

# Journal of Biodiversity and Environmental Sciences (JBES) ISSN: 2220-6663 (Print) 2222-3045 (Online) Vol. 7, No. 1, p. 423-430, 2015 http://www.innspub.net

OPEN ACCESS

# Plum shelf life enhancement by edible coating based on pectin and carboxymethyl cellulose

Sima Panahirad<sup>1</sup>, Nasser Mahna<sup>1\*</sup>, Rahim Naghshiband-Hassani<sup>1</sup>, Babak Ghanbarzadeh<sup>2</sup>, Fariborz Zaare-Nahandi<sup>1</sup>

<sup>a</sup>Department of Horticultural Sciences, Faculty of Agriculture, University of Tabriz, Tabriz, Iran

<sup>b</sup>Department of Food Sciences, Faculty of Agriculture, University of Tabriz, Tabriz, Iran

Article published on July 26, 2015

Key words: Postharvest, Edible coating, Shelf life, Polygalacturonase, Prunus domestica L.

#### **Abstract**

The aim of the current study was to explore the effect of carboxymethylcellulose–pectin (CMC-Pec) based coating treatments on some qualitative characteristics of plum fruits (*Prunus domestica* L.) during shelf life period at 19 °C and 65% RH (relative humidity) for eight days. In this sense, 0.5% Pectin with four concentrations of CMC (0, 0.5, 1 and 1.5%) were applied to plum fruits in a completely randomized design and weight loss, firmness, vitamin C (vit C), titratable acidity (TA), total soluble solids (TSS), pH and polygalacturonase (PG) were measured. The results showed that except vit C and firmness, other quality parameters were affected by CMC-Pec based coatings. In general, 0.5% Pec + 0.5% CMC had the best results in terms of all measured parameters and could be suggested as a coating to apply on plum fruits during postharvest periods to improve quality properties.

\*Corresponding Author: Nasser Mahna 🖂 mahna@tabrizu.ac.ir

#### Introduction

Edible films and coatings are considered as a new approach for fruits and vegetables preservation especially through past decades. There are wide ranges of materials which are used to supply edible films and coatings such as lipids, polysaccharides, carbohydrates, proteins and etc. each having many constituents. On top of that, each material includes different characteristics which cause unlike effects on food features (Bourtoom, 2008). Moreover, their usage reduces non-biodegradable packaging ingredients which lessen environmental concerns and deliberate as safe methods (Olivas et al., 2008; Campos et al., 2011). Edible films and coatings mostly act as modified atmosphere and regulate O2 and CO2 transmission between coated fruits and environment, which in turn prevents ripening process that leads to senescence and decay (Bal, 2013). Applications of edible films and coatings to maintain plum quality have been reported in several studies: hydroxypropylmethylcellulose (Navarro-Tarazaga et al., 2008), whey protein (Reinoso et al., 2008), versasheen (Eum and Hwang, 2009), Aloe arborescens and Aloe vera gels (Guillen et al., 2013) and alginate (Valero et al., 2013). There are some other works in other fruits like methylcellulose in apricot (Ayranci and Tunc, 2004).

Plums (Prunus domestica L.) have a short shelf life period due to fast ripening behavior after harvesting and their perishable nature. Plums have 2-6 weeks commercial shelf life depending on cultivar (Abdi et al., 1997). Although maintenance of them in low temperature (o °C) could increase postharvest life, chilling injury symptoms limit its advantages and consequently accelerate fruits quality loss (Crisosto et al., 2004; Manganaris et al., 2008). The changes could influence consumer acceptance as organoleptic characters of fruits are the main factors in this regard (Diaz-Mula et al., 2008). Also, plum fruits contain lots of valuable compounds like vit C, polyphenols, anthocyanin and flavonoids as antioxidants which help human body fight risky diseases (Tomas-Barberan and Espin, 2001, Collin and Harrington,

2002; Scalbert et al., 2005).

