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Estimation of quality of water of river drino (albania) based on analysis of diatoma

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Abstract

The main objective of this paper is to investigate the genera of diatoms, which were identified in the Drino river in the period 2011-2012 in the Gjirokastrë region. A classification of the quality of the water through the bio-indicator diatoms was also carried out. The study area included 5 sampling sites along the Drino River. In total, 116 taxa of diatoms within 33 different genera were found.. There were 59 bio-indicator species. Detailed floristic analysis of the diatoms' flora has not been conducted before in these rivers. The presence of polysaprobic species in the Drino river, such as, *Gomphonema parvulum*, *G.olivaceum*, *Nitzschia palea*, is a clear indication of the pollution of waters coming from a variety of source. According to the bio-indicator saprobic species investigated the waters in the Drino river is classified as II class of bonity, respectively as mesosaprob. Based on this estimation and on the use of the Drino river waters by the local residents, the physical-chemical and microbiological study of these waters in the future is of great interest.

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Introduction

Water is a prerequisite for life and a key factor in virtually all human economic activities. Freshwater rivers are crucial sources of water and it is necessary to study, protect, and improve their ecological status (Zalewski, 2000). Self-purification of water bodies depends on marine life of which algae serve as the primary producers and form the basis of the food chain. In addition to this, algae diversity is a clear indicator of the ecological and hygienic (health) status of the river as well as of the self-purification capabilities (Janauer and Dokulil, 2006). Our approach is based on the bio-indication of algal habitats, which includes not only the biodiversity but also the density of each bio-indicator species. The assessment of aquatic ecosystems is based on algal indicators which are widely used (WFD, 2000), in particular, the assessment of water quality based on the saprobity indicator, obtained from the analysis of algal communities, (Sumita, 1986; Watanabe *et al.*, 1986), but their capabilities are not yet fully understood.

Albania is a country rich in waters, which being near urban centres are under constant risk of being polluted by household and industrial waste. These waters are rich in diatoms many species of which serve as bio-indicators due to their capabilities.

The study of diatoms and the monitoring of the water quality in the main rivers which traverse the coastal lowlands of the Albanian stretch of shoreline of the Adriatic Sea using diatoms has been conducted, (Miho *et al.*, 2006). As regards the Drino river, which is located in the south of Albania and empties into the Vjose river, there are no data concerning the diversity of diatoms, the assessment of water quality and the classification of its waters because of the fact that it is not included in the areas which are equipped with monitoring stations. The river originates in the northwestern part of the Ioannina, in Greece then flows through urban areas. For this reason during the last decades, it has turned into a large reservoir of various pollutants. Consequently, various

considerable modifications in the hydro-chemical composition of the water and of the algae community are taking place. Periphytic diatoms are an excellent indicator of the ecological condition of rivers and streams, due to their ability to react fast to changes in nutrient concentration. Rivers abound with diatoms which are primary producers and are found in all habitats (Round, 1991). The aim of this study is to identify the diatoms found in the Drinos river waters, which creates a data basis that will enable the determination of the water pollution level on the basis of the presence of diatoms.

Material and methods

The samples were collected from 5 sampling sites along the Drino river during the four seasons: spring 2011, summer 2011, autumn 2012 and winter 2012. Water samples were collected in 500 ml glass bottles, 10 cm beneath the water surface, using standard methods (Hindak, 1978). Conductivity, pH, salts, TDS (Total Dissolved Salts), were measured on site using portable instruments (HACH), O₂ was measured with portable instruments, such as, oxygenometer (Hana Instrument) and nutrients (N, P, Si) were analyzed by standard methods (DEV, 1981). Epilithon was brushed from the stones using a toothbrush and the upper layer of epilithon was drawn up via a vacuum suction system and then pipetted (Sladeckova, 1962). Epiphyton was sampled from the substrate and placed in the plastic bottles. The diatoms were examined using a Leica microscope, with a digital camera Fujifilm, which photographed the algae directly from the sample.

Diatoms cleaning

Cleaning of diatoms' frustules and the preparation of slides and their determination was done according to standard methods (Krammer and Lange-Bertalot, 1986-2001). Diatoms' identification was done according to the keys: *Bacillariophyta*: (Krammer and Lange-Bertalot, 1986, 1988, 1991a, 1991b).

Study area and sampling sites

Sampling sites are located in:

1. Near Hormovë village, located about 20 km from Gjirokastra city.
2. Near Andon Poci village, located about 12 km from Gjirokastra city.
3. The River Bridge which stands at the entrance to the town of Gjirokastrer, it is highly polluted due to human activity.
4. The Bridge of Kordhoca and
5. Near Glina village.

The Drino river is a tributary of the Vjosa river with a length of 84.6 km, and with a catchment area of 1324 km². Many streams, such as, Sotira, Suha, Nimsa and Kardhiq empty into the Drino river.

Many villages, such as, Glina, Suha, Libohova, Lunxheria, Lazarati, Humelica, Kardhiqi, Hormova, the villages of the Dropulli commune, and the town of Gjirokastrer are located along the Drino valley (Fig.1).



Fig. 1. Study area and sampling stations in river Drino (1-Hormove, 2-Andon Poci, 3-River Bridge in Gjirokastrer city, 4- Kordhoca Bridge, 5- Glina Bridge).

Results and discussion

A total of 116 species of diatoms were identified from the quantitative analysis of the samples taken during the four seasons (spring, summer, autumn and winter) (Table 5).

We They are presented according to the seasons in which they were identified: spring 2011, 66 species (Table 1); summer 2011, 91 species (Table 2); winter 2012, 60 species (Table 5) and autumn 2012, 69 species (Table 4). During spring 66 species were identified within 24 genera, where 6 species within the genera *Navicula* and *Gomphonema* were dominant, followed by 5 species within the genera *Cocconeis* and *Nitzschia*, 4 species within the genus

Surirella and 3, 2 or 1 species within the other genera.

