

Journal of Biodiversity and Environmental Sciences (JBES) ISSN: 2220-6663 (Print) 2222-3045 (Online) Vol. 9, No. 1, p. 420-425, 2016 http://www.innspub.net

RESEARCH PAPER

OPEN ACCESS

Effects of water stress on seed yield and essential oil content of dill genotypes

Kazem Ghassemi-Golezani*1, Limoo Rezaeipour1, Saeideh Alizadeh-Salteh2

¹Department of Plant Eco-physiology, Faculty of Agriculture, University of Tabriz, Tabriz, Iran. ²Department of Horticulture, Faculty of Agriculture, University of Tabriz, Tabriz, Iran

Article published on July 31, 2016

Key words: Dill, Essential oil, Genotype, Seed yield, Water stress.

Abstract

Water stress may alter essence content of medicinal plants. Thus, a split plot experiment based on RCB design with three replications was conducted in 2015, to evaluate the effects of different irrigation treatments (I_1 , I_2 , I_3 , I_4 : irrigation after 70, 100, 130 and 160 mm evaporation, respectively) on seed yield and essential oil content of three dill (*Anethum graveolens* L.) genotypes (Isfahan, Malayer, Varamin). Irrigation treatments and genotypes were allocated to the main and sub-plots, respectively. The essential oil of seeds was extracted by hydro distillation. Means of seeds per plant and seed weight were decreased due to water deficit, which led to significant reduction in seed yield per unit area under severe water stress. Although essential oil percentage of dill seeds increased with decreasing water availability, the highest essential oil yield per unit area was obtained under mild (I_2) and moderate (I_3) water stress. However, severe water deficit significantly reduced essence yield as a result of a large reduction in seed yield per unit area. Malayer with the greatest number of seeds per plant and Varamin with the largest seeds were the high yielding cultivars, with no significant difference in seed yield per unit area. Consequently, seed essence yield of Varamin and Malayer genotypes was about 25% higher than that of Isfahan genotype.

*Corresponding Author: Kazem Ghassemi-Golezani 🖂 golezani@gmail.com

Introduction

Medicinal and aromatic plants are very importance because of the increasing demand for their products. Dill (*Anethum graveolens* L.) is an annual and sometimes biennial herb that is used as a vegetable, a carminative, an aromatic and an antispasmodic (Hornok, 1992; Sharma, 2004) and also as an inhibitor of sprouting in stored potatoes (Score *et al.*, 1997). Dill seed is usually sown at early spring for seed and essential oil production. The dill fruit is a schizocarp with paired carpels that split apart at maturity to release two pericarps, commonly referred to as "seed" (Bouwmeester, 1998).

Constituents of dill include essential oils, fatty acids, proteins, carbohydrates, fiber, ash and mineral elements such as calcium, potassium, magnesium, phosphorous, sodium, vitamin A and niacin (Kaur and Arora, 2010). The dill vegetative tissues, flowers and seeds contain essential oil, most of which accumulated in two latter organs (Ghassemi-Golezani *et al.*, 2008, 2011, 2015). The main constituents of dill essential oil are carvone and limonene (Bailer *et al.*, 2001; Singh *et al.*, 2005; Callan *et al.*, 2007).

In a large part of the agricultural areas in the world, water deficit is limiting crop productivity (Micheletto et al., 2007). Photosynthesis and cell growth are the primary processes which are affected by water stress (Munns et al., 2006). The flowering stage appear to be the most sensitive stage to drought stress (Nayyar et al., 2006). Water stress during flowering and seed filling caused 50-80% reductions in seed yield due to restrictions in photosynthesis (Leport et al., 1999). Coincidence of water stress with reproductive stages reduces duration of flowering and seed filling and consequently lowers the number of seeds per plant, mean seed weight and seed yield per unit area (Ghassemi-Golezani and Mazloomi-Oskooyi, 2008; Ghassemi-Golezani et al., 2010). The effect of water deficit on growth and yield depends on function of genotype, duration of stress, weather conditions, growth and developmental stages of crops (Robertson and Holland, 2004).