The polysaccharides as coating materials for fruits have been applied extensively in the past few years. They have benefits of availability, low cost, and biodegradability (Zhou et al., 2008). Besides, their physico-chemical properties can also be enhanced by modifying them. Cellulose is a naturally occurring polymer, found abundantly, and usually present as a linear polymer of anhydroglucose (Kester and Fennema, 1986). Several cellulose derivatives such as methyl cellulose (MC), carboxymethyl cellulose (CMC), and hydroxypropylmethyl cellulose (HPMC) are widely produced commercially (Bourtoom, 2008). The coatings and films based on these cellulose ethers are commonly transparent, flexible, odorless, tasteless, water-soluble, and resistant to O2 and CO2 (Nisperos-Carriedo et al., 1992). Edible coatings based on cellulose derivatives have also been used to delay ripening in some climacteric fruits like mango, papaya and avocado (Baldwin et al., 1999; Maftoonazad and Ramaswamy, 2005). Sodium-CMC coating extended storage life of pineapple fruit up to 5 weeks at 10°C and 90-95% RH, whereas the control fruit had a storage life of only 28 days (Nimitkeatkai et al., 2006). The application of sodium-CMC coating was also found able to postpone ripening of mango up to 32 days at 13°C and 85% RH (Carrillo-Lopez et al., 2000). One more material based on polysaccharides is Pectin to use as edible coating that is non-toxic, anionic biopolymer; water-soluble and biodegradable with great properties in prevention of volatile compounds particularly O2 transmission. They exist in cell wall of plants and based on their resource have various characters (Medeiros et al., 2012). Oms-Oliu et al. (2008) reported effectiveness of pectin based edible coating on shelf life extension of fresh-cut melon. Avocado fruits coated with pectin based edible coating showed better results as compared with the controls in case of disease, respiration, qualitative parameters, color and texture (Maftoonazad et al., 2007).

To our knowledge, little is known about the effect of

these materials as edible coating especially in combination in plum fruits during shelf life period. Thus, the current study was carried out to elucidate their efficiency in this regard. Changes of weight loss, firmness, pH, vitamin C (vit C), titratable acidity (TA), total soluble solids (TSS) and polygalacturonase (PG) were monitored in plum fruits subjected to edible coatings based on combination of Pec and CMC.

### Materials and methods

Plant materials, coatings preparation and treatments

Plum fruits (Prunns domestica L.) Cultivar 'Golden drop' were hand-harvested at physiologically mature stage from a commercial orchard in Shabestar, East Azerbaijan, Iran. Then, they were transported to the laboratory immediately, washed and air-dried for 2 hours on paper towel and screened based on uniformity of shape, size and peel color. Fruits were divided into 5 groups of 40 fruits in each. Fruits were dipped in different concentrations of aqueous solutions of 0.5 Pec + 0, 0.5, 1 and 1.5 % CMC plasticized with 0.3% w/v glycerol for 60 sec in which 0.5 Pec + 0 CMC were considered as control. All fruits were then air dried for approximately 60 min and stored at 19°C and 65% RH for ten days. Experiment design was completely randomized design (CRD) with three replications for each treatment.

To prepare coatings, 0.5% Pectin with different amounts of (0, 0.5, 1 and 1.5 %) CMC mixed in distilled water and the coatings solutions were mixed for 40 min by stirring at 60 °C. After complete dissolving and getting the clear liquid, glycerol 3% w/v was added and again the solutions were mixed for 30 min by stirring at the same temperature. Then they stored at room temperature to get cold.

**Qualitative** attributes evaluation Weight loss, firmness, TA, TSS, Vit C, pH Weight loss was determined initially and in each sampling during storage and calculated as: weight

loss =  $(Wo - Wf)/Wo \times 100$  in which Wo was the initial sample weight and Wf, the final sample weight. The results were reported as weight loss percentage. Firmness was measured on two sides of each fruit, after the removal of the skin fruit, by an Effegi penetrometer (Model FT-011) with an 8 mm diameter flat probe. Total acidity (TA) was measured by titration with 0.1N NaOH up to pH 8.1 and expressed as percentage of malic acid equivalent per 100 g fresh weight. Total soluble solids (TSS) were determined by measuring the refractive index of juice using a digital refractometer (Model PAL-1) at 20 °C and . Vit C content (mg/100g) of the samples was determined by the titramitric method (AOAC, 2000) in which visual titration method of reduction dichlorophenolindophenol dye was used. Three replicates were assessed for each measurement. The pH was recorded with a pH meter (Hana instrument, Italy).