The highest number of algae was identified on site 1 during spring (table 1), with 53 species, followed by the third site with 40 species, the second site with 37 species, fifth site with 34 species and fourth site with 28 species. There were 39 bio-indicator species, where 18 species within the genus *betamesosaprob* were dominant, followed by 9 species within the genus *alphamesosaprob*, 6 species within the genus *oligo-betamesosaprob*, 4 species within the genus *beta-alphamesosaprob* and 2 species within the genus *oligosaprob*. 91 species within 30 genera were identified during summer, where 18 species within

the *Nitzschia* genus were dominant, followed by 13 species within the genus *Navicula*, 7 species within the genus *Surirella* and 6 species within the genus

Gomphonema; 4, 3, 2, and 1 species within their respective genera were also identified.

Table 1. Determined diatoma in river Drino during the spring season 2011.

Nr. N66	Total number of diatoma	Level of Saprobity	Localities				
			1	2	3	4	5
1	<i>Achnanthes hungarica</i> (Grunow) Grunow	o	1		1		
2	<i>Amphora lybica</i> Ehrenberg	β		1		1	1
3	<i>A. normani</i> Rabenhorst	o	1		1		
4	<i>Aneumastus stroesei</i> (Ostrup) Mann			1	1		1
5	<i>Cocconeis pediculus</i> Ehrenberg	o- β	1				
6	<i>C. placentula</i> Ehrenberg	β	1	1			
7	<i>C. placentula</i> var. <i>lineata</i> (Ehrenberg) Cleve		3	1	5	3	
8	<i>Caloneis amphisbaena</i> (Cleve)	β - α	1				1
9	<i>Cymbela austriaca</i> (Grunow)	β	3	1	1		
10	<i>C. affinis</i> Kützing	o- β	5	5	3		1
11	<i>C. ventricosa</i> Kützing	β	1		1		1
12	<i>C. minuta</i> Hilse et Rabenhorst		1				
13	<i>C. naviculiformis</i> (Auerswald) Cleve	β	3	1	3		1
14	<i>Craticula cuspidata</i> (Kützing) Mann		3	3		1	
15	<i>Cymatopleura solea</i> (Brébisson) W. Smith	β - α		3	1		1
16	<i>Diatoma elongatum</i> var. <i>tenuis</i> (C. Agardh) Van Heurck	α	1		1		1
17	<i>D. moniliforme</i> Kützing		1	1			
18	<i>D. vulgare</i> Bory	β	3	1	1		1
19	<i>Epithemia addnata</i> (Kützing) Brébisson		1				3
20	<i>Fragilaria capucina</i> Desmazières	o- β	3	1			
21	<i>F. ulna</i> (Nitzsch) Lange-Bertalot		3	1	1		1
22	<i>Gomphonema carolinense</i> Hagelstein				1		1
23	<i>G. grovei</i> M. Schmidt		1		1	1	
24	<i>G. micropus</i> Kützing			1	1		
25	<i>G. minutum</i> (C. Agardh)				1	1	1
26	<i>G. olivaceum</i> (Hornemann) Brébisson	β	5		4		
27	<i>Gyrosigma acuminatum</i> (Kützing) Rabenhorst	β				1	
28	<i>G. scalproides</i> (Rabenhorst) Cleve		1		1		1
29	<i>Hantzschia amphioxys</i> (Ehrenberg) Grunow	α	1				1
30	<i>Luticola geoppertiana</i> (Bleish) Mann		3	3	1		
31	<i>L. mutica</i> (Kützing) D. G. Mann		3	1		1	3
32	<i>Melosira varians</i> Agardh	β	1	3		3	
33	<i>Navicula cryptocephala</i> Kützing	α	3	1	5	1	1
34	<i>N. gracilis</i> Ehrenberg	β - α		1			3
35	<i>N. exigua</i> Muller	β	3		5		
36	<i>N. lanceolata</i> (Agardh) Ehrenberg		1	1		1	
37	<i>N. radiosa</i> Kützing	o- β	1			3	
38	<i>N. rhynchocephala</i> Kützing	α	1	1			3
39	<i>N. viridula</i> (Kützing) Ehrenberg	α	5		5	1	1
40	<i>Nitzschia. acicularis</i> (Kützing) W. Sm.	α	3		1		1
41	<i>N. acula</i> Hantzsch in Rabenhorst	α	1	1		3	
42	<i>N. capitellata</i> Hustedt		3	3	1	1	
43	<i>N. closterium</i> (Ehrenberg) W. Sm.			3		1	1
44	<i>N. constricta</i> (Kützing) Ralfs		1	1	1	1	
45	<i>N. gracilis</i> (Hantzsch)			3	1		
46	<i>N. hungarica</i> (Grunow)	α	5		3	5	
47	<i>N. fonticola</i> Grunow	o- β	1	1	3	5	
48	<i>N. levidensis</i> (W. Smith) Grunow		3			1	1
49	<i>N. litoralis</i> Grunow			1		1	
50	<i>N. palea</i> (Kützing) W. Smith	α	1		1		1
51	<i>N. paleaceae</i> (Grunow)		3	1	1		
52	<i>N. recta</i> Hantzsch	β - α	1	1			1
53	<i>N. stagnorum</i> (Rabenhorst)	β	1	1		3	
54	<i>N. sigmoidea</i> (Nitzsch) W. Smith	β			1	1	1
55	<i>N. termalis</i> var. <i>Minor</i> (Hisle)		1			1	1

