

Journal of Biodiversity and Environmental Sciences (JBES) ISSN: 2220-6663 (Print) 2222-3045 (Online) Vol. 9, No. 4, p. 213-218, 2016 http://www.innspub.net

OPEN ACCESS

Haemorrhagic septicaemia in bovine of Balochistan, Pakistan

Zahoor Ahmed¹, Muhammad Kamran Taj^{*1}, Ferhat Abbas¹, Deedar Ahmed¹, Ghulam Mohammad¹, Imran Taj¹, Aziz Ullah¹, Sana Arif², Attiya Aslam², Mriam Bilal², Asia Ali³

¹Microbiology Department (CASVAB), University of Balochistan, Quetta, Balochistan, Pakistan ²Zoology Department Sardar Bahadur Khan Women's University, Quetta, Balochistan, Pakistan. ⁸Computer Science Department Sardar Bahadur Khan Women's University, Quetta, Balochistan Pakistan.

Article published on October 30, 2016

Key words: Pasteurella, Multocida, Haemorrhagic septicaemia, Balochistan, Bovine

Abstract

Total 200 blood samples were collected in which 76% were *Pasteurella multocida* positive and 24% were negative. While district wise results showed that Jaffer Abad, Nassir Abad and Jhal Magssi were the most effected regions of Balochistan. However age wise distributed in bovine was 22% in nine months, 35.5% in one year and 18.5% in two years. The incidence of Haemorrhagic septicaemia was more in buffalo (54%) as compared to cattle (22%). The pH and temperature is the most important factor which effects the growth of *Pasteurella multocida*. Our result revealed that *Pasteurella multocida* grows from 25°C to 45°C while pH results showed growth from 6 to 8 pH. The antibiotic result showed that *Pasteurella multocida* was sensitive to most of classes of antibiotic which were used in study whereas Metronidazole class showed multi drug resistance against *Pasteurella multocida*. In lab animal trial fallowing sign such as nasal discharge and laziness were observed in 24 hr while postmortem result revealed that trachea, lung and heart were hemorrhagic.

*Corresponding Author: Muhammad Kamran Taj 🖂 kamrancasvab@yahoo.com

Introduction

Pasteurella species are small, gram-negative bacilli that colonize mucous membranes of wild and domestic animals but are usually absent from the normal flora in humans (Ataei *et al.*, 2009). Most species can act as primary or opportunistic pathogens in their hosts and are responsible for significant losses to livestock (Blackall *et al.*, 2000).

Human infections are chiefly associated with some form of animal contact and occur predominantly following inflicted injuries (Unchitti *et al.*, 1992). This microorganism is pathogenic for a wide variety of mammals and birds (Ekundayo *et al.*, 2008). *Pasteurella multocida* cause Haemorrhagic septicaemia is cattle and buffalo (Kwon and king, 2003). Haemorrhagic septicaemia is one of the most common economically significant animal diseases occurring in both developed and developing countries (Jesse *et al.*, 2013).

This disease is particularly endemic in Asia and Africa but outbreaks have also occurred in Europe and North America (Rimler et al., 2000). Pakistan has 31.8 million cattle population and a population of 29.0 million buffalo heads (Munir et al., 2007). Haemorrhagic septicaemia is the most common and notable infectious disease of dairy animals in Pakistan, having mortality rate of about 70% and is causing several hundred million dollars annual losses to animal production (May et al., 2001). The highest prevalence of Haemorrhagic septicaemia HS was recorded in Khanewal district and highest disease importance was recorded in Faisalabad district of Punjab (Farooq et al., 2011). Pasteurella multocida is extensively involved in respiratory syndrome of commercial dairy farms all over the world particularly in countries having hot humid environment (Waheedullah et al., 2009).

P. multocida is extensively involved in respiratory syndrome of bovine all over the world. In Pakistan it is the most important economic disease of livestock that has caused huge economic losses in Pakistan. Haemorrhagic septicaemia is the most common and foremost infectious disease of dairy animals (cattle and buffalo) in Balochistan,

having high mortality rate and is causing several hundred million dollars annual losses to animal production. So therefore isolation of *Pasteurella multocida* from commercial dairy farm of Balochistan will significantly help scientists of Balochistan to control and develop good vaccine against Haemorrhagic septicaemia.

