Prevalence of urinary tract infection in both outpatient department and in patient department at a medical college setting of Bangladesh

Arunima Moue¹, Syed A.Q.M. Aktaruzzaman², Nasrin Ferdous³, Md. Rabiul Karim³ M.M.R. Khalil⁴, Ashish Kumar Das^{4*}

Anwer Khan Modern Medical College, Dhanmondi, Dhaka-1205, Bangladesh

²Bangladesh University of Professionals, Mirpur, Dhaka-1221, Bangladesh

^sDepartment of Biochemistry and Molecular Biology, Rajshahi University, Bangladesh ^sInternational Centre for Diarrheal Diseases Research, Bangladesh, Mohakahli, Dhaka-1212, Bangladesh

Key words: Urinary Tract Infection, Uropathogens, OPD, IPD, Antibiotic sensitivity.

http://dx.doi.org/10.12692/ijb/7.5.146-152

Article published on November 30, 2015

Abstract

Urinary tract infection remains one of the most common infections, both in the community and in the hospital. The causative pathogen profile varies from region to region, but Escherichia coli (E. coli) remain the most common causative pathogen. Organism responsible for the hospital acquired infection may have tendency to develop multiple drug resistance. This study was carried out to identify the causative organism for UTI among outpatient department (OPD) and inpatient department (IPD) patients of Anwer Khan Modern Medical College and Hospital, Dhaka, Bangladesh and also to see the antibiotic sensitivity pattern of the isolate according to age and sex. A total of 376 urine specimens received over the six months study period, 79.5 % (299) of the urine samples were culture positive. The female was more prone to UTI which was 79% (239) rather than male follows 21% (60). IPD patients showed 55.5% (166) positive culture compared to as OPD patients as 44.5% (133). The age variation according to sex was found for causing UTI. Therefore, 21-30 years aged female group showed 48.5% and 41-50 years aged male group had 46.7% UTI. The most common bacterial isolate was E. coli 46.8% (140) followed by Enterococcus faecalis 25.9% (77) Pseudomonas aeruginosa 11.4% (34), Staphylococcus saprophyticus 8% (24). E. coli was highly sensitive to Piperacillin (89.24%), Amikacin (85.24%) followed by Imipenem (80.27%). It was also observed that the samples responded effectively to Ampicillin, Amikacin, Gentamycin, Ciprofloxacin, Vancomycin and Linezolid. High degree of resistance was shown for Cefriaxone, Cefepime, Cefrioxne, Norfloxacin, and Cefoxitin on the basis of microbial species. The aim of this study was to raise awareness of UTIs and to expand services for prevention and treatment for UTI. To do this effectively, however, it may be necessary to improve the quality of health care provided at the community-level.

* Corresponding Author: Ashish Kumar Das 🖂 ashish.jajabor@gmail.com

Introduction

Urinary tract infection (UTI) is most common infectious presentation in hospital acquired and community acquired infections since long time (Peleget et al., 2010). There are an estimated 150 million urinary tract infections per annum worldwide and cost the global economy in excess of 6 Billion US dollars (Gonzalez et al., 1999). A limited and predictable spectrum of organisms is responsible urinary tract infections. Most of the UTI are caused by Gram negative bacteria like Escherichia coli, Klebsiella spp., Pseudomonas Proteus spp., aeruginosa, Acinetobacter, Serratia and Morganella margani. UTI also caused by Gram positive bacteria like Enterococcus, **Staphylococcus** especially coagulase negative staphylococci and Streptococcus agalactiae (Mohamed et al., 2012). UTI is much more common in women than in men due to anatomical and physiological reason; by virtue of its position urinogenital tract is more vulnerable to bacterial infections caused by both internal and external flora (Shanthi et al., 2012). Among both outpatients and inpatients, Escherichia coli are the primary urinary tract pathogen, accounting for 75% to 90% of both sides - hospital acquired. UTIs are often treated with different broad spectrum antibiotics when one with a narrow spectrum of activity may be appropriate because of concerns about infection with resistant organisms. Fluor-quinolones are preferred as initial agents for empiric therapy of UTI in area where resistance is likely to be of concern (Schaeffer, 2002). This is because they have high bacteriological and clinical cure rates, as well as low rates of resistance, among most common uropathogen (Goldstein, 2000). The extensive uses of antimicrobial agents have invariably resulted in the development of antibiotic resistance, which, in recent years, has become a major problem worldwide (Kumar et al., 2006). The Infectious Diseases Society of America also recommends that physicians obtain information on local resistance spectrum of organisms because urinary tract infections and that ongoing surveillance are conducted to monitor changes in susceptibility of uropathogens (Warren et al., 1999).

