

## International Journal of Agronomy and Agricultural Research (IJAAR)

ISSN: 2223-7054 (Print) 2225-3610 (Online) http://www.innspub.net Vol. 12, No. 5, p. 100-117, 2018

## **RESEARCH PAPER**

OPEN ACCESS

Improvement in nutrient contents of maize (Zea mays L.) by sulfur modulation under salt stress

Alia Riffat\*, Muhammad Sajid Aqeel Ahmad

Department of Botany, University of Agriculture, Faisalabad, Pakistan

Article published on May 30, 2018

Key words: Sulfur, Salt tolerance, Maize, Growth, Nutrient.

### Abstract

A key factor under saline conditions is the disturbance of osmotic potential that results in imbalance of nutrients in plants, while the application of sulfur not only improves the growth and nutrient status, but also improves the salt tolerance in plants. Thus, a study was carried out to determine the role of sulfur in salt tolerance of crop plants. The seeds of maize were sown in plastic pots filled with sand. Three levels of salinity (25, 50, 75 mM) and five levels of sulfur (20, 40, 60, 80, 100 mM) were applied at sowing time. Various growth parameters and nutrient contents were studied. The results showed that sulfur at 60 and 80 mM improved shoot and root length, fresh and dry weights, nutrient contents (K<sup>+</sup>, Ca<sup>2+</sup>, NO<sub>3</sub><sup>-</sup>, PO<sub>4</sub><sup>3-</sup>, SO<sub>4</sub><sup>2-</sup>, Ca<sup>2+</sup>/Na<sup>+</sup>, K<sup>+</sup>/Na<sup>+</sup>) and lowered Na<sup>+</sup> ions at all levels of salinity. For the determination of variation in salt tolerance potential, the phylogenetic tree was constructed by NTSys PC. Distance matrix showed that Agaitti 2003 and Pearl Basic showed high improvement in salt tolerance by sulfur application and showed more improvement in growth and nutrient contents in maize under salt stress conditions. While sulfur application has not much improved growth and nutrient contents in Pak Afgoi 2003 and Hybrid 1898 showing salt sensitivity as compared to other cultivars studied. In curx, sulfur application (60, 80 mM) has pronounced role in developing salt tolerance potential in maize cultivars by improving plant growth and nutrient contents.

\* Corresponding Author: Alia Riffat  $\boxtimes$  aliariffat@hotmail.com

#### Introduction

Among other abiotic stresses, salt stress has significant harmful effects on agricultural crops (Rengasamy, 2006; Godfray et al., 2010). In the world, salinity has affected 20 % of total cultivated area and 33 % of agricultural lands (Jamil et al., 2011). Salt stress affects many physiological and biochemical processes in the plants. Excess of salts disturb the uptake of water (Kocheva and Georgiev, 2003), that leads to the damages to the cellular organs (Bewley and Black, 1994), reduction in chlorophyll synthesis (McDonald, 2000), and lipid peroxidation (Soeda et al., 2005). The accumulation of salts in the soil horizon, replace and deplete the essential nutrients required for plant growth. As a result, plants become unable to tolerate salt stress (Epstein and Bloom, 2005). Jamil et al. (2011) reported that 50 % of arable land would be salinized in 2050. Therefore, such methods should be developed that induce salt tolerance in plants.

To overcome salt stress conditions, an economical way is to supply the essential nutrients to the plant, so the plant can properly carry out the physiological and biochemical processes inside the plant cell and tolerate the damages caused by salinity. Many inorganic nutrients, plant hormones and osmoprotectants are used for this purpose (Epstein and Bloom, 2005). Among the macronutrients, sulfur has significant contribution in salt tolerance as it helps the plants to uptake nutrients essential for growth and development (Fismes et al., 2000). Therefore, sulfur can improve plant health and vigor by reducing the toxic effects of salt stress. Sulfur is also a very necessary component of proteins, pantothenic acid, vitamin B1, acetyl CoA and biotin (Taiz and Zeiger, 2006). For the proper growth, plants require sulfur in ample quantity. Moreover, sulfur fertilization is low cost and produce fine quality of crop yield (Fismes et al., 2000).

Maize is very essential cereal all over the world after wheat and rice with respect to areas of cultivation and productivity (Sandhu *et al.*, 2007). Maize is enriched with vitamins (thiamine, niacin, riboflavin, pantothenic acid, pyridoxine), and nutrients essential for diet. It also contains oils, resin, salt, fibres and mucilage substances (Kumar and Jhariya, 2013). However, maize is very sensitive to salinity (Mass 1986). Therefore, its production has been severely affected due to salt stress. Maize responds to sulfur fertilization very fast (Ghosh *et al.*, 2000) because sulfur plays a pivotal role in developing salt tolerance in maize by regulating the plant metabolism. This study focuses on the improvement in salt tolerance in maize by sulfur application that not only regulates nutrient uptake and transport but also have significant role in growth and development of maize cultivars. Also, various maize cultivars are classified according to their salt tolerance potential based on growth and nutrient accumulation under saline conditions.

#### Materials and methods

#### Plan of study

A pot experiment was conducted for determining the effect of sulfur in improving growth attributes and ionic contents under saline condition. The experiment was conducted in the environmental biology and plant ecology lab of the Department of Botany, University of Agriculture, Faisalabad. The seeds of maize cultivars (varieties: Sadaf, MMRI, Pearl Basic, Agaitti 2003, Saiwal 2002, Pak Afgoi 2003 and hybrids: Yusafwala Hybrid, Hybrid 1898) were acquired from Maize and Millete Institute Sahiwal, Pakistan. The seeds were sown in pots made up of plastic and filled with thoroughly washed sand.

#### Treatment application

For the application of sulfur potassium sulphate (20, 40, 60, 80, 100 mM) was applied and sodium chloride (25, 50, 75 mM) was used for salinity treatment at sowing time. One set was kept without any treatment named as control (0 mM NaCl, 0 mM S). Hogland's nutrient solution was applied for irrigation during the study. The harvest of plants was taken after 14 days of treatment application for determining the changes in growth and ion accumulation.

#### Shoot and root length

The length of shoot and root was determined with the help of scale and average length of shoot and root of four plants of every replicate was calculated.

#### Shoot and root fresh weight

For determining the shoot and root fresh weight, electrical balance was used. The mean value of four plants of every replicate was calculated and average was taken.

#### Shoot and root dry weight

Shoot and root material was packed in paper envelops and kept in oven at 65°C for nearly two days. By using electrical balance the mean value were calculated.

#### Ionic contents

0.5g dried plant material was weighed and kept in digestion flasks. To each flask 5 ml of concentrated sulfuric acid was incorporated and kept overnight at room temperature. Then flasks were kept in digestion block for heating at 350°C for a period of 30 minutes. For adding H<sub>2</sub>O<sub>2</sub> all flasks were removed from incubation block. 1 ml of H2O2 was added in each flask and again placed in the incubation block. For complete digestion of the plant material, in the digestion flasks, H<sub>2</sub>O<sub>2</sub> was added in same manner. The plant material became digested in the acid solution and clear solution was obtained and then cooled at room temperature. Then distilled water was used for maintaining 50 ml volume of the extract. Then filtered and filled in the bottles labelled with proper treatment. This solution was used for the determination of the ionic contents (Wolf, 1982). Flame photometer (Jenway PFP-7) was used for the determination of Na+, K<sup>+</sup> and Ca<sup>2+</sup> contents in plant material. The standards (from 10, 20 to 100 ppm) of Na<sup>+</sup> K<sup>+</sup> and Ca<sup>2+</sup> ions were prepared and the curves were drawn. A comparison was done of the values of the standard curve and values from the flame photometer and real values were calculated in mg/g dry weight.

