



Ca⁺² and Mg⁺² of pistachio seedling in response to soil compaction under different water levels

Ghazaleh Azizi*, Adel Reyhanitabar, Davoud Zarehaghi, Nosratollah Najafi

Department of Soil Science, Faculty of Agriculture, University of Tabriz, Tabriz, Iran

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Abstract

In order to study the effects of four levels of soil compaction and six different moisture levels and to determine non limiting water range a nested experiment with three replications was conducted to evaluate changes in Ca⁺² and Mg⁺² concentration, uptake content and translocation factor of pistachio seedling (*Pistachio vera L.*). The air dried soil was passed through soil 4.75 mm sieve, and transferred into 36 PVC cylinders, the soils of cylinders were compacted in order to prepare four levels of soil bulk density (1.35, 1.5, 1.65 and 1.8 g cm⁻³). After transferring the pistachio seedlings into soil cylinders and their establishment, six different volumetric water contents, from saturation to permanent wilting point, for each compacted soils were applied. Ca⁺² and Mg⁺² concentration and content in shoot and root and translocation factor of these elements were less under high levels of soil compaction (1.65 and 1.8 g cm⁻³) than that of low levels of soil compaction (1.35 and 1.5 g cm⁻³). Concentration and translocation factor of Ca⁺² significantly enhanced with increasing water deficit, but the amount of Ca⁺² in shoot and root was reduced. Concentration of Mg⁺² in root and amount of Mg⁺² in shoot and root were declined with increasing water stress. Under all soil compaction levels, translocation factor of both elements were declined under water deficit conditions.

*Corresponding Author: Ghazaleh Azizi ✉ azizi.ghazaleh@yhao.com

Introduction

Soil compaction is the reduction of soil volume due to external factors and this reduction lowers soil productivity and environmental quality. The threat of soil compaction is greater today than in the past because of the dramatic increase in the size of farm equipment. Compaction is the process of densifying a soil mechanically and influencing thus physical properties of the soil. Physical properties influence all biological and many chemical processes in the soil (Kemper *et al.*, 1971).

Soil compaction influences agricultural sustainability through its effects on soil properties and crop development. Soil layers may become compacted naturally as a consequence of their textural composition, moisture regime, or the manner in which they were formed in place. Soil compaction is bringing an undesirable consequence of mechanization, which must be avoided (Hillel, 1980).

Compaction affects nutrient availability and uptake through a number of mechanisms. Aeration affects availability of nutrients involved in redox reactions, such as nitrogen, manganese and sulphur and the growth and function of roots (Lipiec and Stepniewski, 1995). The transport of nutrients in the soil is affected; compaction normally increases mass flow transport (Kemper *et al.*, 1971) and the diffusion coefficient at a given gravimetric water content (Bhadoria *et al.*, 1991). Compaction also increases root-to-soil contact, which may facilitate nutrient uptake (Veen *et al.*, 1992), but generally reduces root growth through its effect on aeration and mechanical resistance.

Soil compaction reduces root elongation (Taylor and Ratliff, 1969) and can also cause reductions in shoot growth (Schuurman, 1965). The shoot growth responses attributed to messages produced by compact soil include a reduction in mature cell sizes in leaves (Beemster and Masle, 1996) and a reduction in leaf number (Mulholland *et al.*, 1999). The clearest evidence of compact soil reducing shoot growth is

obtained when plants are grown in entirely compact soil.

Water stress is considered to be one of the most important environmental factors that limit plant production. Status of soil and water is one of the most important factors that affected on root growth and the genetic analysis of maize roots. Seedling survival in drought prone environments may depend upon the species' ability to compensate for the negative effect of low water potentials in the soil and atmosphere by adjusting root and shoot morphological and physiological patterns (Morgan, 1984).

The concept of an index of optimum soil water content for plant growth, as related to soil physical properties was introduced by Letey (1985) and identified as "non-limiting water range" (NLWR). Later, Silva *et al.* (1994) developed the NLWR concept quantitatively, renaming it as the least limiting water range (LLWR). For a given soil type, the LLWR incorporates the limitations of soil aeration, matric suction and soil penetration resistance

for root growth as a function of a single variable (i.e. soil bulk density). Crop response to soil compaction depends on the interaction among crop, soil type, water content, and compaction degree (Lipiec and Simota, 1994). In this context, soil moisture is the main factor and the Least Limiting Water Range (LLWR) relates soil moisture with aeration porosity (10%) and penetration resistance (2 MPa), which are both dependent upon the degree of soil compaction and represent the upper and lower limit of the LLWR (Silva *et al.*, 1994).

