



RESEARCH PAPER

OPEN ACCESS

Evaluation of the discriminatory power of plant dna barcodes rbcl & matk between species of fabaceae

Saadullah^{1*}, Zaheer-ud-Din Khan¹, Muhammad AShaq², Zaib-u-Nisa¹

¹*Department of Botany, GC University, Lahore Pakistan*

²*Biodiversity Institute Ontario (BIO), University of Guelph, Canada*

Key words: DNA barcoding, Fabaceae, rbcl, matK, Identification.

<http://dx.doi.org/10.12692/ijb/8.5.75-86>

Article published on May 18, 2016

Abstract

For species identification, DNA barcoding is a novel diagnostic technique, the aim of which is to contribute towards wide range of ecological and conservation studies. In this study, we evaluated the capacity of the core DNA barcodes rbcl and matK for identifying 62 plant specimen belonging to 22 species of Fabaceae. All plant specimen were collected from district Dera Ghazi Khan, Punjab, Pakistan. In this research work, highest mean pairwise interspecific distance showed by matK and lowest for rbcl. Based on 'best match' and 'best close match' analysis function of TaxonDNA, both matK and rbcl was best with almost 73% correct identification, while based on 'all species barcodes' analysis, rbcl and matK gave the lowest percentage of correct species identifications 64.51% & 51.11%, respectively. By using MEGA5, for each marker neighbor joining (N/J) trees based on Kimur-2-parameter (K2P) were produced. In tree based analysis, species were considered to be discriminated, that form separate clusters in the tree with a bootstrap support >50%. In this study both plant DNA barcodes in combination (rbcl +matK) showed best discriminatory power between different species of Fabaceae.

*Corresponding Author: Saadullah ✉ saadullahkhan313@yahoo.com

Introduction

As we monitor the biological consequences of world climate change and try to preserve species diversity from accelerating habitat destruction. We understand very little about the diversity of plant life and animals, residing in lots of unique ecosystems on the earth. Scientists agreed that the every year price of extinction has expanded from approximately one species per million to a 100-1,000 per million. This means that lots of plant and animals are being lost every year. Maximum of those species that have not yet been identified. By using classical taxonomy its not possible to catalog natural diversity earlier than it disappears.

With more than 18000 species belonging to 650 genera, Fabaceae is the world's 3rd largest family of angiosperm and 2nd largest family of medicinal plants including 490 species which have been used for conventional drug treatments and a few are endangered because of overexploitation. Adulterants are frequently added in drugs from many important medicinal plant species. Entirely based on conventional morphological characteristics it's very difficult to discriminate among medicinal species and their adulterants. A vast range of molecular techniques have been used to overcome this problem which increased the performance and resulted in unique concept of "DNA barcoding" as a way to identify and classify species. In 2003, this novel concept of DNA barcoding was put forward by a Canadian scientist Dr Paul D.N. Hebert. With this new concept of DNA barcoding a small segment of DNA from a specific region of genomes can be a distinguishing feature for all organisms. As a Linnaean binomial is an abbreviated label for the morphology of a species, this short sequence is also an abbreviated label for the genome of the species. With the help of advanced technology, faster sequencing with minimum cost and advanced on line digital public library of sequences, this abbreviated label of genome (barcode key) will revolutionize all aspects of life (Ausubel, 2009). By using DNA barcoding markers, usually plant species are inherently more difficult to discriminate than animal species (Fazekas *et al.*, 2009). After evaluation of

different barcode loci, two plastid markers *rbcl* and *matK* was approved as standard barcodes for plants (CBOL Plant Working Group, 2009). Now DNA barcoding has turn out to be a global initiative and it has standardized the molecular identifications by the use of globally agreed protocols and segments of DNA (Hebert *et al.*, 2003; CBOL Plant Working group, 2009).

In Pakistan 82 genera with 587 species belonging to Fabaceae are reported. In Pakistan, yet no progress has been made towards the development of DNA based biodiversity inventories as a result all areas are still under collected. Primarily based on conventional botanical sampling and identification, in Pakistan taxonomists were able to identify a total of 6000 species that are being degraded at a fast pace and lots of species may face a major risk of extinction (Zabta, 2010).

Aim of this study was to test whether the *rbcl* and *matK* regions are powerful marker for identification and authentication of species of the family Fabaceae. In this research work as a tool for rapid and dependable taxonomic identity, for plant species of family Fabaceae, a bi-locus barcodes of two plastid markers (*rbcl* and *matK*) were generated. All wild plant species were collected from district Dera Ghazi Khan (70 38E and 30 03N), Punjab, Pakistan.

Materials and methods

All wild plant specimen belonging to fabaceae had been collected from District Dera Ghazi Khan (70 38E and 30 03N), Punjab, Pakistan. For tissue sampling a minimum one and maximum six specimen from the fresh leaves of each collected plant specimen have been preserved in air tight plastic bags with small amount of silica gel. Every sample of plant tissue was about 0.5 cm² in size. All plant specimen were identified and herbarium specimens were prepared as per Jain and Rao's (1977) manual and deposited as voucher specimen in Dr. Sultan herbarium of GC university, Lahore. In this study, we analyzed 62 and 45 sequences of *rbcl* and *matK* belonging to 22 and 20 species respectively. Out of total 23 collected species, 19 species have both

rbcl and matK barcodes. Three species like *Acacia nilotica* (5 sequences); *Bauhinia variegata* (1 sequence) and *Prosopis juliflora* have no matK sequences while there was no rbcl sequence of *Abutilon indicum* (Table-1).