#### Phosphate

The phosphate contents were determined by following the procedure given by Yoshida (1976). First reagents were prepared. *Molybdate-vanadate* solution was prepared by dissolving 25g ammonium molybdate in 500 ml water. Separately 1.25g ammonium vanadate was dissolved in 500 ml of 1 N HNO<sub>3</sub>. Then equal volumes of two solutions were mixed to get Molybdate-vanadate solution.

Nitric acid (2N) was prepared by taking 10 ml of HNO<sub>3</sub> and volume was made up to 100ml with distilled water. For determination of phosphate contents, 0.5g plant material was homogenised and boiled in 5ml distilled water for 1h. Following filtrated volume was maintained to 50ml with distilled water. 1ml of the extract, 2 ml of 2N HNO3 and 1ml of distilled water were mixed, vortexed and cooled at room temperature for 20 minutes. The absorbance was noted at 420 nm on spectrophotometer (UV-1100) by using blank water. For the preparation of standards, stock solution of 25 mg/l PO43- was prepared by dissolving 0.11g of monobasic phosphate (KH<sub>2</sub>PO<sub>4</sub>) in 1L water. Then a series of solution (2.5, 5, 7.5, 10, 12.5, and 15 mg/l PO<sub>4</sub><sup>3</sup>) was prepared by adding1, 2, 3, 4, 5 and 6 ml of 25 mg/l PO<sub>4</sub><sup>3-</sup> and volume was maintained to 8 ml.

#### Nitrate

Nitrate contents in the plant tissue were determined by using the method given by Kowalenko and Lowe (1973). Firstly 0.01 % TCA was prepared. 0.247 g chromotropic acid dosodium salt (CTA) was mixed in 100 ml of conc. H<sub>2</sub>SO<sub>4</sub>. 0.01 % stock was prepared by maintaining 10 ml of CTA stock to 100 ml with H<sub>2</sub>SO<sub>4</sub>. For nitrate determination, 0.5g dried plant sample was boiled in 5 ml distilled water for 1 h, filtered and volume was maintained to 50 ml with distilled water. Then 3 ml extract was mixed in 7 ml working CTA solution and vortexed. The absorbance was measured at 430 nm on spectrophotometer (UV-1100) by using water as blank. For the preparation of standard solution, stock was prepared by dissolving 0.7216 g of KNO3 in 1L water. Standard series (10, 20, 30, 40, 50 and 100 mg/l NO<sub>3</sub>-) was prepared by diluting the stock solution.

#### Sulfate

Sulfate contents were found by following the method of Tandon (1993). Two reagents were prepared; Barium chloride/Polyvinyl alcohol and acid mixture. Barium chloride/Polyvinyl alcohol was made by dissolving 60g of BaCl<sub>2</sub>.2H<sub>2</sub>O in 500 ml distilled water. Separately, 2g of polyvinyl alcohol was mixed in 400 ml water and both solutions were mixed after cooling. Finally volume was made to 100 ml with distilled water. Acid mixture was made by mixing 50 ml glacial acetic acid, 20 ml 85 % orthophosphoric acid and 6 ml concentrated  $H_2SO_4/H_2O$  (1:1000) and 800 ml distilled water. The volume was maintained to 1L using distilled water. For sulphate determination, 5 ml of tested solution, 5 ml of acid mixture, and 5 ml of Barium chloride/Polyvinyl alcohol were mixed thoroughly, vortexed, allowed to stand for 30 seconds and absorbance was noted at 420 nm using spectrophotometer (UV-1100).

#### Statistical analysis

The study was conducted in Completely Randomized Design (CRD) in three factor factorial with three replicates. Microsoft excel was used for the preparation of graphs. Co-stat software was used for the performance of Analysis of Variance (ANOVA). For the determination of the variation in the salt tolerance ability of cultivars ameliorated by sulfur, cluster analysis was done by NTSysPC software (v2.10m). Phylogenetic tree was constructed on the basis of distance coefficient by using SAHN (Sequential Agglomerative Hierarchic and Non-over lapping) procedure.

#### Results

Effect of sulfur on growth parameters under salinity The use of sulfur revealed a significant enhancement in shoot and root length, fresh and dry weights of all studied maize cultivars (Fig. 1-6). It was revealed from statistically significant V × S interaction (Table 1). Generally, sulfur improved the maize growth by developing salt tolerance at all levels (20, 40, 60, 80, 100 mM). However, 100 mM sulfur was not much efficient in this respect in improving the maize growth but is significant as compared to control (Fig. 1-6). This result was evident from statistically significant  $Sa \times S$  interaction (Table 1). The application of sulfur improved the maize growth by lowering the toxic effects of salt stress. This was evident from statistically significant V × Sa × S interaction (Table 1). Among maize cultivars used in this study, Agatti 2003 showed salt tolerance by reducing the harmful effects of higher levels of sulfur and salinity. In comparison Pak Afgoi 2003 proved salt sensitive as its growth was much lowered by higher levels of salinity and sulfur.

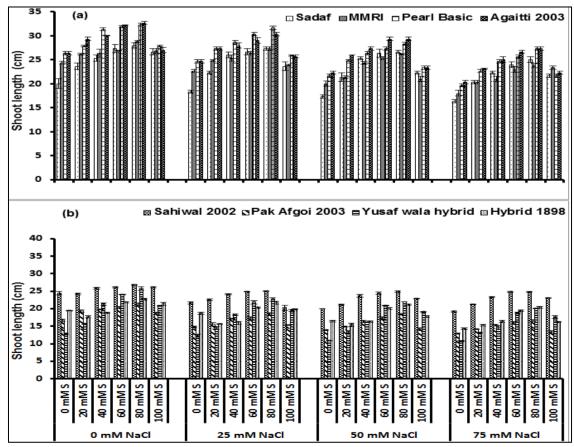


Fig. 1. Effect of different levels of sulfur (S) and NaCl on shoot length of different maize cultivars (Zea mays L.).