As little information is available on the effect of compaction on the growth of plants and Ca^{+2} and Mg^{+2} concentration, uptake content and translocation factor in root and shoot of pistachio under the least limiting water range (LLWR), thus an investigation was carried out to monitor the effect of different soil compaction and water levels (LLWR) on seedling growth and these treats in pistachio plants.

Materials and methods

Plant material and treatments

In order to study the changes in Ca⁺² and Mg⁺² of pistachio seedling (*Pistachio vera L.*) a nested experiment with four levels of soil compaction at six moisture levels with three replications was conducted in university of Tabriz. Requirement soil of this experiment were obtained from Pistachio orchard of East Azarbaijan Research Center for Agriculture and

Natural Resources. Some soil physical and chemical properties have been presented in Table 1. The air dried soil was passed through soil 4.75 mm sieve, and transferred into 72 PVC cylinders (diameter 15.24 and height 50 cm), the soils of cylinders were compacted in order to prepare four levels of soil bulk density (1.35, 1.5, 1.65 and 1.8 g cm⁻³). For proper establishment of seedling, 5 cm of the upper part of cylinders filled by topsoil.

Table 1. Some physical and chemical characteristics of experimental soil.

Texture Class	pH	EC	O.M	CCE	N	P	K	Fe	Mn	Zn	Cu
		dS m ⁻¹		%				mg kg ⁻¹			
Sandy loam	7.6	1.3	0.97	18	0.05	5	325	2.5	3.94	2.14	0.6

Pistachio seed were planted in plastic container and after 25 days, seedling transferred to cylinders with different compaction density. Then, soil moisture in all of cylinders was kept at field capacity up to 7 days. After seedlings establishment, six different volumetric water contents, from saturation to permanent wilting

point, for each compaction level were applied. Moisture levels in six ranges determined as Dasilva *et al.*, (1994), which shown in Table 2. For controlling the moisture content in cylinders used from time domain reflectometry (TDR) every each two days.

Table 2. Six ranges of moisture applied in four levels of soil compaction density.

Compaction levels (g cm ⁻³)	Θ _{fc}	Θ _{pwp}	Moisture levels					
			ML1	ML2	ML3	ML4	ML5	ML6
1.35	24.5	10	39-49	24-39	19-24	14-19	10-14	7-10
1.5	24	11	33-43	25-33	19-25	14-19	11-14	7-11
1.65	29	12.2	33-38	28-33	22-28	17-22	12-17	8-12
1.8	31	13.3	27-32	23-27	19-23	15-19	13-15	8-13

The following formulas were used to calculate the amount of water needed.

$$V = aD (\Theta_{v1} - \Theta_{v2})$$

V, water volume requirements (cm³); a, cross section of cylinders (cm²); D, soil depth (cm); Θ_{v2}, upper limiting range of moisture for each treatment (cm³/cm³) and Θ_{v1}, volumetric moisture amount reading by TDR (cm³/cm³).

Treatments measurement

After 90 days, seedling were cut from the surface of the soil and then the shoot and root of plants dried. The concentration of Ca⁺² and Mg⁺² in shoot and root of plants was measured by flame atomic absorption spectrometry (Waling *et al.* 1989). The amount of Ca⁺² and Mg⁺² was calculated by concentration × dry mater and transfer factor was also calculated by shoot

concentration of Ca⁺² or Mg⁺²/root concentration of Ca⁺² or Mg⁺² (Waling *et al.*, 1989; Rowell, 1994).

All the data were analyzed on the bases of experimental design, using SPSS software. The means of each trait were compered according to Duncan multiple range test at p≤0.05.

Results and discussion

Analysis variance of the date showed that the effects of soil compaction and moisture in each compaction on Ca⁺² concentration of root, Ca⁺² content of shoot and root and translocation factor were significant. Also, the effects of soil compaction on Ca⁺² concentration of shoot was significant. Also, soil compaction had significant effect on Mg⁺² concentration of shoot and root, uptake content of

Mg⁺² in shoot and root and Mg⁺² translocation factor. The effects of moisture in each levels of soil

compaction was only not significant on Mg⁺² concentration of pistachio shoot (Table 3).

Table 3. Analysis of variance of the effects of different soil compaction and moisture levels on Ca⁺² and Mg⁺² concentration, uptake content and translocation factor of pistachio plants.

Source	df	Ca ⁺²				Translocation Factor	Mg ⁺²				
		Concentration		Uptake content			Concentration		Uptake content		Translocation Factor
		shoot	root	shoot	root		shoot	root	shoot	root	
Compaction	3	7.76	25.39	25.9	16.6	1.732	0.05	0.01 ^{ns}	2.40	2.92	0.143
Moisture (compaction)	20	1.56 ^{ns}	4.033*	4.5*	19.1*	0.175*	0.014	0.056*	0.44*	0.54*	0.065*
Error	48	0.89	0.51	0.16	3.31	0.042	0.01	0.007	0.02	0.04	0.032

*,** Significant at p<0.05 and p<0.01, respectively.