Increasing antimicrobial resistance complicates uncomplicated UTI treatment by increasing patient morbidity, costs of reassessment and retreatment and use of broader-spectrum antibiotics (Aypak et al., 2009). Patterns of antibiotic resistance in a wide variety of pathogenic organisms vary even over short periods of time. Periodic evaluation of antibacterial activity is needed to update this information (Gupta et al., 2002). For effective treatment and control of UTI in a particular area/hospital, a good knowledge of the antibiotic sensitivity pattern of the causative agents in that area/hospital is of ultimate importance (Uwaezuoke et al., 2006). Furthermore, baseline estimates of the magnitude of the problem and the extent of antimicrobial resistance among the nosocomial pathogens are the minimum essential prerequisites for any hospital infection control program (Kamat et al., 2009).

This study was carried out to determine the prevalent uropathogens according to age and sex at a medical college hospital among outpatient department (OPD) and inpatient department (IPD) and their antibiotic sensitivity pattern to commonly used antibiotics in order to provide a database for reference. We also compared the antibiotic sensitivity pattern of the bacterial isolates between outpatients and inpatients which had positivity of culture. In the present scenario, where the antibiotic resistance pattern is changing, our study aims at outlining the recommendations for empirical treatment of UTI. Moreover, the data would also help authorities to formulate antibiotic prescription policies.

Methods and materials

The present study is a retrospective study, which was carried out in the clinical Microbiology laboratory of Anwer Khan Modern Medical College and Hospital which is located at Dhaka City, Bangladesh. The duration of the study was six months period from January 2014 to June 2014. A total of 376 samples with or without signs to symptoms of UTI who attended the outpatient department (OPD) and inpatient department (IPD) of our hospital were recruited for this study. They consist of 188 OPD and 188 IPD patients. A significant positive culture was taken that was 299 out of 376 samples which belong both OPD and IPD. Unsterile specimen, time delayed specimen for culture (after 30 min of collection), which were not kept refrigerated at 4°C and inadequate sample for urine culture were excluded from the study.

Freshly voided, clean-catch midstream urine was collected from each patient into sterile screw-capped universal container. The specimen was labeled and transported to the microbiology laboratory for processing within 2 hours. Semi quantitative urine culture was done using a calibrated loop. A loopful (0.001 mL) of well mixed un-centrifuged urine was inoculated onto the surface of MacConkey and blood agar media. All plates were then inoculated at 37°C aerobically for 24 hours. The plates were then examined macroscopically for bacterial growth. A significant growth is considered if the number of colony is >105 colony forming unit (cfu)/ml. Colonial appearance and morphological characters of isolated bacteria was noted and isolated colonies were subjected to preliminary tests like Gram staining, motility by hanging drop, catalase test and oxidase test. These preliminary tests were followed by biochemical reactions for identification of the isolated organism. And the isolated organisms subjected for antibiotic susceptibility testing.

Antimicrobial Susceptibility Testing

Antimicrobial susceptibility testing was done by Kirby-Bauer disk diffusion method by using Mueller Hinton agar plates. Commercially available HiMedia discs were used. The bacterial suspension was made by inoculating 4-5 well isolated identical colonies in peptone water. After 2 hours of incubation the turbidity was standardized by using 0.5 McFarland standards. By using a sterile swab a lawn culture was made on the Mueller-Hinton agar plates. The antibiotic discs were placed and inoculated plates were incubated at 37°C. The results were read after overnight incubation and compared with standard chart. The following drugs were used for Antibiotic sensitivity test, [According to CLSI guidelines].

