International Journal of Biosciences | IJB | ISSN: 2220-6655 (Print), 2222-5234 (Online) http://www.innspub.net Vol. 9, No. 1, p. 303-323, 2016

RESEARCH PAPER

OPEN ACCESS

Generation of reactive oxygen species and their impact on the health related parameters: A critical review

Muhammad Usman Khan¹, Nuareen Aziz Qurashi², Muhammad Saleem Khan^{3*}, Farhat Jabeen³, Ali Umar¹, Junaid Yaqoob⁴, Muhammad Wajid⁵

¹Department of Chemistry, University of Sargodha, Pakistan ²Department of Zoology, Government College Women University, Faisalabad, Pakistan ³Department of Zoology, Government College University, Faisalabad, Pakistan ⁴Department of Chemistry, University of Okara, Pakistan ⁵Department of Biological sciences, University of Okara, Pakistan

Key words: Reactive oxygen species, Oxidative stress, Sources, Impact, Health disorders.

http://dx.doi.org/10.12692/ijb/9.1.303-323

Article published on July 30, 2016

Abstract

Reactive oxygen species (ROS) are highly reactive molecules produced during cellular metabolism and due to some environmental factors. These species have ability to induce damage in vital molecules. The body has several antioxidant systems to cope with but imbalance of oxidants and antioxidants creates a condition commonly known as oxidative stress. The excess production of ROS is the main cause of oxidative stress. In last forty years, oxidative stress was considered as major factors for pathological disorders and ageing. In body, ROS have dual role either beneficial or harmful in the biological systems. They either act as beneficial (NO[•]) in the form of signaling or harmful producing the health disorders. Mitochondrion is the major site for ROS production. The other sources include peroxisomes, endoplasmic reticulum, membrane and cytosol. ROS are studying for their pathogenesis of obesity, diabetes, cancer, inflammation, cardiovascular disease, neurodegenerative disorders and aging. In this review, sources, causes and consequences of superoxide radical, singlet oxygen, ozone, hydrogen peroxide, hydroxyl radical, peroxynitrite, peroxyl and alkoyl radicals and reactive nitrogen species are critically analyzed and discussed in details

* Corresponding Author: Muhammad Saleem Khan 🖂 samiikhan@yahoo.com

Introduction

The oxidative stress may be the disturbance between ROS and ability of antioxidant system to remove them in the biological systems. Jones (2006) defined it as disorder of redox signaling and control. According to Sies (1997) the term oxidative stress means serious imbalance between antioxidant and production of ROS (Datta et al., 2015). ROS increase dramatically during the oxidative stress and causes lipid peroxidation, intact with nucleic acid, lipid and protein and causes loss of membrane integrity, functional changes and mutation. All these factors contribute to health disorder (Kataria et al., 2010) which will be discussed latter in this review. In body it actually has two roles. It is either beneficial or harmful. In moderate concentration, superoxide anion, nitric oxide (NO·) and other reactive oxygen species play a critical role in signaling. In higher concentration due to over production of ROS causes the oxidative stress that leads to the pathological conditions including cardiovascular diseases, cancer (Sosa et al., 2013), diabetes mellitus (Yang et al., 2011), inflammation, neurodegenerative diseases and ageing (Oyinloye et al., 2015).

Body has several enzymatic and non-enzymatic system including catalase (CAT), superoxide dismutase (SOD), GSH-Px and glutathione-Stransferase (GST), glutathione peroxidase, vitamin E components and glutathione (Memisoğullari *et al.*, 2003; Shafaq, 2012). Several synthetic and elements such as cerium also mimic the natural antioxidant scavenging the free radicals and ameliorated the oxidative stress (Khan *et al.*, 2015). Therefore, the aim of this review was to critically analyze the available literature for source types of reactive oxygen species and health disorder due to oxidative stress.

General concept about the free radicals and Oxidative stress

Oxygen is the most common source of free radicals in the biological systems. Over production of ROS inhibit the body normal function and cause damage to both cells and tissues (Wickens, 2001). It is basically produced from cellular substance of endogenous organelles include mitochondria, peroxisomes, cytochrome P_{450} and inflammatory cells (Inoue *et al.*, 2003). The exogenous sources are environmental sources including the ionization radiations, ultraviolet rays and pollutants (Klaunig *et al.*, 2010; Krumova and Cosa, 2016).

ROS primarily attacks on the cell membrane because it reacts with fatty acid of membrane and form the lipid peroxide. The accumulation of lipid peroxide leads to the production of maloiadehyde a potential carcinogen agent. This lipid peroxidation cause permanent loss of membrane elasticity and fluidity and leads to cell rupture(Klaunig et al., 2010; Krumova and Cosa, 2016). The 2nd target of free radicals is body protein. The ROS oxidize the cross link between amino acid and permanent loss of enzymes and connective tissue function (Stadtman, 1995; Wang et al., 2014). It also target the protein synthesis due to inhibition of photosystem II (Nishiyama *et al.*, 2011). The DNA is the 3rd major target of ROS in the cell. The ROS breaks the DNA after inaction and also causes cross linkage of the molecule (Chen et al., 2014).

Sources and production of ROS Endogenous sources

Mitochondria of the cell is the major site for ROS production and both complex I and Complex II found to be established for mitochondrial ROS generation (Dröse and Brandt, 2012). Many enzymes are also responsible for the generation of ROS including xanthine oxidase (Agarwal *et al.*, 2011), NADPH oxidase (Bylund *et al.*, 2010), α-ketoglutarate dehydrogenase complex (Ambrus *et al.*, 2011), dihydrolipoamide dehydrogenase (Zhang *et al.*, 2011; Kareyeva *et al.*, 2012) and d-amino acid oxidases (Fang *et al.*, 2002).

NADPHs oxidase peroxisomes present in the membrane are also the source of free radicals, which consume O_2 and generate H_2O_2 under normal condition. The H_2O_2 in the peroxisome is converted to water with the help of catalase enzyme. Oxidative stress occurs when the damaged peroxisomes unable

to convert it into water and release the H_2O_2 into cytoplasm directly. Under few conditions, H_2O_2 also reacts with O_2 . and form highly oxidizing and toxic compound (OH.) in the Fenton and Haber-Weiss reaction (Fransen *et al.*, 2012).

Gastrointestinal tract is another site for free radical synthesis. O_2 is generated from xanthine oxidase and then it is converted to H_2O_2 in a reaction catalyze by gluthione peroxidase or catalase. H_2O_2 produced by neutrophils is utilized by meloperoxidase produce hypochlorite (OC¹⁻) ions which is very reactive with short reaction time and makes the membrane impermeable (te Velde *et al.*, 2008). The generated ROS in the gastrointestinal tract oxidize the protein, damage the DNA and protein creating diseased condition in colon (Sanders *et al.*, 2004).

Mitochondrial ROS production

The electron transport chain consisted of four complexes. The electrons move to complex I by NADPH and complex II by FADH₂. Then the electrons move to complex III and finally to IV depositing to the molecular oxygen and forming H₂O. however in some cases, electron leak prematurely to O_2 before reaching complex IV and formed superoxide instead of water in complex I to III (Muller *et al.*, 2007). It is estimated that about 1 to 2% of all consumed O_2 formed superoxide, and this production is increases independently in aging (Aung-Htut *et al.*, 2013; Breitenbach *et al.*, 2014).

Peroxisomal production

Investigation in the last decades revealed that endoplasmic reticulum and peroxisomes produce more ROS than mitochondria (Fransen *et al.*, 2012). Peroxisomes are filled with variety of enzymes like oxidoreductase/flavoenzymes that are considered involved in hydrogen peroxide. These enzymes are involved in oxidation of fatty acid or D-amino acid catabolism producing the hydrogen peroxide (Fransen *et al.*, 2012). Some studies also found peroxisomes not only involved in the hydrogen peroxide but also superoxide production. The production of superoxide is mainly due to xanthine oxidase during ischemia reperfusion injury (Cantu-Medellin and Kelley, 2013). Beside the production of ROS, peroxisome also generated the RNS by action of hemeprotein nitric oxide synthase that catalase the oxidation of L-arginine to nitric oxide (Stuehr *et al.*, 2001; Luis, 2011).

ROS production in Endoplasmic reticulum

The main contributors in the ROS production are the member of cytochrome P_{450} with the combination of disulfide isomers protein (PDI) and endoplasmic reticulum oxidoreducation EROI-1. In folding process, PDI protein induces disulfide formation in receptor protein. The isomers reduce this process and regenerated it by oxidio-reduction of ERO1. The ERO1 transferred the electrons to molecular oxygen through FAD. However, incomplete transfer led to superoxide production (Bhandary *et al.*, 2012; Benham *et al.*, 2013).

The family of P450 is found mainly in the ER and does the takes of xenobiotic detoxification by increasing solubility. In the process, electrons transferred from NADH to cytochrome P $_{450}$ leading to xenobiotic hydroxylation. Sometimes the transfer of electrons results in the formation of superoxide radicals (Bae *et al.*, 2011).

Membranes and Cytosol production of ROS

Membrane also produces ROS due to activity of NADPH oxidases. Electrons move on form NADPH to FAD, two heme type to finally O_2 forming the superoxide (Rinnerthaler *et al.*, 2012).

