



## RESEARCH PAPER

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## Productivity of maize through salt tolerant *Rhizobia* strains under salt stress conditions

Shahab Ahmad Khosa<sup>\*1</sup>, Zahir A Zahir<sup>2</sup>, Khalid Saifullah Khan<sup>1</sup>

<sup>1</sup>Department of Soil Science, PMAS-Arid Agriculture University, Rawalpindi, Pakistan

<sup>2</sup>Institute of Soil and Environmental Sciences, University of Agriculture, Faisalabad, Pakistan

**Key words:** Inoculation, Maize, *Rhizobia* strains, Salinity

<http://dx.doi.org/10.12692/ijb/11.1.204-211>

Article published on July 27, 2017

### Abstract

A study was conducted to investigate the potential of *Rhizobia* to improve the growth and productivity of maize under saline conditions. Maize seeds were inoculated with four pre-isolated *Rhizobia* strains (CRI-34, CRI-29, S-43 and LSI-25) along with un-inoculated control. Two salinity levels (6 dSm<sup>-1</sup> and 12 dS<sup>-1</sup>) along with original EC maintained in the pots using NaCl salt. Maize seeds were sown in the pots. Results indicated that *Rhizobia* has the potential to induce salt tolerance in plants under saline stress conditions. Significant increase in growth and yield of maize was recorded. Among the *Rhizobia* strains LSI-25 performed better in almost all the yield and growth parameters. Under LSI-25 inoculated seeds, 35% increase in growth and yield was recorded followed by S-43 with 24%, CRI-29 with 14% and CRI-34 with 11%. *Rhizobia* strains increased the crop growth and yield by maintaining the nutrient balance in maize plants.

\* **Corresponding Author:** Shahab Ahmad Khosa ✉ [khosa.shahab@gmail.com](mailto:khosa.shahab@gmail.com)

## Introduction

Salinity is the major problem throughout the world limiting the agricultural productions especially in arid and semi-arid regions (Munns, 2002). Almost 800 million hectares of land is salt affected throughout the world. Soil salinity effects the plant growth through osmotic stress, production of ethylene, nutrient imbalance and disturbance with photosynthesis (Sairam and Tyagi, 2004). The ability of rhizobia is influenced by environmental factors which can be mainly categorized by physical factors, chemical factors and nutrient deficiencies (Giller, 2001). Soil salinity reduces plant growth, nutrient uptake especially phosphorus (P) uptake due to precipitations of phosphate ions with Ca ions in saline soils (Grattan and Grieve, 1999). Salinity causes toxic effects in plants like ionic effect and osmotic stress. Under saline soils, ionic effect including uptake in nitrogen (N) and intrusion in transportation of necessary ions within the plants and extreme lowering of photosynthesis activity in stressed plants while under osmotic stress, effects on reduction in cell wall extension and cellular expansion which leads to the stunted plant growth. The ability of plant to survive and retain their growth under saline conditions is identified as salt tolerance. Development of salt-tolerant crops is not an economic approach for sustainable agriculture whereas the yield and growth of maize can be improved by inoculation of seeds with efficient and competitive strains of rhizobia as it is the feasible way of production improvement (Ben Romdhane *et al.*, 2008). So, understanding the mechanism inducing salt tolerance in plants may be useful in developing new strategies to facilitate growth under saline conditions.

Rhizobia are considered as PGPR in non-legumes where they benefit plant growth. Plant growth promoting rhizobacteria stimulate the growth and colonize the plant roots (Arshad and Frankenberger, 2012). Plant growth promoting rhizobacteria able to produce various compounds like phytohormones, siderophores, organic acids and phosphorous solubilization and are also able to produce antibiotics that tackle the harmful bacteria.

Plant growth promoting rhizobacteria can affect the plant growth and development in a positive way by the synthesis of biologically active substances. The net effect of microbe-plant association on growth of plant could be positive, negative, or neutral. These bacteria are considered as efficient competitors with other microbes in root region (Kennedy, 2005; Nadeem *et al.*, 2006). They produce indole- acetic acid, cytokinins, gibberalins, and substances that inhibit the ethylene production. Plant growth can be facilitated by increasing the natural resistance of host against pathogens and by synthesizing the inhibitory substances (Cazorla *et al.*, 2007). Keeping in view the importance of maize and problems of salinity, present study was conducted to evaluate the potential of rhizobia for promoting growth and yield of maize under saline conditions in a pot trial.