DNA extraction, amplification and sequencing

Under a project "GCUDG" on BOLD, extraction of DNA was carried out from silica gel dried leaves. At the Canadian Centre for DNA Barcoding (CCDB), university of Guelph, DNA extraction, PCR amplification and sequencing was done. By following standard protocol, the labeled tubes of 96- well box were loaded with tissue samples taken from silica gel dried leaves (http://ibol.org/wpcontent/uploads/2011/04/Sample_Submission_Package-Plant.pdf). By using the semi-automated method for plant DNA extractions, tissue lysis and DNA extraction have been executed (http://www.ccdb.ca/CCDB_DOCS/CCDB_DNA_Ext_raction-plant_life.pdf) (Ivanova *et al.*, 2008). According to the standard protocol of CCDB, PCR products for rbcl and matK were obtained (http://www.ccdb.ca/CCDB_DOCS/CCDB_Amplification-flora.pdf). The primers rbcl- F (ATGTCACCACAAA CAGAGACTAAAGC) (Levin *et al.*, 2003) and rbcl-R (GTAAAATCAAGTCCACCRGC) (Kress & Erickson 2007) had been used for the sequencing of 552 bp rbcl barcode while the 773 bp long matK barcode was acquired with the matK-KIM primers, MatK- 1RKIM-f (CCCAGTCCATCTGGAAATCTTGGTTC) and MatK- 3FKIM-r (GTACAGTACTTTTGTGTTTACGAG) described at http://www.ccdb.ca/CCDB_DOCS/CCDB_PrimerSet_s-plants.pdf. Purification and bi-directional sequencing of PCR products was also carried at CCDB (Hajibabaei *et al.*, 2006).

Sequence alignments and molecular analysis

By using CODONCODE aligner, assembling and editing of sequences of both barcodes was done (CodonCode company, Dedham, MA, U.S.A.). Separate analysis of nucleotide sequences of each barcode of matK and rbcl was done. Using default

parameters under the profile alignment option on MEGA5, multiple sequence alignments were carried out with ClustalW (Tamura *et al.*, 2011). For 'Neighbor joining' cluster analysis of both barcodes (matK and rbcl), 1 consensus barcode of all species was obtained by using the 'Consensus Barcode Generator' function of TaxonDNA because in the data set there were more than one sequence for most of the species (Meier *et al.*, 2006). Among taxa, patterns of sequence divergence were visualized by means of neighbor-joining (NJ) analysis which was performed on MEGA5 (Tamura *et al.*, 2011). For assessment of node support, bootstrap test with 500 replicates was carried out on MEGA5, (Felsenstein, 1985). Sequence distances were computed with Kimura 2-Parameter (K2P) evolutionary model (Kimura, 1980).

From aligned sequence data, the accuracy of species assignments of the samples were tested by using 'best match', 'best close match' and 'all species barcodes' functions of TaxonDNA. 'Best match' "best close match" and all species barcode' of each sequence was determined by its comparison with all other sequences in the aligned data set. For each locus or combined loci in the dataset, distribution of pairwise interspecific and intraspecific distances was analyzed by the 'pairwise summary' function of TaxonDNA (Meier *et al.*, 2006).

Results and discussion

PCR amplification and bidirectional sequencing of rbcl and matK markers

The important criteria for the assessment of utility of DNA barcodes is success of PCR amplification along with sequence recoverability from both gene regions rbcl and matK. The amplification success in both barcodes rbcl and matK was 91% (65/67) and 68% (45/67) respectively. The aligned sequence length of rbcl was 505-552 bp and 724-846 bp of matK (Table-1). In bidirectional sequencing recovery, for most of the PCR amplicon 552 bp long target sequence of rbcl mostly showed no variation in sequence length while matK showed significant variation in its sequence length. Our results supported the earlier studies of

Kress *et al.*, 2005 and Kress and Erickson, 2007 in which high PCR amplification and sequencing success with no variation in sequence length of *rbcl* was reported. Moreover, Maia *et al.*, 2012 reported 100% PCR amplification and sequencing success in *rbcl*. This research work also support the previous

research work of Zhang *et al.*, 2012 which reported highly variable PCR success rate of *matK*, ranged between 40% to 97%. Although, in this study no repeat sequences were documented in *matK* as documented by Fazekas *et al.*, 2010 which influenced the sequencing quality.

Table 1. List of collected plant specimen with maximum number of sequences and base pairs of both DNA barcodes *matK* and *rbcl*.

Serial Number	Plant name	Maximum Number of Sequences	No. of Bases in <i>matK</i> Barcodes	No. of Bases in <i>rbcl</i> Barcodes
1	<i>Abutilon indicum</i>	5	795	Nil
2	<i>Acacia brownii</i>	1	810	552
3	<i>Acacia modesta</i>	1	789	551
4	<i>Acacia nilotica</i>	5	Nil	552
5	<i>Albizia lebbeck</i>	3	784	528
6	<i>Argyrolobium roseum</i>	2	787	552
7	<i>Astragalus adscendens</i>	1	782	510
8	<i>Astragalus onobrychis</i>	5	784	552
9	<i>Astragalus purshii</i>	1	724	552
10	<i>Atylosia aphylla</i>	1	764	552
11	<i>Bauhinia variegata</i>	1	Nil	552
12	<i>Cassia occidentalis</i>	1	494	552
13	<i>Crotalaria medicaginea</i>	1	783	505
14	<i>Crotalaria pumila</i>	2	784	521
15	<i>Dalbergia sissoo</i>	5	846	552
16	<i>Indigofera hechstetteri</i>	1	802	552
17	<i>Indigofera sessiflora</i>	2	794	552
18	<i>Indigofera suffruticosa</i>	2	792	529
19	<i>Lathyrus aphaca</i>	5	846	552
20	<i>Medicago polymorpha</i>	6	846	540
21	<i>Melilotus officinalis</i>	6	781	552
22	<i>Prosopis cineraria</i>	5	846	552
23	<i>Prosopis juliflora</i>	5	Nil	552

Table 2. Summary of the pairwise intraspecific and interspecific distances in the barcode loci of different species of Fabaceae.

