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RESEARCH PAPER

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Macropropagation using cuttings from yakal (*Shorea astylosa* Foxw) wildlings under various rooting hormones and level of concentrations

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Abstract

Shorea astylosa Foxw is a species of Dipterocarpaceae family. Productions of planting stock of dipterocarp species using seeds confront various problems. Flowering occurs at intervals of 3-10 yr. Propagation of wildlings cuttings was used in this study with levels of hormone, IBA, NAA, and Hormex. It was laid in $2 \times 2 \times 5$ factorial experiment in CRD. Two segments were collected, topmost and middle parts. Hormones were used in levels; 0 ppm, control, 500 ppm, 1000 ppm, 1500 ppm, Hormex, 10mL per gallon of water, replicated three times. The study conducted from May 22 to October 4, 2019, in clonal nursery of Ecosystem Research, Development Bureau, DENR, Maharlika, Bislig City, Surigao del Sur. The results revealed that using IBA in the topmost segment produces more leaves than the middle segment, 1000 ppm of IBA has higher average survival rate of 93%. The average survival of topmost and middle segments has a significant difference with 78% and 61%, respectively. Significant differences observed between topmost and the middle segments, the highest average survival of topmost for IBA was in treatment 3 with 93% survival. For NAA, the highest was in treatment 1 with 96% average survival. For middle, highest survival for IBA was in treatment 1 with 90% survival and, for NAA, it was in treatment 1 with 73% average survival. These results implied that even without applying rooting hormone, cutting segments of yakal wildlings produces higher survival rate than other treatment levels with hormone application.

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Introduction

Dipterocarpaceae is a family of 17 genera and approximately composed of 500 species worldwide. The largest genera are *Shorea* (196 species), *Hopea* (104 species), *Dipterocarpus* (70 species), and *Vatica* (65 species) (Ashton, 2005; Kettle, 2010). *Shorea astylosa* Foxw is a species of plant in the Dipterocarpaceae family. It is endemic in the Philippines, which is known as yakal in Filipino language. A major portion of our dipterocarp forest has been devastated and transformed into grassland and brush land.

It has reported that the forest destruction is mainly caused by land conversion for the settlement, agricultural development, shifting cultivation, logging, forest fire, and to some extent mining, energy projects, and pest and disease treatment (Fernando 2001). The tree is threatened by destruction of its habitat. The plant is classified as 'Critically Endangered' in the IUCN Red List of Threatened Species (2013).

Sakai *et al.* (2006) mentioned that the productions of planting stock of dipterocarp species using seeds confront various problems. Flowering occurs at intervals of 3–10 yr. and flowering evolved to satiate seed predators and/or to facilitate pollination. Flowering is thought to be triggered by droughts that occur during periods from La Niña to El Niño.

This results to the irregularity in the supply of seeds due to irregular flowering and fruiting, short viability period of the collected seeds, low quality of the seeds and the lack of seed storage and handling facilities (FDPM, 1998). Due to these problems, efforts in the reforestation of natural forests have been hindered. The scaling up of the domestication of native timber trees is constrained by the limited availability of planting materials and low-quality germplasm (Tolentino *et al.*, 2002; Gregorio *et al.*, 2005).

Vegetative propagation of *S. astylosa* and other dipterocarp species by stem cuttings is an important alternative for production of high quality and uniform planting stock for reforestation programs. Stem

cuttings offer several advantages over seeds. It is also inexpensive and easier to practice than other vegetative propagation methods, such as tissue culture (FDPM, 1998).

As an alternative production strategy, the objective of this study is to determine the potentials of propagating wildling stem cuttings of *S. astylosa* when applied with different levels of IBA, NAA and commercially available rooting hormone, HORMEX, on the recommended application on the rooting and survival of macro propagated *S. astylosa*.

The irregularity in the supply of seeds due to irregular flowering and fruiting of *S. astylosa* justifies the use of wildlings as alternative source of vegetative propagation. The collection of wildling cuttings must be viewed within the wider context of sourcing high quality germplasm for reforestation efforts and species conservation since it is classified as critically endangered. However, protocol for vegetative propagation of *S. astylosa* using cuttings from wildlings is not yet establish hence this study.

Materials and methods

Study Site

The study was conducted at the clonal nursery of Ecosystem Research and Development Bureau, Department of Environment and Natural Resources, barangay Maharlika , Bislig City, Surigao del Sur with geographical coordinates of 9°23' 3.92" N, 125°58' 41,72" E (Fig. 1).

Experimental Design

This experiment was laid-out in a $2 \times 2 \times 5$ factorial experiment in Complete Randomized Design (CRD). Two segments of the wildlings were collected as cutting materials, the topmost part and the middle part. IBA, NAA rooting hormones were used in varying levels; 0 ppm, as control, 500 ppm, 1000 ppm, and 1500 ppm, and the commercially recommended application of Hormex, 10mL per gallon of water, and replicated three times. A total of 600 cuttings were observed in the clonal nursery with 10 cuttings per experimental unit (Fig. 2).



Fig. 1. Map of the Study Area.

C2	C2	C1	C1	C1	C2	C1	C2	C1	C1	C2	C2	C2	C2	C2
H1	H1	H2	H2	H1	H1	H1	H1	H1	H1	H2	H1	H1	H2	H1
L3	L4	L3	L2	L5	L5	L1	L3	L3	L4	L5	L4	L1	L2	L2
C1	C2	C1	C1	C2	C2	C2	C2	C1	C2	C1	C1	C2	C2	C2
H2	H2	H2	H2	H2	H1	H1	H2	H1	H2	H1	H1	H1	H2	H2
L5	L1	L2	L4	L3	L5	L1	L3	L2	L4	L3	L1	L2	L5	L4
C1	C2	C2	C2	C2	C1	C1	C1	C1	C2	C1	C2	C2	C1	C2
H2	H2	H2	H2	H2	H1	H2	H1	H2	H1	H2	H2	H2	H1	H1
L3	L2	L5	L3	L4	L5	L5	L4	L1	L3	L1	L2	L1	L2	L5
C1	C2	C1	C1	C1	C1	C1	C1	C2	C1	C2	C1	C2	C1	C1
H2	H2	H2	H2	H1	H1	H2	H2	H1	H1	H1	H1	H1	H2	H1
L3	L1	L2	L1	L5	L3	L4	L5	L2	L2	L1	L4	L4	L4	L1

Fig. 2. Experimental lay-out of a 2 x 2 x 5 factorial experiment involving two cuttings (Top most segment and Middle segment).