In cytosol, ROS produced as byproducts of arachihonic acid metabolism. Cyclooxygenase and Lipoxygenase enzymes used the arachiodic acid as substrate and produce H_2 prostaglandin and leukotriene respectively. Both enzymes have ability to produce superoxide in the presence of NADPH and NADH (Kukreja *et al.*, 1986; Whicher and Evans, 2012). The level of arachiodic acid is generally low but increase in the skin due to skin inflammatory diseases including psoriasis apoptotic dermatitis leading to aging (Ziboh *et al.*, 2000; Whicher and Evans, 2012).

Additionally, iron of the cell and organelles react with oxygen forming ROS in the cytosol. In reaction which is Haber-Weiss reaction, the ferric iron reacts with superoxide forming the ferrous iron. In other reaction, which is Fenton reaction, ferrous reacts with hydrogen peroxide regenerating the ferric iron and very reactive hydroxyl radical (OH[•]) and hydroxide (OH[•]). Both radicals are harmful (Chemizmu and Fentona, 2009). The skin acts as interface between environment and body the exogenous ROS production (Poljšak and Dahmane, 2012; Chen *et al.*, 2014).

Exogenous sources

Ionization radiations

Ionization radiations are believed to be involved in all the steps of carcinogenesis including initiations, promotion and progression (Little *et al.*, 2008). The damage done by these radiations include apoptosis, gene mutation and cancer (Riley, 1994; Kadhim *et al.*, 2013). The biological effects of ionization radiations are due to ROS which rapidly produced in radiolysis (Tulard *et al.*, 2003).

Nano-materials mediated ROS generation

Nanomaterial fullerenes and metals ions in nature induce oxidative stress (Bonner, 2007; Asghar et al., 2016). The factors of nanoparticles induce oxidative stress are cell inaction, prooxodant functional group on nanoparticles and redox cycling on the surface of nanoparticles (Huang et al., 2010). However, several studies revealed the active role of reactive particle surface in the generation of ROS (Schins, 2002; Khan et al., 2015; Shakeel et al., 2015; Khan et al., 2015b; Asghar et al., 2016). Nanoparticles produces the free radicals when oxidant and free radicals bound to the active surface of particles e.g. SiO and SiO₂ present in the quartz particles generated the OH- and O2.species (Knaapen et al., 2004). The other molecules such as nitrogen dioxide and ozone also generated the oxidative stress on surface of nanoparticles and induce oxidative stress (Buzea et al., 2007).

The free radicals are generated on the nanoparticles due to one of following reasons.

a. Free radicals are produce when the nanomaterials are dissolved in the aqueous suspension (Fubini and Hubbard, 2003; Asghar *et al.*, 2015). Upon dissolving nanoparticles release the metals ions and induce the oxidative stress (Knaapen, *et al.*, 2004). For example, quartz particles produce the H₂O₂ and O₂- in the aqueous suspension.

b. Chemical and metal compounds on the surface of nanoparticles also enhanced the ROS production and oxidative stress (Wilson *et al.*, 2002).

c. Some transition metals like chromium and copper etc. found engaged in generation of ROS in Haber-Weiss and Fenton-type reaction.

d. Some metals nanoparticles such as Co, Ni etc. also activate the intracellular pathways of MARK and NF- α B for inducing the intracellular free radicals (Nel *et al.*, 2006).

e. Nanoparticles also activated the mitochondrial mediated ROS production. As major site, once nanoparticles get enter in to mitochondria, they impaired the electron transport chain, activate the NADPH like enzymes, depolarization of mitochondrial membrane and initiating the ROS production (Xia *et al.*, 2006).

f. Internalization of nanoparticles also imitated the immune response including neutrophils and macrophages contributing to the ROS production (Risom *et al.*, 2005).

g. Small particles having high surface to volume ratio reported to involved in the ROS production due to large number of active sites. The large surface area of surface particles are more exposed to reaction than interior molecules (Nel *et al.*, 2006). All these factors contribute to ROS generation and finally oxidative stress.

h.

Mostly xenobiotic are the exogenous sources of ROS productions and oxidative stress. These sources might

be barbiturates, chlorinated compounds, phorbol esters and metal ions (Hamid *et al.*, 2016). The exogenous sources produced the ROS by metabolism of primary radicals or by activating the endogenous sources. The resultant oxidative stress induces the DNA damage, lipid peroxidation and also modulating the antioxidant system (Klaunig and Kamendulis, 2004; Pu *et al.*, 2006). Some types of ROS discovered some of them discuss in table 1.

Table 1. Reactive species and their source and place of synthesis.

Reactive species	Place or source of synthesis
Hydrogen peroxide	Large number of reactions in the body
Superoxide anion	Mitochondria, cardiovascular system
Nitric oxide	Intestinal sub mucosa and some other cells by nitrogen oxide synthase
Nitrogen dioxide	During atmospheric pollution of dioxide
Peroxyl radicals	During oxidative damage of DNA, protein and sugar etc.
Ozone	In atmosphere pollutants
Peroxynitrite	Reaction of NO and superoxide in the body
Hydroxyl	In Fenton reaction
Alkoxyl	free radical chain reaction

Superoxide radical (O_2^{-})

This reactive species is produced in the mithchondrial complex I, III and consider part of intracellular signaling (Murphy, 2009). The evidence supported that the superoxide involved in the intracellular signaling cascade by four ways.

Superoxide radical usually generated due to reduction of O_2 . It is reactive than O_2 often associated with inflammatory pathway by the activation and regulation of inflamasome and inflammatory cytokines (Goetz and Luch, 2008; Zhou *et al.*, 2011). With long half-life involves in the inactivation of catalase, GPX and oxidation of glutathione in the absence of scavengers. Different studies revealed the role of superoxide in the development of disease state including cancer (Ambrosone *et al.*, 1999), cardiovascular (Collin *et al.*, 2007), inflammation (Afonso *et al.*, 2007) and neurodegenerative diseases (Waris and Ahsan, 2006).

The intracellular enzyme SOD is an antioxidant metaloenzyme and actively involved in scavenging the super oxide radicals. It has different isoforms with different metal ions (Copper, manganese and zinc) at active site (Zelko *et al.*, 2002). In animal models, the SOD protects the brain, heart and liver from ischemic

and alcohol induced injury (Wheeler *et al.*, 2001). The mutation in SOD causes degradation of motor neuron and induced paralysis or death, susceptibility to type 2 diabetes, Alzheimer s disease and cancer (Tamai *et al.*, 2006; Wheatley-Price *et al.*, 2008). Over expression of SOD also causes the oxidative stress (Kowald *et al.*, 2006).

Singlet oxygen

It is not a free radical species with ample energy and no unpaired electrons but very reactive than O_2 with 10-5 s half-life. It is generated in the electronic excitation of molecular oxygen, termination of peroxyl radical in peroxidase- mediated reaction (Davies *et al.*, 1999). It was first observed in 1924 and found out the more reactive form of oxygen. The singlet oxygen targets the protein, nucleic acid, sterols and lipids consequence in the skin cancer. B-carotene and ascorbic acid actively involved in the scavenging of the singlet oxygen (Young and Woodside, 2001).

However, some studies also supported the beneficial role of singlet oxygen in the photodynamic therapy of carcinogenic cells. In the process of treatment carcinogenic cells accumulates the light sensitive agent during irradiation that produce the singlet oxygen and ROS in most of the cases. The singlet oxygen and ROS causes the cell death by cytotoxicity and inducing the apoptosis (Juarranz *et al.*, 2008; Plaetzer *et al.*, 2009). Chemically, several compounds are used for scavenging the singlet oxygen including histidine, azide and 2-phenylisobenzofuran in laboratory (Foote *et al.*, 2012).

Ozone (O₃)

Ozone is acidic smelling, irritating, colorless gas and a form of elemental oxygen. It is also a powerful oxidizing agent than molecular oxygen. Its low concentration (0.5 ppm) causes the lung damage. Its few hours explore initiate inflammation, damage to macrophages of pulmonary tracts and this way decreasing the resistance to infection. It also causes the irritation of eyes and oxidation of lipid and protein. The inhaled O_3 reacts with ascorbates urate and GSH of the body. These compounds are scavengers of O_3 (Halliwell and Gutteridge, 2015).

Hydrogen peroxide (H_2O_2)

Like singlet oxygen, hydrogen peroxide (H_2O_2) is also not a free radical species and showed relatively stable state in most of the studies (Park and Imlay, 2003). However it gained much interest due to its ability of generating the ROS. Hydroxyl radical is its most important ROS. In biological systems, the H_2O_2 is produced in one of the following processes; (a) reduction of the superoxide by SOD generated the H_2O_2 (b) action of amino acid oxidase, glycolate oxidase and urate oxidase on their respective substrates also generates the H_2O_2 (Benov, 2001).

The H_2O_2 also showed the direct effect on intracellular signaling cascades by transduction of signals, genetic mutation of catalase enzyme and up regulation of SOD (mn) and SOD (Cu). Chen *et al.* (2014) suggested the proliferation of endothelial cells after the treatment with H_2O_2 . Catalase and Gpx (cellular enzymes) actively involved in scavenging of H_2O_2 and conversion into the H_2O (Young and Woodside, 2001).

Hydroxyl radical (OH-)

Hydroxyl radical is considered the most toxic species produced by reduction of H_2O_2 (Park *et al.*, 2004). Due to very short half-life (10-9 sec), the hydroxyl radical immediately reacts with biomolecules after the formation. Fenton- type reaction with Iron (II) and zinc (I) and Haber-Weiss reactions are the primary sources of cellular hydroxyl radicals.