Amikacin (30µg), Gentamycin (10µg), Ciprofloxacin (5µg), Ceftriaxone (30µg), Norfloxacin (10µg), Ampicillin (10µg), Imipenem (10µg), Cefoxitin (30g). Cefepime (30µg), ceftazidime (30µg), Piperacillin (100µg). Erythromycin (15µg), Clindamycin (2µg), Oxacillin (1µg), Linezolid (15µg) and Vancomycin(30µg).

Statistical Analysis

The statistical analysis of data was performed using Graph Pad Prism-5 for Windows (Graph Pad Software, San Diego, CA, USA). The data was analyzed using One Way Analysis of variance (ANOVA) followed by Bonferroni testing. After analyzing the values were used an online software (www.socialscistatistics.com).

Results

A total of 376 urine samples were received, of which 188 (50%) were from OPD patients and rest of the 188(50%)) from IPD. Growth was present in 79.5% (299 out of 376) samples whereas 20.5% samples had no longer growth. The patients ranged from ages 1 year to > 50 years, with a mean age of 46.24 years. The prevalence in female was 79.0 % and the prevalence in male was 20.0%, which is shown in Table: 02. Male and female culture positivity was 22.6% (30 of 133) and 77.5% (103 of 133) in OPD and 18.0 % (30 of 166) and 81.9% (136 of 166) respectively in IPD samples. In OPD male patient positive culture predominantly found high from 16.7 % to 46.7 % in the age range in between 31 to > 50 years which has the relevancy with IPD male patient which range was 13.3 % to 46.7 % in the similar age group. On the other hand, for both in OPD and IPD female patient positive culture found high in age range from 21 to 40 years. Therefore it seems that among females UTI was commonly seen in the age group of 21-40 years and in males it was common between 41-60 years. The age wise distribution of samples and their positivity is shown in table 2. As a whole, the female patients in OPD expressed the highest culture positivity which was 48. 5 % but in the meantime both OPD and IPD male patient showed similar positive culture characteristics like 46.7 %. Fifteen patients presented a mixed infection with two organisms (10 in OPD and 5 in IPD). Thus, a total of 299 urine isolates were obtained, of which 166 (55.5%) were IPD sample isolates and 133(70.7%) were OPD sample isolates. *E. coli* was the most commonly isolated urinary

pathogen (48.1 %), followed by *Enterococcusfaecalis* (26.3%), *Pseudomonas aeuroginosa* (11.3%), *Staphylococcus saprophyticus* (7.5%) and *Klebsiella spp.* (3.8 %). The isolation rates of other organisms are shown in table 1.

Table 1. OPD and IPD organism causing UTI.

Organism	0	PD isolates cau	sing UTI	IPD isolates causing UTI				
	Number	of Percenge		Number	of Percentage	Mean±SD	P-Value	
	growth	(%)		growth	(%)			
Escherichia coli	64	48.1	19±3.5	76	45.8	23.7±5.9	0.047	
Enterococcus faecalis	35	26.3	11.5±4.6	42	25.3	15.0±3.4	0.878	
Pseudomonas aeruginosa	15	11.3	6.8±2.0	19	11.4	9.6±3.5	0.575	
Staphylococcus saprophyticus	10	7.5	4.7±1.8	14	8.4	7.2±2.7	0.653	
Klebsiella	5	3.8	3.1±1.4	8	4.8	5.0 ± 1.9	0.882	
Pneumoniae								
Proteous mirabilis	3	2.3	2.0±1.0	5	3.0	3.5±1.0	0.890	
Acinetobacter spp	1	0.8	NA	2	1.2	2.0±0.9	0.956	
Total	133	100		166	100			

Legend: P> 0.005 (not significant) and P<0.005 (highly significant), SD= Standard Deviation, NA= Not Applied, UTI= Urinary Tract Infection, OPD= Outpatient Department, IPD=In Patient Department.

