



RESEARCH PAPER

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Meal worm (*Tenebrio molitor*) as potential alternative source of protein supplementation in broiler

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Abstract

Protein is major nutrient costing higher value compared to other counterparts. A number of attempts have been made since long to find substitutes that could act as an effective and cheaper source of protein in poultry diet. Currently there has a great interest to assess the potential of different types insect meal in poultry nutrition as protein source. A preliminary study was undertaken to examine the potential benefits of locally produced meal worm supplementation in broiler birds on feed intake, body weight gain, feed conversion ratio (FCR), dressing percentage, antibody titer against ND and mortality. At the end birds from different experimental groups were slaughtered, dressed and cooked for meat sensory qualities. Meal worm larvae were grown at laboratory level, killed with 5% concentrated salt solution and oven dried 40°C for 48-72 hrs. Meal worm is rich in protein 45.83%, lysine 4.51% and methionine 1.34%. A total of 120 day-old broiler chicks were randomly allotted to four replicated (n=3; 10 birds/replicate) dietary treatments i.e. WM-1, WM-2, WM-3 that received 50, 100, 150g meal worm, respectively with WM-0 served as control. Supplementation of meal worm did not affect feed intake, however significantly improved body weight gain of birds in all supplemented groups by 84 to 185 g in absolute. FCR of WM-0 was poor (2.01) and significantly improved in WM-3 group (1.75). Carcass yield was improved in all meal worm supplemented groups compared to control. It was interesting to note that sensory meant qualities including heamagglutination antibody titer against Newcastle disease and mortality among different groups was not significantly altered. It could be deduced from present findings that meal worm has the potential to improve bird performance with affecting meat quality. However, further research work is warranted to replace soybean with worm meal to examine it impact as alternative protein in minimizing feed cost without compromising bird performance.

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Introduction

Feed is the major segment (60-70%) in the cost of poultry production (Abidin *et al.*, 2011). There is still a great increase has been seen in the price of poultry feed ingredients specially soybean meal in the last decade. This has led nutritionist to search alternative sources of protein that are cheaper and comparable in term of nutritive value to other commonly used protein meals used in poultry diet (Okah and Onwujiariri, 2012). A number of efforts have been made previously to minimize poultry cost using locally available alternative protein sources (Bhuiyan, 1998; Awoniyi *et al.*, 2004; Ojewola *et al.*, 2005). Recently there has been a great interest to grow insect and process them as an alternative feed ingredient due to its high nutritive profile and essential amino acids also due to high price of fish meal its deteriorated quality (Akpodiete and Inoni, 2000; Etuk *et al.*, 2003; Karimi, 2006).. Research suggests that insect based feeds are comparable with fishmeal and soybean meal (Huis *et al.*, 2013). There is however inconsistent and equivocal information available on the nutritive value of mealworm and its implications as poultry feed ingredient.

Mealworm (*Tenebrio molitor*) are common feedstuffs for pet animals including small mammals and reptiles and are a formidable source of insect protein with about 44-70% in their body tissues (Romas-Elorduy, 1987). Agriculturally, edible insects such as mealworm have great potential because they contain superior grade of proteins, lipids, carbohydrates and vitamins (De Foliart, *et al.*, 2009) that may provide a possible alternative source of nutrition for poultry broilers. Studies that explored the potential of *Tenebrio molitor* for massive human and animal consumption have also shown that ground mealworm emit no ammonia excretion (Oonincx, *et al* 2010). Insect rearing could be one of the ways to enhance food and feed security (van Huis *et al.*, 2013). They grow and reproduce easily, have high feed conversion efficiency and can be reared on bio-waste streams. One kg of insect biomass can be produced from on average 2kg of feed biomass (Collavo *et al.*, 2005).

Insects can feed on waste biomass and can transform this into high value food and feed resource. A desk study (Veldkamp *et al.*, 2012) has demonstrated that it is technically feasible to produce insects on a large scale and to use them as alternative sustainable protein rich ingredient in pig and poultry diets, particularly if they are reared on substrates of bio-waste and organic side streams.