Secondary metabolites in the medicinal and aromatic plants are strongly influenced by genotypes and environmental conditions (Yazdani *et al.*, 2002). Environmental stresses bring about a wide range of responses in medicinal plants from the changes in growth and yield to the changes in secondary metabolites (Ghassemi-Golezani et al., 2008, 2011). This may be associated with the function of secondary metabolites as self-defense components against stress conditions. In other words, the stress conditions accelerate the biosynthesis of essential oils (El-Din et Among the different environmental al., 2009). constraints, drought is an important abiotic factor limiting plant productivity (Ghassemi-Golezani et al., 2014, 2015). Depending on the plant growth stages, drought stress influences morphology, anatomy, physiology and biochemistry of plants (Upadhyaya and Panda, 2004). The adaptability and responses to water stress depend on duration and magnitude of stress and developmental stage of plant (Kramer, 1983). The experiments with Cymbopogons have demonstrated that water stress alters the oil biogenetic capacity without any change in the oil gland count, as observed in the excised systems subjected to short-term stress conditions (Baher, 2002). It was found that essential oil content of dill organs increased with decreasing water availability during plant growth and development, but essence yield declined as a result of a large decrease in organs yields (Ghassemi-Golezani et al., 2015).

However, when water stress occurred during reproductive stages of dill, both essence percentage and yield enhanced due to stress and the highest essence yield per unit area was recorded under mild water deficit (Ghassemi-Golezani *et al.*, 2008).

Although the secondary metabolite production is believed to be stimulated by stressful environment, the extent of this effect is not unique. Therefore, this research was carried out to assess the effects of water limitation on seed yield and essential oil content of dill genotypes.

Materials and methods

Location and experimental design

This experiment was conducted in 2015 at the Research Farm of the Faculty of Agriculture, University of Tabriz, Tabriz, Iran (Latitude 38°05'N, Longitude 46°17'E, Altitude 1360 m above sea level) to evaluate the effects of different irrigation intervals on seed and essential oil production of dill. The climate is characterized by mean annual precipitation of 245.75 mm per year and mean annual temperature of 10°C. The experiment was arranged as split plot based on RCB design in four replications with irrigation treatments (I_1 , I_2 , I_3 and I_4 : irrigation after 70, 100, 130 and 160 mm evaporation from class A pan, respectively) in main plots and dill genotypes (Isfahan, Malayer, Varamin) in sub-plots. Each plot consisted of six rows of 4 m length with 25 cm apart.

Seed sowing and seedling establishment

Seeds were treated with Benomyl at a rate of 2 g kg⁻¹ and were sown by hand on 4 may, 2015, in 1.5 cm depth of a sandy loam soil. Sowing density was 80 seeds m⁻². All plots were irrigated immediately after sowing. After seedling establishment, plants in each plot were thinned to keep 56 plants m⁻² and irrigations were carried out according to the treatments. Weeds were controlled by hand during crop growth and development as required.

Seed yield and essence extraction

At maturity, plants from 1 m² of each plot were harvested and seeds per plant, 1000 seed weight and seed yield per unit area for each treatment at each replicate were determined. A sub-sample of 20 g seeds from each plot was mixed with 500 ml distilled water and essential oil was extracted by hydrodistillation for 3 hours, using a Clevenger apparatus at 250°C (Darzi *et al.*, 2012).

Analyses of variance

Analyses of variance of the data were carried out by SPSS software and means of each trait were compared by Duncan test at $p \le 0.05$. Excel software was used to draw Fig.s.

Results and discussion

Analysis of variance of the data (Table 1) showed that the effects of irrigation intervals on seeds per plant, 1000 seed weight and seed yield per unit area, essential oil percentage and essential oil yield were significant. Genotypic differences also had significant effects on 1000 seed weight and seed yield, but not on seeds per plant and essential oil percentage and yield. The interaction of irrigation \times genotype was only significant for 1000 seed weight (Table 1).

Table 1. Analyses of variance of the effects of water supply on yield components, seed yield, essential oil content and yield of dill genotypes.