Ca⁺² and Mg⁺² concentration in shoot

Concentration of Ca⁺² in shoot of pistachio seedling under high soil compaction (1.65 and 1.8 g cm⁻³) was lower than that of low soil compaction. There was no significant difference in this treat between 1.35 g cm⁻³ and 1.5 g cm⁻³ and also between 1.65 g cm⁻³ and 1.8 g cm⁻³ levels of soil compaction (Fig. 1a). Barraclough and Wier (1988) also indicated that with increasing of

soil compaction Ca⁺² concentration of wheat seedlings declined. Concentration of Mg⁺² in shoot declined with increasing soil compaction from 1.35 to 1.65 (g cm⁻³). The least of this treat was showed in severe soil compaction levels (1.8 g cm⁻³) (Fig. 1b). Barraclough and Wier (1988) reported that with increasing soil compaction, concentration of Mg⁺² in wheat and barley seedling reduced.

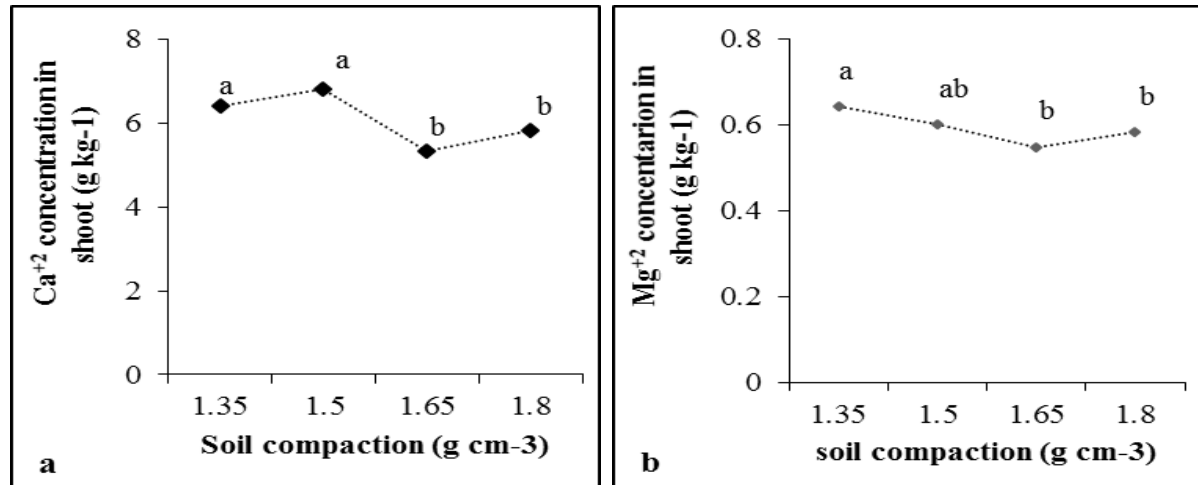


Fig. 1. Ca⁺² and Mg⁺² concentration in shoot of pistachio in response to different soil compaction.

Ca⁺² and Mg⁺² concentration in root

Under low soil compaction (1.35 g cm⁻³) the highest Ca⁺² concentration in root of seedling was showed in moisture levels three (ML3), which difference between ML3, ML4 and ML5 was not statistically significant. Also, there was no significant difference in Ca⁺² concentration of seedling under ML2 and ML6. The least of this treat was obtained from ML1 (Fig. 2a).

The highest concentration of Ca⁺² of root in low levels of soil compaction (1.35 and 1.5 g cm⁻³) was in least limiting water range (LLWR). However, under high levels of soil compaction (1.65 and 1.8 g cm⁻³) was out of LLWR. Maximum concentration of Mg⁺² in root under 1.35 (g cm⁻³) of soil compaction was showed in LLWR, in contrast, under 1.5 (g cm⁻³) of soil compaction was recorded in ML1 (had no significant

difference with LLWR). Under 1.65 (g cm⁻³) of soil compaction the highest concentration of Mg⁺² was showed in ML2. Concentration of Mg in shoot was similar in all soil moisture under 1.8 (g cm⁻³) of soil compaction (Fig. 2b).

