

Journal of Biodiversity and Environmental Sciences (JBES)

ISSN: 2220-6663 (Print), 2222-3045 (Online)

http://www.innspub.net Vol. 6, No. 1, p. 633-641, 2015

RESEARCH PAPER

OPEN ACCESS

Palynostratigraphy and palynofacies of the sanganeh formation in Qarah-Su section (NE, Iran)

Narges Shokri*, Ebrahim Ghasemi-Nejad¹, Ali-Reza Ashouri

Department of Geology, Faculty of Sciences, Ferdowsi University of Mashhad, Mashhad, Iran Department of Geology, University College of Sciences, University of Tehran, Tehran, Iran

Key words: Kopeh-Dagh Basin, Lower Cretaceous, Palynofacies, Palynostratigraphy, Dinoflagellate cysts, Iran.

Article published on January 31, 2015

Abstract

Palynomrphs extracted from rock samples of the Sanganeh Formation cropping out in the central Kopeh-Dagh Basin indicate a late Aptian-early Albian age for the formation. Eighty samples recovered from the shale and marl beds were processed and analysed for palynomorphs. Two palynological zones and tree types of Palynofacies have been identified. The Palynofacies and dinoflagellate assemblages indicate deposition in neritic to shallow marine environments.

^{*}Corresponding Author: Narges Shokri ⊠ shokri_narges@yahoo.com

Introduction

The Kopeh-Dagh basin is located in northeast of Iran with WNW-ESE trend. This sedimentary basin contains a great gas field which is in common with Iran, Turkistan and Afghanistan. The recent topography and morphology has formed during the Alpine tectonic phase (Aghanabati, 2004). This basin is especially interesting for geologists because it is located between the two super continents of Aurasia and Gondowana.

Palynology is regularly been used in such fields as biostratigraphy, paleoclimatology pale environmental reconstruction which has more recently been developed. Palynofacies analysis is an interdisciplinary approach. The entire organic content of the slides are assumed as sedimentology components that reflect original conditions of depositional environments (Tyson, 1995; Götz et al., 2008). The use of Palynofacies analysis for environmental interpretation allows determination of depositional environments (e.g. salinity, oxygenation) (Tyson, 1993, 1995; Batten, 1996). We try to interpret paleoenvironment of the Sanganeh Formation via determining such parameters as distribution of dinocysts as littoral and neritic group, the Peridinioid to Gonyaulacoid ratio (P/G), and palynofacies types; Also determination of the relative age of the study beds based on dinoflagellate cyst assemblages.

The Sanganeh Formation is one of the lower Cretaceous rock units in the Kopeh-Dagh Basin in north eastern Iran and south eastern Turkmenistan. Its type section thickness is 770m at the Sanganeh village, north-east of Mashhad (Afshar-Harb, 1994). The thickness increases from east to west (200-2000m). This Formation is overlain by the Aitamir Formation Sandstones and underlain by the Sarcheshmeh Formation. Although, it is overlain unconformably by the Kalat Formation (Maasstrichtian) in the west part of the basin. Overally, the age of the Sanganeh Formation has been designated as Albian (Kalantari, 1969) based on planktonic foraminifera and Late Aptian age based on benthic foraminifera (Motamedalshariati et al., 2010). Based on nannofossils, Mahanipour et al., (2011) suggested an early Aptian for this Formation. On the other hand, Raisossadat (2006) attributed the Sanganeh Formation to the late Early Aptian to Early Albian age based on ammonite fossils recovered in different sections.

The studied section is located at the Qarah-Su village at N36° 58' 14.04", E59° 40' 46", north-east of Mashhad (Fig. 1). At the studied section, the Sanganeh Formation is composed of dark grey shale, marl, limestone and siltstone. Its thickness is 580 m which contains some structures such as concretion, Septarian nodules (sometimes with ammonite core) and cone-in-cone. The aim of this paper is study palyno stratigraphy and palyno facies of the Sanganeh formation in Qarah-Su section (NE Iran) and find types of Palynofacies and indicate deposition in neritic to shallow marine environments.