## Materials and methods

A pot trial was conducted on maize in the wire house of the Institute of Soil and Environmental Sciences, University of Agriculture Faisalabad to evaluate the efficiency and potential of Rhizobium under salt stress conditions to improve the growth and yield of maize.

Details of experiment are given below:

### *Acquisition of Rhizobia isolates*

Pre-isolated Rhizobium strains were attained from the Soil Microbiology and Biochemistry Laboratory, Institute of Soil and Environmental Sciences, University of Agriculture, Faisalabad. These strains included the following:

CRI-34 (*Mesorhizobium ciceri*)

CRI-29 (*Mesorhizobium ciceri*)

LSI-25 (*Rhizobium leguminosorum*)

S-43 (*Rhizobium phaseoli*)

### *Preparation of inoculums*

Fresh inoculums of each strain were developed using sterilized yeast extract mannitol broth. After three days of incubation of inoculated broth (at 28°C and 100rpm) with respective isolate (CRI-34, CRI-29, LSI-25 and S-43), optical density was measured and dilutions were made if needed to have uniform population.

### *Seed inoculation*

Maize seeds were inoculated with respective bacterial strains. For this purpose, slurry was used which was prepared with peat, broth culture and sugar solution 10% in the ratio 5:4:1. In case of un-inoculated control, peat treated with sterilized broth was used for seed coating.

### *Pot trial*

Soil was obtained from the research area of Institute of Soil and Environmental Sciences, University of Agriculture Faisalabad. Salt (Sodium Chloride) was added to soil. The recommended level of chemical fertilizers (N: P: K @ 20:60:60kg ha<sup>-1</sup>) was applied at the time of sowing. The soil was analyzed for physicochemical characteristics before filling the pot at 10kg pot<sup>-1</sup>.

Three salinity levels of 1.41 dS m<sup>-1</sup> (Control original soil), 6 dS m<sup>-1</sup>, and 12 dS m<sup>-1</sup> were developed by using calculated amount of NaCl in each pot. Seeds of maize were sown at the rate of five inoculated seeds pot<sup>-1</sup>. Each treatment was replicated thrice. Pots were arranged in completely randomized design at ambient light and temperature. Good quality irrigation water which meets the irrigation quality criteria for crop was used to irrigate the pots. After germination, uniform plant population of two plants pot<sup>-1</sup> was maintained by thinning. Plant parameters were studied including plant height, root length, grain yield, 1000 grain weight, cob fresh weight, cob dry weight, N P K contents in straw and grain. Soil textural class was determined by using international soil classification system. After 90 days, at maturity, plants were harvested and data about growth and yield parameters were recorded.

### *Statistical Analysis*

Analysis of variance (ANOVA) technique was used from Steel *et al.*, (1997) and Duncan's Multiple Range (DMR) test was applied to see the significance of difference among the treatment mean (Duncan, 1955).

### **Results**

Inoculation of LSI 25 was prominent for improving root length of the crop with an increase of 35%

followed by S-43 with 24%, CRI-29 with 14% and CRI-29 with 11% improvement. Plant height of maize is extensively affected by salinity where 12 dS m<sup>-1</sup> maximally reduced the plant height followed by 6 dS m<sup>-1</sup>. Plant height was reduced up to 25% at 12 dS m<sup>-1</sup> and 6 dS m<sup>-1</sup> decreased it up to 15% compared to control treatment. Mainly inoculation of LSI-25 remained important for improving plant height of the crop with an increase of 28% followed by CRI-34 with 10% improvement, while other inoculated isolates (CRI-29 and S-43) improved the plant height but remained statistically at par with un-inoculated control. Strain LSI-25 significantly improved the root length up to 35%, plant height up to 28%. Strain S-43 significantly improved the cob length up to 25%. 12 dS m<sup>-1</sup> maximally reduced the 1000 grain weight compared to control treatment.