This result was confirmed by Trought and Drew (1980) on wheat seedling. Under all soil compaction

levels, increasing of water deficit had no significant effect on root Ca⁺² concentration, as this trait increased under 1.65 g cm⁻³ level of soil compaction. This may be associated with reduction of root dry weight and increasing of Ca⁺² concentration under water stress. Chahkhoo (2010) also indicated that under water deficit stress, the concentration of Ca⁺² in roots of pistachio increased.

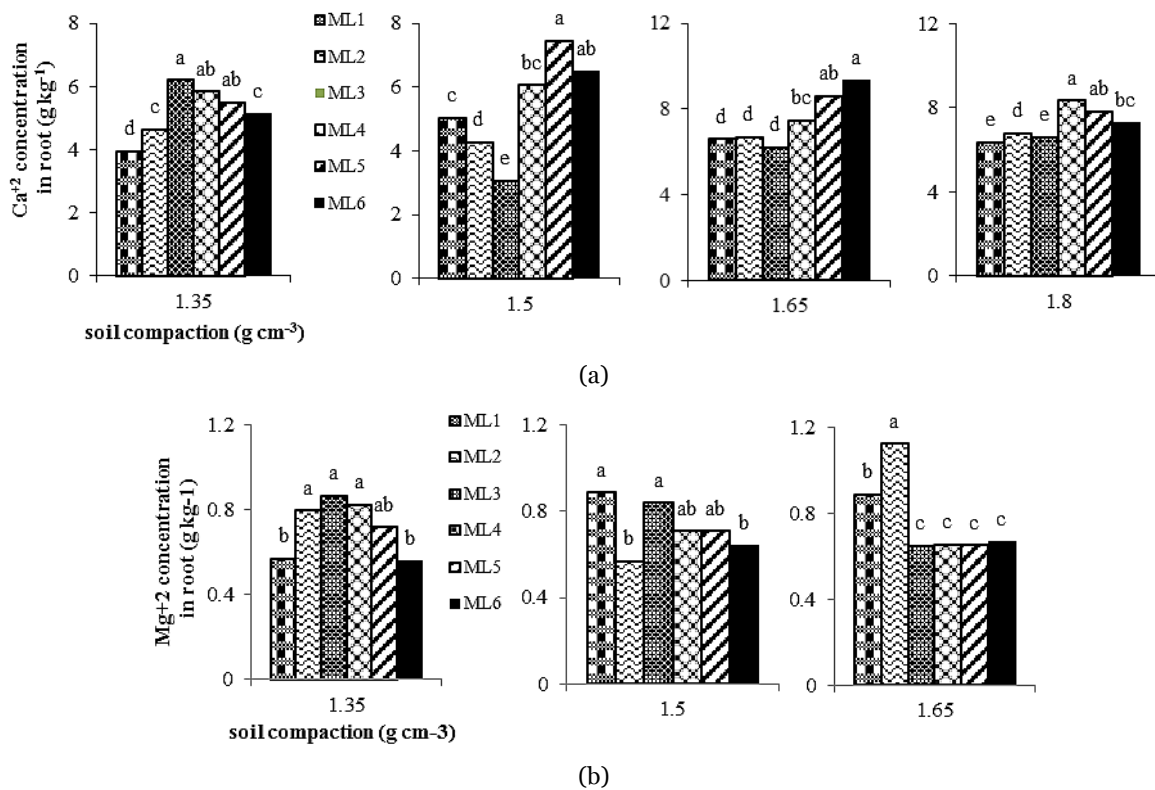


Fig. 2. Changes of Ca⁺² and Mg⁺² concentration (g kg⁻¹) in root of pistachio seedlings under different levels of soil compaction and moisture levels (ML).

Ca⁺² and Mg⁺² content in shoot

The amount of Ca⁺² uptake in shoot of pistachio seedling in 1.35 and 1.65 g cm⁻³ levels of soil compaction under ML1, ML2, ML3 and ML4 was statistically similar and more than that of ML5 and ML6. In soil compaction of 1.65 g cm⁻³ from ML1 up to ML3 the amount of Ca⁺² uptake was increased. In contrast, under ML4 to ML6 this treat gradually declined. Under severe level of soil compaction (1.8 g cm⁻³), uptake of Ca⁺² in shoot of seedlings increased up to ML2 and then significantly reduced (Fig. 3a). Between the highest amount of Ca⁺² in shoot and LLWR there was no significant difference under 1.35

and 1.5 of soil compaction levels. The highest amount of shoot Ca⁺², only under severe compacted soil (1.8 g cm⁻³) was in LLWR (Fig. 3a). Amount of Mg⁺² uptake at ML3 and ML4 (in LLWR), ML1 and ML2 under 1.35 and 1.5 g cm⁻³ of soil compaction was not statistically similar. The highest amount of Mg⁺² in shoot of pistachio under 1.8 (g cm⁻³) of soil compaction was in LLWR but, under 1.65 (g cm⁻³) of soil compaction was out of LLWR (Fig. 3b).