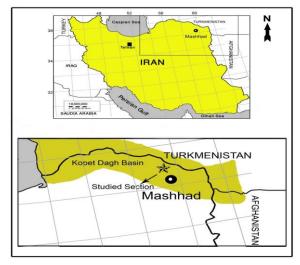


Fig. 1. Location map of the Qarah-Su village (Studied section).

Materials and methods

Sampling method

A total of 80 rock samples were collected from the shale and marl beds of the Sanganeh Formation in the Qarah-Su village, central Kopeh-Dagh basin, Iran (Fig. 1). The samples were processed for palynomorphs. The precise location of each sample is given in Fig.2.

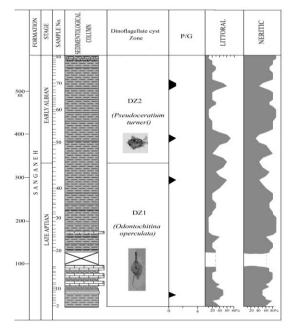


Fig. 2. Relative abundance of dinoflagellate cyst paleoenvironmental groups at Qarah-Su section. P/Gratio of abundance of peridinioid/ gonyaulacoiddino flagellates.

Analytical techniques

After washing and drying, the sediment samples were prepared for palynological analysis following standard extraction techniques (Poulsen et al., 1990; Travers, 2007) involving hydrochloric (HCl) and hydrofluoric acid (HF) treatment in order to dissolve carbonate and siliceous contents. The concentration of palynomorphs was filtered by decantation over a 10µm nylon mesh sieve, and the samples were centrifuged to concentrate the residues. The residues were not oxidized or stained. Three to five slides were prepared from each sample. The slides were investigated under a light microscope with magnifications of 200× -1000×, to identify and count the palynomorphs. All slides and residues are stored in the paleontology group, Department of Geology, Faculty of Sciences, Ferdowsi University of Mashhad, Iran.

Results and discussions

Palynostratigraphy and Dinoflagellate assemblages: The Qarah-Su section has shown a moderate density and diversity of dinoflagellate cysts with more than 37dinoflagellate cyst species belonging to 19 genera recorded.

Two bio zones have been identified based on the stratigraphic ranges of dinoflagellate cysts recorded.

DZ1: This dinozone corresponds to the total-range occurrence of the dino flagellate cyst species Odontochitinao perculata, which is considered to be diagnostic of it (Evans, 1966). It encompasses the interval from the base to 330 m with and is also marked with presence of Cribroperidinium, Cyclonephelium distinctum, Oligosphaeridium pulcherrimum, Paleoperidinium cretaceum, Spinifer itesramosus, Subtilisphaera and Coroniferaoceanica; however Oligosphaeridium is the dominated genus (Fig. 3). O. operculatais reported from Aptianstrata in New Zealand (Wilson, 1984) and Australia (Helby et al.,1987, 2004; Helby and Mcminn,1992, Oosting et al, 2006).

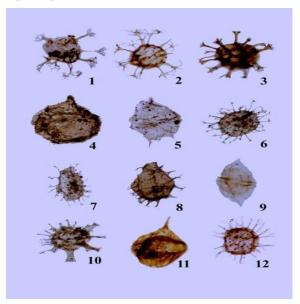


Fig. 3. Oligosphaeridium pulcherrimum (Deflandre and Cookson, 1955) Davey and Williams 1966. X500; 2, Oligosphaeridium complex (White, 842), Davey and Willams, 1966. X500;3, Kleithria s, Haeridium sp., 4, Cyclonephelium sp., X500; 5, Paleoperidinium cretaceum Pocock 1962. Χ 500; Surcolosphaeridium longifurcatum (Firtion, 1952) Davey et al, 1966b. X 500;, 7, Cleistosphaeridium sp., X500;8, Spiniferites sp., X500;9, Subtilisphaera sp., X 500;10, Florentinia daenei (Davey and Williams, 1966) Davey and Verdier 1973. X 500; 11, Cribroperidinium orthoceras (Eisenack, 1958) Davey, Emend. Sarjeant, 1958. X500; Systematophora sp., X500.