Materials and methods

Study area and samples collection

The samples were collected from 10 districts of Balochistan (Jaffer Abad, Nassir Abad, Jhal Magsi, Sohbatpur, Sibi, Kachhi, Lehri, Harnai, Barkhan, and Quetta. The research was conducted in center for advanced studies in vaccinology and biotechnology (CASVAB) university of Balochistan. Five milliliter blood was collected aseptically from the jugular vein of each diseased cattle and buffalo showing typical signs of Haemorrhagic septicaemia (clinically positive cases) and also from animals died due to Haemorrhagic septicaemia (Varte *et al.*, 2014).

Isolation and identification

The isolation of organism was done from blood on brain heart infusion blood agar (BHIBA) and tryptose yeast extract agar. The isolates were identified on the basis of cultural, morphological and biochemical characteristics (Cheesbrough *et al.*, 2006).

Antimicrobial susceptibility test

Mueller hinton agar using Disc Diffusion Bauer technique and Mc. Farland turbidity (0.5) was used for standardized antibiotic sensitivity test were performed on standard method following clinical and laboratory standards institute (CLSI, 2008) protocol. Isolates organism were considered as sensitive, and resistant to a particular antimicrobial agent on the basis of inhibitory zone (Sugun *et al.*, 2013).

Pathogenicity of isolated bacteria in mice

0.5 ml growth suspension of isolated having 1×10^9 CFU/mL was *subcutaneously* injected in to the mice. The dead mice postmortem was carried out to observe localized lesion on different part of body and observed disparate organs (Sanchez *et al.*, 2004).

Result

Total 200 blood samples were collected in which 76% were *Pasteurella multocida* positive and 24% were negative as shown in Fig. 1. The district wise result reveled that, 14% samples were positive from Jaffer Abad, 13.5% were from Nassir Abad, 13% were from Jhal Magsi, 13% were from Sohbatpur, 8.5% were from Sibi, 07% were from Kachhi, 04% were from Lehri, 1.5% were from Harnie, 01% were from Barkhan, and 0.5% were from Quetta, as shown in Fig. 2. The age wise distributed was 15% in nine months 19.5% in one year and 11% in two years in buffalo while 07% in nine months, 16%. In one year and 7.5% in two years in cattle as shown in Fig. 3.

Results about species wise incidence of Haemorrhagic septicaemia exposed that the incidence of Haemorrhagic septicaemia in buffalo was 54% and in cattle was 22%. The buffalo of all districts were more effect as shown in Fig. 4.



Fig. 1. Overall percentages of positive and negative samples of *Pasteurella multocida*.



Fig. 2. *Pasteurella multocida* positive samples from different districts of Balochistan.



Fig. 3. Positive samples age wise from different districts of Balochistan.



Fig. 4. *Pasteurella multocida* in cattle and buffalo from different districts of Balochistan.

Temperature and pH effect on the growth of Pasteurella multocida

Temperature and ph effect on the growth of *Pasteurella multocida*. The pH and temperature is one of the most important environmental factors that affect many aspects of the biological system of *Pasteurella multocida*. The *Pasteurella multocida* grows from 37°C to 45°C while pH result showed growth from 6 - 8 pH as shown in Table 1.

Table 1. Growth of *Pasteurella multocida* ondifferent temperature and pH.

Temperature °C						
0	-					
4	-					
10	-					
15	-					
20	+					
25	+					
30	+					
35	+					
37	+					
40	+					
45	+					
50	-					
	pH					
2	-					
3	-					
4	-					
5	-					
6	+					
7	+					
8	+					

Biochemical tests

Pasteurella multocida was identified by gram staining and through different biochemical tests

(IMIVIC, sugar fermentation tests, catalase test, oxidase test, gelatin liquefaction test, and H_2S production tests) as shown in Table 2.