Experimental Procedure

Preparation of Rooting Hormone

Rooting hormone preparation was conducted in the College of Forestry & Environmental Science, Central Mindanao University – Bukidnon. The preparation of rooting hormone in varying levels of concentration per treatment of IBA, NAA and hormex.

Cuttings Collection and treatment

The source of cuttings was coming from the wildlings of *S. astylosa* and collected in their natural habitat at the laboratory area of ERDB, DENR, Maharlika, Bislig City, Surigao del Sur. The wildlings were not uprooted in the field, only the portion serves as the cutting materials were cut and collected leaving the basal area and root system intact in the ground. The average length of cuttings was 8-10cm and the basal part of cuttings were cut in slant.

Only three to four leaves were retained from cuttings and cut to half of their original size to reduce transpiration. Cuttings were planted in the closed mist rooting chamber vertically with the depth of 2 -3cm and distance of 5cm × 5cm between cuttings, 10cm between treatments. The area for the study was .5 meters × 5.2 meters (Fig. 3).





Fig. 3. Schematic Diagram of cutting set-up in the clonal nursery.

Data collection

Data collection were done at the 160th day of the study. The average number of leaves and roots, length of roots and average survival rate were measured after the termination of the study.

Average number of leaves was determined using the formula: $ANL = \frac{TNL}{TNCT}$

Where, ANL – Average Number of Leaves TNL – Total Number of Leaves TNCT – Total Number of Cuttings/Treatment

Average number of roots was determined using the formula: $ANR = \frac{TNR}{TNCS}$

Where, ANR – Average Number of Roots TNR – Total Number of Roots TNCS – Total Number of Cuttings Survive

The average root length was determined using the formula: $ALR = \frac{TRL}{TNR}$

Where, ARL – Average Root Length TRL – Total Root Length TNR – Total Number of Roots

Survival rate was determine using the equation below: $SR(\%) = \frac{NSC}{TNCP}X$ 100

Where, SR– Survival Rate NSC – Number of Survived Cuttings TNCP – Total Number of Cuttings Planted

Data analysis

The data were analyzed using analysis of variance (ANOVA) and the significant effect of each factor were separated by Tukey's pairwise comparison post hoc (Least Significance Difference) test at a 5% level of significance (Moreira *et al.*, 2009).

Results and discussions

A. Comparison of cutting segments (topmost segment and Middle Segment).

IBA hormone

Average Number of Leaves

The highest average number of leaves of the topmost segment was in treatment 3 with a mean of (1.0), followed by treatment 4 (.92), treatment 2 (.79), treatment 5 (.77) and the lowest was in treatment 1 (.70). For the middle segment, the highest mean was observed in treatment 1 (.63) followed by treatment 3 (.45), treatment 4, (.34) treatment 2 (.32), and the lowest was in treatment 4 (.19) (Table 1). No significant result was observed per treatment on the topmost segment, the same with the middle segment in terms of the average number of leaves.

However, the mean comparison on the average number of leaves between the topmost segment and middle segment had a significant difference with a mean of .84 for the topmost segment and .39 for the middle segment (Table 2). These results shown that the topmost segment treated with IBA significantly produces more leaves than the middle segment. Likewise, these results corroborate to the study of Kassahun and Mekonnen (2011) on the effect of cutting position on the propagation ability of stevia through its effect on leaf number and survival rate varies from top and middle cutting position of a plant.

They further added that this difference in propagation ability of apical and middle cuttings of stevia could be due to high concentration of endogenous root promoting substances in the apical cuttings which arise from the terminal buds and also "more cells" which are capable of becoming meristematic (Hartman and Kester 1983).

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Table 1. IBA: Comparison on the means of the cutting segments on the Average Number of Leaves of cuttings from Yakal wildlings160 DAP.

	Ave. no of	Ave no of
		11,0,10,01
	leaves	leaves
}	.71	.63
3	.80	.32
3	1.0	.45
3	.91	.35
}	•77	.19
		1eaves .71 .80 1.0 .91 .77

CV: 40.65 Mean: .6140

Table 2. IBA: Mean comparison on the cutting segments on average number of leaves of Cutting from Yakal Wildling 160 DAP.

Cuttings	Average	N Group			
Top most segment	.84	15 a			
Middle segment	.39	15 b			

Means with the same letter are not significantly different.

Average Number of Roots

Analysis of variance shows no significant difference was observed per treatment on the topmost segment and middle segment on the Average number of roots. These results mean that both topmost and middle segments produce almost the same performance in terms of producing the number of roots. However, the present findings contrast with the study of Deen and Mohamoud (1996) as cited by Kassahun and Mekonnen (2011), they mentioned that apical cuttings exerted a greater number of roots for rosemary (Rosemarinus officinalis L.). Similar results were also reported by Palanisamy and Kumar (1997) in rooting of neem (Azadirachta indica A. Juss), where cutting from the upper part of the branches is rooted better than the lower ones (Wassner and Ravetta 2000). Leakey (1983) also added that with the propagation of Triplochiton scleroxylon a gradual reduction in rooting percentage was recorded with distance from the apex. In comparison to the present study, many authors (Leakey 1983; Hansen 1986, 1988; Hartman et al., 1990; Jawanda et al., 1991; Al-Sagri and Alderson 1996) reported that the best rooting of cuttings is usually found from the basal portions of shoots. This variation comes due to the variation in accumulation of photosynthetic products, mostly carbohydrates or it could be due to juvenility factors

(Jawanda *et al.* 1991), species, environmental conditions, the season of propagation, degree of maturity, rate of growth (Hansen 1986; Leakey and Coutts 1989).

