



## RESEARCH PAPER

## OPEN ACCESS

## Sequential optimization of *aspergillus flavus* kla-80 for the production of naringinase using stastical experimental designs

Ananda Sindhe, K. Lingappa\*

*Department of Microbiology, Gulbarga University, Kalaburagi, Karnataka, India*

**Key words:** Naringin, Naringenin, *Aspergillus flavus*, Response surface methodology, Central composite design

<http://dx.doi.org/10.12692/ijb/22.5.149-159>

Article published on May 18, 2023

### Abstract

Naringin is a flavanone bitter glycoside which is found in flowers and fruit of the grapefruit, which on further hydrolysis yields naringenin and glucose. As naringin is responsible for the bitterness of the fruit, to neutralize the bitterness by eco-friendly method, microbial method is performed by using naringinase enzyme. In the present study naringinase producing strain *Aspergillus flavus* KLA-80 which was previously isolated from soil sample of Kalyan-Karnataka region, India, and deposited in NCFCC with an accession number of OQ152018 which was maintained in the lab conditions was used for optimization studies. Further to increase the activity of enzyme one factor at a time approach was carried out where the activity was found to be increased from 559 to 720U/mL in a temp of 35°C, pH 4.5, carbon source 1.25% and nitrogen source of 0.15% at an agitation speed of 120rpm respectively. Eventually the production of naringinase from *Aspergillus niger* is optimized by Response Surface Methodology i.e., by CCD (Central Composite Design) method considering four factors at a time (pH, Agitation speed, Galactose, Peptone) as the variables. Based on the statistical method of optimization it is found that the enzyme activity was increased upto 20.41%.

\* Corresponding Author: K. Lingappa ✉ [lingappak1@rediffmail.com](mailto:lingappak1@rediffmail.com)

## Introduction

In the present scenario, fruit juices are gaining importance because of their freshness and its property to increase the immunity of the body. Out of many major types, citrus family fruits are known to be one of the world famous crop as they are rich in particular vitamins, minerals and bioactive compounds. In India, different varieties of citrus fruits grown including mosambi, kinnow, oranges as they belongs to the family of Rutaceae and Plantae kingdom (Tran *et al.*, 2020). As the production of citrus fruit stands at 3rd position after Bannana and Mangao as per the report of National Horticulture for the 2nd advance estimate of the year 2019-20. As these cirtus fruits are rich in phytochemicals which mainly contains naturally occurring biological active compounds including lipids, phenols, alkaloids, diterpens etc (Mishra and kar, 2003). Flavanoids are the know to be a major secondary metabolities of the plants (Chen *et al.*, 2016) as These flavanoids are found to have a natural anti-oxidant properties which surge the body's defense mechanism, which was found to be one of the riverting hail in the recent medicine (Adami *et al.*, 2022). A precise of around 4000 flavonoids has known to be discovered in fruits, herbs and vegetables (Cook 1996). Among these flavnoids, Naringin is a flavanone glycoside which is known to present in citrus and grape fruits, as it bears a recognizable bitter taste to the fruits (Alam *et al.*, 2014). It is a compratively water soluble bitter compound present in fruit membranes namely Albedo and predominatly present in peels followed by seeds (Manish and Veeya, 2022). As these bitterness inturn causes the reduction to quality and the commercial value of the fruits thereby making it as a presistent problem in recent times (Zhang *et al.*, 2022).To make citrus fruit for consumer acceptibilty, the bitterness should be reduced below its threshold level by various physio-chemical methods which has been flourished (Munoz *et al.*, 2022). In physio-chemical method various different absorbents has been tested in which Synthetic natural resin showed highest efficieny (Riberio *et al.*, 2002) and also in event to this ion-exchange resins are also showed reduction in bitterness thereby maintaing the taste and acidity of the juice (Kranz *et al.*, 2010). But one of

the drawbacks of these methods is that it alter the chemical reactions which leads to the mislaying of certain nutrients and also because of it's prohibitive costs which are not recomandable nowadays (Lee and kim, 2003). In order to overcome the drawbacks of these physico-chemical methods, Biological method of conversion of bitter compound is scruntinized i.e., by enzymatic method using naringinase enzyme which has a multi-enzyme activity. (Puri M, 2011). Naringinase was also used in the enhancement of aroma in wine and also in the preparartion of natural glycoside hydrolysed product (Puri *et al.*, 2009).