Barcode loci	Intraspecific distances (%)		Interspecific distances (%)	
	Minimum	Maximum	Minimum	Maximum
<i>rbcl</i>	0.00%	0.45%	0.00%	2.00%
<i>matK</i>	0.00%	0.50%	0.00%	3.00%
<i>rbcl+matK</i>	0.00%	0.07%	0.00%	22.91%

Intra / Inter specific Divergence

By using the criteria of DNA 'barcode gap' the distributions of intra vs. inter-specific variability have been compared. On the basis of 'barcode gap', a species is considered as distinct from it is nearest neighbor if its minimum inter-specific distance between nearest neighbor is greater than its

maximum intra-specific distance. In this study, among the 62 sequences of species belonging to Fabaceae the percent intra-specific divergence ranges from 0.0% to 0.45% and 0.0% to 0.50% for *rbcl* and *matK* respectively (Table-2). In this intra-specific distance analysis, 82.22% and 98%, sequences of *matK* and *rbcl* respectively demonstrated no intra-

specific variation. For *rbcl* inter-specific divergence varies from 0.0% to 2% while 0.0% to 3% was observed for *matK*. In *rbcl*+*matK* combination 0.00% to 0.7% intra-specific and 0.00% to 22.91% inter-specific distance was observed (Table-2). This study included 13 congeneric species from 5 genera for *rbcl*

and 10 congeneric species from 4 genera for *matK*. Pairwise divergences among these congeneric species were considered their ability to distinguish the species. For *rbcl* and *matK*, per genus the number of congeneric species varied between 2 to 4 and 2 to 3 respectively.

Table 3. Identification success based on the 'best match', 'best close-match' and 'all species barcodes' analysis by TaxonDNA.

Barcode Locus & Threshold Value	Best Match			Best Close Match			All Species Barcodes					
	Correct (%)	Ambiguous (%)	Incorrect (%)	Correct (%)	Ambiguous (%)s	Incorrect (%)	No Match	Correct (%)	Ambiguous (%)	Incorrect (%)	No Match	
rbcl		72.58%	11.29%	16.12	72.58%	11.29%	6.45%	9.67%	64.51%	22.58%	3.22%	9.67%
	0.5%	(45)	(7)	(10)	(45)	(7)	(4)		(40)	(14)	(2)	(6)
	1 %	(45)	(7)	(10)	(45)	(7)	(5)	8.06%	(40)	(15)	(2)	(5)
matK		73.33%	0.00%	26.66%	73.33%	0.00%	4.44%	22.22%	51.11%	26.66%	0.00%	22.22%
	0.5%	(33)		(12)	(33)		(2)	(10)	(23)	(12)		(10)
	1 %	(33)	(0)	(12)	(33)	(0)	(3)	(9)	(23)	(13)		(9)

The number of congeneric species pairs formed with *rbcl* sequences were 32 while sixteen congeneric species pairs have *matK* sequences. Out of 13 congeneric species with *rbcl* sequences 10/13 (76%) species and 9/10 (92%) congeneric species with *matK* sequences have been successfully identified. With no inter-specific distance, two congeneric species *Crotalaria pumila* and *Crotalaria medicaginea* remained unidentified with both *matK* and *rbcl* sequences. Generally, in closely related congeneric species barcoding gaps are usually narrow due to which large overlap was observed among the *rbcl* sequences of congeneric species as compared to congeneric species with *matK* sequences. Across all the species (non-congeneric), pairwise divergence in both *rbcl* and *matK* sequences demonstrated clear boundaries between species and differentiated 80.65% and 88.89% species respectively. Results of this research work are almost similar to the work of Zhang *et al.*, 2012 and de Vere *et al.*, 2012 whom have distinguished plant groups above the species or generic levels with the help of barcode gap and distribution of intra and interspecific distances among species.

The above results indicated that *matK* showed more

discriminatory power than *rbcl*. Hollingsworth *et al.*, 2011 have also reported how the discriminatory power of *matK* is slightly greater than *rbcl*. In their research work, Gao *et al.*, 2011 have also reported that among the sequences of fabaceae, *matK* demonstrated more discriminatory power than *rbcl*. Yet, there were exceptions, *rbcl* sequences of *Acacia nilotica*, *Acacia brownii*, both *rbcl* and *matK* sequences of *Crotalaria pumila*, *Crotalaria medicaginea* overlapped and showed zero interspecific distance between one another. Because of absence of barcode gap *Acacia nilotica*, *Acacia brownii*, *Crotalaria pumila* and *Crotalaria medicaginea* did not warrant further analysis to determine barcode gap. Absence of barcode gap in *matK* and *rbcl* have fairly documented by Pettengill and Neel 2010; Fu *et al.*, 2011; Jiang *et al.*, 2011 and Yang *et al.*, 2012 at species level in several plant genera. Except few species with zero inter-specific distance most of the species of Fabaceae have unique *matK* and *rbcl* sequences. In this study, reliable identification of species was provided by barcode gap analysis but in plants a single parameter has not been a sufficient for identification of species. So as an alternative criterion 'best match', 'best close match' and 'all species barcode analysis' was suggested by

Meier *et al.*, 2006.