Average Root Length

Analysis of variance also reveals that no significant difference was observed per treatment on the topmost segment and in the middle segment for the Average root length. These results were different from the results mentioned by Araya (2005) that the difference in rooting due to cutting position can be related to the difference in the chemical composition of the shoots. Hartman *et al.* (1990) also indicated middle cuttings could be a more mature and low meristematic activity to develop roots than the apical cuttings.

Average Survival Rate

The highest average survival rate for the topmost segment was in treatment 3 with (93%), followed by treatment 4 and treatment 5 with the same mean of (80%), followed by treatment 1 (56%). For the middle segment, the highest average survival rate was observed in treatment 1 (90%) followed by treatment 4 (80%), treatment 5 (73%), and the lowest average survival rate was in treatment 2 (60%) (Table 3).

Treatment 1 has significantly lowered average survival rate of (56%) compared to treatment 3 (93%) Treatment 4 (80%) and Treatment 5 (80%) (Table 3).

The findings of Patricio et.al, (2006) in their study on the Macropropagation of *Shorea guiso* using stem cuttings partially corroborate when they mentioned that the application of IBA increased% survival by 16% for cuttings treated with 500 ppm IBA (86%) over untreated (74%). However, it contradicts when they further mentioned that doubling the concentration from 500 ppm of IBA to 1000 ppm did not improve the survival of the cuttings which is not observed in this present study.

This study shows that in treatment 3 (1000 ppm) of IBA with 93% survival rate has a significant difference in terms of survival rate compared to other levels of treatment lowered than 1000 ppm like in treatment 2(500 ppm) with 70% survival rate and the control or no IBA application (treatment 1) with 56% survival rate (Fig. 4).

Table 3. IBA: Comparison of the mean of cutting segments at each level of treatment on the Survival Rate of Yakal wildlings cuttings 160 DAP.

Treatment	Survival (%)				
Levels	Top most	Middle segment			
	segment				
1	56 ^c	90 ^a			
2	$70^{\rm bc}$	60 ^c			
3	93 ^a	70 bc			
4	80 ^{ab}	$80^{\text{ ab}}$			
5	80 ab	73 abc			

Means with the same letter are not significantly different.

CV: 15.14 Mean: 75



Fig. 4. IBA: Standard of error of the mean of cutting segments at each level of treatment on the Survival Rate of Yakal wildlings cuttings 160 DAP. (Values represent mean ± standard error (SE: 6.58).

Table 4. IBA: Comparison of cutting segments at each level of treatment on the average survival rate of cutting from Yakal wildlings 160 DAP.

	Average Survival Rate					
Segment	1	2	3	4	5	
Top most	56 ^b	70 ^a	93 ^a	80 a	80 a	
Middle	90 ^a	60 ^a	70 ^b	80 ^a	73 ^a	
a.c. 1.11				1.01 .1 11	<u>.</u>	

Means with the same letter are not significantly different

No significant difference was observed between treatment 3 and treatment 4 and between treatment 3 and treatment 5 (Fig. 4). This result suggests that using 1000 ppm of IBA is beneficial compared to using 1500 ppm of IBA in terms of the survival rate of cuttings from Yakal wildlings. Likewise, using 10mL/gallon of water of hormex, is more economical than using 500 ppm, 1000 ppm & 1500 ppm of IBA in terms of survival rate for the topmost segment cuttings from Yakal wildlings.

For the middle segment, significant differences in the survival rate were also observed in Treatment 1 with a (90%) survival rate compared to Treatment 2 (60%), and Treatment 3 (70%). A significant difference was also observed between the survival rate of Treatment 2 (60%) and Treatment 4 (80%) survival rate. This result implied that the application of IBA did not affect the survival rate of the middle segment of the cuttings on the basis that Treatment 1 produces a significantly higher survival rate of cuttings compared to treatment 2 and treatment 3. This result supports the findings of Totaan (2019) when he mentioned that it is probable that the application of exogenous auxin to the stem cuttings of Antidesma bunius (Linn) has no effect at any level of concentration because of the interference of high level of endogenous auxin present during the period of collection. He further mentioned that there were reports that the cuttings with a larger diameter and longer length result in better survival and growth under normal conditions (Hannerz et al., 1999, Vigl and Rewald 2014, OuYang et al., 2015).

Table 4 shows significant results were observed between means of the topmost segment and middle segment in treatment 1 with 56% and 90%, respectively. This result implies that the middle segment of cuttings produces a higher survival rate compared to the topmost segment when planted without the application of IBA. But, this finding contradicts the finding of Hartman, *et al.* (1997) as cited by Hansel (1986) when they mentioned that the survival rates varied from 62.4% for middle cuttings to 80.11% for top cuttings.

They also added that for many years' propagation ability has been known to vary between cuttings from different parts of the same plant, especially in woody species and this was correlated with the structure of the stem or difference in the chemical composition of the plant along the stem. However, the topmost segment has a significantly higher survival rate compared to the middle segment in treatment 3 with a means of 93% and 70%, respectively (Table 4). Likewise, using of 1000 ppm of IBA significantly higher in the topmost segment compared to the middle segment.



NAA hormone

Average Number of Leaves

Table 5 presents the average number of leaves in the topmost segment and middle segment 160 DAP.

The highest average number of leaves for the topmost segment was in treatment 1 with a mean of (1.0), followed by treatment 5 (.76), treatment 4 (.72), treatment 3 (.64) and the lowest was in treatment 2 (.61). For the middle segment, the highest mean was observed in treatment 5 (.56) followed by treatment 4 (.46), treatment 2 (.44) treatment 3 (.35) and the lowest was in treatment 1 (.34). No significant result was observed in the comparison between means of levels of treatments in the topmost segment, the same with the means of levels of treatments in the middle segment in terms of a number of leaves (Table 5).