Nahar and Germazpeter (2002) on tomato showed similarly result. With reduction soil moisture, the movement of Ca⁺² form soil to root surface

significantly declined and also as the Ca^{+2} uptake by plant is inactive pathway, thus this linked by plant

transpiration which declined under water deficit conditions (Yu *et al.*, 2007).

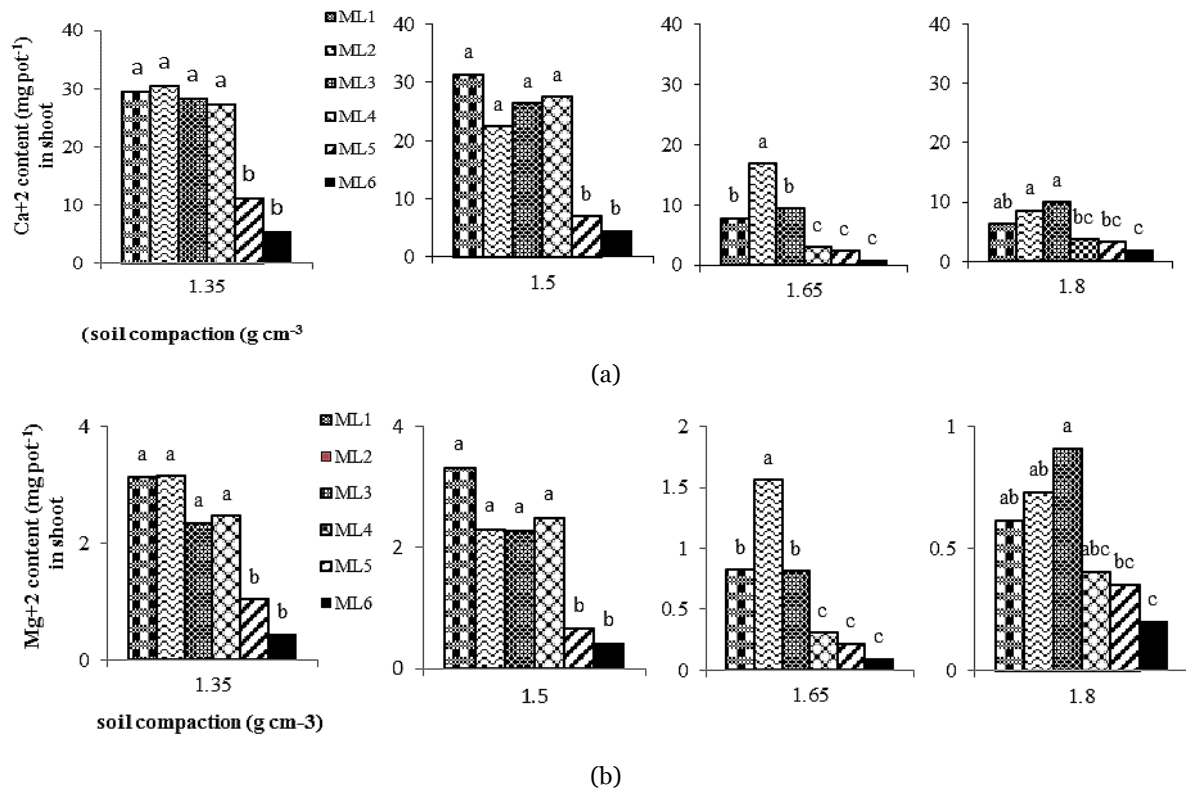


Fig. 3. Changes of Ca^{+2} and Mg^{+2} content (mg pot⁻¹) in root of pistachio seedlings under different levels of soil compaction and moisture levels (ML).

Ca⁺² and Mg⁺² content in root

In lower level of compacted soil (1.35 g cm⁻³) with decreasing moisture availability from ML1 to ML3 the amount of Ca^{+2} uptake by pistachio seedling significantly enhanced. But, under severe moisture conditions from ML3 to ML6 these treat gradually declined. These changes in Ca^{+2} uptake was mostly similar with plants changes under severe soil compaction condition (1.8 g cm⁻³). Maximum content of Ca^{+2} uptake in 1.5 g cm⁻³ level of soil compaction was showed in ML4, which had no significantly differences with ML1, ML2, ML5 and ML6. Under 1.65 g cm⁻³ level of soil compaction condition, the least Ca^{+2} uptake obtained from well moisture condition (ML1) and the highest of this trait showed in ML2 and then gradually up to water stress condition (ML6) declined (Fig. 4a). The highest amount of Ca^{+2} uptake under 1.35, 1.5 and 1.8 g cm⁻³ levels of soil compaction was in LLWR.