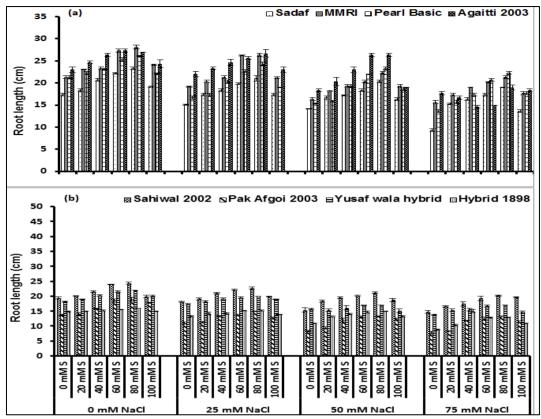
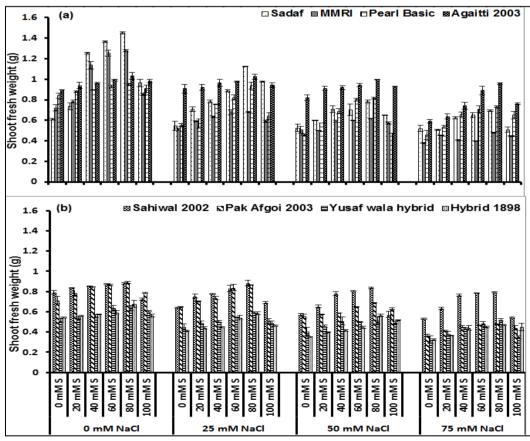
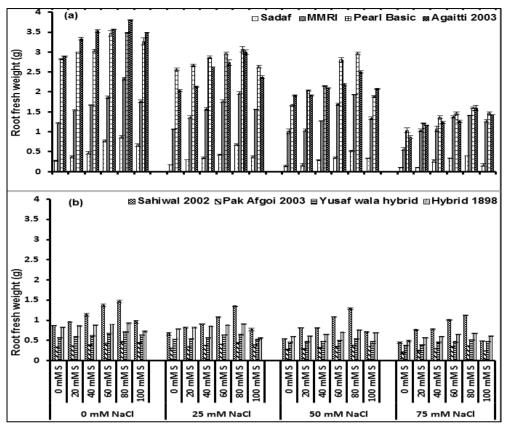


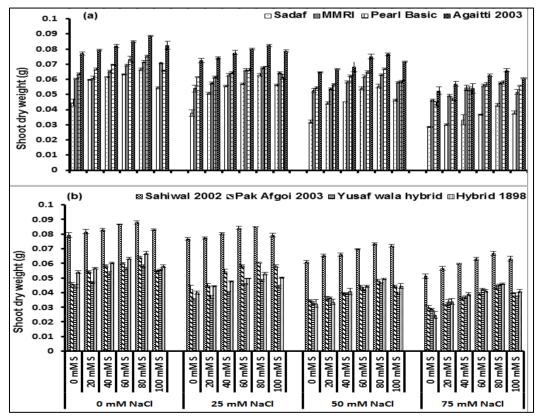
Fig. 2. Effect of different levels of sulfur (S) and NaCl on root length of different maize cultivars (Zea mays L.).



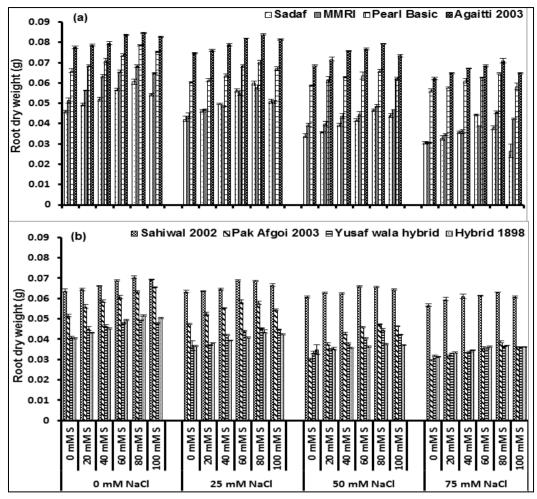
**Fig. 3.** Effect of different levels of sulfur (S) and NaCl on shoot fresh weight of different maize cultivars (*Zea mays* L.).



**Fig. 4.** Effect of different levels of sulfur (S) and NaCl on root fresh weight of different maize cultivars (*Zea mays* L.).



**Fig. 5.** Effect of different levels of sulfur (S) and NaCl on shoot dry weight of different maize cultivars (*Zea mays* L.).



**Fig. 6.** Effect of different levels of sulfur (S) and NaCl on root dry weight of different maize cultivars (*Zea mays* L.).

**Table 1.** Mean squares from analysis of variance (ANOVA) of the data for morphological parameters and nutrien contents of maize subjected to salt stress and sulfur application.

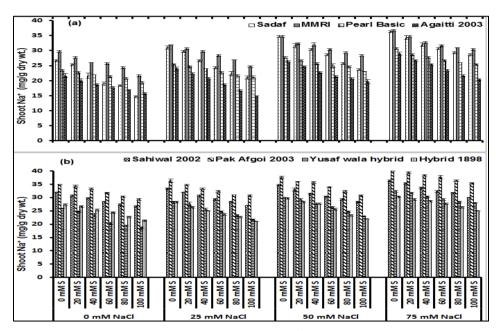
SOV	df	SL	RL	SFW	RFW	SDW	RDW	Leaf Na+ content
Variety (V)	7	1194.04 ***	832.37 ***	1.44 ***	49.45 ***	0.011 ***	0.012 ***	1214.17 ***
Salinity (Sa)	3	479.50 ***	765.15 ***	2.33 ***	14.38 ***	0.011 ***	0.0066 ***	1069.85 ***
Sulfur (S)	5	526.47 ***	344.46 ***	0.71 ***	3.07 ***	0.0024 ***	0.0012 ***	612.96 ***
V × Sa	21	6.56 ***	18.10 ***	0.11 ***	2.14 ***	1.50e-4 ***	1.95e-4 ***	22.58 ***
$V \times S$	35	19.60 ***	7.61 ***	0.024 ***	0.28 ***	2.44e-5 ***	2.15e-5 ***	3 5.11 ***
Sa × S	15	2.36 ***	1.58 ***	0.0085 ***	0.16 ***	1.11e-5 ***	1.68e-5 ***	1.11 ***
$V \times Sa \times S$	105	2.18 ***	3.23 ***	0.018 ***	0.11 *	1.16e-5 ***	8.87e-6 ***	0.93 ***
Error	384	0.40	0.34	0.44	0.091	3.55e-6	1.47e-6	0.38
SOV	df	Leaf K+	Leaf Ca <sup>2+</sup>	Leaf	Leaf	Leaf NO3 <sup>-</sup>	Leaf PO <sub>4</sub> 3-	Leaf SO42-
		content	content	Ca <sup>2+</sup> /Na <sup>-</sup> content	K+/Na+ content	content	content	content
Variety (V)	7	2412.71 ***	54.76 ***	0.19***	$12.3^{***}$	2.57***	20.14***	26123.38***
Salinity (Sa)	3	1239.09 ***	72.70 ***	0.25***	8.89***	3.34***	4.06***	15963.83***
Sulfur (S)	5	122.62 ***	1.89 ***	0.03***	2.84***	17.26***	3.54***	5924.52***
V × Sa	21	2.67 ***	1.78 ***	0.006***	0.21***	0.32***	0.62***	577.36***
$V \times S$	35	0.15 ns	0.21 ns	0.001***	0.07***	0.23***	0.35***	170.61***
Sa x S	15	0.13 ns	0.12 ns	0.002***	0.06***	0.24***	0.15***	73.79ns
$V \times Sa \times S$	105	0.18 ns	0.05 ns	1.57e-4ns	0.011***	0.04***	0.08***	50.62ns
Error	384	0.21	0.41	5.512e-4	0.002	0.014	0.021	47.62

\*, \*\*, \*\*\* = significant at 0.05, 0.01 and 0.001 levels, respectively. ns = non-significant, e=exponent.