56	<i>N.vermicularis</i> (Grunow)	β	1		3	3	
57	<i>Pinnularia microstauron</i> var. <i>Brebisonii</i> (Kützing) Mayer	β	1		1		
58	<i>P.viridiformis</i> (Nitzsch) Ehrenberg		1		1		1
59	<i>Rhoicosphaenia curvata</i> (Grunow)	β	1	1	1		
60	<i>Stauroneis anceps</i> Ehrenberg	β	3	3	1	5	1
61	<i>Surirella ovata</i> Kützing	o- β	5				1
62	<i>S.linearis</i> W.Smith	β	1	1	3	1	
63	<i>S. robusta</i> Ehrenberg		3				1
64	<i>Synedra acus</i> Hustedt		1	1		1	1
65	<i>S. nana</i> Meister				1		1
66	<i>S. ulna</i> Kützing	β	5	1	5		1
Total number of species per locality			53	37	40	28	34

The highest number of algae during summer (table 2) was found on sites 1, 3, 4 with 51 species, followed by site 5 with 40 species, site 2 with 39 species. There were 43 bio-indicator species, where 11 species within the genus betamesosaprob were dominant, followed by 10 species within the genus oligo-betamesosaprob,

9 species within the genus oligosaprob, 6 species within the genus alphamesosaprob, 2 species within the genus xeno-saprob, 1 species within the genus alpha-betamesosaprob, 1 species within the genus xeno-oligosaprob and 1 species within the genus oligo-xenosaprob.

Table 2. Determined diatoma in the river Drino during summer season 2011.

Nr.	Total number of diatoma	Level of Saprobity	LOCALITIES				
			1	2	3	4	5
91							
1	<i>Achnanthes hungarica</i> (Grunow) Grunow	o	1		1		
2	<i>A. coarctata</i> (Brébisson) Grunow	x	1	1			1
3	<i>Achnanthidium minutissimum</i> (Kützing)Czarneck		1		1	1	
4	<i>Amphora lybica</i> Ehrenberg		1	1			1
5	<i>A. normani</i> Rabenhorst	o	1		3	3	
6	<i>Aneumastus stroesei</i> (Ostrup) Mann			1			
7	<i>Cocconeis pediculus</i> Ehrenberg	o- β		1		3	1
8	<i>C. placentula</i> Ehrenberg	β	3				1
9	<i>C. placentula</i> var. <i>lineata</i> (Ehrenberg) Cleve		3	1	5	3	
10	<i>Craticula accomoda</i> (Hustedt) Mann	o- β	1			3	
11	<i>C. cuspidata</i> (Kützing) Mann	o			3	1	1
12	<i>Cyclotella ocellata</i> Pantocsek			1		1	
13	<i>Cymatopleura solea</i> (Brébisson) W.Smith	β			1	1	
14	<i>Cymbella affinis</i> Kützing	o- β	3		3	3	1
15	<i>C. helvetica</i> Kützing	o	3	1	3	3	1
16	<i>C. minuta</i> Hilse ex Rabenhorst		1		3		
17	<i>C. naviculiformis</i> (Auerswald) Cleve	β	3	1	3		1
18	<i>Diatoma ehrenbergii</i> Kützing		3	1	5	3	
19	<i>D. moniliforme</i> Kützing				3		1
20	<i>D. vulgaris</i> Bory	β		1	3	3	
21	<i>Epithemia adnata</i> (Kützing) Brébisson				3		1
22	<i>Fragilaria capucina</i> Desmazières	o- β	3	1			
23	<i>F. ulna</i> (Nitzsch) Lange-Bertalot				1		1
24	<i>F. ulna</i> complex <i>oxyrhynchus</i> Lange-Bertalot		1			1	
25	<i>Frustulia vulgaris</i> (Thwaites) De Toni	o	1	1	1		1
26	<i>Gomphonema carolinense</i> Hagelstein				1		1
27	<i>G.grovei</i> M.Schmidt		1		1	1	
28	<i>G. micropus</i> Kützing			1	1		
29	<i>G. minutum</i> (C.Agardh) C.Agardh				1	1	1
30	<i>G. olivaceum</i> (Hornemann) Brébisson	β	1		1	1	
31	<i>G. parvulum</i> (Kützing) Kützing	β	1	1			1
32	<i>Gyrosigma acuminatum</i> (Kützing)Rabenhorst	β	1		1		