Age in year	OPD					IPD					
	Male in number	Percentage (%)	Female in number	Percentage (%)	Meann±SD	Male in number	Percentage (%)	Female in number	Percentage (%)	Mean±SD	P Value
0-01	1	3.3	4	3.9	5.0±4.8	1	3.3	5	3.7	29.3±20.0	0.989
11-20	1	3.3	7	6.8	5.8±4.9	2	6.7	15	11.0	34.0±18.4	0.045
21-30	3	10	50	48.5	7.0±4.8	4	13.3	58	42.7	38.2±18.2	0.033
31-40	5	16.7	15	14.6	8.3±4.9	4	13.3	22	16.1	30.3±11.1	0.057
41-50	14	46.7	14	13.6	10.0±5.6	14	46.7	19	13.0	32.5±14.8	0.987
>50	6	20	13	12.6	6.0±3.8	5	16.7	17	12.5	22.0±12.0	0.667
Total	30	100	103	100		30	100	136	100		

Table 2. Age and sex distribution of patients with culture.

Legend: SD= Standard Deviation, OPD= Outpatient Department, IPD=In Patient Department.

Discussion

Effective management of patients suffering from bacterial UTIs commonly relays on the identification of type of organisms that caused the disease and selection of an effective antibiotic agent to the organism. Diagnosis of UTIs is a good example of the need for close cooperation between the clinician and the microbiologist. In our study prevalence rate of infection of urinary pathogen was 79.5 %, similar study by (Kattel, 2008) in which 26% of urine specimens showed significant bacterial growth. The prevalence of UTI is more in females when comparing males. This correlates with other study (Momoh, 2005) in which 60.2% were females and 39.8% were males. Women are more prone to UTIs than men because of short urethra and are closer to anus. Among patients with UTI, females were most commonly in the age group between 21-40 years and males were between 41-60 years. This was in consistent with a study of (Nerukar, 2012) in which 52.16% were in the age group 21-40 years, who concluded that most uncomplicated urinary tract infections occurs in women who are sexually active, with far fewer cases occurring in older women, those who are pregnant, and in men. In older men, the incidence of UTI may increase due to prostatic obstruction or subsequent instrumentation. This was in consistent with the study of (Banerjee, 2009). *E.coli* was the predominant bacteria found in our study similar result was found by (Durgesh, 2012) showed that prevalence of *E.coli* was 31.5%. The second isolated pathogen was *Enterococcus faecalis* and count were 26.3% and 25.3 % in OPD and IPD respectively, this correlates with other study of

(Shanthi, 2012) in which 28.1% of *Enterococcus faecalis* was isolated as urinary pathogen (Momoh, 2012). In our study *E.coli* was most resistant to Ampicillin, Penicillin and Gentamycin. It was most sensitive to and Amikacin (85.24%), similar finding were seen in a study by (Razak *et al.*, 2012), who concluded that the organisms showed resistance to older urinary antimicrobial agents such as Ampicillin which indicates that increased consumption of particular antibiotics can be the pathway to its resistance. *Enterobacter spp.* was mostly sensitive to Linezolid (94.49%) and Vancomycin (87.70%). It was resistant to Ceftriaxone and Cefepime as 100%.. The antibiotic sensitivity pattern is shown in the table 3.

Table 3. In vitro antibiotic sensitivity pattern of most frequent isolated.