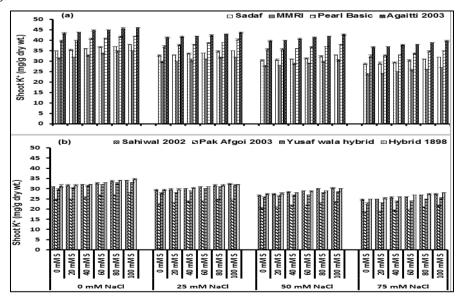
Abbreviations: Shoot length (SL), Root length (RL), Shoot fresh weight (SFW), Root Fresh Weight (RFW), Shoot dry weight (SDW), Root dry weight (RDW), Sodium(Na<sup>+</sup>), Potassium (K<sup>+</sup>), Calcium (Ca<sup>2+</sup>), Nitrate ( $NO_3^{-}$ ), Phosphate ( $PO_4^{3-}$ ), Sulfate ( $SO_4^{2-}$ ).

Effect of sulfur on nutrient contents under salinity

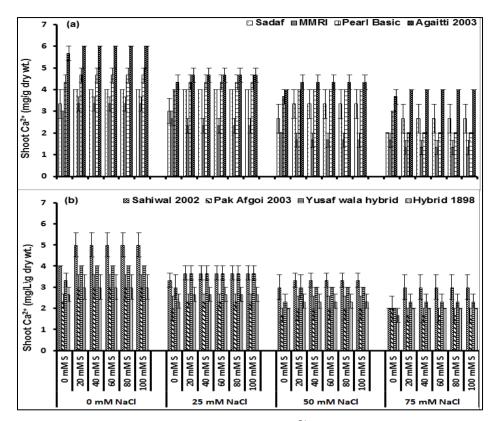
In the present study, application of sulfur improved the calcium, potassium, nitrate, phosphate and sulphate contents and reduced sodium contents as compared to plants with no sulfur application. Results revealed that salinity increased sodium (Na) contents in maize plants. It was evident from statistically significant V × Sa, V × S, Sa × S, and V × Sa × S interactions (Table 1). Sulfur augmentation improved the salt tolerance in maize plants; it was due to the reason that sulfur lowered the sodium uptake (Fig. 7) and significantly improved the uptake of potassium (K<sup>+</sup>) and calcium (Ca<sup>2+</sup>), nitrate (NO<sub>3</sub><sup>-</sup>), phosphate (PO<sub>4</sub><sup>3-</sup>), sulphate (SO<sub>4</sub><sup>2-</sup>), Ca<sup>2+</sup>/Na<sup>+</sup> and K<sup>+</sup>/Na<sup>+</sup> in maize plants (Table 1). Sulfur application at 60 mM and 80 mM levels showed much effective in reducing the effects of salt stress (Fig. 8-14). Overall salt tolerant maize cultivar Agaitti 2003 accumulated less sodium (Na) contents as compared to Pak Afgoi 2003 (Fig. 7). Also the application of sulfur improved the salt tolerance in Pak Afgoi 2003 by improving the growth parameters and ionic contents.



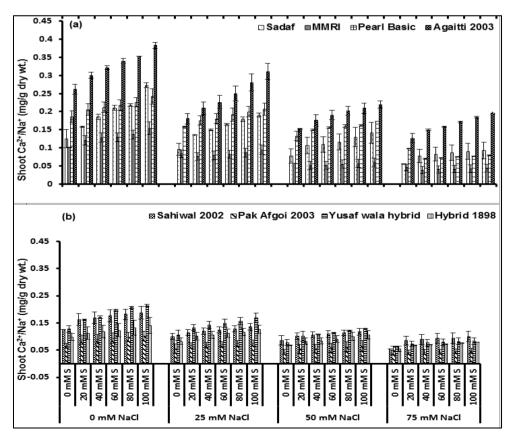
**Fig.** 7. Effect of different levels of sulfur (S) and NaCl on shoot Na<sup>+</sup> contents of different maize cultivars (*Zea mays* L.).



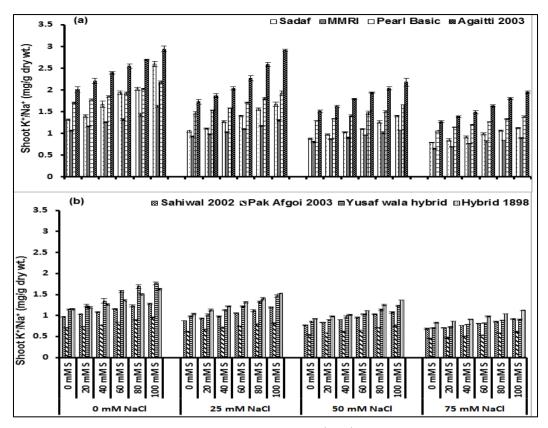
**Fig. 8.** Effect of different levels of sulfur (S) and NaCl on shoot  $K^+$  contents of different maize cultivars (*Zea mays* L.).



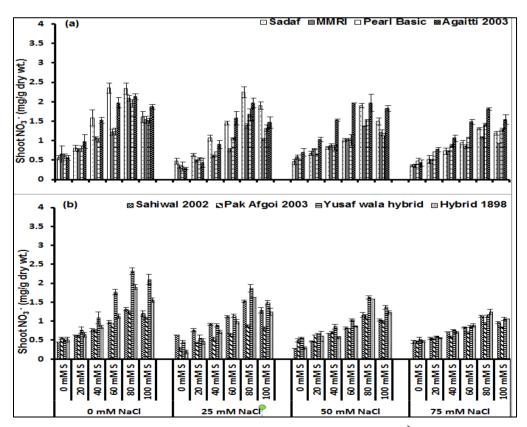
**Fig. 9.** Effect of different levels of sulfur (S) and NaCl on shoot Ca<sup>2+</sup> contents of different maize cultivar (*Zea mays* L.).



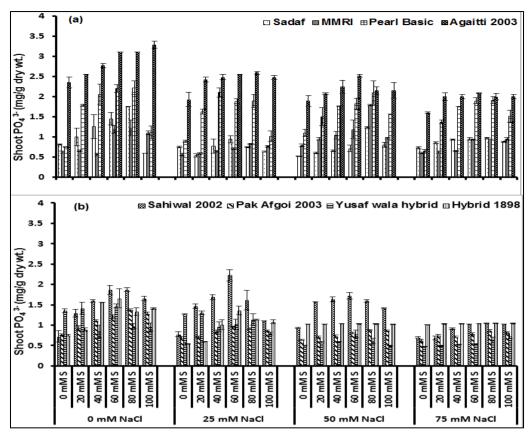
**Fig. 10.** Effect of different levels of sulfur (S) and NaCl on shoot  $Ca^{2+}/Na^{+}$  of different maize cultivar (*Zea mays* L.).



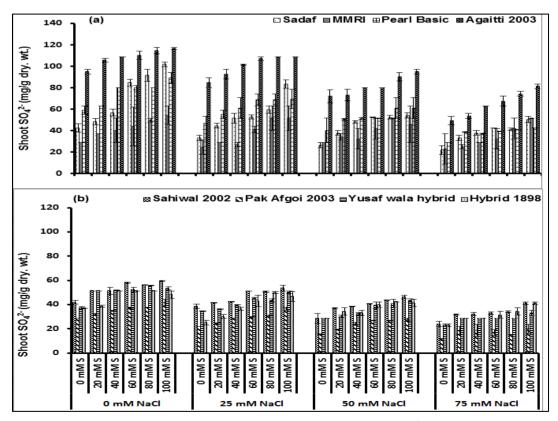
**Fig. 11.** Effect of different levels of sulfur (S) and NaCl on shoot  $K^+/Na^+$  contents of different maize cultivars (*Zea mays* L.)