Although, some authors considered this species as index form for *Huterivian baremian* (Haq et al, 1987; Costa and Davey, 1992) and for upper Baremian (Harding, 1990). The other taxa were recorded from different point associated with this zone are: Subtilisphaera and Achomosphaera from late Aptian (Davey and Verdier, 1974; Skupien, 2003); Kiokansiumpolypes, Cyclonepheliumdistictum and Oligosphaeridium complex in Canada (Kimyai, 2000); Oligosphaeridium complex and Cassiculosphaeridium reticulate with early Aptian in age from Australia, South eastern France and south of Italy (Stover, 1996) and Albian from Egypt (Omran et al., 1990). The dinocysto. complex has a stratigraphic range of Valanginian to upper Albian for Tethys region (Williams and Bujak, 1985).

DZ2. This dinozonecorre sponds to the total-range occurrence of the dinoflagellate cyst species Pseudoceratiumturneri with an early Albian age (Morgan, 1980).

This with biozone also recognized Cassiculosphaeridium reticulata, Achomosphaera, Florentiniamantellii and Cyclonephelium; dominated genera are Systematophora and Cribroperidinium. This biozones encompassed the interval from 330m to the top of the formation.

Some species such as Paleoperidinium cretaceum, Spiniferitesramosus and Coroniferaoceanica were recorded from late Aptian strata and Florentiniamantellii from Albianof northern Western Desert of Egypt (Omran et al., 1990).

Dinoflagellate cysts are useful indicators for original paleoproductivity in surface water and oxygen changes in the bottom waters (Pross, 2001). Diversity of living dinoflagellate assemblages are controlled basically by the availability of nutrients, temperature, stratification, vertical water salinity environmental stress (Pross and Brinkhuis, 2005; Wall et al., 1977).

The Peridinioid to Gonyaulacoid ratio (P/G) is used to determine paleosalinity variations and proximity to shorelines. Peridinioid-dominated assemblages reflect low salinity and nutrient-rich conditions (Jaminski, 1995) related to near shore environments (Lagoonal, Brackish water). In contrast, low values of the ratio, i.e. gonyaulacoid-dominated assemblages indicate open marine environments and are resistant to oxic conditions (Sluijs et al, 2005).

In addition, cavate cysts are adapted to high-energy conditions and shallow marine environments; in contrast, chorate cysts are often inhabitant of lowenergy and deep waters (Ghasemi-Nejadetal, 1999). Therefore, the genera Spiniferites, Florentinia, Achomosphaera, Oligosphaeridium are cyst types with relatively long processes that are considered to be typical for outer neritic to open marine mid-shelf environments (Oosting et al, 2006). Genus Cleistosphaeridium reflects shallow marine and some genera such as Cyclonephelium, Subtilisphaera and Systematophora are associated with marginal marine (brackish, coastal) (Omran et al., 1990). Spiniferites with high percentage of phytoclasts indicate regressive conditions.

Abundance of Cribroperidinium in the more dysoxic intervals in the Votorantim section in Brazil indicates shallower environments (Santos et al., 2013). This genus has been identified in strata deposited in inner neritic environments in southeast France (Wilpshaar and Leerveld, 1994), north-central Spain (Peyrot et al., 2011; 2012), the Neuquén Basin in Argentina (Guler et al, 2013) and the Sergipe Basin in Brazil (Santos et al., 2013).

The few reported dinocysts are represented mainly by Subtilisphaera and Oligosphaeridium. These reflect marginal (Brackish) marine environments (Harding, 1986).

In the study section, the dinoflagellate assemblage composition dominated by Oligospharidium specis indicates inner neritic environment. In some parts of the section (Fig. 2), littoral group such Systematophora, Subtilisphaera and pseudoceratium show a regressive phase.

According to the distribution of dinoflagellate cysts as presented above, Oligosphaeridiumthat reflects the inner neritic environment is the dominated species. Distribution patterns of paleoenvironmentally important dinoflagellate cysts are displayed in Fig. 2. Additionally, the proportion of genus Spiniferites increases upwards. This trend reflects a deepening upwards sedimentation trend. Increase in the P/G ratio (especially Subtilisphaera) indicates increase in nutrient content.