						Paste	urella	multoc	cida							
Gram St	aining	Gram N	Vegativ	e Bact	teria											
Shaj	pe	Coccoba	acilli													
						Bic	ochem	ical Tes	st							
Hemolysis	Growth On MacConkey's	Simmon	Currate Gelatin Hvdrolvsis	Methyl Red	Voges- Proskauer	Motility Test	Urease Test	Lysine Decarboxylas	Lysine	Decarboxylas Growth On Potassium	Hydrogen Sulfide	Nitrate Reduction	Indole Production	Ornithine Decarboxylas	Catalase Test	Oxidase Test
-	-	-	-	-	-	-	-	-	-	+	+	+	+	+	+	+
Sugar Fermentation Tests																
Glucose	Fructose	Galactose	Mannose	Sucrose	Maltose	Trehalose	Arabinose	Xylose	Mannitol	Sorbitol	Inositol	Dulcitol	Salicin	Lactose		
+	+	+	+	+	+	+	+	+	+	+	-	-	-	-		

Table 2. Biochemical tests and sugar fermentation test for confirmation of pathogens.

Antimicrobial test

Twenty-three antibiotics has been used against *Pasteurella multocida* and result showed that *Pasteurella multocida* was sensitive to

classes Penicillin, Tetracycline, Quinolones, Cephalosporin, and Aminoglycosides, while only resistance to class Metronidazole as shown in the Table 3.

Table 3. Antibiotic resistance and sensitivity test against Pasteurella multocida.

			Pasteurella multocida				
	Class	Antibiotics	Zone of inhibition (mm)				
I.	Penicillin	Penicillin-G Amoxycillin	(11mm) (17mm)				
		Ampicillin Cloxacillin	(16mm) (08mm)				
I.	Tetracycline	Oxytetracycline	(22mm)				
I.	Quinolones	Ciprofloxacin Oflaxcin Rifampicin Lincomycin Bacitracin Eprofloyacin	(18mm) (16mm) (16mm) (08mm) (07mm)				
7.	Cephalosporin	Cepharadin Cefepime Cephalexin	(201111) 14mm) (12mm) (14mm)				
7.	Aminoglycosides	Gentamycin Amikacin	(15mm) (16mm) (07mm)				
I.	Maerolides	Erythromycin	(09mm)				
I.	Glycopeptides	Vancomycin	(12mm)				
I.	Polypeptide	Colistin Sulphate	(09mm)				
ζ.	Flagyl	Metronidazole	Resistant				
ζ.	Miscellaneous	Chloramphenicol Sulphamethoxazole Trimethoprim	(19mm) (12mm) (12mm)				

216 | Ahmed et al.

Pathogenic effect in mice

The *Pasteurella multocida* was inoculated in to mice, with direct intra peritoneal injection of the purified bacterial species. After inoculation of pathogens in to mice was observed to checked different clinical symptoms of mice. In 24 hours nasal discharge, laziness, and reluctant to movement was noted in inoculated mice. After 24 hours the inoculated animal was noted as dead in cage. The postmortem of mice showed hemorrhagic lesion on trachea, heart, lungs and kidneys as shown in Fig. 5.



Fig. 5. Pathogenicity of Pasteurellamultocida in mice.

Discussion

Total 200 samples were collected in which 76% were positive and 24% were negative for *Pasteurella multocida*. Districts wise results showed that Jaffer Abad, Nassir Abad and Jhal Magssi, were the most effected regions of balochistan while Harnai, Barkhan and Quetta, were less effect region of balochistan for haemorrhagic septicaemia. The incidence of Haemorrhagic septicaemia in buffalo was 54% and in cattle was 22%. Buffalo was the most effect specie then cattle in all ten districts of the province. The bovine of 1 year (35.5%) was more affected as compared with other age groups.

The effect of different range temperature and pH on *Pasteurella multocida* has been characterized. The result showed that the *Pasteurella multocida* yield was highly depended on temperature low temperature and high temperature didn't allow *Pasteurella multocida* to grow on media. While the tremendous growth of *Pasteurella multocida* was recorded at 35°C to 37°C where as poor growth was observed at 20°C to 25°C.

pH of media was also found affecting *Pasteurella multocida* survival. Our result demonstrate that apart from ideal values of pH been demonstrated the growth of *Pasteurella multocida* in maximal at various other values of pH. Our result showed that the best pH for the growth of *Pasteurella multocida* was from 6 to 8 pH. The antibiogram results revealed that the best choice of drugs for the control of *Pasteurella multocida* was Tetracycline. In animal trial nasal discharge and hemorrhagic lesion were observed at heart, liver and kidney.