Drugs	E.coli	Enterobacter	r S. prophyticus(%)	Klebsiella	Pseudomonas	Proteus
	(%)	spp. (%)		pneumoniae (%)	spp. (%)	spp. (%)
Ampicillin	13.90	16.50	20.60	6.42	67.21	75.01
Amikacin	85.24	65	67.57	37.01	53.10	63
Gentamycin	30.03	31	63.41	43.14	37.26	40.22
Ciprofloxacin	21.21	19.30	15.63	75	34.20	37.30
Ceftriaxone	20.31	-	-	27.75	-	25.01
Clindamycin	ND	62.41	63.41	ND	ND	ND
Norfloxacin	16.35	10.52	34.16	23.90	-	50.21
Cefepime	17.25	-	-	17.85	ND	-
Imipenem	80.27	ND	ND	96.42	30.02	45.33
Cefoxitin	ND	ND	17.07	-	-	-
Linezolid	-	94.49	100	-	-	12.23
Vancomycin	-	87.70	87.80	-	52.01	-
Penicillin	-	18.18	7.01	-	24.69	24.23
Piperacillin	89.24	ND	ND	ND	-	-

Legend: P> 0.005 (not significant) and P<0.005 (highly significant), SD= Standard Deviation, NA= Not Applied, ND: Not Detectable; (-): Antibiotic resistant pattern.

Staphylococcus saphrophyticus was sensitive to Linezolid (100%), Vancomycin (87.80%) and highly resistant to Ceftriaxone and Cefepime like *Enterbacter spp. Klebsiella pneumonia* was most sensitive to Imipenem (96.42%) and highly resistant to Cefoxitin, Linezolid, Vancomycin and Penicillin. *Psedomonas spp.* showed their best sensitivity to Ampicillin (67.21%) Amikacin (53.10%) and resistant to Ceftriaxone, Norfloxacin, Cefoxitin, Linezolid and Piperacillin wherasproteous spp. was resistant to Cefepime and Vancomycin. Ampicillin demonstrated the highest effectiveness about 75.01% to the *Proteous spp.* among all others isolates.

Conclusion

The high incidence rate of 79.5% reported in this study should be of great concern, as not only do UTIs pose a threat to health, but they also impose an

Int. J. Biosci.

economic and social burden due to the stigma associated with these infections. E.coli was the most frequent causative agent in UTI. Higher prevalence of UTI was seen in females (79%) rather than males (21%). Gram negative organisms were most commonly isolated organisms in UTI among which E.coli was the most frequent agent. Urinary pathogens showed resistant to commonly used antibiotics like Cefriaxone, Cefepime, Cefrioxneand Norfloxacin. On the basis of this study we can conclude that the resistance of commonly used antibiotics is very crucial. The antibiotic treatment should be limited to symptomatic UTIs and be initiated after sensitivity testing only. As drug resistance among pathogens in an evolving process, routine surveillance and monitoring studies should be conducted to help physician to start most effective empirical treatment should be carried out in order to prevent the cases becoming symptomatic later with resultant renal damage.

Acknowledgement

Authors are very much thankful to Prof. Dr. Md. Tahminur Rahman, (MBBS, MPhil, PhD), Vice Principal, Anwer Khan Modern Medical College, Dhaka, for providing necessary facilities and support.

References

Anton Peleg Y, David Hooper C. 2010. Hospital-Acquired Infections Due to Gram-Negative Bacteria . New England Journal of Medicine **362**, 1804-13.

Aypak C, Altunsoy A, Düzgün N. 2009. Empiric antibiotic therapy in acute uncomplicated urinary tract infections and fluoroquinolone resistance: A prospective observational study. Annals of Clinical Microbiology and Antimicrobials **8**, 27.

Biswas D, Gupta P, Prasad R, Singh V, Arya M, Kumar A. 2006. Choice of antibiotic for empirical therapy of acute cystitis in a setting of high antimicrobial resistance. Indian Journal of Medical Sciences **60(2)**, 53-8.

Durgesh D. 2012. Prevalence and antibacterial

susceptibility pattern of Urinary Tract Infection causing Human Pathogenic Bacteria. Asian Journal of Biomedical and Pharmaceutical Sciences **2(15)**, 1-3.