Best Close Match and All Species Barcode Analysis

The analysis on the basis of 'best match', 'best close match' and 'all species barcodes' was done by using TaxonDNA (Meier *et al.*, 2006). All above mentioned parameters were employed to test the accuracy of species assignments. By using above parameters closest match of a sequence was determined by its

comparisons with all the other sequences. According to Xiang *et al.*, 2011 and Zhang *et al.*, 2012, large number of researchers have applied these statistics in barcode studies for species assignments. On the basis of frequency distribution of pairwise intra-specific distances similarity threshold is established. The threshold was set at a value below which 95% of all intra-specific were found (Meier *et al.*, 2006).

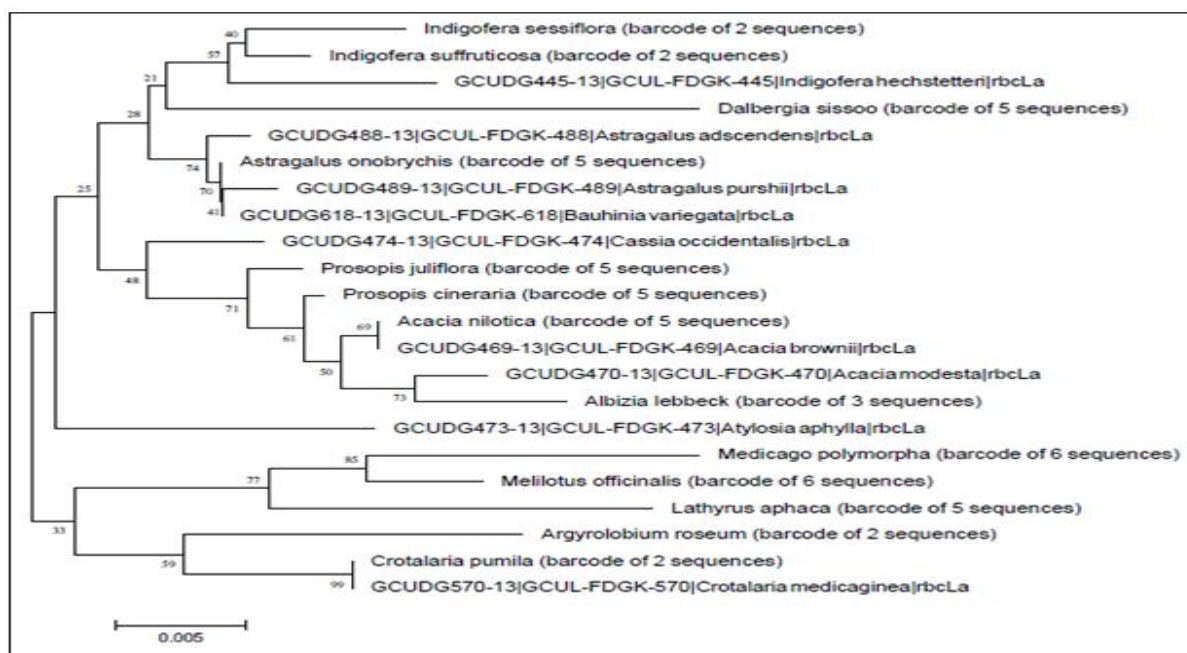


Fig. 1. Phylogenetic Tree of rbcL.

The evolutionary history was inferred using the Neighbor-Joining method. The optimal tree with the sum of branch length = 0.16217251 is shown. The percentage of replicate trees in which the associated taxa clustered together in the bootstrap test (500 replicates) are shown next to the branches. The tree is drawn to scale, with branch lengths in the same units as those of the evolutionary distances used to infer the phylogenetic tree. The evolutionary distances were computed using the Kimura 2-parameter method and are in the units of the number of base substitutions per site. The analysis involved 22 nucleotide sequences. Codon positions included were 1st+2nd+3rd+Noncoding. All positions containing gaps and missing data were eliminated. There were a total of 469 positions in the final dataset. Evolutionary analyses were conducted in MEGA5.

The query sequence is assigned to a species with which it shows the smallest genetic distance. Within this study, the rbcL and matK sequences of individual samples were queried against sequences reference barcode library of GCUDG project on BOLD. A successful identification was considered to be achieved if both sequences were from the same species otherwise mismatched sequences were considered as failure. At 0.5% and 1% threshold

values, % age of correct species identifications according to 'best match' and 'best close match' analysis, were 72.58% and 73.33% in rbcL and matK respectively (Table-3). At both above mentioned threshold values, in 'best close match' analysis "ambiguous" identification were 11.29% and 0.00% in rbcL and matK respectively while incorrect identification have been 16.12% in rbcL and 26.66% in matK. In 'best match' and "best close

match” analysis of rbcL, % age of sequences without any match at 0.5% and 1% were different and exact same situation was observed in matK also (Table-3). In rbcL, at 0.5% and 1% threshold, %age of sequences without any match were 9.67% and 8.06% while in matK %age of sequences without any match at 0.5% and 1% were 25.22% and 20.00% respectively (Table-3). On the basis of ‘all species barcodes’ analysis, the identification success by both barcodes rbcL and matK was lower than ‘best match’ and ‘best close match’ analysis as the correct identifications by rbcL and matK were 64.51% and 51.11% respectively while % age of ambiguous identification and no match by rbcL was different at both threshold, same situation

was observed for matK (Table-3). Based on ‘all species barcodes’, analysis matK with 0.00% of incorrect species identification performed well than rbcL (3.22%) (Table-3). In the ‘best match’, ‘best close match’ and ‘all species barcodes’ analysis, “correct identification” means that the hit in the database with smallest genetic distances is from same species as that of the query; “ambiguous identification” signifies that several hits from our database were found to give the same smallest genetic distance towards the query sequence; “incorrect identification” signifies that the hit having the smallest genetic distance is not from the expected species.