33	<i>G. attenuatum</i> (Kützing) Rabenhorst	β	1	1	1	
34	<i>G. scalpoides</i> (Rabenhorst) Cleve			1	1	
35	<i>Hantzschia amphioxys</i> (Ehrenberg) Grunow	α	1	1		1
36	<i>Hippodonta capitata</i> (Ehrenberg) Lange-Bertalot				1	
37	<i>Luticola goeppertiana</i> (Bleish) Mann		1		1	1
38	<i>L. mutica</i> (Kützing) D.G. Mann			1		1
39	<i>Melosira varians</i> Agardh	β	1	1	1	
40	<i>Meridion circulare</i> (Grev.) C. Agardh	α	1	1		1
41	<i>Navicula capitatoradiata</i> Germain			1	1	1
42	<i>N. cryptotenella</i> Lange-Bertalot			3	3	
43	<i>N. cryptocephala</i> Kützing	α	3	3	1	1
44	<i>N. lanceolata</i> (Agardh) Ehrenberg		3	3	3	
45	<i>N. radiosa</i> Kützing	$\alpha - \beta$		1	3	1
46	<i>N. recens</i> (Lange-Bertalot) Lange-Bertalot	$\alpha - \beta$	1	3	3	1
47	<i>N. rhynchocephala</i> Kützing	α		3		
48	<i>N. species aff radisafallax</i> Lange-Bertalot		1			1
49	<i>N. tripunctata</i> (O.F.Müller) Bory		3		3	1
50	<i>N. trivialis</i> Lange-Bertalot		3	3		
51	<i>N. tuscula</i> Ehrenberg	$\alpha - \beta$		1	1	1
52	<i>N. viridula</i> (Kützing) Ehrenberg	α		1	5	3
53	<i>N. viridula var. rostellata</i> (Kützing) Cleve		3	3		
54	<i>Nitzschia acula</i> Hantzsch in Rabenhorst			1		1
55	<i>N. acicularis</i> (Kützing) W. Sm.			1	1	
56	<i>N. capitellata</i> Hustedt		3	3		
57	<i>N. closterium</i> (Ehrenberg) W.Smit			3	1	1
58	<i>N. constricta</i> (Kützing) Ralfs		1		3	
59	<i>N. commutata</i> Grunow			1	1	
60	<i>N. dissipata</i> (Kützing) Grunow	$\alpha - \beta$	1	1	3	3
61	<i>N. elegantula</i> Grunow in Van Heurck			1	1	3
62	<i>N. eglei</i> Lange-Bertalot			1		1
63	<i>N. filiformis</i> (W. Sm.) Van Heurck	$\alpha - \beta$			1	1
64	<i>N. fonticola</i> Grunow	$\alpha - \beta$	1	1	3	
65	<i>N. levidensis</i> (W.Smith) Grunow		3			1
66	<i>N. litoralis</i> Grunow			1	1	
67	<i>N. linearis</i> (Agardh) W.Smith	$\alpha - \beta$	1			1
68	<i>N. palea</i> (Kützing) W.Smith	α		1	3	
69	<i>N. pusilla</i> Grunow				1	1
70	<i>N. recta</i> Hantzsch	$\beta - \alpha$	1	3		
71	<i>N. sigmoidea</i> (Nitzsch) W.Smith	β		3	3	
72	<i>Pinnularia microstauron</i> (Ehrenberg) Cleve	$\alpha - x$	3	1		1
73	<i>P. microstauron var. brebisonii</i> (Kützing) Mayer	x	3	1		
74	<i>Planothidium ellipticum</i> (Cleve) Round		3	1		
75	<i>P. lanceolatum</i> (Brébisson) Round		3			1
76	<i>Reimeria sinuata</i> (Greg.) Kociolek & Stoermer			1	3	
77	<i>Rhoicosphaenia abbreviata</i> (Agardh) Lange-Bertalot	$x - \alpha$	1	1	1	
78	<i>Sellaphora pupula</i> (Kützing) Mjereschowsky	α		3	1	
79	<i>S. pupula fo. rostrata</i> (Hustedt) Bukhtiyarova		1	3		
80	<i>Stauroneis smithii</i> Grunow	$x - \alpha$	1		1	1
81	<i>S. anceps</i> Ehrenberg		1	1		
82	<i>Surirella angusta</i> Kützing	α		1		1
83	<i>S. brebisonii var. kuetzingii</i> Krammer & Lange-Bertalot			1	3	
84	<i>S. linearis</i> W.Smith	β		3	3	1
85	<i>S. minuta</i> Brébisson in Kützing	α	1	3	3	
86	<i>S. ovalis</i> Brébisson	α	1	1		1
87	<i>S. patella</i> Kützing			1	1	
88	<i>S. robusta</i> Ehrenberg			1	1	
89	<i>Synedra acus</i> Hustedt		1	1	1	1
90	<i>S. nana</i> Meister			1		
91	<i>S. ulna</i> Kützing		1		1	1
Total number of species per locality			51	39	51	40

During autumn 69 species were identified within 29 genera, where 12 species within the genus *Nitzschia* were dominant, followed by 9 species within the genus *Navicula*, 4 species within the genus *Surirella*, and 3, 2, 1 species were identified within the other genera. The highest number of algae during autumn was found (table 3) on site 1 with 42 species, site 2 with 38 species, site 4 with 36 species, site 5 with 35 species, site 3 with 30 species. There were 34 bio-indicator species, where 9 species within the genus oligosaprob and genus betamesosaprob were

dominant, followed by 7 species within the genus oligo-betamesosaprob, 4 species within the alphamesosaprob, 3 species within the xeno-oligosaprob, 1 species within the genus xeno-saprob and genus beta-alphamesosaprob. 60 species within 24 genera were identified during winter, where 6 species within the genus *Navicula* were dominant, followed by 5 species within the genus *Nitzschia*, 5 species within the genus *Cocconeis*, 4 species within the genus *Surirella*, 3, 2, 1 species were identified within the other genera.

Table 3. Determined diatom in the river Drino during autumn season 2012.

Nr. 69	Total number of diatoma	Level of Saprobity	LOCALITIES				
			1	2	3	4	5
1	<i>Achnanthes hungarica</i> (Grunow) Grunow	o	1		1		
2	<i>A. coarctata</i> (Brébisson) Grunow	o	1	1			1
3	<i>Achnanthidium minutissimum</i> (Kützinger) Czarneck		1			1	
4	<i>Amphora lybica</i> Ehrenberg		1	1			1
5	<i>A. normani</i> Rabenhorst	o	1		1	3	
6	<i>Aneumastus stroesei</i> (Ostrup) Mann			1			
7	<i>Cocconeis pediculus</i> Ehrenberg	o- β		1		3	1
8	<i>C. placentula</i> Ehrenberg	β	3				1
9	<i>Craticula accomoda</i> (Hustedt) Mann	o- β	1			3	
10	<i>C. cuspidata</i> (Kützinger) Mann	o			1	1	1
11	<i>Cyclotella ocellata</i> Pantocsek			1		1	
12	<i>Cymatopleura solea</i> (Brébisson) W.Smith	β			1	1	
13	<i>Cymbella affinis</i> Kützinger	o- β	3		3	3	1
14	<i>C. helvetica</i> Kützinger	o	3	1	1	1	1
15	<i>C. naviculiformis</i> (Auerswald) Cleve	β	3	1	3		1
16	<i>Diatoma ehrenbergii</i> Kützinger		3	1		3	
17	<i>D. moniliforme</i> Kützinger				3		1
18	<i>D. vulgaris</i> Bory	β		1	3	3	
19	<i>Epithemia adnata</i> (Kützinger) Brébisson				3		1
20	<i>Fragilaria ulna</i> (Nitzsch) Lange-Bertalot				1		1
21	<i>Frustulia vulgaris</i> (Thwaites) De Toni	o	1	1	1		1
22	<i>Gomphonema carolinense</i> Hagelstein			1	1		1
23	<i>G.grovei</i> M.Schmidt		1			1	
24	<i>G. parvulum</i> (Kützinger) Kützinger	β	1	1			1
25	<i>Gyrosigma acuminatum</i> (Kützinger) Rabenhorst	β	1		1		
26	<i>G. attenuatum</i> (Kützinger) Rabenhorst	β	1			1	
27	<i>G. scalpoides</i> (Rabenhorst) Cleve				1	1	
28	<i>Hantzschia amphioxys</i> (Ehrenberg) Grunow	α	1	1			1
29	<i>Luticola goeppertiana</i> (Bleish) Mann		1			1	1
30	<i>Melosira varians</i> Agardh	β	1	1	1	1	
31	<i>Meridion circulare</i> (Grev.) C. Agardh	o	1		1		1
32	<i>Navicula capitatoradiata</i> Germain			1		1	1
33	<i>N. cryptocephala</i> Kützinger	α	3	3		1	1
34	<i>N. lanceolata</i> (Agardh) Ehrenberg		3			3	
35	<i>N. radiosa</i> Kützinger	o- β		1		3	1
36	<i>N. rhynchocephala</i> Kützinger	α			3		