To understand the importance of mealworm in poultry feed present study was planned to investigate the supplemental impacts of meal worm produce in small scale in labs. Dired mealworms were added to poultry diet at different level to observe the growth performance carcass yield, antibody titer again newcastel disease and meat quality attributes.

Material and methods

This study was conducted in collaboration at the Poultry Research Unit of the University of Agriculture and Veterinary Research Institute, Peshawar, Pakistan.

Bird's husbandry

Before the commencement of study all equipments, gears and premises were cleaned and disinfected. One hundred and twenty day-old chicks were procured from commercial hatchery. These chicks were randomly distributed in replicated (n=3; 10 birds/replicate) groups (n=4) in cleaned floor cages (10x10 sq. ft). Experimental groups were designated as Control (MW-0), MW-1, MW-2 and MW-3 received 1, 2 and 3g dried meal worm per kg of diet. During the course of experiment optimum managemental conditions were provided and birds had an ad libitum access to feed and water. This trial was lasted for 6 weeks. Experimental ration was formulated and mixed in feed mill unit in mash form. Meal worm larvae were produced in the research lab using darkling beetles, fed on wheat bran as growth media. Fresh larvae were harvested, killed and dried at 60°C. Ground mealworms were mixed in the feed for the experiment. Prior to mixing dried meal were found to contain permissible level of mycotoxin (5ppb) and no bacterial growth. Experiment was lasted for four weeks.

Lab analyses

Proximate analyses of dried meal worm were carried out using standard procedures of the Association of Official Analytical Chemists (AOAC 2000). Amino acid and minerals contents of mealworm sample were performed by the methods of the Association of Official Analytical Chemists (AOAC, 1990). The PICO TAG method with modification was employed for determining the amino acid from standard curves based on peak area measurement by PICO TAG amino acid analyzer (Deng *et al.*, 2004).

Production traits

Feed intake, body weight gain, feed conversion ratio (FCR), dressing percentage, HI antibody titer against ND and mortality were recorded and feed cost and gross return was determined. Data were statistically analyzed using standard procedure of ANOVA in a completely randomized design using Statistical package SAS (1998). Least significant difference test (LSD) was used for separation of means (Jan *et al.*, 2009).

Findings of the research study

Proximate analysis and amino acid profile

The proximate composition of worm meal including dry matter, crude protein, ash, fats, crude fibre level, the amino acid and mineral profile are given (Table 1). A considerable amount of crude protein (45.83%) and crude fat (34.20%) were recorded. Almost all essential amino acids Methionine (1.34) Lysine (4.51) were observed as in worm meal analyzed in present study (Table 1). No significant ($P \leq 0.05$) difference was seen in the feed intake of different groups Table 2. Weight gain was significantly improved in treated groups. Group WM-3 (1423.3±3.92) had significantly higher weight gain followed by group WM-2 (1346.3±5.48) and group WM-1 (1322.0±2.08). Mean FCR for supplemented groups was significantly better ($P \leq 0.05$) as compared to control group. Overall FCR was poor in group WM-0, control (2.011±0.01) followed by group WM-1 (1.88±0.05) and least value was found in group WM-3 (1.75±0.01).

Table 1. Nutrient profile of dried mealworm used in the experiment.

Ingredients %	Percentage/(mg/g of mineral/kg)	Amino acid profile	(grams/100g of meal worm)
Moisture	5.78	Alanine	4.34 ± 0.3
Ash	2.51	Arginine	2.21 ± 0.2
Crude Protein	45.83	Aspartic acid	3.23 ± 0.7
Crude Fat	34.2	Cystine	3.62 ± 0.2
Crude Fiber	3.97	Glutamic acid	4.75 ± 0.6
		Glycine	2.65 ± 0.2
Calcium	3.82	Histidine	1.65 ± 0.2
Phosphorous	0.07	Isoleucine	4.51 ± 0.3
Iron	0.63	Leucine	5.32 ± 0.2
Zinc	1.02	Lysine	4.51 ± 0.3
Copper	0.123	Methionine	1.34 ± 0.4
Potassium	0.085	Proline	2.34 ± 0.2
		Phenylalanine	1.54 ± 0.4
		Serine	3.45 ± 0.2
		Threonine	1.64 ± 0.3
		Tyrosine	2.32 ± 0.3
		Valine	4.42 ± 0.2

Effect of meal worm supplementation on production performance

Table 2. Effect of meal worm supplementation on feed take, weight gain and FCR.