In order to optimize the enzyme production, classical methods which are used in the early days are found to be a tedious time consuming and also not recomended if larger number of varibales are involved (Sandom 1958).In this regard to enhnace the production optimization of the media is done by RSM method, which is a powerfull and efficient mathematical approach applied for the optimization of fermentation process which also includes medium optimization for enzyme production (Dutt *et al.*, 2009). CCD method is one of the integral part of RSM. As it is a set of statistical techniques and mathematical tools which is applied in the progression of appropriate functional relationship between the response and relatable input variables (Amiri *et al.*, 2019).

As this method has several advantages like reducing the number of experiments, predicting the response, checking the adequacy of the method, requiries shorter time and more economical compared to the classical method (Oskouie *et al.*, 2008). It also helps to study the interaction effects between the parameters and also in determining the optimum conditions are the min advnatages of the RSM (Jitolis *et al.*, 2021).

In the present investigation, submerged fermentation of *Aspergillus flavus* KLA-80 was done for the production of naringinase by one factor at a time method of optimization follwed by Cental Composite Desgin where the synchronous effect of 4 different variables were analysed for the optimal naringinase production.

## Materials and methods

### *Micro-Organism and Culture Media*

An isolate *Aspergillus Flavus* KLA-80 was maintained in our laboratory at 4°C, which was investigated in this study for the naringinase production, was isolated from the soil of Kalyan-Karnataka region (India) which is identified and deposited in the NCCCI, and accession number of OQ152018 is obtained. The strain was maintained in the agar plate containing (g/l<sup>-1</sup>) following components with slight changes as prescribed by (Mukund *et al.* 2013) NH<sub>4</sub>NO<sub>3</sub>,5gm; KCl,0.2gm; KH<sub>2</sub>PO<sub>4</sub>,0.4gm; FeSO<sub>4</sub>.7H<sub>2</sub>O,0.01gm; ZnSO<sub>4</sub>,0.01gm; MnSO<sub>4</sub>,0.01,gm; MgSO<sub>4</sub>,0.2gm; Agar,15gm; Naringin,1gm; at pH 4.5.

### *Preparation of spore suspension*

The spore suspension of the isolated culture KLA-80 was developed from the 7 days old culture grown on Naringin Agar Media in a 10ml of sterile water containing 0.01% of Tween-80 by suspending the spores with a sterile loop (Shivalee *et al.*, 2018).

### *Quantitative assay for naringinase activity*

The activity of Naringinase was done by the method as prescribed by (Davis 1947) with some modifications. The fermented broth was allowed to centrifuge at 3000 rpm for 10min using Cryogen Refrigerator centrifuge and the 0.1ul of supernatant was allowed to mix with 0.9ul of naringin (0.05% of naringin dissolved in 0.1M sodium acetate buffer pH-4) and this mixture is allowed to incubate at 50°C for 60min after incubation 0.1ul of incubated solution is added with 5ml of 90% of diethylene glycol and the 0.1ul of 4N NaOH is added to stop the reaction and the reaction was allowed to incubate at 20 min for the color development and finally, the O.D<sub>420</sub> of residual naringin was measured by using UV-Visible spectrophotometer. All the data was measured three times in parallel. One unit (U) of naringinase activity was defined as the amount of enzyme that could hydrolyze 1μmol of naringin/ min at the assay conditions (Puri and Kalra, 2005).

### *Effect of physical parameters*

Some of the parameters like (temperature, pH, agitation speed, inoculum size and age (days) were

studied to find its influence on the naringinase production in the basal media used.

The effect of temperature on the basal media was assessed by the incubating the reaction mixture at different temperature such as 25,30,35,40,45,50. Naringinase activity was examined for every 24 hrs for the successive 5 days.

pH in the radius of 3-6 were investigated for their effect on basal media for the production of naringinase. The pH of the medium was adjusted using 1N HCl or 1N NaOH. The flasks were incubated at 35°C for 5 days. The samples are withdrawn regularly for the determination of the naringinase activity.

Effect of stirring was carried at different rpm to determine the activity of naringinase i.e at an agitation speed of 0, 60, 80, 100, 120, 140, 160, 180 and 200rpm. The activity was determined at regular interval of time.

The fungi was grown in the basal media and the sowing was carried out at different inoculum size ranging from 0.25-1.50ul/ml. The inoculated flasks were incubated at 35°C for 5 days, further the naringinase activity was investigated at the regular interval of time.