37	<i>N. tripunctata</i> (O.F.Müller) Bory		3	1		3	1
38	<i>N. trivialis</i> Lange-Bertalot		3		3		
39	<i>N. viridula</i> (Kützinger) Ehrenberg	α		1		3	
40	<i>N. viridula</i> var. <i>rostellata</i> (Kützinger) Cleve		3		3		
41	<i>Nitzschia acula</i> Hantzsch in Rabenhorst			1			1
42	<i>N. acicularis</i> (Kützinger) W. Sm.			1		1	
43	<i>N. commutata</i> Grunow				1	1	
44	<i>N. dissipata</i> (Kützinger) Grunow	α - β	1	1	3	3	1
45	<i>N. elegantula</i> Grunow in Van Heurck			1		3	1
46	<i>N. eglei</i> Lange-Bertalot			1			1
47	<i>N. fonticola</i> Grunow	α - β	1	1	3		1
48	<i>N. levidensis</i> (W.Smith) Grunow		3				
49	<i>N. litoralis</i> Grunow			1		1	
50	<i>N. linearis</i> (Agardh) W.Smith	α - β	1	1			1
51	<i>N. recta</i> Hantzsch	β - α	1		3		
52	<i>N. sigmoidea</i> (Nitzsch) W.Smith	β		3		3	
53	<i>Pinnularia microstauron</i> (Ehrenberg) Cleve	α -x	3	1			1
54	<i>P. microstauron</i> var. <i>Brebissonii</i> (Kützinger) Mayer	x	3	1			
55	<i>Planothidium ellipticum</i> (Cleve) Round		3	1			
56	<i>P. lanceolatum</i> (Brébisson) Round		3				1
57	<i>Reimeria sinuata</i> (Greg.) Kociolek & Stoermer			1		3	
58	<i>Rhoicosphaenia abbreviata</i> (Agardh) Lange-Bertalot	x-o	1		1	1	
59	<i>Sellaphora pupula</i> (Kützinger) Mjereschowsky	α		1	3	1	
60	<i>S. pupula</i> fo. <i>rostrata</i> (Hustedt) Bukhtiyarova		1				
61	<i>Stauroneis smithii</i> Grunow	x-o	1	1		1	1
62	<i>S. anceps</i> Ehrenberg		1		1		
63	<i>Surirella angusta</i> Kützinger	o		1			1
64	<i>S. brebissonii</i> var. <i>kuetzingii</i> Krammer & Lange-Bertalot			1		3	
65	<i>S. ovalis</i> Brébisson	o	1		1		1
66	<i>S. robusta</i> Ehrenberg				1	1	
67	<i>Synedra acus</i> Hustedt		1	1		1	1
68	<i>S. nana</i> Meister			1	1		
69	<i>S. ulna</i> Kützinger		1			1	1
Total number of species per locality			42	38	30	36	3
							5

The highest number of algae during winter (table 4) was identified on site 2 with 27 species, site 1 with 26 species, site 4 with 25 species, site 3 with 18 species, site 5 with 15 species. There were 21 bio-indicator species, where 9 species within the genus

betamesosaprob were dominant, followed by 5 species within the genus oligosaprob, 4 species within the genus oligo-betamesosaprob, 2 species within the genus alphamesosaprob, 1 species within the genus beta-alphamesosaprob.

Table 4. Determined diatom in the river Drino during winter season 2012.

Nr.	Total number of diatoma		LOCALITIES				
			1	2	3	4	5
60		Level of Saprobity					
1	<i>Achnanthes hungarica</i> (Grunow)	o	1				
2	<i>A. clevei</i> var. <i>clevei</i> Grunow		1			1	
3	<i>A. ventralis</i> (Krasske) Lange-Bertalot		1	1	1		1
4	<i>Amphora lybica</i> Ehrenberg		1	3		3	
5	<i>A. normani</i> Rabenhorst	o		1		1	1
6	<i>Cocconeis pediculus</i> Ehrenberg	α - β	3	3		3	
7	<i>C. placentula</i> Ehrenberg	β	3	5		1	