**Fig. 12.** Effect of different levels of sulfur (S) and NaCl on shoot nitrate  $(NO_3^{-})$  contents of different maize cultivars (*Zea mays* L.).



**Fig. 13.** Effect of different levels of sulfur (S) and NaCl on shoot phosphate  $(PO_4^{3-})$  contents of different maize cultivars (*Zea mays* L.).



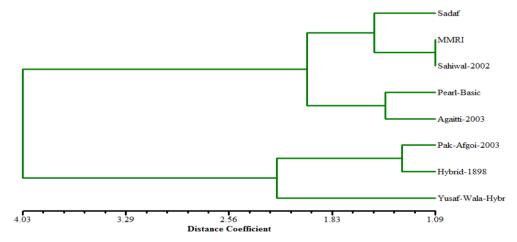
**Fig. 14.** Effect of different levels of sulfur (S) and NaCl on shoot sulfate  $(SO_4^{2-})$  contents of different maize cultivars (*Zea mays* L.).

Determination of variation in salt tolerance of maize cultivars

The data of all studied growth attributes and ionic contents of maize cultivars was fed to NTSysPC software (v2.10m) for constructing phylogenetic tree created on SAHN method. The tree was formed on the basis of distance coefficient (DC). Less distance between two groups means higher variation in the salt tolerance ability modulated by sulfur. And more distance in group means that sulfur application has induced less variation in salt tolerance ability of maize cultivars.

# Determination of variation based on growth parameters

The cluster analysis clearly divided all maize cultivars in two groups (DC=4.03) showing higher variation in maize growth (Table 2). Salt tolerant group (Sadaf, MMRI, Sahiwal-2002, Pearl Basic, Agatti-2003) and salt sensitive group (Pak Afgoi 2003, Hybrid-1898, Yusaf Wala Hybrid) (Fig. 15). In salt tolerant group, Agaitti 2003 and Pearl Basic branched at higher distance (DC=1.5) reflecting their higher salt tolerant ability by sulfur application. MMRI and Sahiwal branched at equal distance (DC=1.09) categorised as moderately salt tolerant and sadaf at lower distance (DC=1.645) ranked as salt sensitive variety. In salt sensitive group, Yusaf wala hybrid was branched at most distance (DC=2.385) categorized as moderately salt sensitive. Other two cultivars i-e Pak Afgoi 2003 and Hybrid 1898 branched at least distance (DC=1.3) showing salt sensitive cultivars (Table 2). Overall, among all cultivars, Agaitti 2003 and Pearl Basic responded to sulfur application by improving growth under salt stress condition while Pak Afgoi 2003 and Hybrid 1898 at least improved by sulfur fertilization.



**Fig. 15.** The clustering analysis based on phylogenetic distance formed by NTS Sys PC for growth of maize (*Zea mays* L.) cultivars ameliorated by sulfur under saline conditions.

## Determination of variation based on nutrient contents

Sulfur application improved the beneficial ionic contents (K<sup>+</sup>, Ca<sup>2+</sup>, NO<sub>3</sub><sup>-</sup>, PO<sub>4</sub><sup>3-</sup>, SO<sub>4</sub><sup>2-</sup>, Ca<sup>2+</sup>/Na<sup>+</sup>, K<sup>+</sup>/Na<sup>+</sup>) and lower down the sodium contents (Na<sup>+</sup>) by developing salt tolerance potential in all maize cultivars. However, the tree representing ionic contents showed variable response to sulfur application (Fig. 16). Two groups were observed in this tree (DC=7.36). Salt tolerant group (Agitti 2003, Pearl Basic) and salt sensitive group (Sadaf, Yusaf wala hybrid, Hybrid 1898, MMRI, Sahiwal 2002, Pak Afgoi 2003).

In salt tolerant group Agaitti 2003 and Pearl Basic branched at higher distance (DC=3.29) showing highly salt tolerant. In salt sensitive group, MMRI and Sahiwal 2002 branched at equal distance (DC=1.43) categorized as least salt sensitive.

Yusaf wala hybrid and Hybrid 1898 branched at least but equal distance (DC=1.8) ranked as moderately salt sensitive, Sadaf branched at lower distance (DC=3) showed salt sensitive variety and Pak Afgoi 2003 branched at least distance coefficient (DC=5.6) so ranked as highly salt sensitive variety (Table 2).

Growth param	eters			Ionic contents				
Group	Cultivars	Distance	Salt	Group	Cultivars	Distance	Salt tolerance	
		coefficient (DC)	tolerance ability			coefficient (DC)	ability	
Salt tolerant	Agaitti 2003	1.5	Highly salt	Salt	Agaitti	3.29	Highly salt	
	_	U U	tolerant	tolerant	2003		tolerant	
	Pearl Basic	1.5	Highly salt		Pearl Basic	3.29	Highly salt	
	MMRI	1.09	tolerant Moderately	Salt	MMRI	1.43	tolerant Least salt	
	MINICI	1.09	salt tolerant	sensitive	MININI	1.43	sensitive	
	Sahiwal 2000	1.09	Moderately salt tolerant		Sahiwal	1.43	Least salt	
	a 1.6				2002	0	sensitive	
	Sadaf	1.645	Least salt tolerant		Yusafwala Hybrid	1.8	Moderately salt sensitive	
Salt sensitive	Yusafwala Hybrid	2 385	Moderately		Hybrid	1.8	Moderately salt	
buit sensitive			salt sensitive		1898	1.0	sensitive	
	Pak Afgoi 2003	1.3	Highly salt		Sadaf	3	Highly salt	
	Hybrid 1898		sensitive			- (	sensitive	
	Hybrid 1898	1.3	Highly salt sensitive		Pak Afgoi 2003	5.6	Highly salt sensitive	
						S	adaf	
					MMRI			
					1			
						s	ahiwal-2002	
					Ч			
Г		-				Γ <sup>Y</sup>	usaf-Wala-Hybr	
							lybrid-1898	
						-1	lyblid-1898	
						P	ak-Afgoi-2003	
						P	earl-Basic	
_						A	gaitti-2003	
7.36	5	5.88	4.40		2.92	1.43		
			Distance Coeffici	ient				

**Table 2.** The clustering based on growth attributes and ionic contents of maize (*Zea mays* L.) cultivars as ameliorated by sulphur application under salinity.

**Fig. 16.** The clustering analysis based on phylogenetic distance formed by NTS Sys PC for ionic contents of maize (*Zea mays* L.) cultivars ameliorated by sulfur under saline conditions.