Naringinase activity was measured by incubating production media which was seeded with different inoculum age starting from 12hr, 18hr, 24hr upto 5 days. The activity was recorded at regular intervals. The experiments were carried in the 500ml of conical flask containing 100ml of the desired medium (Mukesh *et al.*, 2012).

### *Effect of carbon and nitrogen sources*

Numerous carbon sources such as glucose, lactose, rhamnose, sucrose, starch, maltose and galactose were evaluated to determine its effects on the naringinase production by supplementing them with a carbon source for the basal production media.

Different organic and in-organic sources chosen for the study were peptone, beef extract, yeast extract,

malt extract,  $\text{NaNO}_3$  and  $\text{NH}_4\text{NO}_3$ . As these nitrogen sources were selected to determine effect on naringinase either it will be having a negative or positive effect. The basal media was supplemented with metal salts such as,  $\text{LiSO}_4$ ,  $\text{FeSO}_4$ ,  $\text{CaCl}_2$ ,  $\text{KCl}$ ,  $\text{NaCl}$ ,  $\text{MnSO}_4$  and  $\text{ZnSO}_4$  (Sukhvir *et al.*, 2016).

#### *Experimental design and statistical analysis*

Optimization of cultural media for the enhanced production is always been a tedious task. To overcome such kind of interpretation, RSM method has been designed, in which CCD (Central Composite Design) is one of the integral part of the RSM. In which four independent factors selected for naringinase production are agitation speed, carbon sources, nitrogen sources and pH. As these individual independent variables has two levels -1 and +1. The factors were set at four different variables coding levels of the factors refer to transforming the analyzed real value into co-ordinates inside. The experimental design includes a set of 30 experiments. The analysis of variance (ANOVA) was applied to determine the effect of inoculum percentage and regression analysis to predict naringinase activity. The consequences of all terms in the polynomial were judged analytically by computing the F-value at a probability (p) of 0.001, 0.01 or 0.05. The regression coefficients were then used to make statistical calculation to generate contour maps from the regression models (Lee *et al.*, 2006).

#### **Results and discussion**

Production of naringinase from microbial source and making it economically cheap for the use of industries has been a worth praising achievement in the field of fermentation technology.

#### *Physical parameters*

Environmental conditions have its effect on the production of extracellular enzyme. Some physical parameters like temperature, pH, agitation speed, inoculum size and age were studied for investigating their effects on the naringinase production.

As mentioned in the below Table 1, The optimal incubation temperature for the production of naringinase was found to be 35°C as this result was

expected since the tested isolate is a mesophilic in nature. A slight decrease in an enzyme production was observed thereafter at 40°C and so on. In relation to this most of the isolated micro-organisms showed optimal naringinase production in the range of 25 to 40°C. It reported that *Aspergillus niger* van Tieghem MTCC 2425 showed optimum activity at 29°C (Borkar *et al.*, 2020). Whereas *Aspergillus tubingensis* UA13 produces significant amount of naringinase at 35°C (Xia *et al.*, 2021). High temperatures also been reported to produce naringinase as reported by *Aspergillus oryzae* 11250 which shows optimal temperature of 45°C (Zhu *et al.*, 2017) respectively

Nearly all the naringinase isolated till now has found to have pH in the range of 4-6. Various pH ranging from 3-6 were tested in this investigation, where the pH 4.5 was found to produce highest naringinase activity as mentioned in below table 1, when compared to other pH.

The naringinase produced from *Bacillus methylotrophicus* (Mukund *et al.*, 2013) at pH 6 was found to produce highest naringinase activity of 8U/L. At pH 4.7 *Aspergillus niger* 2425 (Borkar *et al.*, 2020) found an optimal activity for the naringinase production. On the other hand Immobilized Naringinase isolated from *Aspergillus niger* also showed optimum activity at the pH4 (Gupta *et al.*, 2021).

Agitation leads to better movement of oxygen and mass transfer in different phases as it is also known to balance homogenous physical and chemical environment in the media by ceaseless stirring. In the present investigation isolated naringinase showed an highest activity at 120rpm. Further agitation of 150rpm showed highest enzyme activity for the enzyme isolated from the novel strain *Bacillus methylotrophicus* (Mukund *et al.*, 2013). The naringinase isolated from *Aspergillus niger* VBo7 (Vinothkumar *et al.*, 2010) showed highest activity of 17.12IU/ml at an agitation speed of 180rpm. The bacteria *Staphylococcus xylosus* MAK2 (Puri *et al.*, 2010) showed highest activity of 8.9IU/ml at an agitation speed of 300rpm.