Mainly inoculation of CRI-34 remained prominent for improving 1000 grain weight of the crop with an increase of 27% followed by LSI-25 with 24%, S-43 with 26% and CRI-29 with 29% improvement. Mainly inoculation of LSI-25 remained prominent for improving cob yield dry of the crop with an increase of 45% as compared to control. Inoculation of CRI-34 remained prominent for improving cob yield fresh of the crop with an increase of 31% than control. All the inoculated isolates improved the grain yield but remained statistically at par with un-inoculated control. All the inoculated isolates significantly improved the % N in grain and % in straw as compared to control treatment. CRI-34 remained prominent for improving % P in grain of the crop with an increase of 25% and S-43 remained prominent for improving % P in straw of the crop with an increase of 30% as compared to control.

Under normal i.e. original salinity inoculation remained beneficial for improving the % K in grain and straw of the crop but most of the isolates remained statistically similar to un-inoculated control but LSI-25 was significantly different from inoculated as well as respective un-inoculated control. Overall, LSI-25 and CRI-34 remained most prominent strains for improving growth and productivity of maize under saline conditions.

**Table 1.** Physico-chemical properties of soil.

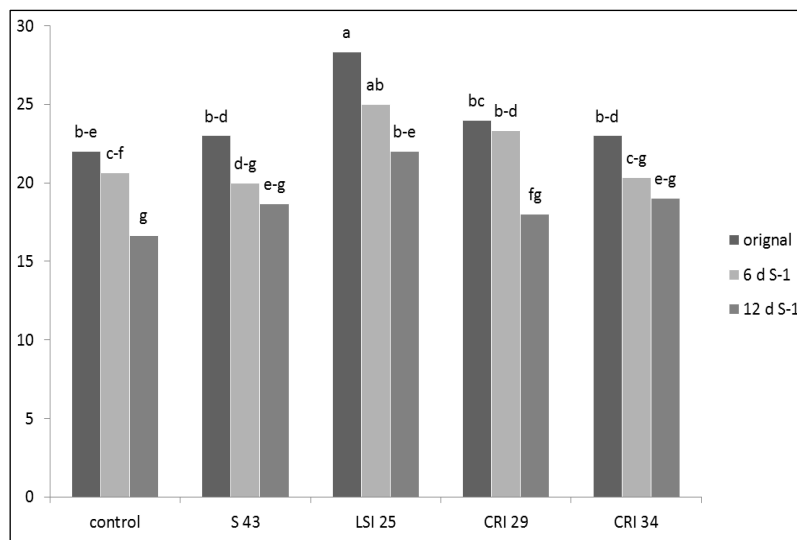
| Characteristics        | Value                   |
|------------------------|-------------------------|
| Particle size analysis |                         |
| Sand                   | 29.2%                   |
| Silt                   | 57.80%                  |
| Clay                   | 13%                     |
| Texture                |                         |
| Chemical analysis      |                         |
| Organic matter         | 0.46%                   |
| Saturation percentage  | 34.98%                  |
| pH                     | 9.93                    |
| EC                     | 4.48 dS m <sup>-1</sup> |
| Organic Matter         | 0.73%                   |
| Available phosphorous  | 6.3 ppm                 |
| Extractable potash     | 500 ppm                 |
| Na <sup>+</sup>        | 46.48meqL <sup>-1</sup> |
| Ca + Mg                | 2.73meqL <sup>-1</sup>  |
| K <sup>+</sup>         | 500 ppm                 |
| SAR                    | 37%                     |
| ESP                    | 18                      |