8	<i>C.placentula</i> var. <i>lineata</i> (Ehrenberg) Cleve		3		1		1	
9	<i>Craticula accomoda</i> (Hustedt) Mann			1		1		
10	<i>Centronella reichelti</i> (Voigt)		1			1		
11	<i>Cyclotella ocellata</i> Pantocsek			1	1			
12	<i>Cymatopleura solea</i> (Brébisson) W.Smith	β- α	1	1		1		
13	<i>Cymbella affinis</i> Kützing	o- β	5	3		3		
14	<i>C. helvetica</i> Kützing		1	1			1	
15	<i>Diatoma ehrenbergii</i> Kützing		3	1		1		
16	<i>D. moniliforme</i> Kützing		1	3	1	1		
17	<i>D. vulgaris</i> Bory	β	3	3		3		
18	<i>Epithemia adnata</i> (Kützing) Brébisson		1			1		
19	<i>F. ulna</i> (Nitzsch) Lange-Bertalot		1	3		1		
20	<i>F.ulna</i> complex <i>oxyrhynchus</i> Lange-Bertalot			1	1			
21	<i>Frustulia vulgaris</i> (Thwaites) De Toni	o	1			1	1	
22	<i>Gomphonema microporus</i> Kützing		1	1				
23	<i>G.minutum</i> (C.Agardh)		1	3		1		
24	<i>G.olivaceum</i> (Hornemann) Brebisson	β	1	1		1		
25	<i>G.longiceps</i> var. <i>subclavatum</i> Grunow		1			1		
26	<i>G.parvaulum</i> var. <i>exlissimum</i> Grunow		1				1	
27	<i>G.tenue</i> Fricke		1	1	1			
28	<i>Gyrosigma acuminatum</i> (Kützing)	β	1	1		1		
29	<i>G. attenuatum</i> (Kützing) Rabenhorst	β	1	1	1	1		
30	<i>G. scalproides</i> (Rabenhorst) Cleve		1	1				
31	<i>Hantzschia amphioxys</i> (Ehrenberg) Grunow	α	1	1		1		
32	<i>H.elongata</i> (Hantzch) Grunow					1		
33	<i>Luticola goeppertiana</i> (Bleish) Mann		1		1	1	1	
34	<i>Melosira varians</i> Agardh	β	1			1		
35	<i>Meridion circulare</i> (Grev.) C. Agardh	o	1	1		1		
36	<i>Navicula cryptotenella</i> Lange-Bertalot			3	1	3		
37	<i>N. lanceolata</i> (Agardh) Ehrenberg		3	5		3		
38	<i>N. radiosa</i> Kützing	o- β	3	3	1		1	
39	<i>N. tripunctata</i> (O.F.Müller) Bory		3	1		3	1	
40	<i>N. trivialis</i> Lange-Bertalot		1			3		
41	<i>N. viridula</i> var. <i>rostellata</i> (Kützing) Cleve		3	3			1	
42	<i>Nitzschia acula</i> Hantzsch in Rabenhorst	α	3	3	1	3		
43	<i>N.capitellata</i> Hustedt			3		3		
44	<i>N.constricta</i> (Kützing)		3	3		3		
45	<i>N. dissipata</i> (Kützing) Grunow	o- β	1		1	3		
46	<i>N.exilis</i> Kützing			1			1	
47	<i>Pinnularia microstauron</i> (Ehrenberg) Cleve	o			1	1		
48	<i>P.microstauron</i> var. <i>Brebisonii</i> (Kützing) Mayer		3	3	1	3		
49	<i>Planothidium ellipticum</i> (Cleve) Round					1		
50	<i>P. lanceolatum</i> (Brébisson) Round		1	3		1		
51	<i>Reimeria sinuata</i> (Greg.) Kociolek & Stoermer		3			3	1	
52	<i>Rhoicosphaenia abbreviata</i> (Agardh)Lange-Bertalot	β	1	1	1	1		
53	<i>Sellaphora pupula</i> (Kützing)		3	3		1		
54	<i>S. pupula</i> fo. <i>rostrata</i> (Hustedt) Bukhtiyarova		1		1	1	1	
55	<i>Stauroneis smithii</i> Grunow			1		1	1	
56	<i>Surirella angusta</i> Kützing	β	3	3		3		
57	<i>S. brebissonii</i> var. <i>kuetzingii</i> Krammer			3	1	3		
58	<i>S.linearis</i> W.Smith		1				1	
59	<i>S.minuta</i> Brébisson in Kützing			1		1		
60	<i>Synedra ulna</i> (Nitzsch) Ehrenberg	β	3	5	1	3		
Total number of species per locality			21	47	42	18	46	16

Benthic algal communities are usually rich species and therefore provide a lot of information which is ideal for the monitoring of the environment because

of their relatively short life cycle, allowing for a rapid response to environmental changes. At the same time they also reveal the cumulative effect that the

environmental changes has had over a longer time period (Janauer and Dokulil, 2006). Because of this, bio-indicator algae are included in the EC-Water Framework Directive (WFD, 2000) for the

assessment and monitoring of water quality. The complete database includes taxa of 116 diatoms, but only 21 diatoms species within 15 genera are found in all seasons (Table 2).

Table 5. Determined diatom in the river Drino during four season: spring, summer, autumn and winter.