#### Discussion

Results showed that salt stress reduced shoot and root length. It might be due to high concentration of Na<sup>+</sup> and Cl<sup>-</sup> in the rooting zone (Zhang *et al.*, 2010) that lower water potential in growing media, resulting in loss of cell turgor. This causes reduction in photosynthetic rate, cell division and cell elongation that ultimately reduces plant length (Zekri, 1991; Ali-Dinar *et al.*, 1999; Ebert *et al.*, 2002). The reduction in root and shoot length by imposition of salinity has been reported in various studies (Werner and Frankelstein, 1995; Okcu *et al.*, 2005; Atak *et al.*, 2008; Asaadi, 2009). However, application of sulfur improved root and shoot length in the current study. The increase in root length by the application of sulfur might be due to the reason that sulfur enhances the cell division and cell elongation in the meristematic region of the plant. Therefore, application of sulfur keeps root system very healthy (Chandel *et al.*, 2002). Diepenbrock (2000) reported that sulfur maintains health of root which in turn transports the nutrient to the upper parts of the plant that results in gaining in the fresh biomass of the plant. Bejandi *et al.* (2009) reported that sulfur application significantly enhanced the shoot length in soyebean.

Salinity reduced fresh and dry weight of all maize cultivars. The findings of current investigation are in accordance to earlier studies (Netondo et al., 2004; Ashraf et al., 2008; Akhzari et al., 2012; Mantri et al., 2012). In various plants the reduction in fresh and dry biomass by salt stress has been reported e.g. Zea mays L., Raphanus sativus L., Kyllinigia peruviana L., Bruguiera gymnorrhiza and Brassica campestris L. (Cicek and Cakirlar, 2002; Jamil et al., 2007; Ha et al., 2008; Rui et al., 2009; Memon et al., 2010). Sulfur has very significant role in formation of proteins and a number of metabolites necessary for increase in fresh and dry weights of plants (Ali et al., 1990; Zhao et al., 1993). Previous studies revealed that use of sulfur improved shoot and root fresh and dry weights (Gilbert et al., 1984).

Salinity causes reduction in water potential, imbalance in nutrient composition and transport and ultimately disrupting ionic homeostasis in plants (Chinnusamy et al., 2005; Parida and Das, 2005; Genc et al., 2007; Zhang et al., 2013). While sulfur application enhances the nutrient availability in plants under salt stress conditions which has been supported by previous studies (Aslam et al., 2001; Prasad, 2003; Ali et al., 2008). In this study salt stress increased the Na<sup>+</sup> contents and reduced the K<sup>+</sup> contents in all studied maize cultivars. Under salt stress conditions, Na<sup>+</sup> contents become very high that compete with K<sup>+</sup> ions and reduces its uptake. This reduction in K<sup>+</sup> uptake causes the reduction in K<sup>+</sup>/Na<sup>+</sup> ratio (Kaya *et al.*, 2007). In this study, using sulfur, the harmful effects of salt stress were reduced by reducing the Na<sup>+</sup> and improving K<sup>+</sup> contents in maize plants. This property is due to the reason that sulfur has such metabolites that modulate the physiological and molecular process under saline conditions. This modulation creates salt tolerance in crop plants (Khan et al., 2014). Reich et al. (2016) found that the use of sulfur improved the potassium contents in plants. It also improves the K<sup>+</sup>/Na<sup>+</sup> ratio inside the plant. Hence, developing salt tolerance, as the rise in K<sup>+</sup>/Na<sup>+</sup> ratio indicates the salt tolerance in plants (Shirazi et al., 2002; Kaya et al., 2002).

Results showed that salinity reduced the Ca2+ ions and Ca<sup>2+</sup>/Na<sup>+</sup> ratio in plants. It was supported by previous studies. Salt stress reduces the Ca2+ ions that reduce the Ca<sup>2+</sup>/Na<sup>+</sup> ratio in the plant (Khan, 2001), while sulfur application improved the Ca2+ and Ca2+/Na+ ratio in maize cultivars. Aulakh and Dev (1978) found positive correlation between sulfur and calcium. Sulfur improves the availability of Ca<sup>2+</sup> ions to the plant that lower the toxic effects of salinity on plant health and vigor (Badr et al., 2002). Improvement in Ca<sup>2+</sup> ions not only exclude Na<sup>+</sup> but also helps the plant to efficiently use nitrogen (Aslam et al., 2001; Mahmood et al., 2009). The adequate amount of sulfur maintains Ca2+/Na+ ratio in the plants that reduce the toxic effects of salinity and helps in establishment of crop plants (Aslam et al., 2001).

In this study it was found that salinity reduced  $NO_{3}$ , PO<sub>4</sub><sup>3-</sup>, and SO<sub>4</sub><sup>2-</sup> contents in plants while sulfur application enhanced the salt tolerance in maize plants by balancing the nutrient concentration, uptake and transport of beneficial ions (NO<sub>3</sub><sup>-</sup>, PO<sub>4</sub><sup>3-</sup>, and SO<sub>4<sup>2-</sup></sub>). These findings have been reported in previous studies. Perez-Alfocea et al. (2015) reported that salt stress reduced the nitrate and total nitrogen contents in tomato plants. However, sulfur application improves the nitrate contents in plants by improving the activities of the enzymes involved in nitrate uptake. Prosser et al. (2001) found that a reduction in sulfur lowered the nitrate reductase activity in maize and spinach (Prosser et al., 2001). Salinity also reduces the phosphate contents in maize plants. It was supported by findings of Aslam et al. (1996), who found that salinity reduces the phosphate contents in rice. Similarly, Turhan and Atilla (2004) evaluated that by increasing the NaCl level from 500 to 2000m g/L the phosphorus contents were decreased. The application of sulfur in low quantity improved the phosphorous uptake in the plant which improved the nutrient deficiency under stress conditions. It was related to earlier researches. Randhawa and Arora (2000) reported that the sulfur and phosphorous has significant positive interaction. The sulfate contents were reduced by high level of salinity in comparison to the plants with no salt application.

It has been reported that the NaCl salinity reduced the  $SO_4^{2-}$  contents in *Brassica rapa*. Moreover, sulfur contents were increased by application of sulfate salts (Reich *et al.*, 2017). For the determination of relative improvement in salt tolerance ability by sulfur application, cluster analysis was constructed and distance coefficients (DC) was constructed by SAHN method. This procedure is very helpful in determination of cultivar variation to salt tolerance modulated by sulfur. In previous studies, such methods have been used for various germination and growth experiments (Saboora *et al.*, 2006; Ali *et al.*, 2014).

#### Conclusion

In conclusion, although applied salinity decreased the growth of maize plants, the use of sulfur was found efficient in alleviation of harmful effect of salt stress. Sulfur improves availability of macronutrients in the plants which are key regulators in alleviation of salt stress. The effectiveness of the use of applied level of S was from 60 to 80 mM concentration. So, sulfur could be applied at about 60 mM to 80 mM for improving nutrient contents and ultimately growth of maize under saline conditions.

#### Acknowledgement

The authors acknowledge the partial financial assistance provided by University of Agriculture Faisalabad under UAF Ph.D. Research Grant (2907/ORIC) for completion of this study.

#### References

Akhzari D, Sepehry A, Pessarakli M, Barani H. 2012. Studying the effects of salinity, aridity and grazing stress on the growth of various halophytic plant species (*Agropyron elongatum, Kochia prostrate* and *Puccinellia distans*). World Applied Sciences Journal **17**, 1278-1286.

Ali A, MachadoVS, Hamill AS. 1990. Osmoconditioning of tomato and onion seeds. Scientia Horticulturae **43**, 213-224. DOI: 10.1016/0304-4238 (90)90093-t.