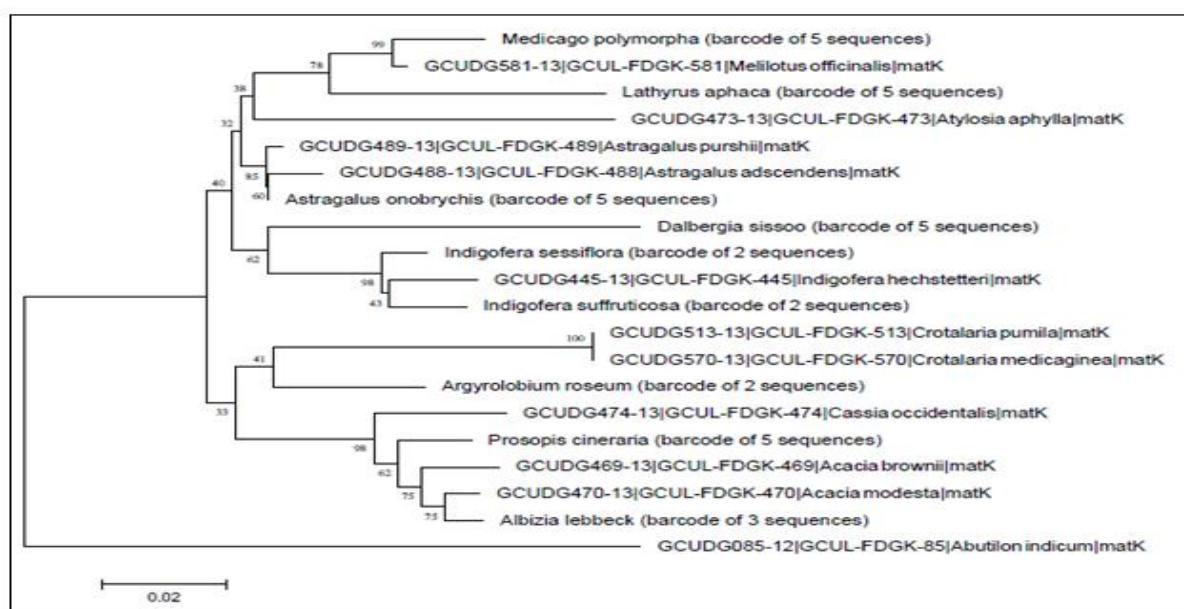


Fig. 2. Phylogenetic Tree of matK.

The evolutionary history was inferred using the Neighbor-Joining method. The optimal tree with the sum of branch length = 0.58950767 is shown. The percentage of replicate trees in which the associated taxa clustered together in the bootstrap test (500 replicates) are shown next to the branches. The tree is drawn to scale, with branch lengths in the same units as those of the evolutionary distances used to infer the phylogenetic tree. The evolutionary distances were computed using the Kimura 2-parameter method and are in the units of the number of base substitutions per site. The analysis involved 20 nucleotide sequences. Codon positions included were 1st+2nd+3rd+Noncoding. All positions containing gaps and missing data were eliminated. There were a total of 348 positions in the final dataset. Evolutionary analyses were conducted in MEGA5.

In this research work, among tested loci there was no consistency in species identification success, as in ‘best match’ and ‘best close match’ analysis, matK with 73.33% identification success showed slightly better performance than rbcL (72.58%) while in ‘all

species barcodes’ analysis rbcL with 64.51% performed better than the matK 51.11% (Table-2). In 2010, Mattio and Payri in their study on DNA barcoding of *Sargassum* species have reported that the species identification success was different

among different loci. In a study by Xiang *et al.*, 2011, matK emerged as the best barcode for identification of species in the genus *Holcoglossum*. In contrast, according to Newmaster *et al.*, 2008, matK did not show good results in identification of species of Myristicaceae and identification success was not more than 48.6%. As being a plant DNA barcode the performance of rbcL hasn't been very promising in many plant groups. In 2010, Ren *et al.*, have reported only 10% identification success of *Alnus*

species by rbcL. For identification of closely related species of *Lysimachia* L. (Myrsinaceae), Zhang *et al.*, in 2012 have reported very poor performance by rbcL which ranged from 25.7 % – 32.3%. Little and Stevenson, in 2007 and Ferguson in 2012 have criticized the distance based method because for distinguishing taxonomic groups the determination of a single universal threshold of genetic distance is extremely difficult.

