing moderately asymmetric karyotype. Stebbins (1971) mentioned that the asymmetric karyotype indicate advance nature and from that point of view the studied plant is comparatively advanced nature from the different members of this species.

Karyotype symmetry and asymmetry index

In this study, the Total form percentage (TF%) was found 41.56% (Table 3). Karyotype symmetry index (Syi%) was 71.11% (Table 3). Karyotype symmetry index values decreased with increasing asymmetry. On the other hand, karyotype asymmetry index (AsK%) was 58.44% (Table 3). The value of karyotype asymmetry index increases with the increasing asymmetry. Thus the above findings indicating the moderately asymmetric nature of *Colocasia esculenta* var. *esculenta*.

These karyotypic features were not available in previous reports. So, these features could be helpful to identification of this species.

Therefore, the interphase nuclei, prophase chromosome and metaphase chromosome of Colocasia esculenta var. esculenta can be characterized cytogenetically with the above features.

References

Acevedo-Rodríguez P, Strong MT. 2005. Monocots and Gymnosperms of Puerto Rico and the Virgin Islands. Contributions from the United States National Herbarium **52**, p 415.

Alam SkS, Deen SS. 2002. Karyotype and isozyme analysis in three forms of *Colocasia esculenta* (Araceae). Bangladesh Journal of Botany **31(2)**, 95-98.

Alam SkS, Kondo K, Tanaka R. 1993. A chromosome study of eight orchid species in Bangladesh. La Kromsomo **11-71-72**, 2456-2464.

Ara H, Partha P, Hassan A. 2000. *Colocasia falax* Schott. (Araceae) - A new angiospermic record for Bangladesh. Bangladesh Journal of Plant Taxonomy **7(2)**, 85-87.

Barrau J. 1957. Taro (an annotated bibiography). South Pacific Commission Quarterly Bulletin **3**, 31-32.

Begum R, Alam SkS. 2004. Karyomorphological study in two orchid species. Dhaka University Journal of Biological Science **13(1)**, 99-101.

Int. J. Biosci.

Bonna IJ, Alam SkS, Sultana SS. 2018. Cytogenetical Characterization of *Acalypha indica* L. in Bangladesh. Dhaka University Journal of Biological Science **27(2)**, 183-189.

Cattani MV, Papeschi AG. 2004. Nucleolus organizing regions and semi-persistent nucleolus during meiosis in *Spartocera fusca* (Thunberg) (Coreidae, Heteroptera). Hereditas **140**, 105-111.

Coates DJ, Yen DE, Gaffey PM. 1988. Chromosome variation in taro, *Colocasia esculenta*: implications for origin in the Pacific. Cytologia **53**, 551-560.

Deen SS, Alam SkS. 2002. Comparative study in two forms of *Xanthosoma violaceum* (Araceae) through karyotype and isozyme analysis. Bangladesh Journal of Botany **31(1)**, 45-47.

Denham T. 2011. Early Agriculture and Plant Domestication in New Guinea and Island Southeast Asia. Current Anthropology **52(4)**, 379-395.

FAOSTAT. 2000. FAO statistical database: agricultural production of primary crops. Available from http://apps.fao.org/ default.htm. Accessed July 2001.

Fawzia R, Alam SkS. 2011. Fluorescent Karyotype Analysis in Four Varieties of *Solanum melongena* L. Cytologia **76(3)**, 345-351.

Ghani A. 1998. Medicinal plants of Bangladesh. Asiatic society of Bangladesh, Dhaka p 460.

Heng L. 1979. Araceae. Fl. Reipubl. Popularis Sin. 13(2), 1-210.

Huang SF, Zhao ZF, Chen ZY, Chen SJ Huang XX. 1989. Chromosome counts on one hundred species and infraspecific taxa. Acta Botanica Austro Sinica **5**, 161-176.

Hussain MU, Islam M, Afroz M, Sultana SS, Alam SkS. 2016. Karyotype and RAPD analysis of male and female *Coccinia grandis* L. from Bangladesh. Cytologia **81(3)**, 349-355. **Ivancic A, Lebot V.** 1999. Botany and genetics of New Caledonian wild taro, *Colocasia esculenta*. Pacific Science **53**, 273-285.