Ali MN, Yeasmin L, Gantait S, Goswami R, Chakraborty S. 2014. Screening of rice landraces for salinity tolerance at seedling stage through morphological and molecular markers. Physiology and Molecular Biology of Plants **20**, 411-423. DOI: 10.1007/s12298-014-0250-6. Ali R, Khan MJ, Khattak RA. 2008. Response of rice to different sources of sulfur (S) at various levels and its residual effect on wheat in rice-wheat cropping system. Soil and Environmental Sciences **27**, 131-137.

**Ali-Dinar HM, Ebert G, Ludders P.** 1999. Growth, chlorophyll content, photosynthesis and water relations in guava (*Psidium guajava* L.) under salinity and different nitrogen supply. Gartenbauw is Senschaf **64**, 54-59.

**Asaadi AM.** 2009. Investigation of salinity stress on seed germination of *Trigonella foenum-graecum*. Research Journal Biological Sciences **4**, 1152-1155.

Ashraf M, Athar HR, Harris PJC, Kwon TR. 2008. Some prospective strategies for improving crop salt tolerance. Advance Agronomy **97**, 45-109.

Aslam M, Flowers TJ, Qureshi RH, Yeo AR. 1996. Interaction of phosphate and salinity on the growth and yield of rice (*Oryza sativa* L.). Journal of Agronomy and Crop Science **176**, 249-258. DOI: 10.1111/j.1439-037X.1996.tb00469.x

Aslam MI, Mahmood H, Qureshi RH, Nawaz S, Akhtar J, Ahmad Z. 2001. Nutritional role of calcium in improving rice growth and yield under adverse conditions. International Journal of Agriculture and Biology **3**, 292-297.

Atak M, Kaya MD, Kaya G, Kaya M, Khawar KM. 2008. Dark green colored seeds increase the seed vigor and germination ability in dry green pea (*Pisum sativum* L.). Pakistan Journal of Botany **40**, 2345-2354.

DOI: 10.1007/s10531-010-9833.

**Aulakh MS, Dev G.** 1978. Interaction effect of calcium and Sulfur on the growth and nutrient composition of alfalfa (*Medicago sativa* L. pers.), using<sup>35</sup>S. Plant Soil **50**,125-134.

**Badr Z, Ali A, Salim M, Niazi BH.** 2002. Role of Sulfur for potassium/sodium ratio in sunflower under saline conditions. Helia **25**, 69-78.

**Bejandi TK, Sedghi M, Sharifi RS, Namvar A, Molaei P.** 2009. Seed priming and Sulfur effects on soybean cell membrane stability and yield in saline soil. Pesq agropec bras Brasília **44**, 1114-1117.

**Bewley JD, Black M.** 1994. Seeds: physiology of development and germination. New York: Plenum 1994.

**Chandel RS, Sudhakar PC, Singh K.** 2002. Direct and residual effect of Sulfur on Indian mustard (*Brassica juncea* L.) in rice (*Oryza sativa* L.). Indian Journal of Agricultural Sciences **72**, 230-232.

**Chinnusamy V, Jagendorf A, Zhu JK.** 2005. Understanding and improving salt tolerance in plants. Crop Science **45**, 437-448.

**Cicek N, Cakirlar H.** 2002. The effect of salinity on some physiological parameters in two maize cultivars. Bulgarian Journal of Plant Physiology **28**, 66-74.

**Diepenbrock W.** 2000. Yield analysis of winter oilseed rape (*Brassica napus* L.): A review. Field Crop Research **67**, 35-49.

DOI: 10.1016/S0378-4290 (00)00082-4.

Ebert G, Eberle J, Ali-Dinar H, Lüdders P. 2002. Ameliorating effects of  $Ca(NO_3)_2$  on growth, mineral uptake and photosynthesis of NaCl-stressed guava seedlings (*Psidium guajava* L.). Scietia Horticulturae **93**, 125-135.

**Epstein E, Bloom AJ.** 2005. Mineral Nutrition of Plants: Principles and Perspectives, 2nd Edn. Sunderland: Sinauer Associates Inc.

**Fismes J, Vong PC, Guckert A, Frossard E.** 2000. Influence of Sulfur on apparent N-use efficiency, yield and quality of oilseed rape (*Brassica napus* L.) grown on a calcareous soil. European Journal of Agronomy **12**, 127-41.

**Genc Y, McDonald GK, Tester M.** 2007. Reassessment of tissue Na<sup>+</sup> concentration as a criterion for salinity tolerance in bread wheat. Plant Cell Environment **30**, 1486-1498.

DOI: 10.1111/j.1365 -3040.2007.01726.x.

**Ghosh PPK, Hati KM, Mandal KG, Misra AK, Chaudhary RS, Bandyopadhyay KK.** 2000. Sulfur nutrition in oilseed based cropping systems. Fertilizer News **45**, 27-40.

**Gilbert MA, Robson AD.** 1984. The effect of Sulfur supply on the root characteristics of subterranean clover and annual ryegrass. Plant and Soil 77, 377-380.

Godfray HCJ, Beddington JR, Crute IR, Haddad L, Lawrence D, Muir JF, Pretty J, Robinson S, Thomas SM, Toulmin C. 2010. Food security: the challenge of feeding 9 billion people. Science **327**, 812-818.

Ha E, Ikhajiagba B, Bamidele JF, Ogic-odia E. 2008. Salinity effects on young healthy seedling of *kyllingia peruviana* collected from escravos, Delta state. Global Journal Environment Research **2**, 74-88.

Jamil A, Riaz S, Ashraf M, Foolad MR. 2011. Gene expression profiling of plants under salt stress. Critical Review in Plant Sciences **30**, 435-458.

Jamil M, Rha ES. 2007. Gibberellic acid (GA3) enhances seed water uptake, germination and early seedling growth in sugar beet under salt stress. Pakistan Journal of Biological Sciences **10**, 654-658. DOI: 10.3923/pjbs.2007.654.658.

Kaya C, Kirnak H, Higgs D, Saltali K. 2002. Supplementary calcium enhances plant growth and fruit yield in strawberry cultivars grown at high (NaCl) salinity. Scientia Horticulture **93**, 65-74. DOI: 10.1016/S0304-4238(01)00313-2.

**Kaya C, Tuna AL, Ashraf M, Altunlu H.** 2007. Improved salt tolerance of melon (*Cucummis melo* L.) by the addition of proline and potassium nitrate. Environment and Experimental Botany **60**, 397-403. DOI: 10.1016/j.envexpbot.2006.12.008.

**Khan MA, Ungar IA.** 2001. Alleviation of salinity stress and the response to temperature in two seed morphs of *Halopyrum mucronatum* (Poaceae). Australian Journal Botany **49**, 777-783. Khan NA, Khan MIR, Asgher M, Fatma M, Masood A. 2014. Salinity Tolerance in Plants: Revisiting the Role of Sulfur Metabolites. Journal of Plant Biochemistry and Physiology **2**, 120. DOI: 10.4172/2329-9029.1000120.

Kocheva K, Lambrev P, Georgiev G, Goltsev V, Karabaliev M. 2004. Evaluation of chlorophyll fluorescence and membrane injury in the leaves of barley cultivars under osmotic stress. Bioelectrochemistry **63**, 121-124. DOI: 10.1016/j.bioel echem.2003.09.020.

Kowalenko CG, Lowe LE. 1973 Determination of nitrates in soil extracts. Soil Science Society of America Proceedings **37**, 660.