Treatment	df	Seeds per plant	1000 seed weight	Seed yield	Essential Oil	Essential oil yield
Replicate	2	57969.9 ^{ns}	0.001 ^{ns}	69.68 ^{ns}	0.031 ^{ns}	0.022 ^{ns}
Irrigation (I)	3	1585465.2*	0.532^{**}	13770.79**	0.974**	2.376^{*}
Ea	6	190000.0	0.014	987.88	0.035	0.321
Genotype (G)	2	101608.6 ^{ns}	0.145**	2635.57^{*}	0.024 ^{ns}	0.518 ^{ns}
I×G	6	39805.7 ^{ns}	0.041*	196.33 ^{ns}	0.009 ^{ns}	0.078 ^{ns}
Eb	16	123325.3	0.025	493.83	0.063	0.264
CV	-	22.12	12.24	17.934	15.475	28.371

ns, *, **: No significant and significant at $P \le 0.05$ and $P \le 0.01$, respectively.

Mean number of seeds per plant and 1000 seed weight were decreased as a result of water limitation, which led to significant reduction in seed yield per unit area under severe water stress (Table 2). Reduction in seed number was due to a decrease in flower formation and an increase in flower (Fang *et al.*, 2009; Ghassemi-Golezani *et al.*, 2012). Irrigation disruption during seed filling can decrease filling duration and photosynthate mobilization to seeds, thereby decreasing seed weight (Ghassemi-Golezani *et al.*, 2009). In general, water stress during vegetative growth has the greatest impact on plant biomass, while during reproductive development it has the most limiting effect on seed yield (Ghassemi-Golezani *et al.*, 2008).

Malayer genotype produced about 1% and 12% more seeds per plant, compared with Varamin and Isfahan. However, these differences in seeds per plant were not statistically significant. In contrast, Varamin had the largest seeds, followed by Malayer and Isfahan (Table 2). The superiority of Varamin in seed weight was more evident under I_1 and I_2 and considerably reduced under I_3 and I_4 (Fig. 1). The lowest seed yield per unit area was recorded for Isfahan genotype, due to the production of the least number of seeds per plant with the smallest sizes. Malayer with the greatest number of seeds per plant and Varamin with the largest seeds produced the highest seed yield per unit area, with no significant difference between them (Table 2). Variation in yield components and seed yield among dill genotypes were directly related with their genetic constitution, which classified Varamin and Malayer as high yielding genotypes.

Table 2. Means of the yield components, seed yield and essential oil content and yield of dill for different irrigation treatments and genotypes.

Treatments	Seeds per plant	1000 seed weight (g)	Seed Yield g/m²	Essential oil (%)	Essential oil yield g/m²
Irrigations					
I1	1591 a	1.622 a	144.20 a	1.1778 c	1.6889 b
I2	1954 a	1.270 b	138.77 a	1.6056 b	2.2309 a
I3	1806 a	1.237 b	125.29 a	1.7511 ab	2.1909 ab
I4	999.2 b	1.038 c	59.49 b	1.9544 a	1.1353 c
Genotypes					
G1	1609 a	1.397 a	127.18 a	1.5708 a	1.8942 a
G2	1487 a	1.177 c	99.94 b	1.6458 a	1.5752 a
G3	1667 a	1.301 b	124.96 a	1.66 a	1.9651 a

Different letters in each column indicate significant difference at $P \le 0.05$. I_1 , I_2 , I_3 , I_4 : irrigation after 70, 100, 130 and 160 mm evaporation, respectively. G_1 , G_2 , G_3 : Varamin, Isfahan and Malayer genotypes, respectively.

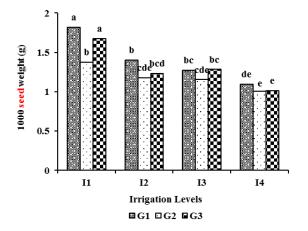


Fig. 1. Mean 1000 seed weight of dill genotypes under different irrigation treatments

Different letters indicate significant difference at $P \le 0.05$. I1, I2, I3, I4: irrigation after 70, 100, 130 and 160 mm evaporation, respectively.

Essential oil percentage increased with decreasing water availability. However, the differences in essential oil percentage between I_2 and I_3 and also between I_3 and I_4 were not significant (Table 2). In contrast, seed essential oil yield were considerably improved when plants were subjected to mild (I_2) and moderate (I_3) water deficit, but

it was significantly diminished under severe water stress (I_4) (Table 2). Seed essential oil yield of all dill seeds genotypes were statistically similar (Table 1).