Group	Feed intake	Body Weight Gain	Feed conversion ratio (FCR)
	Mean ± SE	Mean ± SE	Mean ± SE
WM-1	2491.7±3.52	1322.0 ^c ±2.08	1.88 ^b ±0.05
WM-2	2485.7±1.45	1346.3 ^b ±5.48	1.84 ^c ±0.07
WM-3	2485.0±4.35	1423.3 ^a ±3.92	1.75 ^d ±0.01
WM-0	2498.3±4.05	1238.3 ^d ±1.66	2.01 ^a ±0.01
P-value	0.0897	0.0000	0.0000

Means followed by different superscripts are significantly different $\alpha=0.05$.

Dressing percentage was calculated on last day of the experiment. The statistical analysis showed that dressing percentage was significantly ($P < 0.05$) higher for higher supplemented group WM-3 (66.10 ± 0.25) as compared to WM-0 (63.28 ± 0.13).

At the end of experiment antibody titer against Newcastle disease virus for all experimental groups showed non-significant result. This indicates that meal worm supplementation had no adverse effects on hemagglutination antibody titer.

Numerically it was higher in group WM-3 (6.56 ± 0.88) followed by group WM-1 (4.66 ± 0.33) and least was found in group WM-2 (4.45 ± 0.33). The mortality in control group was higher as compared to supplemented groups (Table 3).

Table 4. Organoleptic Study of broiler meat fed with meal worm.

Group	Taste	Tenderness	Juicines	Color	Flavor
	Mean \pm SE				
WM-1	2.35 ± 0.08	2.41 ± 0.03	2.35 ± 0.36	2.45 ± 0.07	2.54 ± 0.17
WM-2	2.33 ± 0.36	2.37 ± 0.15	2.33 ± 0.12	2.44 ± 0.26	2.54 ± 0.36
WM-3	2.33 ± 0.37	2.37 ± 0.48	2.32 ± 0.06	2.47 ± 0.13	2.53 ± 0.15
WM-0	2.44 ± 0.48	2.42 ± 0.38	2.37 ± 0.38	2.49 ± 0.51	2.57 ± 0.18

Economics of meal worm supplementation (PKR)

Economics of meal worm (MW) supplementation was calculated in terms of feed cost, meal worm cost, operational cost, total expenses, gross return and net profit. The economics is shown table 4.9. Gross return and net profit was significantly higher ($P < 0.05$) for

Table 3. Dressing percentage antibody titer and mortality of broiler fed with meal worm.

Group	Dressing Percentage	Antibodies titer against ND	Mortality
	Mean \pm SE	Mean \pm SE	Mean \pm SE
WM-1	$63.277^c \pm 0.31$	4.66 ± 0.33	3.33 ± 0.33
WM-2	$64.167^b \pm 0.21$	4.45 ± 0.33	3.33 ± 0.33
WM-3	$66.108^a \pm 0.25$	6.56 ± 0.88	0.00 ± 0.00
WM-0	$63.282^c \pm 0.13$	5.33 ± 0.88	6.66 ± 0.66

Means followed by different superscripts are significantly different $\alpha = 0.05$

Organoleptic study of broiler meat fed with meal worm

It was observed that organoleptic properties evaluation i.e. taste, tenderness, juiciness, flavor and color were not influenced by the addition of mealworm in the diet of broilers. They all were not significantly ($P < 0.05$) different among the experimental groups (Table 4).

group WM-3 (43.55 ± 0.40) followed by group WM-1 (41.95 ± 0.49), WM-2 (38.66 ± 0.83) and lowest return (35.71 ± 0.33) was WM-0 control group. Highest gross return and net profit in group B may be attributed to the quantity and quality (amino acid composition) of protein in the meal worm (Table 5).

Table 5. Economics of meal worm supplementation (PKR).