**Kumar D, Jhariya NA.** 2013a. Nutritional, medicinal and economical importance of corn: A mini review. Research Journal of Pharmaceutical Sciences **2**, 7-8.

Mahmood IA, Salim M, Ali A, Arshadullah M, Zaman B, Mir A. 2009. Impact of calcium sulphate and calcium carbide on nitrogen use effi ciency of wheat in normal and saline sodic soils. Soil and Environment **28**, 29-37.

Mantri N, Patade V, Penna S, Ford R, Pang E. 2012. Abiotic Stress Responses in Plants: Present and Future. In: Abiotic stress responses in plants: metabolism, productivity and sustainability. Ahmad P, Prasad MNV (Ed) Springer, New York pp. 1-19.

**Mcdonald MB.** 2000. Seed priming. In: Black M, Bewley, JD (Ed). Seed technology and its biological basis. Sheffield: Sheffield Academic 2000. p. 287-325.

**Memon SA, Hou X, Wang LJ.** 2010. Morphological analysis of salt stress response of pak Choi. Electronic Journal of Environmental Agricultural and Food Chemistry **9**, 248-254.

**Netondo GW, Onyango JC, Beck E.** 2004. Sorghum and salinity. I. Response of growth, water relations, and ion accumulations, and ion accumulation to NaCl salinity. Crop Sciences **44**, 797-805.

DOI: 10.2135/cropsci2004.7970.

**Okcu G, Kaya MD, Atak M.** 2005. Effects of salt and drought stresses on germination and seedling growth of pea (*Pisum sativum* L.). Turkish Journal of Agricultre and Forestry **29**, 237-242.

**OParida AK, Das AB.** 2005. Salt tolerance and salinity effect on plants: A review. Ecotox Environ Saf **60**, 324-349. DOI: 10.1016/j.ecoenv.2004.06.010.

,5

**Perez-Alfocea F, Estan MT, Santa Cruz A, Maria, Bolarin C.** 2015. Effects of salinity on nitrate, total nitrogen, soluble protein and free amino add levels in tomato plants, Journal of Horticultural Science **686**, 1021-1027.

DOI: 10.1080/00221589. 1993.11516443.

**Prasad B.** 2003. Effect of direct and residual effects of different S fertilizers on groundnut and wheat cropping system on typic haplaquent soils. Plant Nutrition **26**, 997-1008.

DOI: 10.1081/PLN-120020071.

**Prosser IM, Purves JV, Saker LR, Clarkson DT.** 2001. Rapid disruption of nitrogen metabolism and nitrate transport in spinach plants deprived of sulphate. Journal of Experimental Botany **52**, 113-121. DOI: 10.1093/jxb/52.354.113.

**Randhawa PS, Arora CL.** 2000. Phosphorussulfur interaction effects on dry matter yield and nutrient uptake by wheat. Journal of Indian Society of Soil Science **48**, 536-544.

**Reich M, Shahbaz M, Prajapati DH, Parmar S, Hawkesford MJ, De Kok LJ.** 2016. Interaction of sulphate with other nutrients as revealed by H2S fumigation of Chinese cabbage. Frontiers in Plant Science 7, 541.

DOI: 10.3389/fpls.2016.00541.

**Reich M. Aghajanzadeh T, Helm J, Parmar S, Malcolm J, Hawkesford, Luit J, De Kok**. 2017. Chloride and sulphate salinity differently affect biomass, mineral nutrient composition and expression of sulphate transport and assimilation genes in *Brassica rapa*. Plant Soil **411**, 319-332. DOI: 10.1007/s11104-016-3026-7. Rengasamy P. 2006. World salinization with emphasis on Australia. Journal of Experimental Botany 57,1017-1023. DOI: 10.1093/jxb/erj108.

**Rui S Wei, Mu-xiang C, Cheng-jun J, Min W, Boping Y.** 2009. Leaf anatomical changes of *Burguiera gymnorrhiza* seedlings under salt stress. Journal of Tropical and Subtropical Botany **17**, 16-175.

Saboora A, Kiarostami K, Behroozbayati F. Hajihashemi S. 2006. Salinity (NaCl) tolerance of wheat genotypes at germination and early seedling growth. Pakistan Journal of Biological Sciences 9, 2009-2021.

DOI: 10.3923/pjbs.2006.2009.2021.

Sandhu KS, Singh N, Malhi NS. 2007. Some properties of corn grains and their flours I: Physicochemical, functional and chapati-making properties of flours. Food Chemistry **101**, 938-946. DOI: 10.1016/j.foodchem.2006.02.040.

Shirazi M, Khanzada B, Ali M, Islam E, Mujtaba S, Ansari R, Alam S, Khan M, Ali M. 2002. Response of three wheat genotypes grown under saline medium to low/high potassium levels. Acta Physiologia Plantarum **24**, 157-161.

Soeda Y, Kkonings MCJM, Vorst O, Van, Amml, Houwelingen GM, Stiipen CA, Maliepaard J, Kodde RJ, Bino SPC. 2005. Gene expression programs during *Brassica oleracea* seed maturation, osmopriming, and germination are indicators of progression of the germination process and the stress tolerance level. Plant Physiology **13**7, 354-368.

DOI: 10.1104/pp.104.051664.

**Tandon HLS.** 1993. Methods of Analysis of Soil, Plants, Water and Fertilizers. Fertilization Development and Consultation Organisation, New Delhi, India. **Turhan E, Atilla E.** 2004 Effects of sodium chloride applications and different growth media on ionic composition in strawberry plant. Journal of Plant Nutrition **27**, 1663-1666.

DOI: 10.1081/PLN-200026009.

**Werner JE, Finkelstein RR.** 1995. Arabidopsis mutants with reduced response to Nal and osmotic stress. Physiologia Plantarum **93**, 659-666.

**Wolf B.** 1982. A comprehensive system of leaf analysis and its use for diagnosing crop nutrient status. Communications in Soil Sciences and Plant Analysis **13**, 1035-1059.

DOI: 10.1080/00103628 209367332.

**Yoshida S, Foorno DA, Cock JH, Gomez KA.** 1976. Laboratory Manual for Physiological Studies of Rice, 3rd Edn Los Baños: International Rice Research Institute.

**Zekri M.** 1991. Effects of NaCl on growth and physiology of sour orange and Cleopatra mandarin seedlings. Scietia Horticulturae **47**, 305-315. DOI: 10.1016/0304-4238(91)90013-O.

Zhang JL, Flowers TJ, Wang SM. 2010. Mechanisms of sodium uptake by roots of higher plants. Plant Soil **326**, 45-60. DOI: 10.1007/s11104-009-0076-0.

Zhang M, Fang Y, Ji Y, Jiang Z, Wang L. 2013. Effects of salt stress on ion content, antioxidant enzymes and protein profile in different tissues of *Broussonetia papyrifera*. South African Journal of Botany, **85**, 1-9.

DOI: 10.1016/j.sajb.2012.11.005.

**Zhao FJ, Evans EJ, Bilsborrow PE, Syers JK.** 1993. Influence of S and N on seed yield and quality of low glucosinolate oilseed rape (*Brassica napus* L.). Journal of the Science of Food and Agricultre **63**, 29-37. DOI: 10.1002/jsfa.2740630106.