However, the mean comparison on the average number of leaves in the topmost segment and middle segment showed a significant difference with a mean of .75 for the topmost segment and 43 in the middle segment (Table 6).

These results show that the topmost segment of cuttings produces more leaves than the middle segment. Again, this result supports the study of Kassahun and Mekonnen (2011) on the effect of cutting position on the propagation ability of *stevia* through its effect on leaf number varies from top and middle cutting position of a plant.

In their study, they mentioned that the leaf numbers of the seedlings ranged from 5.6 to 8.22 pairs the highest being recorded for top cuttings and the lowest for the middle cutting position.

Table 5. NAA: Comparison on the mean of the cutting segments of cuttings from Yakal wildlings on the Average Number of Leaves 160 DAP

Treatment	Replicate	Cutting	Cutting
		(1)	(2)
		Mean	Mean
1	3	1.0	.34
2	3	.61	.45
3	3	.64	.34
4	3	.72	.46
5	3	.76	.56

CV: 47.01 Mean: .59

Table 6. NAA: Comparison on the mean of cutting segments on ave. number of leaves in of Cuttings from Yakal wildlings 160 DAP.

Average	N Group
.75	15 ^a
.43	15 ^b
	·75 ·43

Means with the same letter are not significantly different.

Average Number of Roots

Analysis of variance shows that no significant difference was observed between levels of treatments in the topmost segment and in the middle segment for the Average number of roots of cuttings.

However, the mean comparison on the average number of roots of the topmost segment and middle segment had a significant difference with a mean of 25% for the topmost segment and .oo in the middle segment (Table 7).

These results show that the topmost segment produces more roots than the middle segment cuttings (Fig. 5). These results further supports the statement of Wassner and Ravetta (2000) as cited by Kassahun and Mekonnen (2011) when they reported that in rooting of Neem (*Azadirachta indica* A. Juss), where cutting from the upper part of the branches rooted better than the lower ones. Hartman *et al.* (1990) as cited by Araya (2005).

Also mentioned that the difference in rooting due to cutting position can be related to the difference in the chemical composition of the shoots.

Middle cuttings could be a more mature and low meristematic activity to develop roots than the apical cuttings. Moreover, Hartmann, *et al.* (2002) and Brardwaj and Mishra (2005) stated that the rooting ability of juvenile cuttings may be ascribed to optimum levels of sugars and the total carbohydrate content and low nitrogen levels while the reduction in rooting potential of cuttings from the stem of mature donors might be due to a decrease in the content of endogenous auxins or an accumulation of inhibitory substances. **Table 7.** NAA: Comparison of the mean of cutting segments on the average number of roots of Yakal wildlings cuttings160 DAP.

Cuttings	Means	N Group
1	.25	15 ^a
2	.00	$15^{\rm b}$

Means with the same letter are not significantly

CV: 220.89 Mean: .12



Fig. 5. NAA: Standard Error of the mean of cutting segments on the average number of roots of Yakal wildlings cuttings160 DAP (Values represent mean ± standard error (SE: .16).

Average Root Length

Analysis of variance shows that no significant difference was observed between levels of treatments in the topmost segment and in the middle segment on the average root length of cuttings. However, the mean comparison on the average root length of the topmost segment and middle segment had a significant difference with a mean of .69 for the topmost segment and .oo for the middle segment (Table 8). These results show that the topmost segment produces root length than the middle segment (Fig. 6). This finding could be supported by some studies clarifying that the apical dominance of the apical shoot could affect rooting due to the variation of endogenous auxin or/and cytokinin contents with respect to the axillary shoots (Pandeliev et al., 1990, Torres 2004). In addition, stem cuttings within the juvenile stage have less lignified tissues and lower production of rooting inhibitors as compared to stem cuttings collected from mature plants (Leopold and Kriedemann 1975, Hartmann et al., 1997, Castro and Bonfil, 2013).

Survival Rate

The highest average survival rate in the topmost segment was in treatment 1 with (96%), followed by treatment 3 (83%) treatment 5 (76%), followed by treatment 2 (63%). For the middle segment, the highest average survival rate was in treatment 2 (76%) followed by treatment 1 (control) with (73%), treatment 5 (10mL/gal), with (66%) and the lowest was in treatment 4 (1500 ppm) with (43%) (Table 9).

Table 8. NAA: Comparison on the mean of cutting segments on the average root length of cuttings from Yakal wildlings 160 DAP.

Cuttings	Average root	N Group
	length	
1	.69	15 ^a
2	.00	15 ^b
	1	1 101 11 0077

Means with the same letter are not significantly *CV*: 193.44 Mean: .3492



Fig. 6. NAA: Standard Error of the mean of cutting segments on the average root length of cuttings from Yakal wildlings 160 DAP (Values represent mean \pm standard error (SE: .39).

No significant result was observed in treatment means in the topmost segment and treatment mean in the middle segment in terms of survival rate. However, the mean comparison on the average survival rate in the topmost segment and the middle segment has a significant difference with (78%) and (61%) respectively (Table 10). These results showed that the topmost segment had a higher survival rate compared to the middle segment of the cuttings.

Table 9. NAA: Comparison on the treatment means of cutting segments on Average Survival rate of Cuttings from Yakal wildlings 160 DAP.

		Treatment						
Cuttings	1	2	3	4	5			
1	.96	.63	.83	.70	.76			
2	.73	.66	.46	.43	.66			
CV: 28.35 Me	ean: 69							

Table 10. NAA: Comparison on the mean of cutting segments on Average Survival rate of cuttings from Yakal wildlings on the survival rate 160 DAP.