Palynofacies analysis and sedimentary environments

In palynofacies analysis the entire organic content of the slides are investigated. Palynofacies studies are to understand paleoenvironment used hydrocarbon potential of the Sanganeh Formation. The main particles that were used for determination of the sedimentary environment were divided into three main groups: structured debris and phytoclasts, palynomorphs, and amorphous organic matter (AOM).

According to Tyson (1995), the AOM group consists of structureless particles that were observed under light microscopy. AOM quantity directly depends on the relative sea level changes. Their preservation increases in reducting conditions. The phytoclast group consists of structured particles that have terrestrial sources such as plant debris, cuticle, spore and pollen. This group is subdivided into two major subgroups: opaque and translucent. In spite of their high density (Van der Zwan, 1990), they display a high portability and can undergo long distance transport (Bombardiere and Gorin, 2000).

Marine palynomorphs such as dinoflagellate cysts, acritarchs and foraminiferal test linings; show ecological and environmental conditions.

Depositional paleoenvironments and paleoecology factors such as sea level changes, oxygen rate and proximity to the source of terrigenous material can be figured out even in carbonate systems (Götz et al., 2008; Santos et al., 2013) by using these groups, their percentage, and contribution on the slides.

Therefore, at least 300 particles are counted for each slide. The palynofacies composition and relative abundance in the samples are presented (Fig. 4). The AOM -palynomorphs- phytoclasts ternary plot (Fig. 5) show the presence of three palynofacies types in the Sanganeh Formation at the study area.

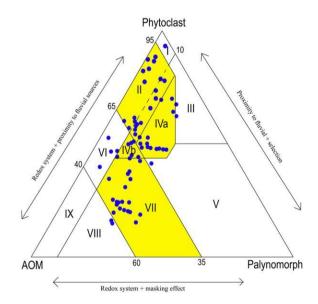


Fig. 4. Ternary diagram plot forQarah-Su (late Aptian-early Albian) section. Key paleoenvironmental fields (Tyson, 1995): I- Highly proximal shelf or basin; II- Marginal dysoxic-anoxic basin; III-Heterolithicoxic shelf (proximal shelf); IV-Shelf to basin transition; V- Mud-dominated oxic shelf (distal shelf); VI- Proximal suboxic-anoxic shelf; VII- Distal dysoxic-anoxic shelf; VIII-Distal dysoxicoxic shelf; IX-Proximal suboxic-anoxic basin.

Palynofacies I (PF-1): Opaque phytoclasts

This facies is characterized by a predominance of phytoclasts (up to 60% of the total particulate organic matter). The phytoclasts are mainly opaque and well preserved. The high values of opaque phytoclasts indicate oxidizing conditions and proximity to terrestrial sources or redeposition of terrestrial organic matter (Tyson, 1989). This facies is classified palynofacies type II as deduced from the AOM-Phytoclast-Palynomorph ternary plot of Tyson (1993) shown in Fig. 5.

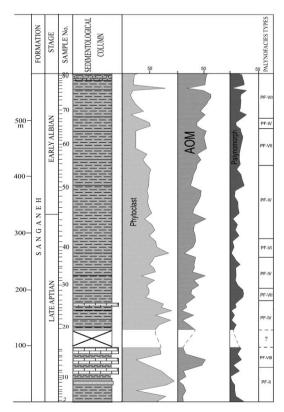


Fig. 5. Stratigraphic distribution of the tree main groups (phytoclast, AOM and palynomorph through theQarah-Susectiion.

Palynofacies II (PF-2): opaque phytoclasts, marine palynomorphs and amorphous organic matter.

This facies is distinguished by the commonness (up to 30%) of marine palynomorphs (specially dinocysts), common AOM (20-32%) and low frequency of opaque phytoclasts (Fig. 4). Dinocysts have good density and preservation but some of them reformed to AOM. This facies is classified as type IV of Tyson (1993).