Inoculum size plays an crucial role in fermentation process as it effects cultural parameters such as growth rate, nutritional utilization and cultural morphology. In this ongoing investigation the isolated strain KLA-80 was cultured in a basal nutrient media with different inoculum size. From the results it is found that naringinase activity gradually increases with increasing its inoculum upto reaching its maximum at an range of 0.25-0.75%. *Aspergillus niger* isolated from rotten pomelo (*citrus maxima*) found to produce highest activity at 0.5% of inoculum size (Sanjay *et al.*, 2020). Whereas naringinase produced from using Amla (Supriya chalta *et al.*, 2022). as substrated showed highest activity of 111870.3U/gm at an inoculum size of 20%.

In case of inoculum age, as this plays a crucial role while using mesophilic micro-organisms due to relatively low growth rate. In the present investigation different inoculum age is tested i.e from 24hrs to 8 days. It is found that highest naringinase activity was observed at 5th day of incubation. *Rhizophus stolonifer* using paddy husk showed highest activity of 1073.97u/g after optimization of the basal medium on the 8th day (Karuppaija *et al.*, 2016). Whereas *Aspergillus niger* MTCC 1344(Puri *et al.*, 2005) showed highest naringinase activity of 968 u/g on the 7th day of fermentation.

**Table 1.** Physical parmeters in relation to enzyme activity.

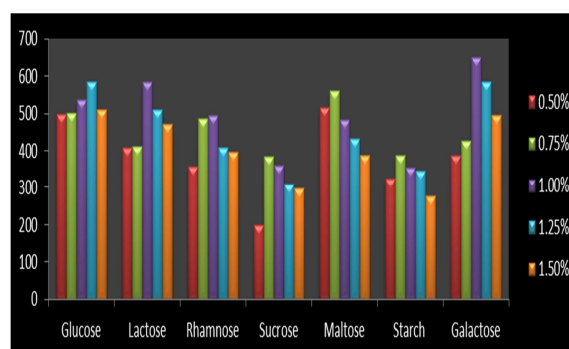
SL	Parameters	Range	Activity (Increment)
1	pH (4.5)	3.5-6	610
2	Temp (35)	30-55	630
3	Agitation speed (180)	60-180	651
4	Inoculum size (0.75)	0.25-0.50%	668
5	Inoculum age (5 <sup>th</sup> day)	24hrs- 7days	685

#### One factor at a- Time-method

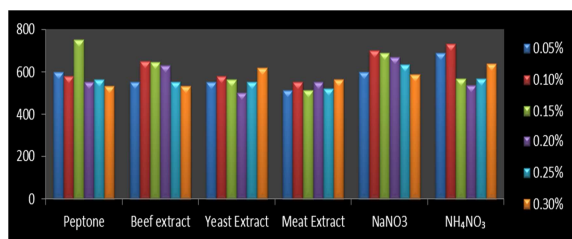
In the present study, after 5 days of fermentation, galactose showed highest naringinase activity when compared to other carbon sources like glucose, lactose, rhamnose, sucrose, starch and maltose. Hence galactose (700U/mL at 1.25%) is found to be a one of the cheap resources and also doesnot have any demerits, it was further selected for media formulations in subsequent experiments. Eventually,

a trial was conducted to determine the optimum nitrogen sources required for the naringinase production. A total of 6 different nitrogen sources are studied in this investigation which includes peptone, beef extract, yeast extract, malt extract, NaNO<sub>3</sub> and NH<sub>4</sub>NO<sub>3</sub>. Among all these 6 nitrogen sources, peptone (752U/mL at 0.15%) was found to be most effective and notably superior than other sources. In this view, it is found that organic nitrogen sources yield higher naringinase activity then in-organic nitrogen sources in sub-merged fermentation. As in the case of *Bacillus methylotrophicus* sucrose (1%) along with yeast extract (0.5%) showed highest activity of 12.05U/L when compared to the basal media (Mukund *et al.*, 2013). In the case of *Rhizopus stolonifer* 1.5% of glucose concentration and 0.5% of ammonium nitrate produced a high amount of naringinase i.e 1073.97U/g which is expentionally more then the unoptimized media (Karuppaija *et al.*, 2016). Whereas *Aspergillus niger* isolated from the soil showed an increase in activity upto 1.8 times with an 0.5% rhamnose and 0.25% peptone (Vinothkumar *et al.*, 2010).