Cuttings	Average Survival Rate	N Group
	(%)	
1	78	15 ^a
2	61	15 ^b

Means with the same letter are not significantly

Comparison between Hormones (IBA and NAA) and levels of concentration Top most segment (cuttings 1)

Average Number of Leaves

The highest average number of leaves in IBA were observed in treatment 3 with a mean of (1.0), followed by treatment 4 (.92), treatment 2 (.79), treatment 5 (.77) and the lowest mean was in treatment 1 (.70). For NAA, the highest mean was observed in treatment 1 (1.0) followed by treatment 5 (.76), treatment 4 (.72) treatment 3 (.64) and the lowest was in treatment 2 (.61). No significant result was observed in the comparison between hormones and levels treatment means for the topmost segment (Table 11).

Average Number of Roots

Analysis of variance shows that no significant difference was observed between means of Hormones (IBA and NAA) and levels of concentration for the topmost segment on the Average number of roots of cuttings.

Average Root Length

Analysis of variance shows that no significant difference was observed between means of Hormones and its levels for the topmost segment on the Average roots length of cuttings.

Average Survival Rate

The highest average survival rate in IBA was in treatment 3 with (93%), followed by treatment 4 and treatment 5 with the same (80%), followed by treatment 2 (70%) and treatment 1 (56%) average survival rate. For NAA, the highest mean survival rate was observed in treatment 1(96%) followed by treatment 3 (83%), treatment 5 (76%), and treatment 2 (63%) (Table 12).

Table 11. Top most segment: Comparison of meansbetween Hormone and levels of treatment on theAverage Number of Leaves of cuttings from Yakalwildlings 160 DAP.

Treatment	Replicate	IBA	NAA
		Mean	Mean
1	3	.70	1.0
2	3	.79	.61
3	3	1.0	.64
4	3	.91	.72
5	3	•77	.76

CV: 32.70 Mean: .79

Table 12. Top most segment: Comparison ofHormone at each level of treatment on the AverageSurvival rate of Yakal wildlings cuttings 160 DAP.

Hormone	Average Survival rate per treatment				
	1	2	3	4	5
1 (IBA)	56 ^b	70 ^a	93 ^a	80 ^a	80 ^a
2 (NAA)	96 ^a	63 ^a	83 ^a	70 ^a	76 ^a

Means with the same letter are not significantly different CV: 14.81 Mean: 77

Significant results were observed in treatment 1 on average survival rates for the topmost segment (Fig. 7). Though both experiment groups of cuttings from the topmost segment were not applied with rooting hormone, these results may be attributed to the report mentioned by Ou Yang et al. (2015) as cited by Totaan (2019) that aside from the effect of stem cutting position, the size of stem specifically the length of cuttings probably influenced the percent survival as observed in their study. He added that there are reports that the cuttings with a larger diameter and longer length result in better survival and growth under normal conditions. David (2003) also added that even if the cuttings were taken properly, they would not grow well if the environmental conditions were not correct.



Fig. 7. Top most segment: Standard Error of Hormone mean at each level of Treatment on the Average Survival rate of Yakal wildlings cuttings 160 DAP (Values represent mean ± standard error (SE: .15).

Table 13 shows that IBA, Treatment 1 has a significantly lower average survival rate of (56%) compared to treatment 3 (93%), Treatment 4, and Treatment 5 with the same of (80%). A significant difference was also observed between Treatment 3 (93%) and Treatment 2 (70%). This result suggests that Treatment 3 of IBA has a higher average survival

rate for the topmost segment compared to other Treatment levels of IBA. For NAA, Table 13 shows that Treatment 1 has a significantly higher average survival rate of (96%) compared to Treatment 2 (63%), Treatment 4 (70%), and Treatment 5 (76%). A significant difference was also observed between Treatment 3 and Treatment 2. This result suggests that the application of IBA and NAA would not affect on the significant survival rate of the topmost segment of cuttings. This result might be due to the accumulation of photosynthetic products, mostly carbohydrates or it could be due to juvenility factors (Jawanda et al., 1991), environmental conditions, the season of propagation, degree of maturity, rate of growth (Hansen 1986; Leakey and Coutts 1989) that supports for the higher survival rate of cuttings from wildlings of Yakal even without the application of rooting hormone, IBA and NAA.

Table 13. Top most segment: Comparison treatment at each level of Hormone on the Average Survival rate of Yakal wildlings cuttings 160 DAP.

	IBA	NAA
Treatment		
1	56 ^c	96 ^a
2	70 bc	63 °
3	93 ^a	83^{ab}
4	80 ^{ab}	$70^{\rm bc}$
5	80 ^{ab}	76 ^{bc}

Means with the same letter are not significantly different

Moreover, the application of 1000 ppm of IBA for the topmost segment has no significant difference when using 1500 ppm. No significant difference was also observed using hormex: 10mL/gal of water compared to 1000 ppm and 1500 ppm.

Middle segment

Average Number of Leaves

The highest average number of leaves for IBA were observed in treatment 1 with a mean of (.62), followed by treatment 3 (.45), treatment 4 (.34), treatment 2 (.32), and treatment 5 (.19). For NAA, the highest mean was observed in treatment 5 (.56) followed by treatment 4 (.46), treatment 2 (.44) treatment 3 (.34), and treatment 1 (.34) (Table 14). No significant result was observed in the comparison between treatment means of IBA and NAA (Fig. 8).



Fig 8. Middle segment: Standard Error between hormone and treatment means on the Average Number of Leaves of Yakal wildlings cuttings 160 DAP. (Values represent mean ± standard error (SE: .15).

Average Number of Roots

Analysis of variance shows that no significant difference was observed between treatment means of IBA and NAA on the Average number of roots. However, the mean comparison on the average number of roots in IBA and NAA showed a significant difference with .19 for IBA and .00 for NAA (Table 15). These results shown that IBA significantly produces more roots compared to NAA. These findings corroborate the findings of Larsen and Guse (1997) and Kesteret al, (1990) when they reported that the most reliable rooting hormone is indolebutyric acid (IBA) while others such as naphthalene acetic acid (NAA) can also be used. Though there were reports that it may also be toxic to young/ succulent cuttings of certain species, IBA is still probably the best hormone for general use because of being non-toxic to plants over a wide range of concentration levels (Kester et al., 1990).