The highest amount of Mg^{+2} uptake under low soil compaction (1.35) was in LLWR. Maximum amount of this trait was recorded in ML1 under 1.5 (g cm⁻³) of soil compaction, and this with ML3 and ML4 (in LLWR) was similar. The highest Mg^{+2} uptake under 1.65 and 1.8 (g cm⁻³) of soil compaction was obtained from ML2 and this under higher compacted soil (1.8) had no significantly difference with ML3 (Fig. 4b). Najafi *et al.*, (1390) indicated that uptake of Mg^{+2} under logging significantly was declined. This results may be associated to decrease in oxygen and consequently in ATP synthesis by plants under logging and disruption in root growth under this condition. With increasing water limiting (from moisture level 3 to 6), the amount of Mg^{+2} uptake reduced under all soil compaction levels.

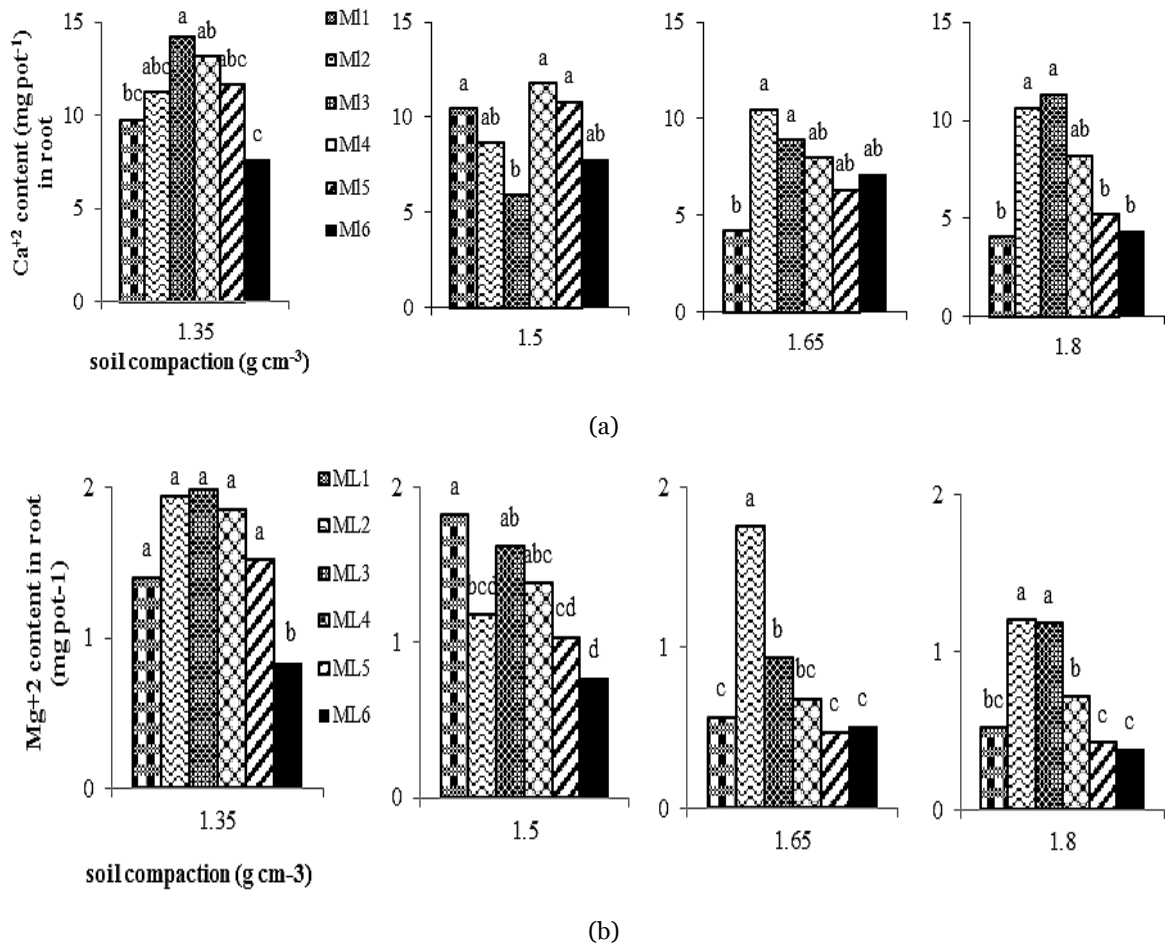


Fig. 4. Changes in Ca²⁺ and Mg²⁺ content in root of pistachio seedling under different levels if soil compaction and moisture levels (ML).