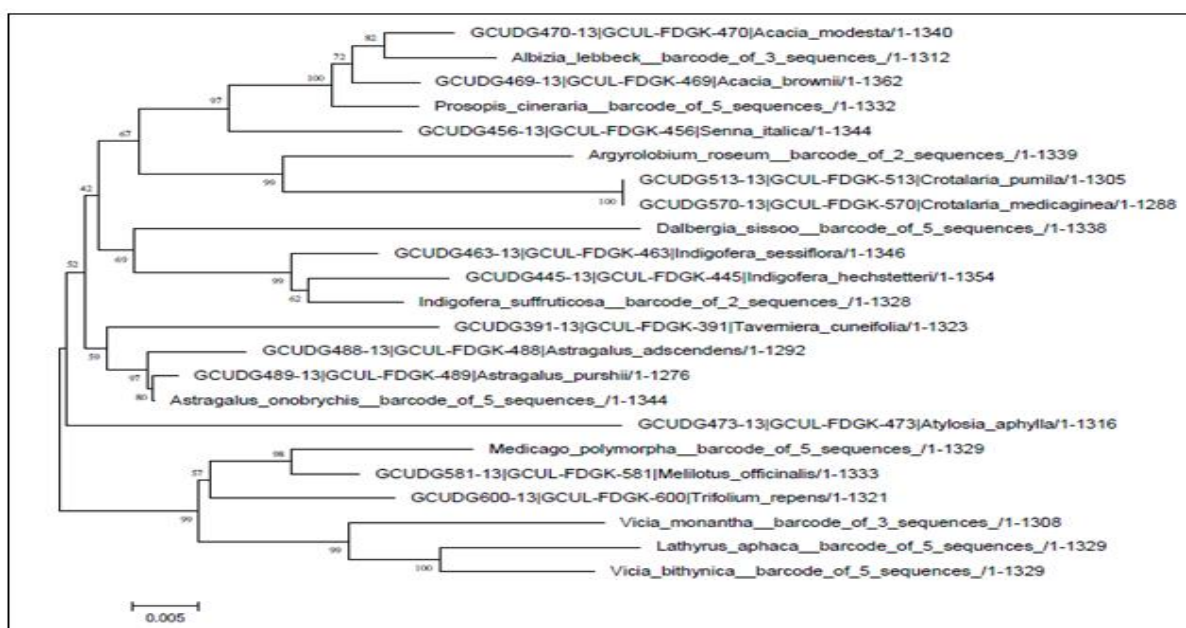


Fig. 3. Phylogenetic Tree of rbcL + matK.

The evolutionary history was inferred using the Neighbor-Joining method. The bootstrap consensus tree inferred from 500 replicates [2] is taken to represent the evolutionary history of the taxa analyzed. Branches corresponding to partitions reproduced in less than 50% bootstrap replicates are collapsed. The percentage of replicate trees in which the associated taxa clustered together in the bootstrap test (500 replicates) are shown next to the branches. The evolutionary distances were computed using the Kimura 2-parameter method and are in the units of the number of base substitutions per site. The analysis involved 23 nucleotide sequences. Codon positions included were 1st+2nd+3rd+Noncoding. All positions containing gaps and missing data were eliminated. There were a total of 1011 positions in the final dataset. Evolutionary analyses were conducted in MEGA5.

The great variation of barcode gap across the different plant groups is a fact, which strongly supports this criticism. Moreover, assigning group specific threshold is not reliable when the estimated intra-group divergence does not represent the entire range of the distribution (Fazekas *et al.*, 2008). In 2012, a study conducted on DNA based evolutionary analysis of *Lespedeza* (Fabaceae), Xu *et al.* have

reported that incongruent signal have been showed by nuclear and plastid markers. So on the basis of above mentioned study, incongruence between sequences of matK and rbcL can not be ruled out.

Neighbor Joining (N/J) Phylogenetic Analysis

In this study, by using MEGA5, three neighbor joining trees were constructed from the aligned consensus

barcode sequences of *rbcl*, *matK* and *rbcl+matK* (Fig-1,2&3). Clustering of species in the neighbor joining tree is mostly occurred on the basis of their genome type. This N/J method has been followed in many floristic barcoding studies (Kuzmina *et al.*, 2012; Saarela *et al.*, 2013). Species forming separate clusters in the tree with a bootstrap support >50% were considered to be discriminated. (Felsenstein, 1985). Bootstrap evaluation with 500 replicates don't care how the tree is correct, its simply offers information about the steadiness of the tree topology (the branching order) and it helps to assess whether the sequence information is good enough to validate the topology (Berry and Gascuel, 1996). Sequence distances were computed using the Kimura 2-Parameter (K2P) model (Kimura 1980). Sequence distances were in the units of the number of base substitutions per site. (Tamura *et al.*, 2011). In each tree the boot strap values are in the form of numbers which are written on each node. Both the trees that were constructed from the sequences of *rbcl* and *matK* were almost similar in topology (branching order) and degree of resolution. On the basis of bootstrap threshold values both tree almost failed to identify all species. In both *rbcl* and *matK* trees bootstrap values ranged between 21% to 48% for 7 to 6 nodes respectively (Fig-1&2). On the other hand, the two locus combination (*rbcl +matK*) provided the most fully resolved and well supported tree. Except one node (42%), all nodes were fully supported with bootstrap value ranged from 52% to 100 (Fig-3). That bi-locus (*rbcl +matK*) tree was most strongly supported tree because it succeeded to discriminate nearly all species (Fig-3). In initial runs, we discovered that on the basis of bootstrap threshold value the individual performance of each plastid markers was very weak and both separate trees of *rbcl* and *matK* failed to discriminate all species (Fig-1& 2). As predicted *rbcl* demonstrated inadequate sequence variation to differentiate among closely related species (Kress *et al.*, 2007; Newmaster *et al.*, 2006). On the basis of PCR recovery *matK* showed poor performance. According to a study by Lahaye in 2008 *matK* showed better recovery rate which indicates that the PCR recovery rate of *matK* can be

improved in future. Phylogenetic tree based methods were criticized due to the fact these trees are not capable to utilize low level of divergence, which is enough for differentiating groups but not for constructing phylogenetic relationships (Fazekas *et al.*, 2008; Saarela *et al.*, 2013).