Table 14. Middle segment: Comparison betweenhormone and treatment means on the Average Numberof Leaves of Yakal wildlings cuttings 160 DAP.

Treatment	Replicate	IBA	NAA
		Mean	Mean
1	3	.62	.34
2	3	.32	•44
3	3	.45	.34
4	3	.34	.46
5	3	.19	.56

CV: 65.33 Mean: .41

Average Root Length

Analysis of variance shows that no significant differences were observed in treatment means in IBA and means in NAA for the average roots length of cuttings. However, the mean comparison on the average mean in IBA and NAA had a significant difference in terms of average roots lengths of the middle segment with a mean of .39 for IBA and .00 in NAA (Table 16). This result shows that IBA produces more root length than NAA.

Table 15. Middle segment: Mean comparison on the average number of roots in IBA and NAA on the Yakal wildlings cuttings160 DAP.

	Average	N Group
IBA (H1)	.19	15 ^a
NAA (H2)	.00	$15^{\rm b}$

Means with the same letter are not significant CV: 254.13 Means: .098

Table 16. Middle segment: Mean comparison on the average root length in Hormone 1(IBA) and hormone 2 (NAA) of Yakal wildlings cuttings160 DAP.

Hormone	Means	N Group
IBA	.39	15 ^a
NAA	.00	15 ^b
	1	1 101 .

Means with the same letter are not significant CV: 230.79 Means: .19

Survival Rate of the middle segment

No significant result was observed in comparison to the treatment mean of IBA and treatment means of NAA in terms of the survival rate of Yakal wildling cuttings. This result reveals that the application of IBA and NAA in the Middle segment had no significant impact on the survival rate of Yakal wildlings cuttings.

Conclusions

The highest average survival rate for the topmost segment was in treatment 1 with (96%) survival rate and followed with the IBA hormone application in treatment 3 with (93%) survival rate. While for the middle segment, the highest survival rate was in treatment 1 with (90%) survival rate. Using IBA hormone in the topmost segment produces more leaves compared to the middle segment of cuttings from Yakal wildlings 160 DAP. Application of IBA of various levels did not influence the survival rate of the middle segment for the reason that Treatment 1 produces a significantly higher survival rate compared to other treatments applied with various levels of IBA. Application of NAA hormone significantly produces a higher average number of leaves, average number of roots, average root lengths, and average survival rate for the topmost segment compared to the middle segment. Application of NAA hormone produces a higher survival rate with 78% on the topmost segment compared to 61% for the middle segment.

Recommendations

Increase the duration of the study by at least 1 year to further evaluate the rooting potential and survival rate of cutting segments from Yakal wildlings.

The topmost segment and middle segment of cuttings could be a potential source of good planting materials even without the application of IBA, NAA & hormex. However, a low concentration from 500 ppm below may be tried. Application of another rooting hormone other than IBA, NAA & hormex is recommended. The majority of forest nurseries in the Philippines are non-mist, in order to evaluate the potential of Yakal wildlings cuttings propagation, it is recommended that a similar study should be conducted in a non-mist set-up.

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References

Adriance GW, Brinson FR. 1955. Propagation of Horticulture Plants. New York:mcGraw- Hill Books, Inc. pp. 119-131.

Agbo CU, Obi IU. 2007. Variability in propagation potentials of stemcuttings of different physiological ages of *Gongronemalatifolia* Benth. World J. Agric. Sci **3(5)**, 576-581.

Arya ST, Tomar R, Tokyt OP. 2005 Effect of plant age and auxin treatment on rooting response in stem cuttings of Prosopis cineraria. Journal of Arid Environments **27(1)**, 99-103.

Ashton PS, Gan YY, Robertson FW. 1984. Electrophoretic and Morphological Comparisons in 10 Rain-Forest Species of Shorea (Dipterocarpaceae), Botanical Journal of the Linnean Society pp. 293-304.

Ashton PS. 1982. Dipterocarpaceae. In: Van Steenis CGGJ (ed) Flora malesiana. Series I. Spermatophyta 9, 237-552.

Ashton PS. 2005 "Dipterocarpaceae," In: E. Soepadmo, L. G. Saw and R. C. K. Chung, Eds., *Tree Flora of Sabah and Sarawak*, Vol. 5, Government of Malaysia, Kuala Lumpur pp. 63-288

Bhardwaj DR, Mishra VK. 2005. "Vegetative propagation of Ulmus villosa: e ffects of plant growth regulators, collection time, type of donor and position of shoo to nadventitiou sroot formation in stem cuttings," New Forests, vol **29, no. 2,** pp. 105-116.

Caspa RG, Biloso A, Akalakou C, Mafolo J, Tsobeng, Kouodiekong L, Tchoundjeu Z. 2014. Nursery substrates and provenances influence rooting performance of juvenile, single-node vine cuttings of *Gnetumafricanum* Welw. (*Gnetaceae*) <u>A</u>frika focus-Volume **27**, Special Agroforestry Issue pp. 7-21 **Caspa RG. Kouodiekong L, Nwegueh AB, Tenku SN, Lahjou JC, Onana J.** 2009. Effects of different substrates on the rooting and shoot development of juvenile stem cuttings of Naucleadiderrichii. (De Wild & T. Durand) Merrill. Int. Biol. Sci **3(5)**, pp. 1124-1132.

Chhun T, Taketa S, Tsurumi S, Ichii M. 2003. The effects of auxin on lateral root initiation and root gravitropism in a lateral rootless mutant Lrt1 of rice (*Oryzasativa* L.). Plant Growth. Regul **39**, 161-170.

David F, Midcap T. 2003. Propagation of Woody Ornamentals by Cuttings. Institute of Food and Agricultural Sciences. University of Florida.

Dick JM, Aminah A. 1994. Vegetative propagation of tree species indigenous to Malaysia. Common For. Rev **73**, 164-171.

Edson JL, David L, Laurenm F. 1991. Propagation of western larch by stem cuttings. Western J. Applied Forestry **6(2)**, 115-125.