Holtzer et al. (1998) have pointed out that water stress can increase, decrease or have no effect on the levels of metabolites, depending upon the plant species. Ghassemi-Golezani et al. (2008, 2015) reported that the essence percentage of dill significantly improved, when plants were subjected to water stress during reproductive stages. Drought stress increases the essential oil content of medicinal and aromatic plants, in order to prevent oxidization within the plant cells (Aliabadi et al., 2009). The results of this research showed that although essential oil percentage of dill seeds increased with increasing irrigation intervals (Table 2), the highest essential oil yield per unit area was obtained under mild (I2) and moderate (I₃) water stress (Table 2). However, severe water deficit significantly reduced essence yield as a result of a large reduction in seed yield per unit area (Table 2).

Although seed essence percentage and yield among dill genotypes was not statistically significant (Table 1), essence yield of Varamin and Malayer genotypes was about 25 % higher than that of Isfahan genotype (Table 2). No significant interaction between irrigation and genotype (Table 1) suggest that the superiority of two previous genotypes to the latter genotype in seed and essence yields were occurred under all irrigation intervals.

Conclusion

The highest essential oil yield per unit area was obtained under mild and moderate water stress, due to less reduction in seeds per plant, seed weight and seed yield and greater oil percentage in comparison with well watering. Seed and essence yields of Varamin and Malayer genotypes were more than those of Isfahan genotype. This superiority was consistent under all irrigation intervals.

References

Aliabadi FH, Valadabadi SAR, Daneshian J, Khalvati MA. 2009. Evaluation changing of essential oil of balm (*Melissa officinalis* L.) under water deficit stress conditions. Journal of Medicinal Plants Research **3**, 329-333.

Baher FZ, Mirza M, Ghorbanli M, Rezaii MB. 2002. The influence of water stress on plant height, herbal and essential oil yield and composition in *Satureja hortensis* L. Flavour and Fragrance Journal 17, 275-277.

Bailer J, Aichinger T, Hackl G, Hueber D, Dachler K. 2001. Essential oil content and composition in commercially dill cultivars in comparison to caraway. Industrial Crops and Products **14**, 229-239.

Bouwmeester HJ, Gershnzon J, Konings M, Croteau R. 1998. Biosynthesis of monoterpene and carvone in the fruit of caraway. I: Demonstration of enzyme activities and their changes with development. Plant Physiology **117**, 901-912.

Callan NW, Duane LJ, Westcott MP, Wrlty LE. 2007. Herb and oil composition of dill (*Anethum graveolens* L.): Effects of crop maturity and plant density. Journal of Industrial Crops and Products **25**, 282-287. **Darzi M, Haj Seyed Hadi MR.** 2012. Effects of the application of organic manure and biofertilizer on the fruit yield and yield components in Dill (*Anethum graveolens*). Journal of Medicinal Plants Research **6**, 3266-3271.

El-Din AAE, Aziz EE, Hendawy SF, Omer EA. 2009. Response of *Thymus vulgaris* L. to salt stress and alar (B9) in newly reclaimed soil. Journal of Applied Sciences Research **5**, 2165-2170.

Fang X, Turner NC, Yan G, Li F, Siddique KHM. 2009. Flower numbers, pod production, pollen viability and pistil function are reduced and flower and pod abortion increased in chickpea (*Cicer arietinum* L.) under terminal drought. Journal of Experimental Botany **61(2)**, 335-345.

Ghassemi-Golezani K, Andalibi B, Zehtab-Salmasi S, Saba J. 2008. Effect of water stress duringvegetative and reproductive stages on seed yield and essential oil content of dill (*Anethum graveolens* L.). Journal of Food, Agriculture and Environment **6**, 282-84.

Ghassemi-Golezani K, Bakhshi J, Dalil B. 2015. Rate and duration of seed filling and yield of soybean affected by water and radiation deficits. Acta Agriculturae Slovenica **105**, 225-232.

Ghassemi-Golezani K, Chadordooz-Jeddi A, Zehtab-Salmasi S. 2014. Effects of seed size and aging on field performance of lentil (*Lens culinaris* Medik.) under different irrigation treatments. Acta Agriculturae Slovenica **103**, 158-166.

Ghassemi-Golezani K, Ghanehpoor S, Mohammadi-Nasab AD. 2009. Effects of water limitation on growth and grain filling of faba bean cultivars. Journal of Food Agriculture and Environment 7, 442-47.