Conclusion

Haemorrhagic septicaemia is an important disease in the tropical area of balochistan Pakistan. This disease is most devastating to smallholder farmers where husbandry and preventive practices are poor and free-range management is common. So proper management & vaccine will provides protection to the bovine of the region.

217 | Ahmed et al.

Acknowledgements

The author is highly grateful to the Director Research LDD Dr. Baig Mohammad Kakar for facilitating the visit and collection of samples from ten districts of the province of Balochistan. Author also wishes to thank the staff of anaerobic laboratory CASVAB Mr. Aurangzaib and Mr. Abdullah who helped to do this research.

Refrence

Ataei S, Burchmore R, Christopher HJ, Finucane A, Parton R, Coote JG. 2009. Identification of immunogenic proteins associated with protection against the hemorrhagic septicemia after vaccination of calves with a live-attenuated aro A derivative of *Pasteurella multocida* B: 2. Res. Vet. Sci **87(02)**, 207-10.

Blackall PJ, Fegan N, Pahoff JL, Storie GJ, Mcintosh GB, Cameron RDA, Boyle DO, Frost AJ, Bara MR, Marr G, Holder J. 2000. The molecular epidemiology of four outbreaks of porcine pasteurellosis. Vet. Microbial **72**, 111-120.

Cheesbrough M. 2006. District laboratory practice in tropical countries. 2nd Edition London, English Language Book Society pp. 100-194.

Ekundayo SO, Odugbo MO, Olabode AO, Okewole PA. 2008. Phenotypic variability among strains of *Pasteurella multocida* isolated from avian, bovine, caprine, leorine and ovine origin. Afr. J. biotech **7**, 1347-1350.

Farooq U, Hussain M, Irshad H, Badar N, Munir R, and Ali R. 2011. Status of hemorrhagic septicemia based on epidemiology in Pakistan. Pakistan Vet **27(2)**, 67-72.

Jesse FFA, Abdinasir YO, Adamu L, Zakaria Z, Abdullah R. 2013. Acute phase protein profile in calves following infection with whole cell, lipopolysa ccharide and outer protein extracted from *Pasteurella multocida* type B: 2. Asian. J. Anim Vet. Adv. **8**, 655-662. Kwon YK Kang MI. 2003. Outbreak of fowl cholera in Baikal teals in Korea. Avian Dis J **47(4)**, 1491-1495.

May BJ, Zhang QT, Kapur V. 2001. Proceedings of the National Academy of Sciences of the United States of America **98(6)**, 3460-5.

Munir R, Shahwar D, Farooq U, Nawaz I, Shahzad I, and Khanum A. 2007. Outer membrane protein profiling of *P. multocida*. Pak Vet J **27(1)**, 1-4.

Rimler RB. 2000. Restriction endonuclease analysis using Hha I and Hpa II to discriminate among group B *Pasteurella multocida* associated with hemorrhagic septicemia. J. Med. Microbial **49**, 81-87.

Sanchez S, Mizan S, Quist C, Schroder P, Juneau M, Dawe D, Ritchie B, and Lee MD. 2004. Serological response to *Pasteurella multocida* Nan H Sialidase in persistently colonized rabbits. Clin Diagn Lab Immunol J **11(5)**, 825-834.

Sugun MY, Musa JA, Moses O, Dugbo O, Muhammad M, Abiyayi E, and Suleiman I. 2013. Isolation and in vitro antibiotic susceptibility of *Pasteurella multocida* from cattle origin. Internal Rese J **4(5)**, 131-134.

Unchitti K, Wongsawangl S, Saitanul K, Thoongsuwan S. 1992. Characteristics of *Pasteurella multocida* isolated from humans, swine and poultry in Thailand. Southeast Asean J. **23(3)**, 520-525.

Varte Z, Dutta TK, Roychoudhury P, Begum J, and Chandra R. 2014. Isolation, identification, characterization and antibiogram of *Pasteurella multocida* isolated from pigs in Mizoram with special reference to progressive atrophic rhinitis. Vet World J. 7, 95-99.

Waheedullah A, Arshad M, Jamal J, Ayub MS, Ali Q. 2009. Differentiation of closely related vaccinal strains of *P. multocida* using PCR. Vet. Scin. J. **4(1)**, 36-35.