Fletcher WW, Kirlwood RC. 1982. Herbicides and Plant Growth Regulators. Great Britain: Granada Publishing, Ltd. P. 810. Delimar Publisher Inc pp. 316-318.

Florido HB, Roxas CA, Cortiguerra FF. 1998. Some Important Philippine Forest Trees Named Before the Turn of the 20th Centur y. Research Information on Ecosystems (RISE). Ecosystem Research and Development Bureau, College.

Fogaca CM, Fett-Neto AG. 2005. Role of auxin and its modulators in the adventitious rooting of Eucalyptus species differing in recalcitrance. Plant Growth.Regul**45**, 1-10.

Forestry Department Peninsular Malaysia (**FDPM**), **Kuala Lumpur**, **August.** 1998. Guidelines to propagate Dipterocarp species by stem cuttings, Malaysia – ITTO project Sustainable Forest management and Development in Peninsular Malaysia, PD 185/91 Rev. 2 (F) – Phase 1. **Goebel K.** 1900. Organography of plants Part 1, Authorized English edition by Isaac Bayley Balfour, Clarendon Press Oxford.

Gregorio NO, Herbohn JL, Harrison SR. 2005. 'Germplasm access and planting stock quality insmallholder forest nurseries in Leyte, the Philippines', in SR Harrison, JL Herbohn, J Suh, EO.

Hansen J. 1988. Effect of cutting position on rooting, auxiliary bud breaks and shoots growth in Stephanotis floribunda. Acta Horticulturae **226**, 159-163.

Haq R. 1992. Effect of light and weed competition on the survival and growth of *Abies pindrow* seedlings of various ages in different soils media in the moist temperate forests of Pakistan. Pak. J. Forestry **42(3)**, 148-162.

Harrison S, Gregorio N. 2006. Designing a Wildlings Collection Strategy to Support Production of Planting Materials for Green Offsets in Southern Mindanao.

Hartman HT, Kester DC, Davies FT, Geneve RL. 2002. Plant Propagation: Principles and Practices (7th edition). Prentice- Hall, 880p. New Jersey.

Hartman HT, Kester DC. 1968. Plant Propagation Principle and Practices Prentice Hall New Delhi India pp. 305-518.

Hartman HT, Kester DC. 1975. Plant Propagation Principles and Prentice Hall of India Private Limited. New Delhi, India pp. 208-211.

Hartman HT, Kester DE, Davies FT, Geneve LR. 1997. Plant Propagation: Principles and Practices (6th Edn), Prince-Hall International, Englewood Cliffs, UK, London 770 pp.

Hartman HT, Kester DE, Davies FT. 1990. Plant Propagation: Principles and Practices (5th Edn), Prince-Hall International Editions, Englewood Cliffs, New Jersey 647 pp. Hartman HT, Kester DE. 1983. Plant propagation: Principles and Practices (4th Edn), Prince-Hall International, London 727 pp.

Hitchcock AE, Zimmerman PW. 1940. Effects' obtained with mixtures of root-inducing and other substances. Cont. Boyce Thompson Inst **11(2)**, 143.

Hudson K. 1997. Overview of cutting propagation. University of Auburn FY 614, 3/7/97.

Itoh AY, Kanzaki TM, Ohkiba T, Palmiotto PA, LaFrankie JV, Kendawang JJ, Lee HL. 2002. Rooting ability of cutting relates to phylogeny, habitat preference and growth characteristics of tropical rainforest trees. For. Ecol.mgmt **168**, 275-278.

Jawanda JS, Singh A, Singh S, Bal JS. 1991. Effect of indolbutyric acid and shoot portion on the rooting of cuttings in Japanese plum. Acta Horticulturae **283**, 189-197

Kassahun BM, Mekonnen SA. 2011. Effect of Cutting Position and Rooting Hormone on Propagation Ability of Stevia (Stevia rebaudiana Bertoni). The African Journal of Plant Science and Biotechnology **6**, 5-8.

Kesari V, Das A, Rangan L. 2010. Effect of genotype and auxin treatments on rooting response in stem cuttings of CPTs of *Pongamia pinnata*, a potential biodiesel legume crop.Curr. Sci **98(9)**, 1234-1237.

Kester DE, Hartmann TH, Davier FT. 1990. Plant propagation: Principles and Practices. 5th edition. Prentice Hall, Singapore p. 647.

Kettle CJ. 2010. "Ecological Considerations for Using Dipterocarps for Restoration of Lowland Rainforest in South- east Asia," *Biodiversity and Conservation*, Vol **19**, **No. 4**, pp. 1137-1151.

Kumar V, Naidu MM, Ravishankar GA. 2006. Developments in coffee biotechnologies–in vitro plant propagation and crop improvement. Plant Cell Tissue Culture **87**, 49-65.



Larsen FE, Guse WE. 1997. Propagating deciduous and evergreen shrubs, trees and vines with stem cuttings. A Pacific Northwest Cooperative Extension Publication, Washington, USA p. 10.

Leakey RRB. 2004 "Physiology of vegetative reproduction," in Encyclopaedia of Forest Sciences, J. Burley, J. Evans, and J. A. Youngquist, Eds pp. 1655-1668, Academic Press, London, UK.

Leakey RRB. Mohammed HRS. 1985. The effect of stem length on root initiation in sequential single node cuttings of Triplochiton sleroxylon K. Schum. Journal of Horticultural Science **60(5)**, 431-437.

Mangaoang and Vanclay J. 2004. ACIAR Smallholder Forestry Project – Redevelopment of Timber Industry Following Extensive Land Clearing, Proceedings from the End of the Project Workshop, 19–21 August 2004, Ormoc City, the Philippines pp. 279-291.

Mesen F, Newton AC, Leakey RRB. 1997. Vegetative propagation of Cordiaalliodora (Ruiz &Pavon) Oken: The effects of IBA concentration, propagation medium and cutting origin. Forest Ecology and management **92**, pp. 45-54.