#### PG activity

The activity was assayed based on the release of decreasing groups produced by PG and measured by spectrophotometer. Sodium acetate buffer (pH=4.5) was used for enzyme extraction. 50µl enzyme extract was mixed with 950 µl Sodium acetate buffer and 1ml of 0.3% polygalacturonic acid then incubated at 30 °C for 45 min. To stop the reaction, 800 µl of 0.1 M borate buffer (pH=9) o°C and 200 µl of 1% cyanoacetamide solution were added to the reaction mixture and boiled for 10 min. After cooling down, the absorbance was measured at 276 nm. A blank was determined in the same way without enzyme addition. The standard curve was built with α-Dgalacturonic acid as reducing sugar. One unit (U) of PG activity was defined as the amount of enzyme that releases 1 µmol of galacturonic acid per min under the assay conditions (Aguiló-Aguayo et al., 2010).

# Statistical analysis

Analysis of variance (ANOVA) carried out with SPSS (version 16, SPSS Inc., Chicago, IL.) software. Differences between means were compared using Duncan's new multiple range test (with significancy at  $P \le 0.05$ ).

#### Results and discussion

The obtained results about TA demonstrated that TA decreased (P≤0.01) (Table 1) as time passed, reached to minimum at the end of storage as a common process. Edible coating containing 0.5%Pec+0.5%CMC significantly (P≤0.01) (Table 1) showed the highest amount, but other concentrations

of CMC in combination with 0.5%Pec surprisingly even acted worse than control (Fig. 1). TA decrease mostly is due to using TA as substrate in respiratory metabolism and changing to sugars. Alginate edible coating delayed TA decrease of plum fruits and coated fruits had higher TA amount (Valero *et al.*, 2013). Plums coated with *Aloe spp.* and versasheen® showed higher content of TA (Eum and Hwang, 2009; Guillen *et al.*, 2013).

**Table 1.** ANOVA for dependent variables for coating treatments, storage time and their interactions for plum fruits.

	Time	Treatment	Time*Treatment	
Firmness	15.386**	0.903 <sup>ns</sup>	2.722*	
Weight loss	79.772**	2.131*	1.252 <sup>ns</sup>	
TSS	7.447**	5.347*	2.416*	
TA	29.642**	32.493**	19.319**	
pН	38.35**	7.11**	$0.651^{\mathrm{ns}}$	
Vit C	8.761**	$0.248^{\mathrm{ns}}$	1.413 <sup>ns</sup>	
PG activity	35.856**	10.587**	6.466**	

Note: \* P≤0.05, \*\* P≤0.01, ns non-significant.

In case of TSS feature, the maximum and minimum amounts of TSS were observed in control (0.5% Pec) and combination of 0.5% Pec with all concentrations of CMC edible coatings ( $P \le 0.05$ ) (Table 1). In our point of view, edible coating represented better results because of less changes in TSS value when it's

compared with day o (Fig. 2). As time passes, TSS amounts increased (P≤0.01) as at the end of the shelf life period the highest amount was noticed. No differences in TSS amount were detected in coated "sapphire" plums and controls (Eum and Hwang, 2009).

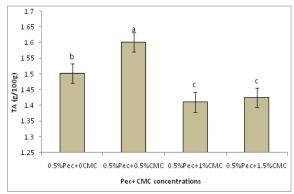
Table 2. effect of coatings on quality parameters and PG activity of plum fruits during storage at 19 °C.