**Table 2.** Effect of rhizobium strains and salinity on maize yield parameters at different salinity levels.

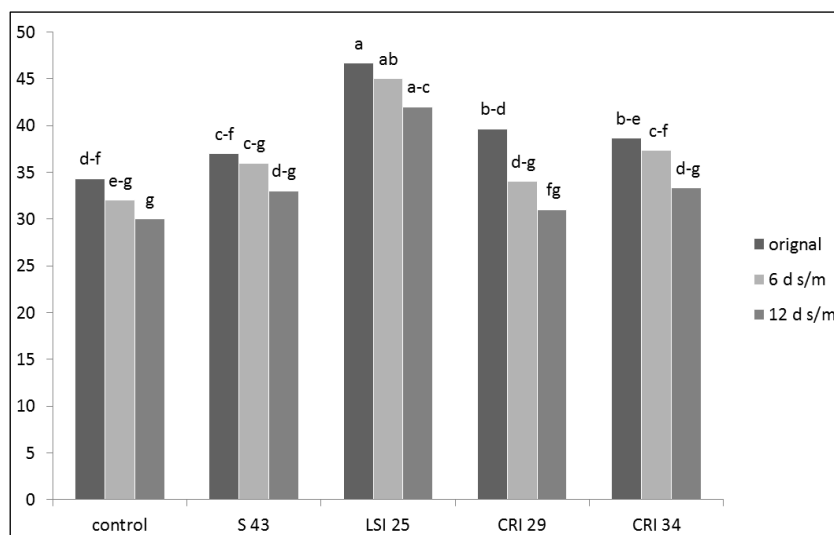
| Salinity level | Rhizobia species | 1000 grain wt (g) | Grain yield (g) | Cob fresh yield (g) | Cob dry yield (g) |
|----------------|------------------|-------------------|-----------------|---------------------|-------------------|
| Control        | Control          | 121hi             | 42cdef          | 36de                | 62def             |
| 6 dSm-1        |                  | 107j              | 35f             | 26f                 | 50g               |
| 12 dSm-1       |                  | 138ge             | 53ab            | 45abc               | 75abc             |
| Control        | S 43             | 128fg             | 49abcd          | 37de                | 72bcd             |
| 6 dSm-1        |                  | 125gh             | 41def           | 34c                 | 66bcd             |
| 12 dSm-1       |                  | 149b              | 58a             | 52a                 | 81ab              |
| Control        | LSI 25           | 145bc             | 55ab            | 46abc               | 73abc             |
| 6 dSm-1        |                  | 136de             | 52abc           | 37de                | 61ef              |
| 12 dSm-1       |                  | 166a              | 53ab            | 47ab                | 83a               |
| Control        | CRI 34           | 141cd             | 47bcde          | 43bcd               | 76abc             |
| 6 dSm-1        |                  | 132ef             | 41def           | 37de                | 63def             |
| 12 dSm-1       |                  | 141cd             | 54ab            | 38cde               | 71cde             |
| Control        |                  | 123ghi            | 47bcde          | 35e                 | 63def             |
| 6 dSm-1        |                  | 119i              | 41cdef          | 31ef                | 61def             |
| 12 dSm-1       |                  | 118i              | 39ef            | 3ef                 | 56fg              |