Palynofacies III (PF-3): marine palynomorphs and amorphous organic matter.

This facies mainly was observed in upper part of the formation. It is characterized by high AOM (45-65%) and common palynomorphs (about 35%). The high AOM preservation is due to reducing basin conditions (Tyson, 1995).It could be caused by higher freshwater runoff (Aksu et al., 1995b, 1999; Abrajano et al., 2002, Kholeif and Ibrahim, 2010). Generally, the high percentage of AOM indicates reducing conditions, dysoxic-anoxic environment (Batten, 1983; Tyson, 1993).

This facie is classified as palynofacies type VII of Tyson (1993), and it just exposed in the middle part of the formation.

Conclusion

Palynomorph and palynofacies analyses of 80 rock samples from the Sanganeh Formation at Qarah-Su section in the Kopeh-Dagh basin, north eastern Iran, revealed that the marine palynomorphs predominated over the continental palynomorphs with a moderate diversity and abundance. The age of the formation was determined on the basis of twopalynological zones were identified the based on dinoflagellate cysts distribution. Odontochitinaoperculata zone indicated the late Aptianan dearly Albian age by Pseudoceratiumturneridino zone.

Based on the distribution of three main palynological groups on the slides, three types of Palynofacies were differentiated. These Palynofacies are based on the classification of Tyson (1993,1995) that show This formation was deposited in a neritic to shallow marine pale environment under dysoxic to suboxic conditions. The percentage of neritic, littoral and P/G group of dinoflagellate cysts also indicate that a distal shelf with some regressive phase.

References

Abrajano T, Aksu AE, Hiscott RN, Mudie PJ. 2002. Aspects of carbon isotope biogeochemistry of Late Quaternary sadiments from the Marmara Sea and Black Sea. Marine Geology 190, 151-164.

Afshar-Harb A. 1994. Geology of KopetDagh. In: Hushmandzadeh A. (Ed.), Treatise on the geology of Iran. Geological Survey of Iran, Tehran, 275pp. (in Persian).

Aghanabati A. 2004. Geology of Iran. Geological survey of Iran. 606pp (in Persian).

Aksu AE, Abrajano J, Mudie PJ, Yasar D. 1999. Organic geochemical and palynological evidence for terrigenous origin of the organic matter in Aegean Sea sapropel S1. Marine Geology 153, 303-318.

Aksu AE, Yasar D, Mudie PJ, Gillespie H. 1995b. Late glacial-Holocene paleoclimatological and paleoceanographic evolution of the Aegean Sea: micropaleontological and stable isotopic evidence. Marine Micropaleontology 25, 1-28.

Batten DJ. 1983. Identification of amorphous sedimentary organic matter by transmitted light microscopy. In: Brooks J. (Ed), Petroleum geochemistry and exploration of Europe. Geological Society Special Publications 12, Blackwell Scientific, Oxford. 275-287.

Batten DJ. 1996. Palynofacies. In: Jansonius J, McGregor DJ (Eds), Palynology: Principles and Applications. American Association of Stratigraphic Palynologists Foundation, Dallas, TX, 1011-1064.

Bombardiere L, Gorin GE. 2000. Stratigraphical and lateral distribution of sedimentary organic matter in Upper Jurassic carbonates of SE France. Sedimentary Geology 132, 177-203.

Costa L, Davey, RJ. 1992. Dinoflagellates of the Cretaceous System. In: Powell AJ.(Ed.), A Stratigraphic Index of Dinoflagellate Cysts. British Micropalaeontological Society Series. Chapman and Hall, 99-153.

Davey RJ, Verdier JP. 1974. Dinoflagellate Cyst from the Aptian type section at Gargas and La Bedoule, France. Palaeontology 17(3), 623-653.

Evans PR. 1966b. Mesozoic stratigraphic palynology of Otway Basin.Bureau Mineral Resources Geology and Geophysics, Australia. Record, 1966/198 (unpublished).

Evans PR. 1966c. Contribution to the Palynology of northern Queensland and Papua. Record of Bureau of Resources, Geology and Geophysics, Mineral 1966/69, 15p.