$Fe^{2+} + H_2O_2 Fe^{3+} + -OH + HO^-$ (Fenton reaction)

 O_2 ·· + H_2O_2 ·OH + O_2 + HO· {Haber- Weiss reaction Haber and Weiss, 1932, (Haber and Weiss, 1934)} It causes the oxidation of protein, lipid and nucleic acid. Base modification, DNA strand breaks and DNA cross linking was also observed after the treatment with hydroxyl radicals (Bar-Or *et al.*, 2001).

Peroxynitrite (ONOO-)

The generated superoxide and nitric oxide in the cell reacts to form proxynitrite (ONOO-) under the inflammatory condition. The peroxynitrate causes the lipid peroxidation and DNA damage. It is also involved in the ageing process because of damage to guanine in telomeres and decreases the production of collagen (Valko *et al.*, 2006; Afonso *et al.*, 2007). The other complications include the vasorestrication due to low availability of nitric oxide. Selenium in the form of selenomethionine and selenocystine found to have the protective role against single strand breaks in DNA due to ONOO- radical.

Peroxyl (RO_2) and alkoyl (RO) radicals

Both radicals are good oxidizing agent since they have ability of accepting electrons (Buettner and Jurkiwicz, 1996). Under the biological medium both reactive species undergo molecular rearrangement to form other radicals. RO• found to initiate lipid peroxidation by two pathways including, fatty acid hydroperoxide (LOOH) independent pathway and LOOH dependent pathways (Aikens and Dix, 1991). The carbon centered radical of RO• has the ability to reacts directly with certain biological molecules like DNA and albumin. RO₂• also induce peroxidation of lipids and damage the protein including lysosomes (Bailey *et al.*, 2003).

Metal based generated ROS

Metals ions such as Cu, Co, Mg, Ni, Zn are biological very important due to their contribution in normal

physiological functioning including, electron transport chain, oxygen transport, catalyst and part of various protein. However, metal ions are toxic too, if mis-regulated during homeostasis.

This condition leads to the oxidative stress due to generation of ROS and onset of many diseases like, anemia, hemochromatosis, Wilson s disease Monks diseases, cancer, diabetes, inflammation and neurodegenerative diseases (Beutler, 2007; Brewer, 2007; Jomova and Valko, 2011). Among the metal ions, copper and iron are more focused and their role in the generation of ROS is more extensively study. It is due to the fact both the metals ions are involved in the generation of OH. (Beutler, 2007; Jomova and Valko, 2011). This hydroxyl radical induce the DNA damage and cell death in the fibroblast of mammalian.

Copper is the 3rd most found metal in the human body after zinc and iron (Brandolini *et al.*, 2002). The copper also generated the hydroxyl radical during the Fenton-type reaction but 50 times faster than iron (Bar-Or *et al.*, 2001; Shen and Anastasio, 2012). The copper concentration is closely maintained to avoid the toxic level (Que *et al.*, 2008). However elevated level is associated with oxidative stress and related disease (Mishra *et al.*, 2008; Que *et al.*, 2008) including Alzheimer' s disease (Zappasodi *et al.*, 2008), cancer (Gupte and Mumper, 2007) Wilson' s disease (Ala *et al.*, 2007) renal diseases (Mishra *et al.*, 2008) and cardiovascular disorders (Leone *et al.*, 2006; Shen and Anastasio, 2012).

External environment polluted with some heavy metals are also the source of oxidative stress and toxicity. For example, Khan *et al.* (2015b) critically reviewed the toxicity and oxidative due to silver nanoparticles in fish model. Hamid *et al.* (2016) investigated the level of murrcury and its impact on the antioxidant system of fish.

Cell has inherent ability to cope with metal based oxidative damage. The cell has various metal binding sites in metaloprotein including transferrin, ceruloplasmin, metallothionein and ferritin (Letavayová *et al.*, 2006; Kontoghiorghes *et al.*, 2008). Some chelating drugs such as, deferiprone, ferrior amino B are used for iron and N. scetyl cysteine amide, tetrathiomolybdale and penicillamine for copper to minize the toxic effects (Zheng *et al.*, 2008).

Reactive nitrogen species (NOS)

The byproduct of nitrogen oxide synthase form the 2nd group of free radicals mainly expressed in intestinal submucosa ad some selected cells (Ghafourifar and Cadenas, 2005). It has usually long half-life due to rapid diffusion into blood where it inactivated by hemoglobin. The nitric oxide has some vital role in the body including neurotransmission, immune-dilation and blood pressure regulation (Matheson et al., 2000). It also prevents the adhesion of leukocytes and toxicity of H2O2 to endothelial cells (Binion et al., 2000). However, the over production of RNS is responsible to impairment of antioxidative system and contributes the damage to large intestine mucous of membrane (Ya Sklyarov et al., 2011). The reaction between O2.- and NO. also leads to the formation of aggressive oxidizing agent the peroxynitrte (ONOO-) that causes the fragmentation of DNA and lipid peroxidation.

Impacts of ROS and RNS Lipid peroxidation

Both RNS and ROS are responsible for lipid peroxidation particularly in the membrane. As the membrane is consisted of polyunsaturated lipids and lipoprotein, the membrane is primary target of lipid peroxidation; a hydroperoxy group is attached or introduced into unsaturated fatty acid with hydrophobic tail and causes the alterations. These structural alterations disturb the hydrophobic lipidlipid interaction and create the hydroperoxy radicals and aldehydes derivatives. The ultimate end product of lipid peroxidation (Malondialdehyde) causes damage to protein by reacting with histidine imidazole group, lysine amino group and sulphydryl group of cysteine (Catalá, 2009). Severe lipid peroxidation was seen in the patients with inflammatory bowel disease depending upon the type

(Kruidenier et al., 2003; Asghar et al., 2016).

Aging

Aging research has targeted on a central finding that dates back to the year 1956. During year, Denham Harman projected that reactive oxygen species (ROS) accumulate over time and are a main contributor to the aging process (Harman, 1955). This idea was broadened sixteen years later by Harman himself identifying mitochondria because the main source of ROS, forming the basis for the mitochondrial radical theory of aging (Harman, 1972). In the last decade, serious doubts arose that ROS are indeed the foremost vital elements that are fueling aging (Hekimi et al., 2011). Intrinsic aging is delineated as results of and corporal genetic factors changes that occur/appear throughout the conventional aging process, whereas extrinsic aging focuses on aging process accelerated by environmental influences (Farage et al., 2008). It had been proposed that solely 3 % of all aging factors have a genetic background (Poljšak and Dahmane, 2012). The deterioration of metabolic processes and normal physiological functions cause aging. According to the free radical theory, the ROS is generated as byproduct of biological oxidation which induces damage to macromolecules ultimately dis-functioning and cell death(Harman, 1955).

Being the major site of intracellular superoxide production and major target of free radicals, mitochondria are closely associated with aging process. Mitochondrial ROS causes damage to mitochondrial constituents including mitochondrial DNA, protein and lipids (Park and Imlay, 2003; Belhadj Slimen et al., 2014). The oxidant induces mutation in the mitochondrial DNA defer rationed mitochondrial normal bioenergetics function leading to the aging. The damages in mitochondrial DNA increase with age which leads to DNA break and somatic mutation. These mutation cause the impairment of respiratory chain complex and increasing the mutation and oxidative damage, energy supply and normal cellular function alters leading to the apoptosis (Judge and Leeuwenburgh,

Effect on DNA

Several chemical reactions that involved the oxygen generated the reactive intermediate that damages the DNA. This damage causes mutations leading to the cancer. The researchers are trying to explore the role of reactive intermediate in the carcinogenesis (Gupta *et al.*, 2013).

Hydroxyl radical produced during Fenton reaction causes oxidation of nuclear DNA. The reaction of ROS with free radicals also leads to deleterious effect on DNA and produced mutagenesis. Most familiar DNA alteration induced by oxidative stress is 8-oxo-2' deoxyguanosine which pair both adenine and cytosine forming the GC and TA transition (Kaiser *et al.*, 2004). This mutation was seen in the skin especially in aging (Sauvaigo *et al.*, 2010). Along with modification in DNA base, ROS also produce double and single DNA breaks (Caldecott, 2008). *Effect on protein*

Oxidative stress induces reversible and irreversible oxidative modifications in the protein. Irreversible modification including tyrosine nitration and carbonylation are associated with oxidative stress and used as biomarkers in diseases and aging (Prokai *et al.*, 2007; Rao and Møller, 2011). Reversible modification includes cysteine modification (Cai and Yan, 2003). It reflects the change in cellular redox state and involved in the singling cascades (Finkel, 2011; Chung *et al.*, 2013).

Cancer

Hydroxyl radical has ability to react with guanosine of nucleotide chain and form 8-oxo-2' deoxyguanosine and high frequencies of this dimer were observed in tumor (Kuppusamy and Zweier, 1989; Ishikawa *et al.*, 2008; Kumar *et al.*, 2008). Basal cell cancer, melanoma and squamous cell cancer were the most frequent cancer types in skin and due to mutation in tumor suppressor gene P_{53} (Brash *et al.*, 1996). Transversion of G to T is the most prominent mutation in P_{53} gene and due to oxidation of guanine

(Waris and Ahsan, 2006).