Miri-Nargesi SM, Sedaghathoor S. 2015. Study on rooting of kiwifruit cultivars (*Actinidiachinensis*) in different substrates and rooting hormones. South Western Journal of Horticulture, Biology and Environment **6(1)**, p. 33-42.

Moreira O, Silvam L, Moura M. 2009. Propagation of the endangered Azorean cherry *Prunus azori ca*using stem cuttings and air layering. *Arquipelago* Life Mar. Sci **26**, 9-14.

Ofori DA, Newton AC, Leaky RRB, Grace J. 1996. Vegetative propagation of Milicia excelsa by leafy stem cuttings: Effects of auxin concentration, leaf area and rooting medium. Forest Ecology and Management **84**, 39-48.

Ofori-Gyamfi E. 1998. Investigation in some factors affecting vegetative propagation of Coffee (*Coffee conaephera*var. Robusta Pierve). M Phil. Thesis, University of Cape Coast, Ghana.

Okunlola, Ibironke A. 2013. The effects of cutting types and length on rooting of Duranta repensin the Nursery. Global journal of human social science, geography, geosciences, environmental and disaster management; Global journal Inc. (USA), Vol (13).

Olosunde OM, Olasantan FO, Olubode OO. 2008. Effect of Growth Media on Rooting of Queen of the Philippine (*Mussaenda philippica* A. Rich) Department of Horticulture, University of Agriculture P.M.B.2240, Abeokuta, Ogun State. Nigerian Journal of Horticultural Science, ISSN 1118-2733 Vol 13, pp. 68-74.

Ono EO, Rodrigues JD, Pinho SZ. 2000. Studies on stem cuttings of kiwi (*Actinidia chinensis* P L. C V Bruno).Brazilian Archives of Biology and Technology [online] **43(1)**.

Overbeek JV. 1951. Use of growth substances in tropical agriculture in plant growth regulators in plant growth substances. Edited by Skoog, Fulke. University of Wisconsin Press.

Paparozzi ET. 2008. Anatomical and physiological changes that occur during rooting of cuttings. In: *Plant Propagation Concepts and Laboratory Exercises.* (Edited by Beyl, C. A. and Trigiano, R. N.), Boca Raton, New York pp. 189-194.

Patricio HP, Castaneto JT, Vallesteros AP, Castaneto ET. 2006. Macropropagation of *Shorea guiso* Using Stem Cuttings, Journal of Tropical Forest Science **18(3)**, 198-201.

Pollisco, Mitzi T. 1997. Some developments on dipterocarp propagation research in the Philippines. In: Proceedings of International Workshop BIO-REFOR. Bangkok, Thailand, (Sangwanit) pp. 34-44.

Ritchie GA. 1991. The commercial use of conifer rooted cuttings in forestry: a world overview. New Forests **5**, 247-275.

Sakai S, Harrisonm RD, Momosem K, Kuraji K, Nagamasum H, Yasunari T, Chong L, Nakashizuka T. 2006. "Irregular droughts trigger mass flowering in a seasonal tropical forests in <u>Asia</u>". American Journal of Botany **93(8)**, 1134-39.



Sakai S, Momose K, Yumoto T, Nagamitsu T, Nagamasu H, Hamid A, Nakashizuka T. 1999. "Plant reproductive phenology over four years including an episode of general flowering in a lowland dipterocarp forest, Sarawak, Malaysia". American Journal of Botany **86(10)**, 141436.

Shiembo PN, Newton AC, Leakey RBB. 1996. Vegetative propagation of Gnetum africanum Welw., a leafy vegetable from West Africa. Journal of Horticultural Science **71(1)**, 149-155.

Suzuki E, Ashton PS. 1996. Sepal and nut size ratio of fruits of Asian Dipterocarpaceae and its implications for dispersal. Journal of Tropical Ecology **12**, pp. 853-870.

Symington CF. 1943. Forester's Manual of Dipterocarps. Malayan forester records no 16. Penerbit Universiti Malaysia. Kuala Lumpur.

Syros T, Yupsanis T, Zafiriadis H, Economou A, 2004. Activity and isoforms of peroxidases, lignin and anatomy, during adventitious rooting in cuttings of EbenuscreticaL. J Plant Physiol **161**, 69-77.

Teklehaimanot Z, Tomlinson H, Lemma T, Reeves K. "Vegetative propagation of Parkia biglobosa (Jacq) Benth, 1996. An undomesticated fruit tree from West Africa," Journal of Horticultural Scienceand Biotechnology, vol **71, no. 2**, pp.205-215,

Tolentino EL, Carandang WM, Roshetko J. 2002. Evaluation of Smallholder Tree Farmers Nursery: Quality Stock Production in Support of the Tree Domestication Program of the Philippines, College of Forestry and Natural Resources. UPLB, Los Baños, Laguna. **Totaan, Darwin E.** 2019. Effects of stem cutting section and indole-3-butyric acid on the vegetative propagation of *Antidesma bunius* (Linn) Spring, Department of Forestry and Agroforestry, College of Agriculture System and Technology, Pampanga State Agricultural University, Magalang, Pampanga, Philippines det21577@gmail.com

Wassner D, Ravetta D. 2000. Vegetative propagation of Grindelia chiloesis (Asteraceae). Industrial Crops and Products **11(1)**, 7-10.

Weaver RT. 1972. Plant Growth Substances in Agriculture. San Francisco, California: W.H. Freeman and Co p. 128.

Wells JS. 1966. Plant propagation practices. The MacMillan Co. New York.

Whiting D, Roll M, Vickerman L. 2014. Colorado Master Gardener Garden notes: Plant Growth Factors-Plant Hormones. Retrieved from http:// www.ext.colostate.edu/mg/gardennotes/145.

Yeboah J, Amoah FM. 2009. The rooting performance of Shea tree (*Vitellaria paradoxa* C.F. Gaertn) cuttings leached in water and application of rooting hormone in different media. J. Plant Sci **4(1)**, 10-14.