Ghasemi-Nejad E, Sarjeant WAS, Gygi R. 1999.Palynology and paleoenvironment of the uppermost Bathonian and Oxfordian (Jurassic) of the Northern Switzeraland sedimentary basin, Memorise Svizzere di Paleontologia 119, 69p.

GötzAE, Feist-BurkhardtS, RuckwiedK. 2008. Palynofacies and sea-level changes in the Upper Cretaceous of the Vocontian Basin, southeast France. Cretaceous Research 29, 1047-1057.

Guler MV, Lazo DG, Pazos PJ, Borel CM, Ottone EG, Tyson RV, Cessaretti N, Aguirre-Urreta MB. 2013. Palynofacies analysis and palynology of the Agua de la Mula Member (Agrio Formation) in a sequence stratigraphy framework, lower Cretaceous, Neuquen Basin, Argentina. Cretaceous Research 41, 65-81.

HardingI C. 1990. Adinocyst calibration of the European Boreal Barremian: Palaeontographica-Abteilung B, 218, 1-76.

Haq BU, Hardenbo lJ, Vail PR. 1987. Chronology of fluctuating sea levels since the Triassic. Science **235**,1156-1167.

Helby R, Mc Minn A. 1992. A preliminary report of Early Cretaceous dinocyst floras from Site 765, Argo Abyssal Plain, Northwest Australia. Proceedings of Ocean Drilling Program, Scientific Results 123, 07-420.

Helby R. 1987. Muderongia and related dinoflagellates of the latest Jurassic to Early Cretaceous of Australia. In: Jell PA. (Ed.), Studies in Australian Mesozoic Palynology. Memorise of Association of Australian Palaeontologists 4, 297-336.

Helby R, Morgan R, Partridge AD. 2004. Updated Jurassic-Early Cretaceous dinocyst Zonation NWS Australia: Geoscience Australia Publication. 2p.

Jaminski J. 1995. The mid-Cretaceous palaeoenvironmental conditions in the Polish Carpathians-a palynological approach. Review of Palaeobotany and Palynology 87, 43-50.

Kalantari A. 1969. Foraminifera from the middleJurassicCretaceous successions of Kopet-Dagh region (N.E. IRAN). Tehran, NIOC. Geology Laboraties, Publicatation. No.3, Ph.D. thesis, London University.

Kholeif SEH, Ibrahim MI. 2010. Palynofacies Analysis of Inner Continental Shelf and Middle Slope Sediments offshore South-eastern Egypt, Mediterranean. Geobios 43, 333-347.

Kimyai A. 2000. Palynology and Biostratigraphy of the Lower Cretaceous sediments in the South Barrow Test well No. 1. Point Barrow, Alaska. Palynology 24, 101-215.

Mahanipour A, Mutterlose J, Kani AL, Adabi MH. 2011. Paleontology and biostratigraphy of early Cretaceous (Aptian) calcareous nannofossils and the δ13C isotope record from NE Iran. Cretaceous Research **32**, 331-356.

Morgan R. 1980. Palynostratigraphy of the Australian Early and Middle Cretaceous. Memorise of the Gelogical Survey of New South Wales. Palaeontology 18, 153p.

Motamedalshariati M, Sadeghi A, Moussavi-Harami 2010. New Foraminifera morphogroups from Sanganeh Formation in TakalKuh section, western Kopeh Dagh basin. Stratigraphy and Sedimentology Researches, 40(3), 137-150.

Omran AM, Soliman HA, Mahmoud MS. 1990. Early Cretaceous palynology of the three boreholes from northern Western Desert (Egypt). Review of Palaeobotany and Palynology 66, 293-312.

Oosting AM, Leerveld H, Dickens GR, Henderson RA, Brinkhuis H. 2006. Correlation of Barremian-Aptian (mid-Cretaceous) dinoflagellate cyst austral realms. Cretaceous Research 27, 792-813.