Translocation Factor of Ca²⁺ and Mg²⁺

Soil compaction levels of 1.35 and 1.5 g cm⁻³ under all moisture conditions had translocation factor more than one. This result clearly indicated that Ca²⁺ translocate from root to shoot of pistachio seedling in these compacted soils. In the lowest soil compaction (1.35 g cm⁻³) maximum translocation factor was showed in ML1, which had no significantly difference with ML2, ML3, ML4 and ML5. Under 1.5 g cm⁻³ levels of soil compaction, the highest amount of translocation factor was indicated in ML3. There was no significant difference in this trait between ML1, ML2, ML4, ML5 and ML6. In higher levels of soil compaction (1.65 and 1.8 g cm⁻³) under all moisture levels this trait was lower than one (Fig. 5a). The Ca²⁺ translocation factor of pistachio plants under both two high levels of severe compaction was lower than

that of the least levels of soil compaction. In all soil compaction levels under water deficit stress (ML6) in comparison to other levels of moisture this trait declined. Transfer factor of Mg²⁺ only under 1.35 (g cm⁻³) of soil compaction was higher than one. At ML3 translocation factor of Mg²⁺ from root to shoot adversely declined. The maximum amount of translocation factor under 1.5 (g cm⁻³) of soil compaction was recorded in ML6 (Fig. 5b). According to Osuagwu *et al.*, (2010) this results is due to the movement of Ca²⁺ from shoot to root, as in these condition Ca²⁺ play a kay roll in tolerance of plants to water stress. Mechanical resistance and poor aeration may restrict root growth, which especially affects the uptake of nutrients (Lipiece and Stepniewski, 1995).

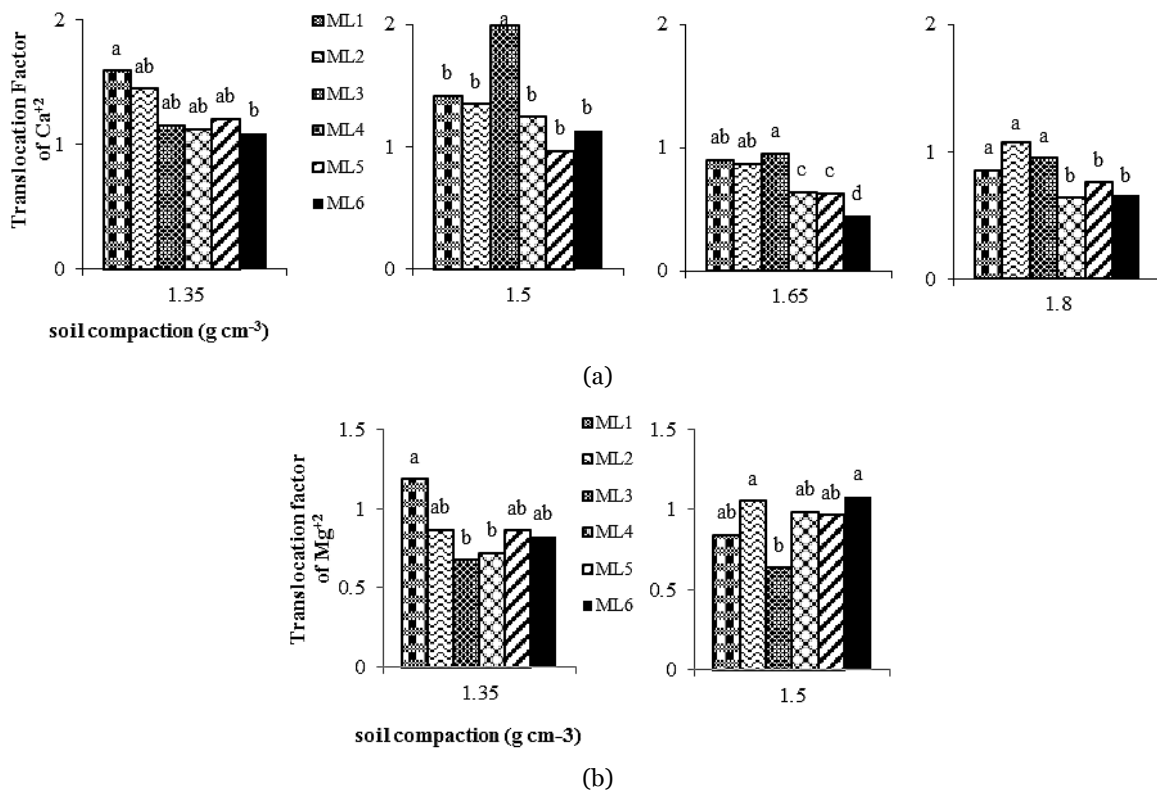


Fig. 5. Changes in Ca²⁺ and Mg²⁺ translocation factor of pistachio seedlings under different levels if soil compaction and moisture levels (ML).