Treatment coatings	Firmness	Weight loss	TA	TSS	pН	Vit C	PG activity
Control (0.5%Pec)	9.14	10.83 <sup>ab</sup>	$1.5^{\mathrm{b}}$	$11.53^{\rm b}$	3.603 <sup>a</sup>	8.4	$0.2085^{c}$
o.5%Pec+o.5%CMC	9.89	9.40 <sup>a</sup>	1.6a	10.76a	3.612a	7.933	$0.2036^{\rm b}$
o.5%Pec+1%CMC	8.72	10.69 <sup>ab</sup>	1.41 <sup>c</sup>	10.3 <sup>a</sup>	$3.681^{\mathrm{ab}}$	8.4	$0.2042^{\mathrm{bc}}$
0.5%Pec+1.5%CMC	9.81	$12.82^{b}$	$1.43^{c}$	10.41 <sup>a</sup>	$3.695^{\mathrm{b}}$	8.467	0.1966 <sup>a</sup>

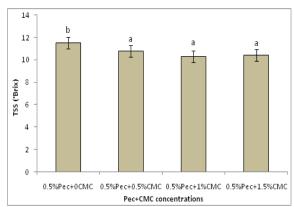
Togrul and Arslan (2004) stated that 1.1% CMC-based edible coating acted better than controls since cause less changes in TSS amounts of mandarin.

The best result in pH property was achieved in controls and similarly 0.5%Pec+0.5%CMC edible coating (Table 2) and others showed pH enhancement (P $\leq$ 0.01) (Fig. 3). During maintenance time of plum fruits pH noticeably increased and at the end of

period reached the top. The pH changes in mandarins coated with a CMC edible coating depended on coating's concentrations. Higher concentrations of CMC caused higher pH and lower concentrations next to beeswax caused lower pH. It was reported that control fruits had higher pH and all fruits showed an increasing trend in pH amount during storage (Togrul and Arslan, 2004).

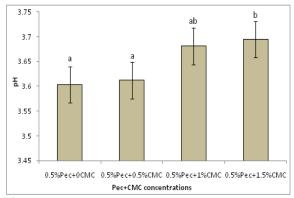


**Fig. 1.** The effect of coating formulations based on Pec+CMC on TA amounts of plum fruits at 19 °C for eight days. (Data are mean+95% interval confidence).

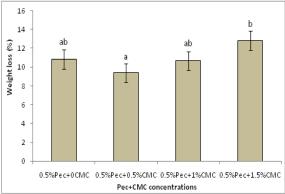


**Fig. 2.** The effect of coating formulations based on Pec+CMC on TSS values of plum fruits at 19 °C for eight days. (Data are mean+95% interval confidence).

Firmness and vit C attributes showed no significant differences between control (0.5% Pec) and other coating treatments (Table 1 and 2). Though, adding CMC to 0.5% Pec caused better results. To identify, regarding firmness, 0.5 and 1% CMC in combination with 0.5% Pec preserved fruit firmness more appropriately and in case of vit C, once more, 1 and 1.5% CMC + 0.5% Pec caused better outcomes and the minimum and maximum vit C values was observed at 0.5% Pec and 0.5% Pec + 1.5% CMC edible coatings. Through maintenance time, vit C content primarily increased and reached the highest at day 4 and then slightly decreased but despite this reduction, its amount at the end of the storage was higher than harvesting time. Fruit firmness declined as time passed and this trend was obvious amongst sampling dates. The least value was measured at the end of storage. Guillen *et al.* (2013) reported no significant differences on firmness between coated and control plum fruits. carboxymethylchitosan-based coating had no effect on vit C content of pear (Zhou *et al.*, 2008).



**Fig. 3.** The effect of coating formulations based on Pec+CMC on pH of plum fruits at 19 °C for eight days. (Data are mean+95% interval confidence).



**Fig. 4.** The effect of coating formulations based on Pec+CMC on weight loss of plum fruits at 19 °C for eight days. (Data are mean+95% interval confidence).