Group	Mean (PKR) \pm SE					
	Feed Cost	MW Cost	Op: Cost	Total Exp	G. Return	Net Profit Net profit
WM-1	$115.83^c \pm 0.17$	$3.73^c \pm 0.77$	50.00 ± 0.0	$169.56^c \pm 0.182$	$211.52^c \pm 0.33$	$41.95^a \pm 0.49$
WM-2	$119.30^b \pm 0.07$	$7.450^b \pm 0.77$	50.00 ± 0.00	$176.75^b \pm 0.078$	$215.41^b \pm 0.87$	$38.66^b \pm 0.83$
WM-3	$123.00^a \pm 0.21$	$11.17^a \pm 0.017$	50.00 ± 0.00	$184.18^a \pm 0.22$	$227.73^a \pm 0.62$	$43.55^a \pm 0.40$
WM-0	$112.42^d \pm 0.18$	$0.00^d \pm 0.00$	50.00 ± 0.00	$162.42^d \pm 0.182$	$198.13^d \pm 0.26$	$35.71^c \pm 0.33$

Means followed by different superscripts are significantly different $\alpha = 0.05$.

Discussion

Proximate analysis, amino acid profile and mineral composition of meal worm

Mealworms (*Tenebrio molitor* L.) are common feedstuffs for animals including a formidable source of insect protein with about 44-70% in their body tissues (Ramos-Elorduy, 1987).

Agriculturally, edible insects such as the yellow mealworm have great potential because they contain superior grade of proteins, lipids, carbohydrates and vitamins (De Folliart *et al.*, 2009) that may provide a possible alternative source of nutrition for poultry broilers.

The potential for insects as livestock feeds may also have an environmental impact as they cost less energy, use less land area and leave less environmental footprints during production (Pimental *et al.*, 1975).

Performance of birds

Meal worm supplementation in basal diet of poultry showed that the overall body weight gain and FCR for supplemented groups was significantly higher ($P \leq 0.05$) as compared control group regardless of feed intake. Nutritional attributes of the insects and insect meal and their use as a component in the diets of ruminants, poultry (both broiler and laying hen) and fish species, potential constraints, if any in using them as alternative feed resources and future research areas.

Furthermore, earlier the mealworm (*Tenbrio molitor*) as a diet for African catfish, *Clarias gariepinus*, was tested for alternative protein source. As a result, catfish showed a good performance of growth and utilization efficiency when it was fed on 80% of mealworm-based diet. In addition our study is supported by (Ng *et al.*, 2001) who suggested that mealworm is highly nutritive diet and acceptable as an alternative protein source.

In very limited experiments, Gould and Moses (1951) force-fed darkling beetle adults and larvae (whole and acerated) one time to chicks and found no adverse effects. Skewes and Monroe (1991) attempted to compare performance of broiler chickens in commercial grow out houses with different population densities of darkling beetle adults and larvae; have no adverse relationships between eating of darkling beetle on chick mortality, feed conversion, condemnation rate, and production cost were detected. Mealworm have great potential because they contain superior grade of proteins, lipids, carbohydrates and vitamins that may provide a possible alternative source of nutrition for poultry broilers.

Dressing percentage antibody titer and mortality

The findings of Khatun *et al.* (2003) found highest dressing percentage for birds fed with silkworm pupae meal instead of fishmeal.

Wijayasinghe and Rajaguru, (1977) also supported our study as they found highest dressing percentage for experimental group fed with 10% silkworm pupae meal (protein source) as compared to those fed with fishmeal.

Again limited research work on this part of the study but many researchers (Tegua *et al.*, 2002; Okah and Onwujiariri, 2012) found better dressing percentages with dietary supplementation of maggot meal or silkworm (insect protein) may be used to obtain better dressing percentage from broiler birds while feeding along conventional protein supplements. Literature is not available on this aspect of the study but although researchers found that protein sources does not adversely affects on antibody titer against ND, however it boost up the immune system of birds.

Taheri *et al.* (2005) used different level of oil extracted propolis as a source of protein in the diet of broiler birds and found significantly higher ($P < 0.05$) antibody titer against ND. Our study supported by Skewes and Monroe (1991) they attempted to compare performance of broiler chickens in commercial grow out houses with different population densities of darkling beetle adults and larvae; have no adverse relationships between eating of darkling beetle on chick mortality. Khatun *et al.* (2003) also supported the findings of this research.