Ghassemi-Golezani K, Mazloomi-Oskooyi R. 2008. Effect of water supply on seed quality development in common bean (*Phaseolus vulgaris*). International Journal of Plant Production **2(2)**, 117-24. Ghassemi-Golezani K, Moradi M, Zehtab-Salmasi S, Alizadeh-Salteh S, Ghassemi S. 2015. Changes in essential oil content of different organs of dill (*Anethum graveolens*) genotypes in response to water Deficit. Azarian Journal of Agriculture **2**, 142-146.

Ghassemi-Golezani K, Mustafavi SH, Shafagh-Kalvanagh J. 2012. Field performance of chickpea cultivars in response to irrigation disruption at reproductive stages. Research on Crops **13**, 107-112.

Ghassemi-Golezani K, Zafarani-Moattar P, Raey Y, Mohammadi A. 2010. Response of pinto bean cultivars to water deficit at reproductive stages. Journal of Food Agriculture and Environment **8(2)**, 801-804.

Ghassemi-Golezani K, Zehtab-Salmasi S, Dastborhan S. 2011. Changes in essential oil content of dill (*Anethum graveolens*) organs under salinity stress. Journal of Medicinal Plants Research **5**, 3142-3145.

Holtzer TO, Archer TL, Norman JM. 1988. Host plant suitability in relation to water stress. In Heinrichs, E.A. (Ed.). Plant Stress Interactions. Willey-Interscience, pp. 111-137.

Hornok L. 1992. Cultivation and Processing of Medicinal Plants. Academic Publication, Budapest 338 p.

Kaur GJ, Arora DS. 2010. Bioactive potential of *Anethum graveolens, Foeniculum vulgare* and *Trachyspermum ammi* belonging to the family *Umbelliferae*-Current status. Journal of Medicinal Plants Research **4**, 087-094.

Kramer PJ. 1983. Water relations of plants. New York: Academic Press.

Leport L, Turner NC, French RJ, Barr MD, Duda R, Davies SL, Tennant D, Siddique KHM. 1999. Physiological responses of chickpea genotypes to terminal drought in a Mediterranean-type environment. European Journal of Agronomy **11(3)**, 279-291. Micheletto S, Rodriguez-Uribe L, Hernandez R, Richins RD, Curry V, Connell MA. 2007. Comparative transcript profiling in roots of (*Phaseolus acutifolius*) and (*Phaseolus* vulgaris) under water deficit stress. Plant Science **173(5)**, 510-520.

Munns R, James RA, Läuchli A. 2006. Approaches to increasing the salt tolerance of wheat and other cereals. Journal of Experimental Botany 57(5), 1025-1043.

Nayyar H, Singh S, Kaur S, Kumar S, Upadhyaya HD. 2006. Differential sensitivity of macrocarpa and microcarpa types of chickpea (*Cicer arietinum* L.) to water stress: association of contrasting stress response with oxidative injury. Journal of Integrative Plant Biology **48(11)**, 1318-1329.

Robertson MJ, Holland JF. 2004. Production risk of canola in the semi-arid subtropics of Australia. Crop and Pasture Science **55(5)**, 525-538.

Score C, Lorenzi R, Ranall P. 1997. The effect of (S)- (+)- carvone treatments on seed potato tuber dormancy and sprouting. Potato Research **40**, 155-161.

Sharma R. 2004. Agro-techniques of Medicinal Plants. Daya Publishing House, New Delhi, pp. 3-10.

Singh G, Maurya S, De Lampasona MP, Catalan C. 2005. Chemical Constituents, Antimicrobial Investigations, and Antioxidative Potentials of *Anethum graveolens* L. Essential Oil and Acetone Extract: Part 52. Journal of Food Science **70**, 208-215.

Upadhyaya H, Panda SK. 2004. Responses of Camellia sinensis to drought and rehydration. Biologia Plantarum **48**, 597-600.

Yazdani D, Jamshidi H, Mojab F. 2002. Compare of essential oil yield and menthol existent in Peppermint (*Mentha piperita* L.) planted in different origin of iran. Journal of Medicinal Plants Institu Jahad daneshgahi **3**, 73-78.