Reference

Barraclough PB, Weir AH. 1988. Effects of a compacted subsoil on root and shoot growth, water use and nutrient uptake of winter wheat. *Journal of Agricultural Science* **110**, 207-216.

Beemster GTS, Masle J. 1996. Effects of soil resistance to root penetration on leaf expansion in wheat composition number and size of epidermal cells in mature blades. *Journal of experimental botany* **47**, 1651-1662.

Bhadoria PBS, Kaselowsky J, Claassen N, Jungk A. 1991. Soil phosphate diffusion coefficients: their dependence on phosphorous concentration and buffer power. *Soil Science Society of America* **55**, 56-60.

Chahkhoo A. 2010. Effect of Nitrogen application on the relative tolerance of pistachio seedling to water stress, M. Sc. Thesis, Department of Soil Sciences, Faculty of Agriculture, University of Vali-e-Asr, farsi Rafsanjan. (In Persian)

Dasilva AP, Kay BD, Perfect E. 1994. Characterization of the least limiting water range of soils. *Soil Science Society of America Journal* **58**, 1775-1781.

Hillel D. 1980. Soil compaction and consolidation. In: *Fundamentals of Soil Physics*, pp. 355-82. Academic Press, New York, USA

Kemper WD, Stewart BA, Porter UK. 1971. Effects of compaction on soil nutrient status. In *compaction of agricultural soils*, ed. K. K. Barnes, W. M. Carleton, H. M. Taylor, R. I. hrockmorton & G. E. van den Berg. American Society of Agricultural Engineers, St Joseph, pp. 178-89.

Letey J. 1985. Relationship between soil physical properties and crop production. *Advances in Soil Science* **1**, 277-294.

- Lipiec J, Stepniewski W.** 1995. Effects of soil compaction and tillage systems on uptake and losses of nutrients. *Soil Tillage Research* **35**, 37-52.
- Morgan JM.** 1984. Osmoregulation and water stress in higher plants. *Annual Review in Plant Physiology* **35**, 299-319.
- Mulholland BJ, Hussain A, Black CR, Taylor IB, Roberst JA.** 1999. Does root-sourced ABA have a role in mediating growth and stomatal response to soil compaction in tomato. *Physiologia plantarum* **107**, 267-276.
- Nahar K, Gretzmacher R.** 2002. Effect of water stress on nutrient uptake, yield and quality of tomato (*Lycopersicon esculentum* Mill.) under subtropical conditions. *Die Bodenkultur* **53**, 45-51.
- Najafi N, Sarhangzadeh A, Ostan S.** 2011. The effects of salinity by NaCl and soil logging on shoot and root concentration of K, Ca, Mg and Na of maize. Fifth Regional Conference on Agricultural Research findings, Kurdistan University, Sanandaj, Iran (In Persian).
- Osugwu GGE, Edeoga HO, Osugwu AN.** 2010. The influence of water stress (drought) on the mineral and vitamin potential of the leaves *Ocimum gratissimum* L. *Recent Research in Science and Technology* **2**, 27-33.
- Rowell DL.** 1994. *Soil Science: Method and Application*. Longman Scientific and Technical, Wiley, UK. P. 350.
- Schuurman JJ.** 1965. Influence of soil density on root development and growth of oats. *Plant and soil* **22**, 352-374.
- Taylor HM, Ratliff LF.** 1969. Root elongation rates of cotton and peanuts as a function of soil strength and soil water content. *Soil Science* **108**, 113-119.
- Trought MCT, Drew MC.** 1980. The development of waterlogging damage in young wheat plants in anaerobic solution culture. *Journal of Experimental Botany* **31**, 1573-1585.
- Veen BW, Van Noordwijk M, de Willigen P, Boone FR, Kooistra MJ.** 1992. Root-soil contact of maize, as measured by a thin-section technique. 3. Effects on shoot growth, nitrate and water-uptake efficiency. *Plant and Soil* **139**, 131-138.
- Waling I, Vark WV, Houba VJG, Van der lee JJ.** 1989. *Soil and Plant Analysis, a series of syllabi*. Part 7. *Plant Analysis Procedures*. Wageningen Agriculture University, Netherland.
- Yu X, Du X, Song L.** 2007. Effects of water stress on the growth and ecophysiology of seedlings of the *Rhus typhina*. *Scientia Silvge Sinicae* **43**, 57-61.