They also did not come across any mortality in all the experimental groups while feeding them different percentages of fish and silkworm pupae meal. Dutta *et al.* (2012) also favors our results. They did not find any mortality case in their research work as our findings revealed. 5% mortality is normal. These findings indicated that worm meal has no toxic affect on birds.

Organoleptic properties

No literature is available regarding organoleptic parameters while feeding the birds on meal worm in ration. Similar study was conducted by Eyo (2006) and Aniebo *et al.* (2011) in fish and reported that maggot meal supplementation in the diet of fish did not affect flavor, juiciness or texture of the fish products.

Similar findings were also reported by Okubanjo *et al.* (2014) who stated that organoleptic study of broiler birds fed with maggot meal did not change taste, tenderness, juiciness, color and flavor of meat.

Economics of meal worm supplementation

Economics was calculated in terms of feed cost, meal worm cost, operational cost, total expenses, gross return and net profit. Statistical analysis showed that gross return and net profit were significantly higher ($P < 0.05$) for group C (43.55 ± 0.40) followed by group A (41.95 ± 0.49), B (38.66 ± 0.83) and lowest return (35.71 ± 0.33) was D control group. The findings of Dutta *et al.*, (2012) are in line to our study who observed that the cost of total feed offered was lowest in where silkworm pupae meal replaced 100% protein of fish meal. This shows that increase of incorporation of silkworm pupae meal in the poultry feed there is corresponding decrease in cost per unit of feed. These findings also agree with the results of some other researchers (Chaudhary *et al.*, 1998; Rahman *et al.*, 1996; Habib and Hasan, 1995; Khatun *et al.*, 2003 and Khatun *et al.*, 2005).

Conclusion

It was concluded from present findings that mealworm could be effectively produced locally and nutrient profile is highly comparable to that of soybean and fish meal. Mealworm supplementation in poultry diet could lead to improved growth performance and efficient utilization of poultry feed. There was no negative impact of mealworm supplementation on chicken meal quality and antibody titer. Further research studies are however warranted to explore its benefits by replacing soybean meal and examining its beneficial implication including reduction of feed cost.

References

Abidin Z, Khatoun A, Numan M. 2011. Mycotoxins in broilers: pathological alterations. Digestibility of Super Worm Meal in Red Tilapia Juvenile. Pak Vet J **32(4)**, 489-493.

Akpodiete OJ, Inoni OE. 2000. Economics of production of broiler chickens fed maggot meal as replacement for fish. Nig. J. Anim. Prod **27**: 59-63 and *Escherichia coli* within the gastrointestinal tract of the lesser mealworm beetle, *Alphitobius diaperinus* (Coleoptera: Tenebrionidae). Poultry Science **88**, 1553-1558
DOI: 10.3382/ps.2008-00553.

Aniebo AO, Erondue ES, Owen OJ. 2009. Replacement of fish meal with maggot meal in African catfish (*Clarias gariepinus*) diets. Revista UDO Agrícola **9**: 666-671.

Awoniyi TAM, Adebayo IA, Aletor VA. 2004. A study of some erythrocyte indices and bacteriological analysis of broiler chickens raised on maggot-meal based diets. Int. J. Poult. Sci **2**, 386-390.

Bhuiyan AKMA, Begum NN, Begum M, Hoq ME. 1989. Survey of potential fish feed ingredients of Bangladesh on the basis of their availability and biochemical composition. Final Report. FRI Research Progress Report 1. Freshwater station, Fri, 70.

Bhuiyan MZ. 1998. Complete replacement of fish meal by full fat soybean and supplementation of lysine and methionine to broilers. M.S. Thesis, Dept. of Poultry Science, Bangladesh Agricultural University Mymensingh.

Bidlingmeyer BA, Cohen SA, Tarvin TL, Frost B. 1987. A new, rapid, high sensitivity analysis of amino acid in food type samples. Journal of Association Official Analytical Chemistry **70**, pp. 241-247.

Bolu SA, Adakeja A. 2008. Effects of poultry offal meal and soyabean meal mixtures on the performance and carcass quality of broiler chicks. Afri. J. of food Agri. Nutri. and Develop **8(4)**, 441-450.