Mutation in P53 increases the risk of cancer due to cell inability of apoptosis. Various studies revealed ROS not only involve in the inducing of tumor but also in its progression. The tumor produces significant amount of H₂O₂ that promote the tumor progression (Waris and Ahsan, 2006). ROS also promotes the release of calcium from locally endoplasmic reticulum store which activates the protein kinase C (PKC). The PKC was found to involve in the cell apoptosis, cell migration, proliferation and reorganization of cytoskeleton (Klaunig et al., 2010). Alteration in the signaling pathway is the most common reason for cancer genesis (de Gruijl et al., 2001). Most of studies revealed the generation of ROS in the melanomas. As melanocyte transformations the melanomas tend to disorganized and promote the ROS. This ROS activate the proto-oncogene pathways (Fruehauf and Trapp, 2008).

Neurodegenerative diseases

Oxidative damage to neurons is the most common reason for most of neurodegenerative diseases. The oxidative stress is responsible for death or dysfunction of neural cell leading to pathogenesis.

4-hydroxy-2,3-nonenal Acrolein, (HNE), F2isoprostanes and malondialdehyde (MDA) are the most important products of lipid peroxidation. Elevated level of HNE is the reason for Alzheimer s disease (Selley et al., 2002). The oxidative stress modifies the DNA base pairing by hydroxylation which increases the level of 8-hydroxyguanine and 8 hydroxy 2 deoxyguanine initiating the AD (Gabbita et al., 1998; Nakabeppu, 2014). In AD the activity of catalase, glutathione peroxide, superoxide dismutase and glutathione reductase increase and use as biomarker of oxidative stress related diseases (Pappolla et al., 1992; Zhao and Zhao, 2013).

Heart Disease

Several lines of evidence demonstrate that oxidative stress plays a very important role in the pathological process and development of cardiovascular diseases, angina pectoris, myocardial infraction, and heart failure (Linke et al., 2005; Little et al., 2008; Csányi and Miller, 2014). Many studies conjointly support the role of OS in disease pathological process of coronary heart disease. Paradoxically, though moderate exercise poses an acute oxidant stress, regular endurance exercise is related to improved cardiovascular operate and a reduction in traditional CHD risk factors. These new findings are consistent with the hypothesis that adaptations elicited by acute exposures to exercise-induced oxidative stress result in long-term vascular protection. This happens through activation of signaling pathways that result in accumulated synthesis of intracellular antioxidants and antioxidant enzymes and shrunken ROS production during exercise (Pandey and Rizvi, 2009). In case of atherosclerosis, several evidences supported the free radicals role in the development of pathogenesis (Steinberg, 1997). In this hypothesis, low density lipoprotein (LDL) that are the main circulating part leave the antioxidant replete plasma, enter to sub-endothelial space of arteries and got oxidized there. The oxidized LDL initiates the process of formation of atherosclerotic lesions. The microphages take these oxidized LDL and release the other factors that stimulate the proliferation of smooth muscles. These oxidized LDL also facilitates cellular adhesion and binding of leukocytes, speed up the plaque formation and causes the stroke and heart attacks (Steinberg, 1997; Singh et al., 2015). Moreover the LDL also showed involvement in the blockage of nutritional antioxidant system (Tribble and Committee, 1999).

as well as hypertension, atherosclerosis, dyslipidemia,

Stroke is the pathological condition and occurs due to the cell death because of oxidative stress in the condition of ischemia (Alexandrova *et al.*, 2004; Bretón and Rodríguez, 2012). There are two types of strokes including ischemic stroke and hemorrhagic stroke. Several evidences support the role and generation of ROS in both conditions leading to oxidative stress. The oxidative stress interrupts the normal flow of blood flow and metabolic pathways which is the leading cause of ischemic strokes (Piantadosi and Zhang, 1996). In the hemorrhagic strokes, the blood borne cells (neutrophils, macrophages/monocytes) accumulates and increases the oxidative stress and eventually causes the disease. *Effect on visceral obesity*

Obesity showed close relationship with metabolic syndrome. It is now recognized that adipose tissue being metabolically active play a critical role in the regulation of homeostasis of energy and pathological effects in obesity related diseases. The role of white adipose tissue medaling inflammation in cardiovascular and diabetes disease is the hot topic of current investigation. Infiltrated adipose tissue by bone morrow produced the macrophage that secrete adipose and cytokinase in the systematic circulation result chronic inflammation (Wellen and Hotamisligil, 2005; Lugogo et al., 2011). Obese persons have higher level of oxidative stress marker and these markers increase with gain of weight (Vincent et al., 2010; Savini et al., 2013). Multiple sources are found associated to obesity including inherent source that increase fat distribution and adiposity. Some other sources are behavioral changes that make a person obese. Increased adipose tissue significantly correlated with increased level of oxidative stress biomarkers (Fujita et al., 2006; Steffes et al., 2006; Bonomini et al., 2015).

Obesity is also associated with several conditions including, insulin resistance, hyper tension, hyperlipidemia and diabetes. Each of these increases the oxidative stress in the obese persons (Shaheen *et al.*, 2007).

Eating balance and rich in antioxidant is necessary to maintained the healthy life and reduce the oxidative stress (Khan *et al.*, 2016). Unfortunately this protection is less effective in obese person having sedentary life style with diet of lower dietary antioxidant and lover vitamin level (Nayak *et al.*, 2000). Obesity increases the chronic oxidative stress that causes the endogen damage found mostly in the cardiovascular system and nonalcoholic hepatic steatosis (Roberts and Sindhu, 2009; Kizhakekuttu and Widlansky, 2010).

Conclusion

Reactive oxygen species and reactive nitrogen species are very reactive species produce during the oxidative metabolisms and due to environmental pollutant sources. These ROS causes oxidative stress in the organisms including the human beings. The oxidative stress causes damage to genetic materials inducing the health disorder and aging. Therefore, it is recommended both natural and synthesis antioxidant should be used to cope with imbalance of oxidatant and antioxidant level.

References

Afonso V, Champy R, Mitrovic D, Collin P, Lomri A. 2007. Reactive oxygen species and superoxide dismutases: role in joint diseases. Joint Bone Spine 74(4), 324-329.

http://dx.doi.org/10.1016/j.jbspin.2007.02.002

Agarwal A, Banerjee A, Banerjee U. 2011. Xanthine oxidoreductase: a journey from purine metabolism to cardiovascular excitation-contraction coupling. Critical reviews in biotechnology **31(3)**, 264-280.

http://dx.doi.org/10.3109/07388551.2010.527823

Aikens J, Dix T. 1991. Perhydroxyl radical (HOO.) initiated lipid peroxidation. The role of fatty acid hydroperoxides. Journal of Biological Chemistry **266(23)**, 15091-15098.

http://dx.doi.org/10.1006/abbi.1993.1455

Ala A, Walker AP, Ashkan K, Dooley JS, Schilsky ML. 2007. Wilson's disease. The Lancet 369(9559), 397-408.

http://dx.doi.org/10.1016/s0140-6736(07)60196-2

Alexandrova M, Bochev P, Markova V, Bechev B, Popova M, Danovska M, Simeonova V. 2004. Dynamics of free radical processes in acute ischemic stroke: influence on neurological status and outcome. Journal of Clinical Neuroscience **11(5)**, 501-506. http://dx.doi.org/10.1016/j.jocn.2003.10.015

Ambrosone CB, Freudenheim JL, Thompson PA, Bowman E, Vena JE, Marshall JR, Graham S, Laughlin R, Nemoto T, Shields PG. 1999. Manganese superoxide dismutase (MnSOD) genetic polymorphisms, dietary antioxidants, and risk of breast cancer. Cancer research **59(3)**, 602-606.

Ambrus A, Torocsik B, Tretter L, Ozohanics O, Adam-Vizi V. 2011. Stimulation of reactive oxygen species generation by disease-causing mutations of lipoamide dehydrogenase. Human molecular genetics, ddr202.

http://dx.doi.org/10.1093/hmg/ddr202

Asghar MS, Quershi NA, Jabeen F, Shakeel M, Khan MS. 2016. Genotoxicity and oxidative stress analysis in the *Catla catla* treated with ZnO NPs. Journal of Biodiversity and Environmental Sciences **8(4)**, 91-104.

Asghar MS, Qureshi NA, Jabeen F, Khan MS, Shakeel M, Noureen A. 2015. Toxicity of zinc nanoparticles in fish: a critical review. Journal of Biodiversity and Environmental Sciences 7(1), 431-439.