Kreike CM, Van EHJ, Lebot V. 2004. Genetic diversity of taro, *Colocasia esculenta* (L.) Schott, in Southeast Asia and the Pacifific. Theoretical and. Applied Genetics **109**, 761-768.

Kuruvilla KM, Dutt B, Roy RP. 1989. Karyomorphological investigations on aroids of North-Eastern Hills. Journal of Cytology and Genetics **24**, 13-22.

Lawrance GHM. 1966. Taxonomy of vascular plants IV. Indian Edition Oxford and IBH Publication Co., New Delhi p 36-40.

Matthews PJ. 2006. Written records of taro in the Eastern Mediterranean. In: Proceedings of the IVth International Congress of Ethnobotany (ICEB 2005) (edErtug ZF), Yeditepe University, Istanbul Turkey p 419-426.

Onwueme I. 1999. Taro cultivation in Asia and the Pacific. Bangkok, Thailand: Food and Agriculture Organization of the United Nations. Regional Office for Asia and the Pacific p 15

Parvin S, Kabir G, Ud-Deen MM, Sarker JK. 2008. Karyotype analysis of seven varieties of taro *Colocasia esculenta* (L.) Schott. from Bangladesh. Journal of Biological Science **16**, 15-18.

Rao N, Henry AN. 1996. The ethno botany of eastern ghats in Andhra Pradesh, India. Botanical Survey of India p 102.

Rattanavalee S, Surapon S, Piyaporn S. 2018. Comparative Karyotype Analysis in Five Strains of *Colocasia esculenta* (L.) Schott (Araceae) in Thailand. *Cytologia* **83(2)**, 169-173.

Sreekumari MT, Mathew PM. 1989. Karyomorphology of six cultivars of taro (*Colocasia esculenta* Schott). New Botanist **16**, 127-135.

Int. J. Biosci.

Sreekumari MT, Mathew PM. 1991. Effect of colchicine treatment in a triploid variety of taro [*Colocasia esculenta* (L.) Schott]. New Botany **18**, 211-215.

Sreekumari MT, Mathew PM. 1989. Karyomorphology of six cultivars of taro (*Colocasia esculenta* Schott). New Botanist **16**, 127-135.

Stebbins GL. 1971. Chromosomal evolution in higher plants. Addison-Wesley publishing company, California, USA p 208.

Subramanian D, Munian M. 1988. Cytotaxonomical studies in south Indian Araceae. Cytologia **53**, 59-66.

Sultana SS, Alam SkS. 2016. Karyomorphology of eleven varieties of *Gossypium hirsutum*. Bangladesh Journal of Botany **45(1)**, 151-159.

Tanaka R. 1971. Type of resting nuclei in Orchidaceae. Bot. Mag. Tokyo **84**, 118-122.

Tanimoto T, Matsumoto T. 1986. Variations of morphological characters and isozyme patterns in Japanese cultivars of *Colocasia esculenta* Schott and C. *gigantea* Hook. Japanese Journal of Breeding **36**, 100-111. **Tijo JH.** 1948. Notes on nucleolar conditions in *Ceiba petandra*. Hereditas **34**, 204-208.

Walter VB, Emery WHP. 1957. Persistent nucleoli and grass systematics. American Journal of Botany **44(7)**, 585-590.

Warasy AA, Alam SkS. 2009. Comparison of Orcein and CMA stained Karyotypes in Three Morphological Forms of *Typhonium trilobatum* L. Cytologia **74(3)**, 311-316.

Zhang CS. 1998. A preliminary study on making plant chromosomal specimens using peppermint oil compound as pretreatment agent. Journal of Wuhan Botanical Research **16(3)**, 280-282.

Zhang GM, Yang ZH. 1984. Studies on chromosome numbers of main cultivars of *Colocasia esculenta* in China. Acta Horticulturae Sinica **11(3)**, 187-190.

Zirkle C. 1928. Nucleus in root tip mitosis in Zea mays. Botanical Gazette **86**, 402-418.