Campbell JM, Loyons DB, Sarazin MJ. 1989. Canadian Beetles (Coleoptera):

Chaudhary RC. 1998. Indonesian Coordinated Rice Testing Network (ICRTN). World Bank-ARMPIL, CRIFC, Bogor, Indonesia 152 pp.

Choudhury K, Das J, Saikia S, Sengupta S, Choudhury SK. 1998. Supplementation of broiler diets with antibiotic and probiotic fed muga silkworm pupae meal. *Indian J. Poultry Sci* **33**, 339-342.

Collavo A, Glew RH, Huang YS, Chuang LT, Bosse R, Paoletti MG. 2005, House cricket small-scale farming. In M.G. Paoletti, ed., *Ecological implications of minilivestock: potential of insects, rodents, frogs and snails* pp. 519-544. New Hampshire, Science Publishers.

Daghir NJ. 1975. Studies on poultry by-product meal in broiler and layer rations. *World Poult. Sci* **31**, 2001-2016.

De Foliart G, Dunkel FV, Gracer D. 2009. The food insects newsletter-chronicle of changing culture. *Salt LakeCity: Aardvark Global Publishing*, Vol (10) p. 414.

Deng SG, Peng ZY, Chen F, Yang P, Wu T. 2004. Amino acid composition and anti-anaemia action of hydrolyzed offal protein from Harengula Zunasi Bleeker *Food Chemistry* pp. 97-102.

Dutta A, Dutta S, Kumari S. 2012. Growth of poultry chicks fed on formulated feed containing silk worm pupae meal as protein supplement and commercial diet. *Online Journal of Animal and Feed Research* vol **2**, 303-307.

Etuk EB, Esonu BO, Udedibe AB. 2003. Evaluation of toasted pigeon pea (*Cajanuscajan*) seeds meal as replacement for soyabean meal and maize in broiler finisher diet.

Eyo AA. 2006. *Fish Processing Technology in the Tropics*. 1st Ed. University of Ilorin. Press. Ilorin Nigeria.

Gould GE, Moses HE. 1951. Lesser mealworm infestation in a broiler house. *Journal of Economic Entomology* **44**, 265.

Habib MAD, Hasan MR. 1995. Evaluation of silkworm pupae as dietary protein source for Asian catfish, *Clariasbatrachus* (L) fingerlings. *Bangladesh Journal of Aquaculture* **17**, 1-7.

Huis AV, Itterbeeck JV, Klunder H, Mertens E, Halloran A, Muirand G, Vantomme P. 2013. Edible insects: Future Prospects for Food and Feed Security. FAO Forestry paper 171. *Journal of Agricultural and Biological Sciences* **4(4)**, 319-331.

Jan MT, Shah P, Hollington PA, Khan MJ, Shohail Q. 2009. *Agriculture research: Design and Analysis*. 1st Ed. Dept of Agronomy, The Uni. Agri. Peshawar, Pakistan.

Karimi A. 2006. The effects of varying fish meal inclusion levels (%) on performance of broiler. *Injuries to Crops, Ornamentals, Stored Products and Buildings*. Research Branch, Agricultural Canada.

Khatun R, Azmal SA, Sarker MSK, Rashid MA, Hussain MA, Miah MY. 2005. Effect of silkworm pupae on the growth and egg production performance of rhode island red (RIR) pure line. *International Journal of Poultry Science* **4**, 718-720.

Khatun R, Howlader MAR, Rahman MM, Hasanuzzaman M. 2003. Replacement of fish meal by silkworm pupae in broiler diets. *Pakistan Journal of Biological Sciences* **6**, 955-958.

Mc Donald P, Edwards RA, Greenhalgh JFD. 2002. *Animal Nutrition*. 6th Edition. Longman, London and New York. 543 pp.

Metcalf CL, Flint WP. 1962. *Destructive and Useful Insects*. 4th Edn., McGraw Hill, Mills and their Control. Research Branch, Agriculture Canada, Ottawa, Ontario, ISBN: 13: 9780660117485.