Aung-Htut MT, Lam YT, Lim Y-L, Rinnerthaler M, Gelling CL, Yang H, Breitenbach M, Dawes IW. 2013. Maintenance of mitochondrial morphology by autophagy and its role in high glucose effects on chronological lifespan of Saccharomyces cerevisiae. Oxidative medicine and cellular longevity 2013.

http://dx.doi.org/10.1155/2013/636287

Bae YS, Oh H, Rhee SG, Do Yoo Y. 2011. Regulation of reactive oxygen species generation in cell signaling. Molecules and cells **32(6)**, 491-509. http://dx.doi.org/10.1007/s10059-011-0276-3

Bailey DM, Davies B, Young IS, Jackson MJ, Davison GW, Isaacson R, Richardson RS. 2003. EPR spectroscopic detection of free radical outflow from an isolated muscle bed in exercising humans. Journal of applied physiology **94(5)**, 1714-

1718.

http://dx.doi.org/10.1152/japplphysiol.01024.2002

Bar-Or D, Thomas GW, Rael LT, Lau EP, Winkler JV. 2001. Asp-Ala-His-Lys (DAHK) inhibits copper-induced oxidative DNA double strand breaks and telomere shortening. Biochemical and biophysical research communications **282(1)**, 356-360.

http://dx.doi.org/10.1006/bbrc.2001.4533

Belhadj Slimen I, Najar T, Ghram A, Dabbebi H, Ben Mrad M, Abdrabbah M. 2014. Reactive oxygen species, heat stress and oxidative-induced mitochondrial damage. A review. International journal of hyperthermia **30(7)**, 513-523. http://dx.doi.org/10.3109/02656736.2014.971446

Benham AM, van Lith M, Sitia R, Braakman I. 2013. Ero1–PDI interactions, the response to redox flux and the implications for disulfide bond formation in the mammalian endoplasmic reticulum. Philosophical Transactions of the Royal Society of London B: Biological Sciences **368(1617**), 20110403. http://dx.doi.org/10.1098/rstb.2011.0403

Benov L. 2001. How superoxide radical damages the cell. Protoplasma **217(1-3)**, 33-36. http://dx.doi.org/10.1007/bf01289410

Beutler E. 2007. Iron storage disease: facts, fiction and progress. Blood Cells, Molecules, and Diseases **39(2)**, 140-147. http://dx.doi.org/10.1016/j.bcmd.2007.03.009

Bhandary B, Marahatta A, Kim H-R, Chae H-J. 2012. An involvement of oxidative stress in endoplasmic reticulum stress and its associated diseases. International journal of molecular sciences **14(1)**, 434-456.

http://dx.doi.org/10.3390/ijms14010434

Binion DG, Rafiee P, Ramanujam KS, Fu S, Fisher PJ, Rivera MT, Johnson CP, Otterson MF, Telford GL, Wilson KT. 2000. Deficient

iNOS in inflammatory bowel disease intestinal microvascular endothelial cells results in increased leukocyte adhesion. Free Radical Biology and Medicine **29(9)**, 881-888.

http://dx.doi.org/10.1016/s0891-5849(00)00391-9

Bonner JC. 2007. Lung fibrotic responses to particle exposure. Toxicologic pathology **35(1)**, 148-153. http://dx.doi.org/10.1080/01926230601060009

Bonomini F, Rodella LF, Rezzani R. 2015. Metabolic syndrome, aging and involvement of oxidative stress. Aging and disease **6(2)**, 109. http://dx.doi.org/10.14336/ad.2014.0305

Brandolini V, Tedeschi P, Capece A, Maietti A, Mazzotta D, Salzano G, Paparella A, Romano P. 2002. Saccharomyces cerevisiae wine strains differing in copper resistance exhibit different capability to reduce copper content in wine. World Journal of Microbiology and Biotechnology **18(6)**, 499-503.

Brash D, Ziegler A, Jonason A, Simon J, Kunala S, Leffell D. 1996. *Sunlight and sunburn in human skin cancer: p53, apoptosis, and tumor promotion.* Paper presented at the The journal of investigative dermatology. Symposium proceedings/the Society for Investigative Dermatology, Inc.[and] European Society for Dermatological Research.

Breitenbach M, Rinnerthaler M, Hartl J, Stincone A, Vowinckel J, Breitenbach-Koller H, Ralser M. 2014. Mitochondria in ageing: there is metabolism beyond the ROS. FEMS yeast research 14(1), 198-212.

http://dx.doi.org/10.1111/1567-1364.12134

Bretón RR, Rodríguez JCG. 2012. Excitotoxicity and oxidative stress in acute ischemic stroke. Stroke **8**, 9. http://dx.doi.org/10.5772/28300

Brewer GJ. 2007. Iron and copper toxicity in

diseases of aging, particularly atherosclerosis and Alzheimer's disease. Experimental Biology and Medicine **232(2)**, 323-335.

Buzea C, Pacheco II, Robbie K. 2007. Nanomaterials and nanoparticles: sources and toxicity. Biointerphases **2(4)**, MR17-MR71. http://dx.doi.org/10.1116/1.2815690

Bylund J, Brown KL, Movitz C, Dahlgren C, Karlsson A. 2010. Intracellular generation of superoxide by the phagocyte NADPH oxidase: how, where, and what for? Free Radical Biology and Medicine **49(12)**, 1834-1845.

http://dx.doi.org/10.1016/j.freeradbiomed.2010.09.0

Caldecott KW. 2008. Single-strand break repair and genetic disease. Nature Reviews Genetics **9(8)**, 619-631.

Cantu-Medellin N, Kelley EE. 2013. Xanthine oxidoreductase-catalyzed reactive species generation: A process in critical need of reevaluation. Redox biology **1(1)**, 353-358.

http://dx.doi.org/10.1016/j.redox.2013.05.002

Catalá A. 2009. Lipid peroxidation of membrane phospholipids generates hydroxy-alkenals and oxidized phospholipids active in physiological and/or pathological conditions. Chemistry and physics of lipids **157(1)**, 1-11.

http://dx.doi.org/10.1016/j.chemphyslip.2008.09.00 4

Chemizmu K, Fentona R. 2009. Fenton reactioncontroversy concerning the chemistry. Ecological chemistry and engineering **16**, 347-358.

Chen W, Balakrishnan K, Kuang Y, Han Y, Fu M, Gandhi V, Peng X. 2014. Reactive oxygen species (ROS) inducible DNA cross-linking agents and their effect on cancer cells and normal lymphocytes. Journal of medicinal chemistry **57(11)**, 4498-4510.

http://dx.doi.org/10.1021/jm401349g

Chung HS, Wang S-B, Venkatraman V, Murray CI, Van Eyk JE. 2013. Cysteine oxidative posttranslational modifications emerging regulation in the cardiovascular system. Circulation research **112(2)**, 382-392.

http://dx.doi.org/10.1161/circresaha.112.268680

Collin B, Busseuil D, Zeller M, Perrin C, Barthez O, Duvillard L, Vergely C, Bardou M, Dumas M, Cottin Y. 2007. Increased superoxide anion production is associated with early atherosclerosis and cardiovascular dysfunctions in a rabbit model. Molecular and cellular biochemistry **294(1-2)**, 225-235.

http://dx.doi.org/10.1007/s11010-006-9263-y

Csányi G, Miller JFJ. 2014. Oxidative stress in cardiovascular disease. International journal of molecular sciences **15(4)**, 6002-6008. http://dx.doi.org/10.3390/ijms15046002

Datta R, Alfonso-García A, Cinco R, Gratton E. 2015. Fluorescence lifetime imaging of endogenous biomarker of oxidative stress. Scientific reports **5**. http://dx.doi.org/10.1038/srep09848

Davies MJ, Fu S, Wang H, Dean RT. 1999. Stable markers of oxidant damage to proteins and their application in the study of human disease. Free Radical Biology and Medicine **27(11)**, 1151-1163. http://dx.doi.org/10.1016/b978-0-444-50957 4.50007-7

de Gruijl FR, van Kranen HJ, Mullenders LH. 2001. UV-induced DNA damage, repair, mutations and oncogenic pathways in skin cancer. Journal of Photochemistry and Photobiology B: Biology **63(1)**, 19-27.

http://dx.doi.org/10.1016/s1011-1344(01)00199-3

Dröse S, Brandt U. 2012. Molecular mechanisms of superoxide production by the mitochondrial respiratory chain Mitochondrial Oxidative

Phosphorylation 145-169 p, Springer.

Fang J, Sawa T, Akaike T, Maeda H. 2002. Tumor-targeted delivery of polyethylene glycolconjugated D-amino acid oxidase for antitumor therapy via enzymatic generation of hydrogen peroxide. Cancer research **62(11)**, 3138-3143. http://dx.doi.org/10.1002/ijc.22982

Farage M, Miller K, Elsner P, Maibach H. 2008. Intrinsic and extrinsic factors in skin ageing: a review. International Journal of Cosmetic Science **30(2)**, 87-95.

http://dx.doi.org/10.1111/j.1468-2494.2007.00415.x

Finkel T. 2011. Signal transduction by reactive oxygen species. The Journal of cell biology **194(1)**, 7-15.

http://dx.doi.org/10.1083/jcb.201102095

Foote CS, Valentine J, Greenberg A, Liebman JF. 2012. Active oxygen in chemistry **2**, Springer Science & Business Media.

Fransen M, Nordgren M, Wang B, Apanasets O. 2012. Role of peroxisomes in ROS/RNSmetabolism: implications for human disease. Biochimica et Biophysica Acta (BBA)-Molecular Basis of Disease **1822(9)**, 1363-1373.

http://dx.doi.org/10.1016/j.bbadis.2011.12.001

Fruehauf JP, Trapp V. 2008. Reactive oxygen species: an Achilles' heel of melanoma? Expert review of anticancer therapy **8(11)**, 1751-1757. http://dx.doi.org/10.1586/14737140.8.11.1751

Fubini B, Hubbard A. 2003. Reactive oxygen species (ROS) and reactive nitrogen species (RNS) generation by silica in inflammation and fibrosis. Free Radical Biology and Medicine **34(12)**, 1507-1516. http://dx.doi.org/10.1016/s0891-5849(03)00149-7

Fujita K, Nishizawa H, Funahashi T, Shimomura I, Shimabukuro M. 2006. Systemic oxidative stress is associated with visceral fat

accumulation and the metabolic syndrome. Circulation Journal **70(11)**, 1437-1442. <u>http://dx.doi.org/10.1253/circj.70.1437</u>

Gabbita SP, Lovell MA, Markesbery WR. 1998. Increased nuclear DNA oxidation in the brain in Alzheimer's disease. Journal of neurochemistry **71(5)**, 2034-2040.

http://dx.doi.org/10.1046/j.14714159.1998.71052034 .x

Ghafourifar P, Cadenas E. 2005. Mitochondrial nitric oxide synthase. Trends in Pharmacological Sciences **26(4)**, 190-195. http://dx.doi.org/10.1016/j.tips.2005.02.005

Goetz ME, Luch A. 2008. Reactive species: a cell damaging rout assisting to chemical carcinogens. Cancer letters **266(1)**, 73-83. http://dx.doi.org/10.1016/j.canlet.2008.02.035

Gupta RK, Patel AK, Shah N, Chaudhary A, Jha UK, Yadav UC, Gupta PK, Pakuwal U. 2013. Oxidative stress and antioxidants in disease and cancer: a review. Asian Pacific journal of cancer prevention: APJCP **15(11)**, 4405-4409. http://dx.doi.org/10.7314/apjcp.2014.15.11.4405

Gupte A, Mumper RJ. 2007. Copper chelation by D-penicillamine generates reactive oxygen species that are cytotoxic to human leukemia and breast cancer cells. Free Radical Biology and Medicine **43(9)**, 1271-1278.

http://dx.doi.org/10.1016/j.freeradbiomed.2007.07.0 03

Haber F, Weiss J. (1934). *The catalytic decomposition of hydrogen peroxide by iron salts.* Paper presented at the Proceedings of the Royal Society of London A: Mathematical, Physical and Engineering Sciences.