**Table 3.** Effect of rhizobium strains on N, P, K in grain and straw at different salinity levels.

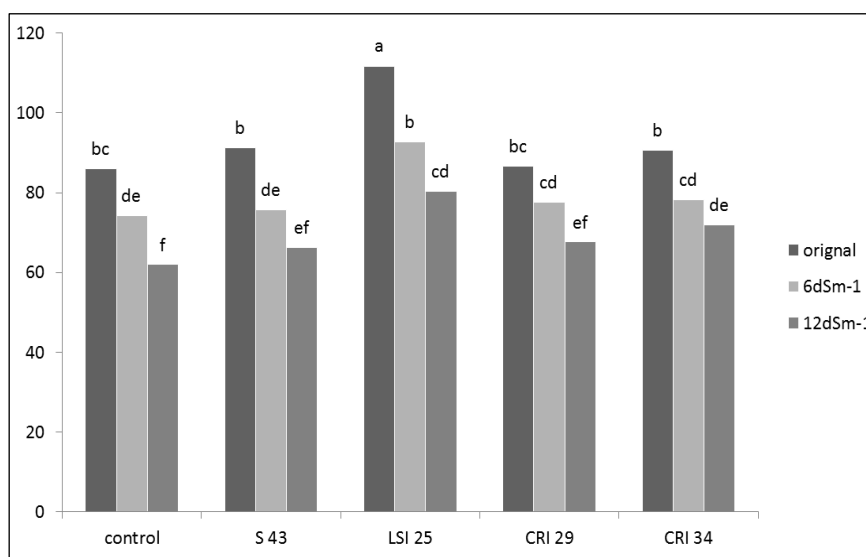
| Salinity Level | Rhizobium species | % N in grain | % N in straw | % P in grain | % P in grain | % K in grain | % K in grain |
|----------------|-------------------|--------------|--------------|--------------|--------------|--------------|--------------|
| Control        |                   | 1.54e        | 1.05b        | 0.46b        | 0.26cde      | 1.48cd       | 0.22bc       |
| 6 dSm-1        |                   | 1.38f        | 0.73d        | 0.26d        | 0.22ef       | 1.38efg      | 0.20bc       |
| 12 dSm-1       |                   | 0.88i        | 0.35e        | 0.13e        | 0.14g        | 1.51i        | 0.18c        |
| Control        | S 43              | 1.69ab       | 1.26a        | 0.55a        | 0.31ab       | 1.61ab       | 0.26ab       |
| 6 dSm-1        |                   | 1.59d        | 1.01b        | 0.33c        | 0.26cde      | 1.49cd       | 0.23abc      |
| 12 dSm-1       |                   | 1.23h        | 0.73c        | 0.18e        | 0.24def      | 1.28gh       | 0.18c        |
| Control        | LSI 25            | 1.72a        | 1.27a        | 0.48b        | 0.27bcd      | 1.69a        | 0.29a        |
| 6 dSm-1        |                   | 1.63c        | 1.08b        | 0.31cd       | 0.24def      | 1.45cde      | 0.21bc       |
| 12 dSm-1       |                   | 1.22h        | 0.90c        | 0.16e        | 0.21f        | 1.22hi       | 0.20bc       |
| Control        | CRI 34            | 1.69ab       | 1.25a        | 0.56a        | 0.29abc      | 1.60ab       | 0.25ab       |
| 6 dSm-1        |                   | 1.66bc       | 1.04b        | 0.33c        | 0.26cd       | 1.55bc       | 0.24abc      |
| 12 dSm-1       |                   | 1.33g        | 0.81cd       | 0.17e        | 0.23def      | 1.39def      | 0.23abc      |
| Control        | CRI 29            | 1.69ab       | 1.21a        | 0.48b        | 0.31a        | 1.52bc       | 0.25ab       |
| 6 dSm-1        |                   | 1.57de       | 0.82cd       | 0.31cd       | 0.25cde      | 1.41def      | 0.22abc      |
| 12 dSm-1       |                   | 1.25h        | 0.36e        |              | 0.21f        | 1.39fgh      | 0.21bc       |



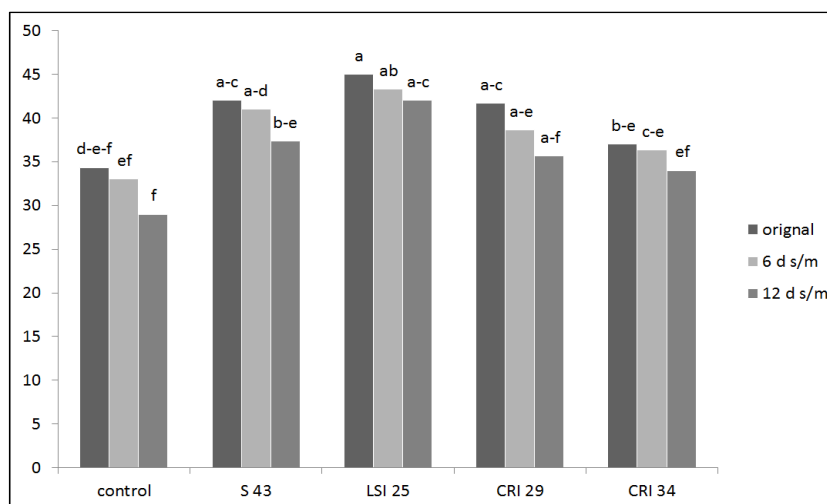
**Fig. 1.** Effect of *Rhizobia* strains on root dry weight of maize at different salinity levels.