Monro HAV. 1969. *Insect Pests in Cargo Ships*. Research Branch, Agriculture Canada, Nergui Ravzanaadii, Seong-Hyun Kim, Won Ho Choi, Seong-Jin Hong, and Nam Jung Kim, 2012. Nutritional Value of Mealworm, *Tenebriomolitor* as Food Source. *International Journal Industrial Entomoly* **1**, 93-98.

Ng W-K, Liew FL, Ang LP, Wong KW. 2001. Potential of mealworm (*Tenebrio* of Infectious Bursal Disease Virus from the Lesser Mealworm), *Alphitobiusdiaperinus* (Panzer). *Poultry Science* **74**, 45-49.

- Ojewola GS, Okoye FC, Ukoha OA.** 2005. Comparative utilization of three animal protein profile, by broiler chickens. *Int. J. Poult. Sci.*, 4970: silkworm caterpillar (*Anaphe infracta*) meal as 462-467.
- Okah and Onwunjiariri.** 2012. evaluated the performance of finisher boilers fed with maggot meals as replacement for fish meal and found out that replacement of a 4.0% dietary fish meal with 50% maggot meal gave superior performance characteristics.
- Okah U, Onwujiariri EB.** 2012. Performance of finisher broiler chickens fed meal worm as are placement for fish meal. *Journal of Agricultural Technology* **8(2)**, 471-477 Ottawa, Ontario pp. 39.
- Okubanjo AO, Apata ES, Babalola OO.** 2014. Carcass and organoleptic qualities of chicken broilers fed maggot meal in replacement for dietary fish meal. *American J. of Res. Communication* **2(4)**, 147.
- Ooninx DGAB, van Itterbeeck J, Heetkamp MJW, van den Brand H, van Loon JJA.** 2010. An exploration on green gas and ammonia production by insects species suitable for animal or human consumption 0014445.
- Pimental D, Dritschilo W, Krummel J, Kutzman J.** 1975. Energy and land constraints in food protein production science **190**, 754-761.
- Pimental D, Dritschilo W, Krummel J, Kutzman J.** 1975. Energy and land constraints in food protein production. *Science* **190**, 754-61.
- Rahman MA, Zaher M, Mazid MA, Haque MZ, Mahata SC.** 1996. Replacement of costly fish meal by silkworm pupae in diet of mirror carp (*Cyprinus carpio* L.). *Pakistan J. Sci. Ind. Res* **39(1/4)**, 64-67.
- Rahman, Jabir MD, Razak SA, Vikineswary S.** 2012. Chemical Composition and Nutrient Determined with Larvae of the Yellow Mealworm, *Tenebrio molitor* L. *J. Nutr* **104**, 1172-1177.
- Ramos-Elorduy J.** 1987. Are insects edible? Man's attitudes toward eating insecta. *Food Deficiency Studies and Perspectives* **20**, 78-83 (UNESCO Principle Regional Office for Asia and the Pacific).
- Ramos-Elorduy J.** 1987. Are insects edible? Man's attitudes towards eating insect. *Food deficiency studies and perspectives* **20**, 78-83.
- Skewes PA, Monroe JI.** 1991. The effects of darkling beetles on broiler performance. *Poultry Sei* **70**, 1034-1036.
- Tacon AGJ, Metian M.** 2008. Global overview on the use of fish meal and fish oil in industrially compounded aquafeeds: Trends and future prospects. *Aquaculture*. **285**, 146-158.
- Tegua A, Beynen AC.** 2005. Alternative feedstuffs for broilers in Cameroon. *Livestock research for rural development* **17(3)**.
- Veldkamp T, van Duinkerken G, van Huis A, Lakemond CMM, Ottevanger E, Bosch G, van Boekel MAJS.** 2012. Insects as a sustainable feed ingredient in pig and poultry diets - a feasibility study. Rapport 638 - Wageningen Livestock Research. <http://www.wageningenur.nl/uploadmm/2/8/o/f26765b9-98b2-49a7-ae43-5251c5b694f6-234247%5B1%5D>.
- Wijayasinghe MS, Rajaguru AS.** 1977. Use of silkworm (*Bombyx mori* L.) pupae as a protein supplement in poultry rations. *J. Natl. Sci. Coun. Sri Lanka* **5**, 95-104.