Halliwell B, Gutteridge JM. 2015. Free radicals in biology and medicine: Oxford University Press, USA. Hamid A, Khan MU, Yaqoob J, Umar A, Ali A, Rehman A, Javed S, Adnan S, Anwar A, Khan MS. 2016. Assessment of mercury load in river Ravi, urban sewage streams of Lahore Pakistan and its impact on the oxidative stress of exposed fish. Journal of Biodiversity and Environmental Sciences **8(4)**, 63-72.

Harman D. 1955. Aging: a theory based on free radical and radiation chemistry. http://dx.doi.org/10.1093/geronj/11.3.298

Harman D. 1972. The biologic clock: the mitochondria? Journal of the American Geriatrics Society **20(4)**, 145-147.

http://dx.doi.org/10.1111/j.1532-5415.1972.tb00787.x

Hekimi S, Lapointe J, Wen Y. 2011. Taking a "good" look at free radicals in the aging process. Trends in cell biology **21(10)**, 569-576. http://dx.doi.org/10.1016/j.tcb.2011.06.008

Huang Y-W, Wu C-h, Aronstam RS. 2010. Toxicity of transition metal oxide nanoparticles: recent insights from in vitro studies. Materials **3(10)**, 4842-4859.

http://dx.doi.org/10.3390/ma3104842

Inoue M, Sato EF, Nishikawa M, Park A-M, Kira Y, Imada I, Utsumi K. 2003. Mitochondrial generation of reactive oxygen species and its role in aerobic life. Current medicinal chemistry **10(23)**, 2495-2505.

http://dx.doi.org/10.2174/0929867033456477

Ishikawa K, Takenaga K, Akimoto M, Koshikawa N, Yamaguchi A, Imanishi H, Nakada K, Honma Y, Hayashi J-I. 2008. ROSgenerating mitochondrial DNA mutations can regulate tumor cell metastasis. Science **320(5876)**, 661-664.

http://dx.doi.org/10.1126/science.1156906

Jomova K, Valko M. 2011. Advances in metalinduced oxidative stress and human disease. Toxicology **283(2)**, 65-87. http://dx.doi.org/10.1016/j.tox.2011.03.001

Jones DP. 2006. Redefining oxidative stress. Antioxidants & redox signaling **8(9-10)**, 1865-1879. http://dx.doi.org/10.1089/ars.2006.8.1865

Juarranz Á, Jaén P, Sanz-Rodríguez F, Cuevas J, González S. 2008. Photodynamic therapy of cancer. Basic principles and applications. Clinical and Translational Oncology **10(3)**, 148-154. http://dx.doi.org/10.1007/s12094-008-0172-2

Judge S, Leeuwenburgh C. 2007. Cardiac mitochondrial bioenergetics, oxidative stress, and aging. American Journal of Physiology-Cell Physiology **292(6)**, C1983-C1992.

http://dx.doi.org/10.1152/ajpcell.00285.2006

Kadhim M, Salomaa S, Wright E, Hildebrandt G, Belyakov OV, Prise KM, Little MP. 2013. Non-targeted effects of ionising radiation— Implications for low dose risk. Mutation Research/Reviews in Mutation Research 752(2), 84-98.

http://dx.doi.org/10.1016/j.mrrev.2012.12.001

Kaiser J, Heidenreich W, Monchaux G, Morlier J, Collier C. 2004. Lung tumour risk in radon-exposed rats from different experiments: comparative analysis with biologically based models. Radiation and environmental biophysics **43(3)**, 189-201.

http://dx.doi.org/10.1007/s00411-004-0251-x

Kareyeva AV, Grivennikova VG, Vinogradov AD. 2012. Mitochondrial hydrogen peroxide production as determined by the pyridine nucleotide pool and its redox state. Biochimica et Biophysica Acta (BBA)-Bioenergetics **1817(10)**, 1879-1885. <u>http://dx.doi.org/10.1016/j.bbabio.2012.03.033</u>

Kataria N, Kataria AK, Pandey N, Gupta P. 2010. Serum biomarkers of physiological defense against reactive oxygen species during environmental stress in Indian dromedaries. HVM Bioflux **2(2)**, 55-60.

Khan M, Quershi N, Jabeen F, Asghar M, Shakeel M. 2016. Analysis of minerals profile, phenolic compounds and potential of Garlic (*Allium sativum*) as antioxidant scavenging the free radicals. International Journal of Biosciences **8(4)**, 72-82. http://dx.doi.org/10.12692/ijb/8.4.72-82

Khan MS, Jabeen F, Asghar MS, Qureshi NA, Shakeel M, Noureen A, Shabbir S. 2015. Role of nao-ceria in the amelioration of oxidative stress: current and future applications in medicine. International Journal of Biosciences **6(8)**, 89-109. http://dx.doi.org/10.12692/ijb/6.8.89-109

Khan MS, Jabeen F, Qureshi NA, Asghar MSSM, Noureen A. 2015b. Toxicity of silver nanoparticles in fish: a critical review. Journal of Biodiversity and Environmental Sciences **6(5)**, 211-227.

Kizhakekuttu TJ, Widlansky ME. 2010. Natural antioxidants and hypertension: promise and challenges. Cardiovascular therapeutics **28(4)**, e20-e32.

http://dx.doi.org/10.1111/j.1755-5922.2010.00137.x

Klaunig JE, Kamendulis LM. 2004. The role of oxidative stress in carcinogenesis. Annu. Rev. Pharmacol. Toxicol. **44**, 239-267.

http://dx.doi.org/10.1201/9780203904787.ch4

Klaunig JE, Kamendulis LM, Hocevar BA. 2010. Oxidative stress and oxidative damage in carcinogenesis. Toxicologic pathology **38(1)**, 96-109.

Knaapen AM, Borm PJ, Albrecht C, Schins RP. 2004. Inhaled particles and lung cancer. Part A: Mechanisms. International Journal of Cancer 109(6), 799-809.

http://dx.doi.org/10.1002/ijc.11708

Kontoghiorghes GJ, Efstathiou A, Ioannou-

Loucaides S, Kolnagou A. 2008. Chelators controlling metal metabolism and toxicity pathways: applications in cancer prevention, diagnosis and treatment. Hemoglobin **32(1-2)**, 217-227. http://dx.doi.org/10.1080/03630260701727119

Kowald A, Lehrach H, Klipp E. 2006. Alternative pathways as mechanism for the negative effects associated with overexpression of superoxide dismutase. Journal of theoretical biology **238(4)**, 828-840.

http://dx.doi.org/10.1016/j.jtbi.2005.06.034

Kruidenier L, Kuiper I, Lamers CB, Verspaget HW. 2003. Intestinal oxidative damage in inflammatory bowel disease: semi-quantification, localization, and association with mucosal antioxidants. The Journal of pathology **201(1)**, 28-36.

http://dx.doi.org/10.1002/path.1409

Krumova K, Cosa G. 2016. Overview of Reactive Oxygen Species. http://dx.doi.org/10.1039/9781849737722-00279

Kukreja R, Kontos H, Hess M, Ellis E. 1986. Superoxide generation by prostaglandin synthetase and lipoxygenase in presence of NADH or NADPH. Paper presented at the FEDERATION PROCEEDINGS.

Kumar B, Koul S, Khandrika L, Meacham RB, Koul HK. 2008. Oxidative stress is inherent in prostate cancer cells and is required for aggressive phenotype. Cancer research **68(6)**, 1777-1785. http://dx.doi.org/10.1158/0008-5472.can-07-5259

Kuppusamy P, Zweier JL. 1989. Characterization of free radical generation by xanthine oxidase. Evidence for hydroxyl radical generation. Journal of Biological Chemistry **264(17)**, 9880-9884. http://dx.doi.org/10.1080/1071576031000107344

Leone N, Courbon D, Ducimetiere P, Zureik M. 2006. Zinc, copper, and magnesium and risks for

all-cause, cancer, and cardiovascular mortality. Epidemiology **17(3)**, 308-314. http://dx.doi.org/10.1097/01.ede.0000209454.4146 <u>6.b7</u>

Letavayová L, Vlčková V, Brozmanová J. 2006. Selenium: from cancer prevention to DNA damage. Toxicology **227(1)**, 1-14. <u>http://dx.doi.org/10.1016/j.tox.2006.07.017</u>

Linke A, Adams V, Schulze PC, Erbs S, Gielen S, Fiehn E, Möbius-Winkler S, Schubert A, Schuler G, Hambrecht R. 2005. Antioxidative effects of exercise training in patients with chronic heart failure increase in radical scavenger enzyme activity in skeletal muscle. Circulation **111(14)**, 1763-1770.