Nr.	Total number of diatoma	Level of Saprobity	Season			
			spring	summer	autumn	winter
1	<i>Achnanthes hungarica</i> (Grunow) Grunow	o	+	+	+	+
2	<i>A.cleveii</i> var. <i>cleveii</i> Grunow					+
3	<i>A. coarctata</i> (Brébisson) Grunow	x		+	+	
4	<i>A.ventralis</i> (Krasske) Lange-Bertalot					+
5	<i>Achnanthidium minutissimum</i> (Kützinger) Czarneck			+	+	
6	<i>Amphora lybica</i> Ehrenberg	β	+	+	+	+
7	<i>A. normani</i> Rabenhorst	o	+	+	+	+
8	<i>Aneumastus stroesei</i> (Ostrup) Mann		+	+	+	
9	<i>Caloneis amphisbaena</i> (Cleve	$\beta - \alpha$	+			
10	<i>Cocconeis pediculus</i> Ehrenberg	o- β	+	+	+	+
11	<i>C. placentula</i> Ehrenberg	β	+	+	+	+
12	<i>C.placentula</i> var. <i>lineata</i> (Ehrenberg) Cleve	β	+	+		+
13	<i>Craticula accomoda</i> (Hustedt) Mann	o- β		+	+	+
14	<i>C. cuspidata</i> (Kützinger) Mann	o	+	+	+	
15	<i>Centronella reichelti</i> (Voigt)					+
16	<i>Cyclotella ocellata</i> Pantocsek			+	+	+
17	<i>Cymatopleura solea</i> (Brébisson) W.Smith	$\beta - \alpha$	+	+	+	+
18	<i>Cymbella affinis</i> Kützinger	o- β	+	+	+	+
19	<i>C. austriaca</i> (Grunow)	β	+			
20	<i>C. helvetica</i> Kützinger	o		+	+	+
21	<i>C.minuta</i> Hilse ex Rabenhorst		+	+		
22	<i>C. naviculiformis</i> (Auerswald) Cleve	β	+	+	+	
23	<i>C.ventricosa</i> Kützinger	β	+			
24	<i>Diatoma elongatum</i> var. <i>tenuis</i> (C. Agardh) Van Heurck	α	+			
25	<i>D. ehrenbergii</i> Kützinger			+	+	+
26	<i>D. moniliforme</i> Kützinger		+	+	+	+
27	<i>D. vulgaris</i> Bory	β	+	+	+	+
28	<i>Epithemia adnata</i> (Kützinger) Brébisson		+	+	+	+
29	<i>Fragilaria capucina</i> Desmazières	o- β	+	+		
30	<i>F. ulna</i> (Nitzsch) Lange-Bertalot		+	+	+	+
31	<i>F.ulna</i> complex <i>oxyrhynchus</i> Lange-Bertalot			+		+
32	<i>Frustulia vulgaris</i> (Thwaites) De Toni	o		+	+	+
33	<i>Gomphonema carolinense</i> Hagelstein		+	+	+	
34	<i>G.grovei</i> M.Schmidt		+	+	+	
35	<i>G.microporus</i> Kützinger					+
36	<i>G.minutum</i> (C.Agardh)		+	+		+
37	<i>G.olivaceum</i> Hornemann) Brebisson	β	+	+		+
38	<i>G.longiceps</i> var. <i>subclavatum</i> Grunow					+
39	<i>G.parvulum</i> (Kützinger) Kützinger	β		+	+	
40	<i>G.parvulum</i> var. <i>exlissimum</i> Grunow					+
41	<i>G.pseudoaugur</i> Lange-Bertalot		+	+		
42	<i>G.tenue</i> Fricke					+
43	<i>Gyrosigma acuminatum</i> (Kützinger) Rabenhorst	β	+	+	+	+
44	<i>G. attenuatum</i> (Kützinger) Rabenhorst	β		+	+	+
45	<i>G. scalpoides</i> (Rabenhorst) Cleve		+	+	+	+
46	<i>Hantzschia amphioxys</i> (Ehrenberg) Grunow	α	+	+	+	+
47	<i>H.elongata</i> (Hantzsch) Grunow					+
48	<i>Hippodonta capitata</i> (Ehrenberg) Lange-Bertalot			+		
49	<i>Luticola geoppertiana</i> (Bleish) Mann		+	+	+	+
50	<i>L.mutica</i> (Kützinger) D.G.Mann		+	+		
51	<i>Melosira varians</i> Agardh	β	+	+	+	+
52	<i>Meridion circulare</i> (Grev.) C. Agardh	o		+	+	+
53	<i>Navicula capitatoradiata</i> Germain			+	+	
54	<i>N. cryptocephala</i> Kützinger	α	+	+	+	
55	<i>N.cryptotenella</i> Lange-Bertalot			+		+