Peyrot D, Barroso-Barcenllia F, Barron E, Comas-Rengifo MJ. 2011. Palaeoenvironmental analysis of Cenomanian-Turoniandinocyst assemblages from the Castilian Platform (northern-Central Spain). Cretaceous Research 32, 504-526.

Pevrot D. Barroso-Barcenilla F. Feist-Burkhardt S. 2012. Palaeoenvironmental controls on late Cenomanian-early Turoniandinoflagellate cyst assemblages from Condemios (Central Spain). Review of Palaeobotany and Palynology 180, 25-40.

Poulsen NE, Gudmundsson L, Hansen JM, Husfeldt Y. 1990. Palynological preparation techniques, a new maceration tank method and other modifications. Geological Survey of Denmark Series 10, 1-22.

Pross J. 2001. Paleo-oxygenation in Tertiary epeiric seas: evidence from dinoflagellate cysts. Palaeogeography, Palaeoclimatology, Palaeoecology 166, 369-381.

Pross J, Brinkhuis H. 2005. Organic-walled dinoflagellate cysts as paleoenvironmental indicators the Paleogene; a synopsis of concepts. Paläontologische Zeitschrift 79, 53-59.

Raisossadat SN. 2006. The ammonite family Parahoplitidae in the Sanganeh Formation of the KopetDagh Basin, north-eastern Iran. Cretaceous Research 27, 907-922.

Santos AS, Helenes J, Carvalho MA. 2013. Palynofacies evidence of dysoxic and upwelling in the Turonian of the Sergipe Basin, Brazil. Cretaceous Research 46, 151-165.

Skupien P. 2003. Dinoflagellate study of the lower cretaceous deposits in the PieninyKlippen Belt.Pochovia section, Slovak Western Carpathiana. Bulletin of the Czech Geological Survey **78**, 67-82.

Sluijs A, Pross J, Brinkhuis H. 2005. From greenhouse to icehouse; organic-walled dinoflagellate cysts as paleoenvironmental indicators in the Paleogene, Earth-Science Reviews **68**, 281-315.

Stover LE. 1996. Mesozoic, Tertiary dinoflagellates, acritarchs and prasinophytes. American Association of stratigraphic Palynologists Foundation **2**,641-750.

Travers A. 2007. Paleopalynology.2nd Edition, Springer, 813 p.

Tyson RV. 1989. Late Jurassic Palynofacies trends, Piper and Kimmeridge Clay Formations, UK onshore and offshore. In: Batten DJ, Keen MC (Eds.), Northwest European Micropaleontology and palynology, British Micropaleontological Society Series. Ellis Horwood, Chichester, 135-172.

Tyson RV. 1993. Palynofacies analysis; in Jenkins, DJ, (Ed.), Applied Micropaleontology.Kluwer Academic Publishers, Dordrecht. 269pp.

Tyson RV. 1995. Sedimentary Organic Matter: organic facies and palynofacies. 615 pp. Chapman & Hall, Londen.

Wall D, Dale B, Lohmann GP, Smith WK. 1977. The environmental and climatic distribution of dinoflagellate cysts in modern marine sediments from regions in the North and South Atlantic Oceans and Adjacent seas. Marine Micropaleontology 2, 121-200.

Wilpshaar M, Leerveld H. 1994. Palaeoenvironmental change in the Early Cretaceous Vocontian Basin (SE France) reflected by dinoflagellate cyst. Review of Palaeobotany and Palynology **84**, 121-128.

Wilson GJ. 1984. New Zealand Late Jurassic to Eocene dinoflagellate biostratigraphy: a summary: Newsletters on Stratigraphy **13**, 104-117.

Williams GL, Bujak JP. 1985. Mesozoic and Cenozoicdinoflagellates. In: Bolli HM, Saunders JB, Perch-Nilsen K. (Eds.), Plankton Stratigraphy, Cambridge University Press, Cambridge, 847-964.

Vander Z, Wan CJ. 1990. Palynostratigraphy and Palynofacies reconstruction of the Upper Jurassic to Lowermost Cretaceous of the Dragen field, offshore Mid Norway. Review of Palaeobotanyand Palynology **62**, 157-186.