Little M, Heidenreich W, Moolgavkar S, Schöllnberger H, Thomas D. 2008. Systems biological and mechanistic modelling of radiationinduced cancer. Radiation and environmental biophysics **47(1)**, 39-47.

http://dx.doi.org/10.1007/s00411-007-0150-z

Lugogo NL, Bappanad D, Kraft M. 2011. Obesity, metabolic dysregulation and oxidative stress in asthma. Biochimica et Biophysica Acta (BBA)-General Subjects **1810(11)**, 1120-1126. http://dx.doi.org/10.1016/j.bbagen.2011.09.004

Luis A. 2011. Peroxisomes as a cellular source of reactive nitrogen species signal molecules. Archives of Biochemistry and Biophysics **506(1)**, 1-11. http://dx.doi.org/10.1016/j.abb.2010.10.022

Matheson PJ, Wilson MA, Garrison RN. 2000. Regulation of intestinal blood flow. Journal of Surgical Research **93(1)**, 182-196. http://dx.doi.org/10.1006/jsre.2000.5862

Memisoğullari R, Taysı S, Bakan E, Capoglu I. 2003. Antioxidant status and lipid peroxidation in type II diabetes mellitus. Cell biochemistry and function **21(3)**, 291-296.

http://dx.doi.org/10.1002/cbf.1025

Mishra OP, Pooniya V, Ali Z, Upadhyay RS, Prasad R. 2008. Antioxidant status of children with acute renal failure. Pediatric Nephrology **23(11)**, 2047-2051.

http://dx.doi.org/10.1007/s00467-008-0875-1

Muller FL, Lustgarten MS, Jang Y, Richardson A, Van Remmen H. 2007. Trends in oxidative aging theories. Free Radical Biology and Medicine 43(4), 477-503.

http://dx.doi.org/10.1016/j.freeradbiomed.2007.03.0 34

Murphy MP. 2009. How mitochondria produce reactive oxygen species. Biochemical Journal **417(1)**, 1-13. http://dx.doi.org/10.1042/bj20081386

Nakabeppu Y. 2014. Cellular levels of 8-oxoguanine in either DNA or the nucleotide pool play pivotal roles in carcinogenesis and survival of cancer cells. International journal of molecular sciences **15(7)**, 12543-12557.

http://dx.doi.org/10.3390/ijms150712543

Nayak D, Karmen C, Frishman W, Vakili B. 2000. Antioxidant vitamins and enzymatic and synthetic oxygen-derived free radical scavengers in the prevention and treatment of cardiovascular disease. Heart disease (Hagerstown, Md.) **3(1)**, 28-45.

http://dx.doi.org/10.1097/00132580-200101000-00006

Nel A, Xia T, Mädler L, Li N. 2006. Toxic potential of materials at the nanolevel. Science **311(5761)**, 622-627.

Nishiyama Y, Allakhverdiev SI, Murata N. 2011. Protein synthesis is the primary target of reactive oxygen species in the photoinhibition of photosystem II. Physiologia Plantarum **142(1)**, 35-46.

http://dx.doi.org/10.1111/j.1399-3054.2011.01457.x

Oyinloye BE, Adenowo AF, Kappo AP. 2015. Reactive Oxygen Species, Apoptosis, Antimicrobial Peptides and Human Inflammatory Diseases. Pharmaceuticals **8(2)**, 151-175.

Pandey KB, Rizvi SI. 2009. Plant polyphenols as dietary antioxidants in human health and disease. Oxidative medicine and cellular longevity **2(5)**, 270-278.

http://dx.doi.org/10.4161/oxim.2.5.9498

Pappolla M, Omar R, Kim K, Robakis N. 1992. Immunohistochemical evidence of oxidative [corrected] stress in Alzheimer's disease. The American journal of pathology **140(3)**, **6**21.

Park JS, Svetkauskaite D, He Q, Kim J-Y, Strassheim D, Ishizaka A, Abraham E. 2004. Involvement of toll-like receptors 2 and 4 in cellular activation by high mobility group box 1 protein. Journal of Biological Chemistry **279(9)**, 7370-7377. http://dx.doi.org/10.1074/jbc.m306793200

Park S, Imlay JA. 2003. High levels of intracellular cysteine promote oxidative DNA damage by driving the Fenton reaction. Journal of bacteriology **185(6)**, 1942-1950.

http://dx.doi.org/10.1128/jb.185.6.1942-1950.2003

Piantadosi CA, Zhang J. 1996. Mitochondrial generation of reactive oxygen species after brain ischemia in the rat. Stroke **27(2)**, 327-332. http://dx.doi.org/10.1161/01.str.27.2.327

Plaetzer K, Krammer B, Berlanda J, Berr F, Kiesslich T. 2009. Photophysics and photochemistry of photodynamic therapy: fundamental aspects. Lasers in medical science 24(2), 259-268.

http://dx.doi.org/10.1007/s10103-008-0539-1

Poljšak B, Dahmane R. 2012. Free radicals and extrinsic skin aging. Dermatology research and practice 2012.

http://dx.doi.org/10.1155/2012/135206

Prokai L, Yan LJ, Vera-Serrano JL, Stevens SM, Forster MJ. 2007. Mass spectrometry-based survey of age-associated protein carbonylation in rat brain mitochondria. Journal of mass spectrometry **42(12)**, 1583-1589.

http://dx.doi.org/10.1002/jms.1345

Pu X, Kamendulis LM, Klaunig JE. 2006. Acrylonitrile-induced oxidative DNA damage in rat astrocytes. Environmental and molecular mutagenesis **47(8)**, 631-638. <u>http://dx.doi.org/10.1002/em.20249</u>

Que EL, Domaille DW, Chang CJ. 2008. Metals in neurobiology: probing their chemistry and biology with molecular imaging. Chemical Reviews **108(5)**, 1517-1549.

http://dx.doi.org/10.1002/chin.200833267

Rao R, Møller IM. 2011. Pattern of occurrence and occupancy of carbonylation sites in proteins. Proteomics **11(21)**, 4166-4173. http://dx.doi.org/10.1002/pmic.201100223

Riley P. 1994. Free radicals in biology: oxidative stress and the effects of ionizing radiation. International journal of radiation biology **65(1)**, 27-33.

http://dx.doi.org/10.1080/09553009414550041

Rinnerthaler M, Büttner S, Laun P, Heeren G, Felder TK, Klinger H, Weinberger M, Stolze K, Grousl T, Hasek J. 2012. Yno1p/Aim14p, a NADPH-oxidase ortholog, controls extramitochondrial reactive oxygen species generation, apoptosis, and actin cable formation in yeast. Proceedings of the National Academy of Sciences 109(22), 8658-8663.

http://dx.doi.org/10.1073/pnas.1201629109

Risom L, Møller P, Loft S. 2005. Oxidative stressinduced DNA damage by particulate air pollution. Mutation Research/Fundamental and Molecular Mechanisms of Mutagenesis **592(1)**, 119-137. http://dx.doi.org/10.1016/j.mrfmmm.2005.06.012 **Roberts CK, Sindhu KK.** 2009. Oxidative stress and metabolic syndrome. Life sciences **84(21)**, 705-712.

http://dx.doi.org/10.1016/j.lfs.2009.02.026

Sanders LM, Henderson CE, Hong MY, Barhoumi R, Burghardt RC, Carroll RJ, Turner ND, Chapkin RS, Lupton JR. 2004. Prooxidant environment of the colon compared to the small intestine may contribute to greater cancer susceptibility. Cancer letters **208(2)**, 155-161. http://dx.doi.org/10.1016/j.canlet.2003.12.007

Sauvaigo S, Caillat S, Odin F, Nkengne A, Bertin C, Oddos T. 2010. Effect of aging on DNA excision/synthesis repair capacities of human skin fibroblasts. Journal of Investigative Dermatology 130(6), 1739-1741.

http://dx.doi.org/10.1038/jid.2010.10

Savini I, Catani MV, Evangelista D, Gasperi V, Avigliano L. 2013. Obesity-associated oxidative stress: strategies finalized to improve redox state. International journal of molecular sciences 14(5), 10497-10538.

http://dx.doi.org/10.3390/ijms140510497

Schins RP. 2002. Mechanisms of genotoxicity of particles and fibers. Inhalation toxicology **14(1)**, 57-78.

http://dx.doi.org/10.1080/089583701753338631

Selley M, Close D, Stern S. 2002. The effect of increased concentrations of homocysteine on the concentration of (E)-4-hydroxy-2-nonenal in the plasma and cerebrospinal fluid of patients with Alzheimer's disease. Neurobiology of aging **23(3)**, 383-388.

http://dx.doi.org/10.1016/s0197-4580(01)00327-x

Shafaq N. 2012. An overview of oxidative stress and antioxidant defensive system.

http://dx.doi.org/10.4172/scientificreports.413

Shaheen M, Echeverry D, Oblad MG, Montoya MI, Teklehaimanot S, Akhtar AJ. 2007.