Conclusion

In literature many stories are available, in one story a barcode performed very well in a specific plant group but failed in another plant group. The equal set of barcodes executed differently in the identical plant group while used individually or in mixtures. Same set of barcodes performed differently in the identical plant groups either used individually or in combination. In this study it was observed that individually both plastid marker *rbcl* and *matK* were not so successful but In contrast, the combination of two loci (*rbcl+matK*) provided correct species identifications for 99% of species of Fabaceae. Thus our results advocated that *rbcl+ matK* combination could make a good contribution in resolving phylogeny at more than one taxonomic levels.

Acknowledgment

We thank all staff at the entomology lab, NIBGE, Faisalabad who provided excellent conditions for our lab work. We extend special thanks to Mr Mehbob, Farrukh Hussnain, Saeed Ahmad, Abdul Razzaq, Haseeb Ahmad for their support and assistance in this project.

References

- Berry V, Gascuel O.** 1996. Interpretation of bootstrap trees, threshold of clade selection and induced gain. *Molecular Biology and evolution* **13(7)**, 999-1011.
<http://dx.doi.org/10.1093/molbev/13.7.999>
- CBOL, Plant Working Group.** 2009. A DNA barcode for land plants. *Proceedings of the National Academy of Sciences USA* **106**, 12794–12797.
<http://dx.doi.org/10.1073/pnas.0905845106>
- De Vere N, Rich TCG, Ford CR.** 2012. DNA

barcoding the native flowering plants and conifers of Wales. PLoS ONE 7, 37945.

<http://dx.doi.org/10.1371/journal.pone.0037945>

Fazekas AJ, Burgess KS, Kesanakurti PR, Percy DM, Hajibabaei M. 2008. Assessing the utility of coding and non-coding genomic regions for plant DNA barcoding. PLoS ONE 3, 1–12.

Fazekas AJ, Kesanakurti PR, Burgess KS, Percy DM, Graham SW, Barrett SCH, Newmaster SG. 2009. Are plant species inherently harder to discriminate than animal species using DNA barcoding markers? Molecular Ecology Resources 9(1), 130–139.

<http://dx.doi.org/10.1111/j.1755-0998.2009.02652.x>

Fazekas AJ, Steeves R, Newmaster SG. 2010. Improving sequencing quality from PCR products containing long mononucleotide repeats. Biotechniques 48, 277–281.

<http://dx.doi.org/10.2144/000113369>

Felsenstein J. 1985. Confidence limits on phylogenies. An approach using the bootstrap. Evolution 39, 783–791.

Ferguson JWH. 2002. On the use of genetic divergence for identifying species. Biological Journal of Linnean Society 75, 509–516.

<http://dx.doi.org/10.1046/j.1095-8312.2002.00042.x>

Fu YM, Jiang WM, Fu CX. 2011. Identification of species within Tetrastigma (Miq.) Planch. (Vitaceae) based on DNA barcoding techniques. Journal of Systematics and Evolution 49, 237–245.

<http://dx.doi.org/10.1111/j.1759-6831.2011.00126.x>

Gao T, Sun Z, Yao H, Song J, Zhu Y, Ma X, Chen S. 2011. Identification of Fabaceae plants using the DNA barcode matK. Planta Medica 77, 92–94.

<http://dx.doi.org/10.1055/s-0030-1250050>

Gonzalez MA, Baralot C, Engel J, Mori SA,

Pétronell P, Riéra B, Roger A, Thébaud C, Chave J. 2009. Identification of Amazonian Trees with DNA Barcodes. PLoS ONE 4(10), 7483.

<http://dx.doi.org/10.1371/journal.pone.0007483>

Hajibabaei M, Janzen DH, Burns JM, Hallwachs Vand, Hebert PDN. 2006. DNA barcodes distinguish species of tropical Lepidoptera. Proceeding of the National Academy of Sciences USA 103, 968–971.

<http://dx.doi.org/10.1073/pnas.0510466103>

Hebert PDN, Cywinska A, Ball SL, De Waard JR. 2003. Biological identifications through DNA barcodes. Proceedings of the Royal Society of London Series B. Biological Sciences 270, 313–321.

<http://dx.doi.org/10.1098/rspb.2002.2218>

Ivanova NV, Fazekas AJ, Hebert PDN. 2008. Semi-automated, membrane based protocol for DNA isolation from plants. Plant Molecular Biology Reporter 26, 186–198.

<http://dx.doi.org/10.1007/s11105-008-0029-4>

Jain SK, Rao RR. 1977. A Handbook of Field and Herbarium Methods. Today and Tomorrow's Printers & Publishers, New Delhi.

Ausubel JH. 2009. A botanical microscope. Proceeding of the National Academy of Sciences USA 106(31), 1256–1257.

<http://dx.doi.org/10.1073/pnas.0906757106>

Jiang YC, Ding L, Zhang R, Yang Y, Zhou, Tang L. 2011. Identification of the genus Epimedium with DNA barcodes. Journal of Medicinal Plants Research 5, 6413–6417.

<http://dx.doi.org/10.5897/JMPR11.545>

Kimura M. 1980. A simple method for estimating evolutionary rates of base substitutions through comparative studies of nucleotide sequences. Journal of molecular evolution 16, 111–120.