**Fig. 2.** Effect of *Rhizobia* strains on root fresh weight of maize at different salinity levels.



**Fig. 3.** Effect of *Rhizobia* strains on plant height of maize at different salinity levels.



**Fig. 4.** Effect of *Rhizobia* strains on root length of maize at different salinity levels.

### Discussion

The result of this study showed that rhizobium strains promoted the growth of maize with different efficiency. Thousand grain weight, cob length, cob weight, total biomass, and straw yield increased significantly over un-inoculated control but the root length and cob yield showed non-significant results under inoculation. On the other hand, inoculation was more effective under high salinity. Some strains were more effective and some were less efficient at different salinity levels as shown in the results. Momentary adjustment in ionic balance may likely be a mechanism of action in salt conditions and *Rhizobium* changes the metabolism of cytoplasmic low molecular weight compounds in response of salinity (Yap and Lim, 1983). Several species of bacteria become accustomed under saline conditions by the intracellular accumulation of low molecular weight organic solutes called osmolytes (Csonka and Hanson, 1991). This accumulation of osmolytes neutralizes the dehydration effect of low water movement in medium but it does not interfere with the structure or function of macro molecules (Smith *et al.*, 1994). Plants under stressed conditions showed significant improvement in growth and yield compared to un-inoculated control. The results from our trial also uncovered that the effect of the rhizobial isolates on growth and yield of maize varied from pot to pot. This variability in the effect of rhizobial inoculants on crop with respect to pot could be explained on the basis of changing soil-plant and micro floral components in any experimental pot.

Hoflich (2000) conducted trials on sandy loam and loamy sand fields and reported considerable improvement in maize growth equal to 10% with R39 on loamy sand field whilst no considerable results regarding maize growth were obtained on the sandy loam field. Due to presence of microorganisms in rhizosphere of maize some macronutrients like N and P concentrations were positively increased. Likewise, Han and Lee (2005) reported that uptake of N and P were significantly improved in soyabean under salt stressed conditions when inoculated with *Rhizobium* when compared with un-inoculated control under salinity. Inoculation by the *Rhizobium* could be responsible for the observed higher N uptake of inoculated plants under both non-saline and saline conditions (Kantar *et al.*, 2003; Oğutcu *et al.*, 2008). From the work of different scientists it was evident that increase in Na<sup>+</sup> contents, caused an increase in Na<sup>+</sup> in general and resulted in high Na<sup>+</sup>/K<sup>+</sup> ratio. As Na<sup>+</sup> in the saline environment is considerably greater than K<sup>+</sup>, therefore it competes with the K<sup>+</sup> uptake effectively through common transport systems (Maathuis *et al.*, 1996; Rains and Epstein, 1967). In the study the improvement in growth under salinity is due to the maintaining the nutrition balance in plant. Decrease in Na<sup>+</sup> concentration may be due to the production of exopolysaccharides (EPSs) by rhizobia. Rhizobial strains are capable of producing exopolysaccharides which have the ability to bind cations including Na<sup>+</sup> (Geddie and Sutherland, 1993). These EPS-producing rhizobia under salt stress conditions have been found to restrict Na<sup>+</sup> uptake by roots (Ashraf *et al.*, 2004).

Protein concentration can be increased and the inhibitory effect of salinity on growth can be minimized by the inoculation with selective strains. Hamdia *et al.*, (2004) also reported significant increase in concentration of protein synthesis in maize crop under salt stressed conditions because of inoculation with *Azospirillum brasilense*.

### Conclusion

Rhizobia inoculation could increase nutrient availability, increase resistance against salt stress and improve crop growth under saline conditions. Therefore, rhizobium inoculation is a good tool in promotion of growth and yield of crops in saline conditions.