Hepatitis C, metabolic syndrome, and inflammatory markers: results from the Third National Health and Nutrition Examination Survey [NHANES III]. Diabetes research and clinical practice **75(3)**, 320-326.

http://dx.doi.org/10.1016/j.diabres.2006.07.008

Shakeel M, Jabeen F, Shabbir S, Asghar MS, Khan MS, Chaudhry AS. 2015. Toxicity of Nano-Titanium Dioxide (TiO2-NP) Through Various Routes of Exposure: a Review. Biological trace element research, 1-36.

http://dx.doi.org/10.1007/s12011-015-0550-x

Shen H, Anastasio C. 2012. A comparison of hydroxyl radical and hydrogen peroxide generation in ambient particle extracts and laboratory metal solutions. Atmospheric environment **46**, 665-668. http://dx.doi.org/10.1016/j.atmosenv.2011.10.006

Sies H. 1997. Oxidative stress: oxidants and antioxidants. Experimental physiology **82(2)**, 291-295.

Singh R, Devi S, Gollen R. 2015. Role of free radical in atherosclerosis, diabetes and dyslipidaemia: larger-than-life. Diabetes/metabolism research and reviews **31(2)**, 113-126.

http://dx.doi.org/10.1002/dmrr.2558

Sosa V, Moliné T, Somoza R, Paciucci R, Kondoh H, LLeonart ME. 2013. Oxidative stress and cancer: an overview. Ageing research reviews 12(1), 376-390.

Stadtman E. 1995. The status of oxidatively modified proteins as a marker of aging. LIFE SCIENCES RESEARCH REPORTS, 129-129. http://dx.doi.org/10.1016/s0891-5849(02)00764-5

Steffes MW, Gross MD, Lee DH, Schreiner PJ, Jacobs DR. 2006. Adiponectin, visceral fat, oxidative stress, and early macrovascular disease: the Coronary Artery Risk Development in Young Adults Study. Obesity **14(2)**, 319-326.

http://dx.doi.org/10.1038/oby.2006.41

Steinberg D. 1997. Low density lipoprotein oxidation and its pathobiological significance. Journal of Biological Chemistry **272(34)**, 20963-20966. http://dx.doi.org/10.1074/jbc.272.34.20963

Stuehr D, Pou S, Rosen GM. 2001. Oxygen reduction by nitric-oxide synthases. Journal of Biological Chemistry **276(18)**, 14533-14536. http://dx.doi.org/10.1074/jbc.r100011200

Tamai M, Furuta H, Kawashima H, Doi A, Hamanishi T, Shimomura H, Sakagashira S, Nishi M, Sasaki H, Sanke T. 2006. Extracellular superoxide dismutase gene polymorphism is associated with insulin resistance and the susceptibility to type 2 diabetes. Diabetes research and clinical practice **71(2)**, 140-145.

http://dx.doi.org/10.1016/j.diabres.2005.05.006

te Velde AA, Pronk I, de Kort F, Stokkers PC. 2008. Glutathione peroxidase 2 and aquaporin 8 as new markers for colonic inflammation in experimental colitis and inflammatory bowel diseases: an important role for H2O2? European journal of gastroenterology & hepatology **20(6)**, 555-560.

http://dx.doi.org/10.1097/meg.0b013e3282f45751

Tribble DL, Committee N. 1999. Antioxidant Consumption and Risk of Coronary Heart Disease: Emphasis on Vitamin C, Vitamin E, and β -Carotene A Statement for Healthcare Professionals From the American Heart Association. Circulation **99(4)**, 591-595.

http://dx.doi.org/10.1161/01.cir.99.4.591

Tulard A, Hoffschir F, de Boisferon FH, Luccioni C, Bravard A. 2003. Persistent oxidative stress after ionizing radiation is involved in inherited radiosensitivity. Free Radical Biology and Medicine **35(1)**, 68-77.

http://dx.doi.org/10.1016/s0891-5849(03)00243-0

Valko M, Rhodes C, Moncol J, Izakovic M, Mazur M. 2006. Free radicals, metals and antioxidants in oxidative stress-induced cancer. Chemico-biological interactions **160(1)**, 1-40. http://dx.doi.org/10.1016/j.cbi.2005.12.009

Vincent HK, Bourguignon CM, Taylor AG. 2010. Relationship of the dietary phytochemical index to weight gain, oxidative stress and inflammation in overweight young adults. Journal of human nutrition and dietetics **23(1)**, 20-29.

http://dx.doi.org/10.1111/j.1365-277x.2009.00987.x

Wang M, Sun S, Neufeld CI, Perez-Ramirez B, Xu Q. 2014. Reactive Oxygen Species-Responsive Protein Modification and Its Intracellular Delivery for Targeted Cancer Therapy. Angewandte Chemie 126(49), 13662-13666.

http://dx.doi.org/10.1002/ange.201407234

Waris G, Ahsan H. 2006. Reactive oxygen species: role in the development of cancer and various chronic conditions. Journal of carcinogenesis **5(1)**, 14.

Wellen KE, Hotamisligil GS. 2005. Inflammation, stress, and diabetes. The Journal of clinical investigation **115(5)**, 1111-1119. http://dx.doi.org/10.1172/jci25102

Wheatley-Price P, Asomaning K, Reid A, Zhai R, Su L, Zhou W, Zhu A, Ryan DP, Christiani DC, Liu G. 2008. Myeloperoxidase and superoxide dismutase polymorphisms are associated with an increased risk of developing pancreatic adenocarcinoma. Cancer **112(5)**, 1037-1042. http://dx.doi.org/10.1002/cncr.23267

Wheeler MD, Nakagami M, Bradford BU, Uesugi T, Mason RP, Connor HD, Dikalova A, Kadiiska M, Thurman RG. 2001. Overexpression of manganese superoxide dismutase prevents alcoholinduced liver injury in the rat. Journal of Biological Chemistry **276(39)**, 36664-36672.

http://dx.doi.org/10.1074/jbc.m105352200

Whicher J, Evans SW. 2012. Biochemistry of inflammation 18, Springer Science & Business Media.

Wickens AP. 2001. Ageing and the free radical theory. Respiration physiology **128(3)**, 379-391. http://dx.doi.org/10.1016/s0034-5687(01)00313-9

Wilson MR, Lightbody JH, Donaldson K, Sales J, Stone V. 2002. Interactions between ultrafine particles and transition metals in vivo and in vitro. Toxicology and applied pharmacology **184(3)**, 172-179.

http://dx.doi.org/10.1006/taap.2002.9501

Xia T, Kovochich M, Brant J, Hotze M, Sempf J, Oberley T, Sioutas C, Yeh JI, Wiesner MR, Nel AE. 2006. Comparison of the abilities of ambient and manufactured nanoparticles to induce cellular toxicity according to an oxidative stress paradigm. Nano letters **6(8)**, 1794-1807.

http://dx.doi.org/10.1021/nl061025k

Ya Sklyarov A, Panasyuk N, Fomenko I. 2011. Role of nitric oxide-synthase and cyclooxygenase/lipooxygenase systems in development of experimental ulcerative colitis. Journal of Physiology and Pharmacology **62(1)**, 65. http://dx.doi.org/10.7124/bc.00008e

Yang H, Jin X, Lam K, Wai C, Yan S-K. 2011. Oxidative stress and diabetes mellitus. Clinical Chemistry and Laboratory Medicine **49(11)**, 1773-1782.

Young I, Woodside J. 2001. Antioxidants in health and disease. Journal of clinical pathology **54(3)**, 176-186.

Zappasodi F, Salustri C, Babiloni C, Cassetta E, Del Percio C, Ercolani M, Rossini PM, Squitti R. 2008. An observational study on the influence of the APOE-ɛ4 allele on the correlation between 'free'copper toxicosis and EEG activity in Alzheimer disease. Brain research 1215, 183-189. http://dx.doi.org/10.1016/j.brainres.2008.03.066

Zelko IN, Mariani TJ, Folz RJ. 2002. Superoxide dismutase multigene family: a comparison of the CuZn-SOD (SOD1), Mn-SOD (SOD2), and EC-SOD (SOD3) gene structures, evolution, and expression. Free Radical Biology and Medicine **33**(**3**), 337-349. http://dx.doi.org/10.1016/s0891-5849(02)00905-x

Zhang Q, Zou P, Zhan H, Zhang M, Zhang L, Ge R-S, Huang Y. 2011. Dihydrolipoamide dehydrogenase and cAMP are associated with cadmium-mediated Leydig cell damage. Toxicology letters **205(2)**, 183-189.

http://dx.doi.org/10.1016/j.toxlet.2011.06.003

Zhao Y, Zhao B. 2013. Oxidative stress and the pathogenesis of Alzheimer's disease. Oxidative medicine and cellular longevity 2013. http://dx.doi.org/10.1155/2013/316523 Zheng Y, Li X-K, Wang Y, Cai L. 2008. The role of zinc, copper and iron in the pathogenesis of diabetes and diabetic complications: therapeutic effects by chelators. Hemoglobin **32(1-2)**, 135-145. http://dx.doi.org/10.1080/03630260701727077

Zhou R, Yazdi AS, Menu P, Tschopp J. 2011. A role for mitochondria in NLRP3 inflammasome activation. Nature **469**(7**329**), 221-225. <u>http://dx.doi.org/10.1038/nature10156</u>

Ziboh VA, Miller CC, Cho Y. 2000. Metabolism of polyunsaturated fatty acids by skin epidermal enzymes: generation of antiinflammatory and antiproliferative metabolites. The American journal of clinical nutrition **71(1)**, 361s-366s.