Kress WJ, Wurdack KJ, Zimmer EA, Weigt

- LA, Janzen DH.** 2005. Use of DNA barcodes to identify flowering plants. *Proceeding of the National Academy of Sciences USA*. **102**, 8369–8374. <http://dx.doi.org/10.1073/pnas.0503123102>
- Kress WJ, Erickson DL.** 2007. A two-locus global DNA barcode for land plants: the coding *rbcl* gene complements the non-coding *trnH-psbA* spacer region. *PLoS ONE* **2(6)**, 508. <http://dx.doi.org/10.1371/journal.pone.0000508>
- Kuzmina ML, Johnson K, Barron HR, Hebert PDN.** 2012. Identification of the vascular plants of Churchill, Manitoba, using a DNA barcode library. *BioMed Central Ecology* **12**, 25. <http://dx.doi.org/10.1186/1472-6785-12-25>
- Lahaye RM, Van Der Bank, Bogari.** 2008. DNA barcoding the floras of biodiversity hotspots. *Proceeding of the National Academy of Science USA* **105**, 2923–2928. <http://dx.doi.org/10.1073/pnas.0709936105>
- Little DP, Stevenson DW.** 2007. A comparison of algorithms for the identification of specimens using DNA barcodes, Examples from gymnosperms. *Cladistics* **23**, 1–21. <http://dx.doi.org/10.1111/j.1096-0031.2006.00126.x>
- Levin RA, Wagner W, Hoch PC.** 2003. Family-level relationships of Onagraceae based on chloroplast *rbcl* and *ndhF* data. *American Journal of Botany* **90**, 107–115. <http://dx.doi.org/10.3732/ajb.90.1.107>
- Maia VH, Mata CS, Franco LO, Cardoso MA, Cardoso SRS.** 2012. DNA barcoding Bromeliaceae: achievements and pitfalls. *PLoS ONE* **7(1)**, 29877. <http://dx.doi.org/10.1371/journal.pone.0029877>
- Mattio L, Payri C.** 2010. Assessment of five markers as potential barcodes for identifying *Sargassum* subgenus *Sargassum* species (Phaeophyceae, Fucales). *Cryptogamie Algologie* **31**, 467–485.
- Meier R, Shiyang K, Vaidya G, Ng PKL.** 2006. DNA barcoding and taxonomy in Diptera: a tale of high intraspecific variability and low identification success. *Systematic Biology* **55(5)**, 715–728. <http://dx.doi.org/10.1080/10635150600969864>
- Newmaster SG, Fazekas AJ, Ragupathy S.** 2006. DNA barcoding in land plants: Evaluation of *rbcl* in a multigene tiered approach. *Canadian Journal of Botany* **84**, 335–441. <http://dx.doi.org/10.1139/b06-047>
- Newmaster SG, Fazekas J, Steevea RAD, Janovec J.** 2008. Testing candidate plant barcode regions in the Myristicaceae. *Molecular Ecology Resources* **8**, 480–490. <http://dx.doi.org/10.1111/j.1471-8286.2007.02002.x>
- Peter Hollingsworth M, Sean Graham W, Damon Little P.** 2011. Choosing and Using a Plant DNA Barcode. *PLoS ONE* **6(5)**, 19254. <http://dx.doi.org/10.1371/journal.pone.0019254>
- Pettengill J.B, Neel MC.** 2010. An evaluation of candidate plant DNA barcodes and assignment methods in diagnosing 29 species in the genus *Agalinis* (Orobanchaceae). *American Journal of Botany* **97**, 1391–1406. <http://dx.doi.org/10.3732/ajb.0900176>
- Ren BQ, Xiang XG, Chen ZD.** 2010. Species identification of *Alnus* (Betulaceae) using nrDNA and cpDNA genetic markers. *Molecular Ecology Resources* **10**, 594–605. <http://dx.doi.org/10.1111/j.1755-0998.2009.02815.x>
- Saarela JM, Sokoloff PC, Gillespie LJ, Consaul LL, Bull RD.** 2013. DNA barcoding the a Canadian Arctic Flora: core plastid barcodes (*rbcl*+*matK*) for 490 Vascular Plant Species. *PLoS ONE* **8**, 36. <http://dx.doi.org/10.1371/journal.pone.0077982>
- Tamura K, Peterson D, Peterson N.** 2011. MEGA5: molecular evolutionary genetics analysis

using maximum likelihood, evolutionary distance, and maximum parsimony methods. *Molecular Biology and Evolution* **28**, 2731–2739.

<http://dx.doi.org/10.1093/molbev/msr121>

Xiang XG, Hu H, Wang W, Jin XH. 2011. DNA barcoding of the recently evolved genus *Holcoglossum* (Orchidaceae: Aeridinae): a test of DNA barcode candidates. *Molecular Ecology Resources* **11**, 1012–1021.

<http://dx.doi.org/10.1111/j.1755-0998.2011.03044.x>

Xu B, Wu N, Gao XF, Zhang LB. 2012. Analysis of DNA sequences of six chloroplast and nuclear genes suggests incongruence, introgression, and incomplete lineage sorting in the evolution of *Lespedeza* (Fabaceae). *Molecular Phylogenetics and Evolution* **62**, 346–358.

<http://dx.doi.org/10.1016/j.ympev.2011.10.007>

Yang JB, Wang YP, Moller M, Gao LM, Wu D. 2012. Applying plant DNA barcodes to identify species of *Parnassia* (Parnassiaceae). *Molecular Ecology Resources* **12**, 267–275.

<http://dx.doi.org/10.1111/j.1755-0998.2011.03095.x>

Zabta Shinwari K. 2010. Medicinal plants research in Pakistan. *Journal of Medicinal Plants Research* **4**(3), 161-176.

Zhang CY, Wang FY, Yan HF, Hao G, Hu CM. 2012. Testing DNA barcoding in closely related groups of *Lysimachia* L. (Myrsinaceae). *Molecular Ecology Resources* **12**, 98–108.

<http://dx.doi.org/10.1111/j.1755-0998.2011.03076.x>