DrAlka Nerurkar. 2012. Bacterial pathogens in urinary tract infection and antibiotic susceptibility pattern. Journal of Pharmaceutical and Biomedical Sciences **21(12)**.

Gonzalez CM, Schaeffer AJ. 1999. Treatment of urinary tract infection: what's old, what's new, and what works. World Journal of Urology **6**, 372-382.

Gupta K, Hooten TM, Stamm WE. 2001. Increasing antimicrobial resistance and the management of uncomplicated community-acquired urinary tract infections. Annals of Internal Medicine. **135**, 41–50.

Goldstein FW. 2000. Antibiotic susceptibility of bacterial strains isolated from patients with community-acquired urinary tract infections in France. Multicentre Study Group. European Journal of Clinical Microbiology and Infectious Diseases **19**, 112-117.

Gupta V, Yadav A, Joshi RM. 2002. Antibiotic resistance patterns in uropathogens. Indian Journal of Medical Microbiology **20**, 96–8.

Hari Kattel P. 2008. Bcteriology of Urrinary Tract Infection among patients attending Tribuvan University Teaching Hospital Kathmandu, Nepal. Journal of Nepal association for Medical Laboratory Sciences **9(1)**, 25-29.

Kumar MS, Lakshmi V, Rajagopalan R. Related Articles. 2006. Occurrence of extended spectrum beta-lactamases among Enterobacteriaceae spp. isolated at a tertiary care institute. Indian Journal of Medical Microbiology **24(3)**, 208-11.

Kamat US, Fereirra A, Amonkar D, Motghare DD, Kulkarni MS. 2009. Epidemiology of hospital acquired urinary tract infections in a medical college

hospital in Goa. Indian Journal of Urology **(25)**, 76–80.

Momoh ARM. 2011. The antibiogram types of Escherichia coli isolated from suspected urinary tract infection samples. Journal of Microbiology and Biotechnology Research **1(3)**, 57-65.

Mohamed Shaaban T, Hassan Ghozlan A, Marwa Maghraby ME. 2012. Susceptibility of Bacteria Infecting Urinary Tract to Some Antibiotics and Essential Oils. Journal of Applied Pharmaceutical Sciences **02(04)**, 90-98.

Nicolle LE. 2001. Epidemiology of urinary tract infection.European Journal of Clinical Microbiology and Infectious Diseases **18**, 153–166.

Razak SK, Gurushanthappa V. 2012. Bacteriology of urinary tract infections and antibiotic susceptibility pattern in a tertiary care hospital in South India. International Journal of Medical Science and Public Health **1**, 109-112.

Banerjee S. 2009. The study of urinary tract infections and antibiogram of uropathogens in and around Ahmadnagar, Maharashtra. The Inrernet Journal of Infectious Diseases **9(1)**.

Schaeffer AJ. 2002. The expanding role of fluoroquinolones. American Journal of Medicine 113 (1A), 45S-54S.

Shanthi J. 2012. Incidence, distribution and antibiogram of uropathogens isolated from patients with urinary tract infections. Advanced Applied Science Research **3(6)**, 3410-3414.

Stamm WE, Norrby SR. 2001. Urinary tract infections: disease panorama and challenges. Journal of Infectious Diseases **183(1)**, S1-S4.

Tankhiwale SS, Jalgaonkar SV, Ahamad S, Hassani U. 2004. Evaluation of extended spectrum beta lactamase in urinary isolates. Indian Journal of Medical Research **120**, 553-556.

Uwaezuoke JC, Ogbulie JN. 2006. Antibiotic sensitivity pattern of urinary tract pathogens in port-Harcourt, Nigeria. Journal of *Applied* Sciences and Environmental Management **10**, 103–7.

Warren JW, Abrutyn E, Hebel JR, Johnson JR, Schaeffer AJ, Stamm WE. 1999. Guidelines for antimicrobial treatment of uncomplicated acute bacterial cystitis and acute pyelonephritis in women. Journal of Clinical Infectious Diseases **29**, 745–75.