Weight loss of plum fruits increased (P≤0.01) during storage at 19 °C (Table 1). However, obtained data demonstrated that 0.5%Pec+0.5% CMC-based edible coating significantly (P≤0.05) decreased weight loss in comparison to control and controversy 0.5%Pec+1.5% CMC- based edible coating increased it (Fig 4, Table 2). Effectiveness of coatings in preventing of fruit weight loss has been previously reported by Valero *et al.* (2013) and Guillen *et al.* (2013). Also, in some cases coatings enhanced weight loss (Navarro-Tarazaga *et al.*, 2008) that overall could describe our results.

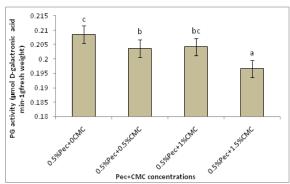


Fig. 5. The effect of coating formulations based on Pec+CMC on PG activity of plum fruits at 19 °C for eight days. (Data are mean+95% interval confidence).

PG activity decreased till 4 days of storage and then increased. PG activity affected by a combination of Pec and CMC edible coatings in which 0.5%Pec+1.5%CMC o.5%Pec+o.5%CMC and significantly reduced the enzyme activity compared to control (Fig 5). Since PG is one of the cell wall hydrolysis enzymes and its activity increased during ripening and after harvesting, a reduction in its activity could prevent softening.

## Conclusion

Regarding to positive effects of edible coating based on Pec-CMC on qualitative attributes of plum fruits, its application during postharvest life could be considered as a safe and stimulating method. Results of current study suggested that 0.5% Pec + 0.5% CMC could be applied in this regard.

# References

Abdi N, Holford P, McGlasson WB. 1997. Effects of harvest maturity on the storage life of Japanesetype plums. Australian Journal of Experimental Agriculture 37, 391–397.

Aguiló-Aguayo I, Soliva-Fortuny R, Martín-Belloso O. 2010. Color and viscosity of watermelon juice treated by high-intensity pulsed electric fields or heat. Innovative Food Science and Emerging Technologies 11, 299-305.

Ayranci E, Tunc S. 2004. The effect of edible coating on water and vitamin C loss of apricots (Armeniaca vulgaris Lam.) and green peppers (Capsicum annuum L.). Food Chemistry 87, 339-342.

Bal E. 2013. Postharvest application of chitosan and low temperature storage affect respiration rate and quality of plum fruits. Journal of Agricultural Science and Technology 15, 1219-1230.

Baldwin EA, Burns JK, Kazokas W, Hagenmaier RD, Bender RJ, Pesis E. 1999. Effect of two edible coatings with different permeability characteristics on mango (Mangifera indica L.) ripening during storage. Postharvest Biology and Technology 17, 215-226.

Bourtoom T. 2008. Edible films and coatings: characteristics and properties. International Food Research Journal 15(3), 237-248.

Campos CA, Gerschenson LN, Flores SK. 2011. Development of edible films and coatings with antimicrobial activity. Food **Bioprocess** and Technology 4, 849-875.

Carrillo-Lopez A, Ramirez-Bustamante F, Valdez-Torrez JB, Rojas-Villegaz R, Yahia EM. 2000. Ripening and quality changes in mango fruits as affected by coating with an edible film. Journal of Food Quality **23(5)**, 479-486.

Collins AR, Harrington V. 2002. Antioxidants; not the only reason to eat fruit and vegetables. Phytochemistry Reviews 1, 167-174.

Crisosto CH, Garner D, Crisosto Bowerman E. 2004. Increasing 'Blackamber' Plum (Prunus salicina Lindell.) Consumer Acceptance. Postharvest Biology and Technology 34, 237-244.

Diaz-Mula HM, Zapata PJ, Guillen F, Castillo S, Martinez-Romero D, Valero D, Serrano M. 2008. Changes in physicochemical and nutritive parameters and bioactive compounds

development and on-tree ripening of eight plum cultivars: a comparative study. Journal of the Science of Food and Agriculture 88, 2499-2507.

Eum HA, Hwang DK. 2009. Influence of edible coating on quality of plum (Prunus salicina Lindl. ev. 'Sapphire'). European Food Research Technology **229**, 427-434.