56	<i>N.exigua</i> (Muller)	β	+			
57	<i>N. lanceolata</i> (Agardh) Ehrenberg		+	+	+	+
58	<i>N.gracilis</i> Ehrenberg	$\beta - \alpha$	+			
59	<i>N. radiosa</i> Kützing	$\alpha - \beta$	+	+	+	+
60	<i>N.recens</i> (Lange-Bertalot)			+		
61	<i>N. rhynchocephala</i> Kützing	α	+	+	+	
62	<i>N. recens</i> Lange- Bertalot			+		
63	<i>N. tripunctata</i> (O.F.Müller) Bory			+	+	+
64	<i>N. trivialis</i> Lange-Bertalot			+	+	+
65	<i>N.tuscula</i> Ehrenberg	$\alpha - \beta$		+		
66	<i>N. viridula</i> (Kützing) Ehrenberg	α	+	+	+	
67	<i>N. viridula</i> var. <i>rostellata</i> (Kützing) Cleve			+	+	+
68	<i>Nitzschia acula</i> Hantzsch in Rabenhorst	α	+	+	+	+
69	<i>N. acicularis</i> (Kützing) W. Sm.	α	+	+	+	
70	<i>N.capitellata</i> Hustedt		+	+		+
71	<i>N.closterium</i> (Ehrenberg) W.Sm.		+	+		
72	<i>N. commutata</i> Grunow			+	+	
73	<i>N.constricta</i> (Kützing) Ralfs		+	+		+
74	<i>N. dissipata</i> (Kützing) Grunow	$\alpha - \beta$		+	+	+
75	<i>N. elegantula</i> Grunow in Van Heurck			+	+	
76	<i>N. eglei</i> Lange-Bertalot			+	+	
77	<i>N.exilis</i> Kützing					+
78	<i>N.filiformis</i> (W.Sm.) Van Heurck	$\alpha - \beta$		+		
79	<i>N.fonticola</i> Grunow	$\alpha - \beta$	+	+	+	
80	<i>N.gracilis</i> Hantzsch		+			
81	<i>N.hungarica</i> (Grunow)	α	+			
82	<i>N. levidensis</i> (W.Smith) Grunow		+	+	+	
83	<i>N. litoralis</i> Grunow		+	+	+	
84	<i>N. linearis</i> (Agardh) W.Smith	$\alpha - \beta$		+	+	
85	<i>N.palea</i> (Kützing) W.Smith	α	+	+		
86	<i>N.paleaceae</i> Grunow		+			
87	<i>N.pusilla</i> Grunow			+		
88	<i>N. recta</i> Hantzsch	$\beta - \alpha$	+	+	+	
89	<i>N. sigmoidea</i> (Nitzsch) W.Smith	β	+	+	+	
90	<i>N.stagnorum</i> (Rabenhorst)	β	+			
91	<i>N.termalis</i> var. <i>minor</i> (Hisle)		+			
92	<i>N.umbonata</i> (Ehrenberg) Lange-Bertalot					
93	<i>N.vermicularis</i> (Grunow)	β	+			
94	<i>Pinnularia microstauron</i> (Ehrenberg) Cleve	β	+	+	+	+
95	<i>P.microstauron</i> var. <i>brebissonii</i> (Kützing) Mayer	$\alpha - x$		+	+	+
96	<i>P.viridiformis</i> (Nitzsch) Ehrenberg		+			
97	<i>Planothidium ellipticum</i> (Cleve) Round			+	+	+
98	<i>P. lanceolatum</i> (Brébisson) Round			+	+	+
99	<i>Reimeria sinuata</i> (Greg.) Kociolek & Stoermer			+	+	+
100	<i>Rhoicosphaenia abbreviata</i> (Agardh) Lange-Bertalot	β		+	+	+
101	<i>R.curvata</i> Grunow	β	+			
102	<i>Sellaphora pupula</i> (Kützing) Mjereschowsky	α		+	+	+
103	<i>S. pupula</i> fo. <i>rostrata</i> (Hustedt) Bukhtiyarova			+	+	+
104	<i>Stauroneis smithii</i> Grunow	$x - \alpha$		+	+	+
105	<i>S. anceps</i> Ehrenberg	β	+	+	+	
106	<i>Surirella angusta</i> Kützing	β		+	+	+
107	<i>S. brebissonii</i> var. <i>kuetzingii</i> Krammer & Lange-Bertalot			+	+	+
108	<i>S.linearis</i> W.Smith	β	+	+		+
109	<i>S.minuta</i> Brébisson in Kützing	α		+		+
110	<i>S. ovalis</i> Brébisson	α		+	+	
111	<i>S.ovata</i> Kützing	$\alpha - \beta$	+			
112	<i>S.patella</i> Kützing			+		
113	<i>S. robusta</i> Ehrenberg		+	+	+	
114	<i>Synedra acus</i> Hustedt		+	+	+	
115	<i>S. nana</i> Meister		+	+	+	
116	<i>S. ulna</i> Kützing	β	+	+	+	=
Total number of species per season			66	91	69	60

The most common species were *Cocconeis placentula* var. *lineata*, *Nitzschia acula*, *Gomphonema parvulum*, *Rhoicosphaenia abbreviata*, *Naviculales*

lanceolata, *Nitzschia dissipata*, *Naviculales radiosa* and *Naviculales gregaria*. These diatoms are known as cosmopolitan, widely distributed in inland waters,

and considered as indicators of eutrophic conditions (Van Dam *et al.*, 1994). In this study were analyzed the 'ecological' species which are responsible for the total amount of Phosphorus (TP), which is, a response to the presence of other species as well. The biological species usually have symmetrical unimodal distribution (concentrated in a single site) along the environmental gradient (Whittaker, 1967), at least those which are unaffected by the gradient factor at the source.

In our study during the period 2011-2012 were identified 116 species of diatoms within 36 genera; 26 species within the genus *Nitzschia* were dominant, while the other genera are represented with a smaller number of species; 15 species within the genus *Navicula*, 10 species within the genus *Gomphonema*, 8 species within the genus *Surirella*, 6 species within the genus *Cymbella*. Out of 116 identified species, 59 are bio-indicator species which belong to the xenosaprobic till alphamesosaprobic level of saprobity. 23 species belong to the Betamesosaprobic level, and dominate the other level of saprobity. The other levels of saprobity have the lowest number of bio indicator species, such as, oligo-betamesosaprobic and alpha- mesosaprobic level of saprobity have 10 species, oligosaprobic level of saprobity has 8 species, beta-alphamesosaprobic level of saprobity has 5 species, xeno-oligosaprobic level of saprobity has 2 species, xenosaprobic level of saprobity has 1 species.

Conclusion

During the study period (2011-2012) were identified 116 algae species of plankton and periphyton within 36 genera. 26 species within the genus *Nitzschia* were the most dominant, followed by 15 species within the genus *Navicula*, 10 species within the genus *Gomphonema*, 8 species within the genus *Surirella*, 6 species within the genus *Cymbella*, 4 species within the genus *Achnanthes*, *Cocconeis*, *Fragilaria*, *Gyrosigma*, *Pinularia* and *Synedra* were identified as having 3 taxa, whereas the other genera were identified with one or two species.. Furthermore, there is a high diversity of diatoms in each season.

There are 59 bio-indicator species. The betamesosaprobic bio-indicator (20 bio-indicator species) were dominant. 11 species which belong to oligo-betamesaprob (o- β) level of saprobity were found, followed by alphamesosaprob bio-indicator (10 species), oligosaprob bio-indicator (9 species), beta-alphamesosaprob bio-indicator (4 species) and alfa-betamesosaprob bio-indicator (1 species).

The presence of bio-indicator diatoms shows that the Drinos river water pollution level is considerable. This water pollution is caused by urban residues. Appreciating the uses of this river water for different needs and in order to get more thorough results, its physical-chemical and microbiological study is necessary. We No bio-indicator species which belong to polisaprob (ρ) level of saprobity were not found. According to the bioindicators species, investigate waters classified in second-II-class of bonity respectively at beta-mesosaprob level.

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