### References

- Arshad M, Frankenberger, Jr WT.** 2012. Ethylene: Agricultural Sources and Application. Kluwer Academic Publishers, New York.
- Ashraf MY, Khan AH, Azmi A.R.** 2004. Cell membrane stability and its relation with some physiological process in wheat. *Acta. Agron. Hung* **4**, 183-191.
- Ben Romdhane SM, Aouani M, Trabelsi De Lajudie P, Mhamdi R.** 2008. Selection of high nitrogen fixation rhizobia for semi-arid Tunisia. *Journal of Agronomy Crop Science* **194(6)**, 413-420.
- Cazorla FMD, Romero A, Perez-Garcia BJJ, Lugtenberg, de Vicente A, Bloemberg G.** 2007. Isolation and characterization of antagonistic *Bacillus subtilis* strains from the avocado rhizosphere displaying biocontrol activity. *J. Appl. Microbiol* **103**, 1950-1959.
- Csonka LN, Hasnon AD.** 1991. Prokaryotic osmoregulation: genetic and physiology. *Annu. Rev. Plant Physiol* **45**, 569-606.
- Duncan DB.** 1955. Multiple range and multiple F tests. *Biometrics* **11**, 1-42.
- Giller KE.** 2001. Nitrogen fixation in tropical cropping systems. Cabi Wallingford.
- Gratten S, Grieve CM.** 1999. Salinity-mineral nutrient relations in horticultural crops. *Sci. Hort* **78**, 127-157.
- Hamdia MA, Shaddad MAK, Doaa MM.** 2004. Mechanism of salt tolerance and interactive effect of *Azospirillum brasilense* inoculation on maize cultivars grown under salt stress conditions. *Plant Growth Regul* **44**, 165-174.
- Han HS, Lee KD.** 2005. Physiological responses of soya bean-inoculation of *Brady rhizobium japonicum* with PGPR in saline soil conditions. *Research Journal of Agricultural and Biological Sciences* **1**, 216-221.
- Hoflich G.** 2004. Colonization and growth promotion of non-legumes by Rhizobium bacteria. In: p. 827-830. *Microbial Biosystems: New Frontiers*. C.R. Bell, M. Brylinsky and P. Johnson-Green (eds.). Proc. of the 8th Int. Symp. on Microbial Ecology. Atlantic Canada Soc. Microbial Ecol., Halifax Canada.
- Kantar F, Elkoca E, Ogutcu H, Algur OF.** 2003. Chickpea yields in relation to *Rhizobium* inoculation from wild chickpea at high altitudes. *J. Agron. Crop Sci* **189**, 291-297.
- Kennedy AC.** 2005. Rhizosphere. P. 242-262. In: D.M. Sylvia, J.J. Fuhrmann, P.G. Hartel and D.A. Zuberer (Eds.), *Principles and Applications of Soil Microbiology*, 2nd Ed. Pearson Prentice Hall: Upper Saddle River NJ.
- Maathuis FJ, Verlin MD, Smith FA, Sander D, Fernandez JA, Walker NA.** 1996. The physiological relevance of Na-coupled K-transport. *Plant Physiol* **112**, 1609-1616.
- Mayak S, Tirosh T, Glick BR.** 2004. Plant growth-promoting bacteria confer resistance in tomato plants to salt stress. *Plant Physiol. Biochem* **42**, 562-575.
- Nadeem SM, Zahir ZA, Naveed M, Arshad M, Shahzad SM.** 2006. Variation in growth and ion uptake of maize due to inoculation with plant growth promoting rhizobacteria under salt stress. *Soil Environ* **25**, 78-84.

**Oğutcu H, Algur OF, Elkoca E, Kantar F.** 2008. The determination of symbiotic effectiveness of *Rhizobium* strains isolated from wild chickpea collected from high altitudes in Erzurum. Turk .J. Agric. For **32**, 241-248.

**Sairam RK, Tyagi A.** 2004. Physiology and molecular biology of salinity stress tolerance in plants. Curr. Sci **86**, 407-421.

**Steel RGD, Torrie JH, Dicky DA.** 1997. Principles and Procedures of statistics-A Biometrical Approach (3rd Ed.) Mc Graw-Hill Book International Co. Singapore.

**Winicov I.** 1998. New molecular approaches to improving salt tolerance in crop plants. Ann Bot. **82**, 703-710.

**Yap SF, Lim ST.** 1983. Response of *Rhizobium* sp. UMKL 20 to sodium chloride stress. Arch. Microbiol **135**, 224-228.