Guillen F, Diaz-Mula HM, Zapata PJ, Valero D, Serrano M, Castillo S. 2013. Aloe arborescens and Aloe vera gels as coatings in delaying postharvest ripening in peach and plum fruit. Postharvest Biology and Technology 83, 54-57.

Kester JJ, Fennema OR. 1986. Edible films and coatings. A review. Food Technology 40(12), 47-59.

Maftoonazad N, Ramaswamy HS, Moalemiyan M, Kushalappa AC. 2007. Effect of pectin-based edible emulsion coating on changes in quality of avocado exposed to Lasiodiplodia theobromae infection. Carbohydrate Polymers 68, 341-349.

Maftoonazad N, Ramaswamy HS. 2005. Postharvest shelf-life extension of avocados using methylcellulose-based coating. LWT- Food Science and Technology 37, 617-624.

Manganaris GA, Vicente AR, Crisosto CH. 2008. Effect of pre-harvest and post-harvest conditions and treatments on plum fruit quality. CAB Reviews: Perspective in Agriculture, Veterinary Science, Nutrition and Natural Resources 3(9).

Medeiros BG, Pinheiro AC, Carneiro-da-Cunha MG, Vicente AA. 2012. Development and characterization of a nanomultilayer coating of pectin and chitosan – Evaluation of its gas barrier properties and application on 'Tommy Atkins' mangoes. Journal of Food Engineering 110, 457-464.

Navarro-Tarazaga ML, Sothornvit R, Pérez-Gago MB. 2008. Effect of plasticizer type and amount on hydroxypropyl methylcellulose-beeswax edible film properties and postharvest quality of coated plums (cv. Angeleno). Journal of Agriculture and Food Chemistry 56(20), 9502-9509.

Nimitkeatkai H, Srilaong V, Kanlavanarat S. 2006. Effect of edible coating on pineapple fruit quality during cold storage. ISHS Acta Horticulturae 712, 643-648.

Nisperos-Carriedo MO, Baldwin EA, Shaw PE. 1992. Development of an edible coating for extending postharvest life of selected fruits and vegetables. Proceedings at the annual meeting of Florida State Horticultural Society 104, 122-125.

Olivas GI, Davila-Avina JE, Salas-Salazar NA, Molina FJ. 2008. Use of edible coatings to preserve the quality of fruits and vegetables during storage. Stewart Postharvest Review 3, 6.

Oms-Oliu G, Soliva-Fortuny R, Martı'n-Belloso O. 2008. Using polysaccharide-based edible coatings to enhance quality and antioxidant properties of fresh-cut melon. LWT 41, 1862-1870.

Reinoso E, Mittal GS, Lim LT. 2008. Influence of whey protein composite coatings on plum (Prunus domestica L.) fruit quality. Food and Bioprocess Technology 1, 314-325.

Scalbert A, Manach C, Morand C, Remesy C, Jimenez L. 2005. Dietary polyphenols and the prevention of diseases. Critical Reviews in Food Science and Nutrition 45, 287-306.

Togrul H, Arslan N. 2004. Carboxymethylcellulose from sugar beet pulp cellulose as a hydrophilic polymer in coating of mandarin. Journal of Food Engineering **62**, 271–279.

Tomas-Barberan FA, Espin JC. 2001. Phenolic compounds and related enzymes as determinants of quality in fruits and vegetables. Journal of the Science

of Food and Agriculture 81, 853-876.

Valero D, Diaz-Mula HM, Zapata PJ, Guillen F, Martnez-Romero D, Castillo S, Serrano M. 2013. Effects of alginate edible coating on preserving fruit quality in four plum cultivars during postharvest storage. Postharvest Biology and Technology 77, 1-6.

Zhou R, Mo Y, Li Y, Zhao Y, Zhang G, Hu Y. 2008. Quality and internal characteristics of Huanghua pears (Pyrus pyrifolia Nakai, cv. Huanghua) treated with different kinds of coatings during storage. Postharvest Biology and Technology 49, 171-179.