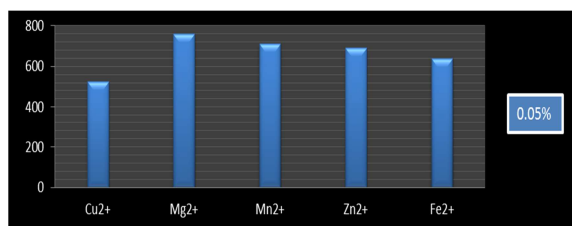
Metal ions are acts as stimulator for the growth of micro-organsims as its in co-operation in the media is necessary as it effects the enzyme production. In the present investigation, metal ions of concentration 0.05% is used where Mg<sup>2+</sup> showed highest activity of 760U/mL whereas Mg<sup>2+</sup> is followed by Cu<sup>2+</sup>>Mn<sup>2+</sup>>Zn<sup>2+</sup>. In case of *Aspergillus Flavus* Isolated From Decaying Citrus Maxima Fruits showed realtive activity of 133.913% at an 0.1% of Na<sup>2+</sup> concentration(Srikantha *et al.*, 2017)



**Fig. 1.** Naringinase activity recorded for various combinations of carbon source (0.25-1.50%).



**Fig. 2.** Naringinase activity recorded for various combinations of nitrogen source (0.05-0.30%).



**Fig. 3.** Naringinase activity recorded for various combinations of metal ions (0.05%).

RSM

Subsequently optimizing the values of variables by “One-factor-at-a-time-method“, 4 most important variables were selected for Response Surface Methodology by CCD method.

Response surface methodology is known to be one of the recommended statistical methods which is used to optimize the optimum level of fermentation conditions which helps to analyze the influence of several variables at a time along with the estimation of their interaction on the production of naringinase. The influence of 4 process parameters for fermentation was analyzed by Central Composite Design with 4 factors (Kalil *et al.*, 2000). The design of the experiment are represented in below Table 2.

**Table 2.** Design of RSM.

Std	Run	Factor 1 A:pH	Factor 2 B: Agitation spe... rpm	Factor 3 C: Galactose %	Factor 4 D: Peptone %	Response 1 Naringinase U/mL
24	1	4.75	120	0.875	0.425	775
27	2	4.75	120	0.875	0.175	867
8	3	6	180	1.5	0.05	589
26	4	4.75	120	0.875	0.175	867
9	5	3.5	60	0.25	0.3	700
5	6	3.5	60	1.5	0.05	538
6	7	6	60	1.5	0.05	521
21	8	4.75	120	-0.375	0.175	812
1	9	3.5	60	0.25	0.05	579
13	10	3.5	60	1.5	0.3	619
20	11	4.75	240	0.875	0.175	648
23	12	4.75	120	0.875	-0.075	651
19	13	4.75	0	0.875	0.175	618
10	14	6	60	0.25	0.3	766
15	15	3.5	180	1.5	0.3	593
18	16	7.25	120	0.875	0.175	433
4	17	6	180	0.25	0.05	646
17	18	2.25	120	0.875	0.175	408
25	19	4.75	120	0.875	0.175	867
22	20	4.75	120	2.125	0.175	686
12	21	6	180	0.25	0.3	654
14	22	6	60	1.5	0.3	613
16	23	6	180	1.5	0.3	582
7	24	3.5	180	1.5	0.05	619
2	25	6	60	0.25	0.05	611
11	26	3.5	180	0.25	0.3	644
29	27	4.75	120	0.875	0.175	867
3	28	3.5	180	0.25	0.05	621
28	29	4.75	120	0.875	0.175	867

The consequence of the model was analyzed by ANOVA method. The tools used for the determination of significance were *F* value and *p* value and it is also used to understand relationship between process

variable and co-efficient. If the *p* value is smaller and *F* value is higher than the corresponding co-efficient (Mukund *et al.*, 2013). Then the *F*-value was computed as the mean square of residual and the

mean square of regression and if the  $p$  value is less than 0.05 indicates that the terms used for the model

are significant, depending upon the importance of particular variable in the study.

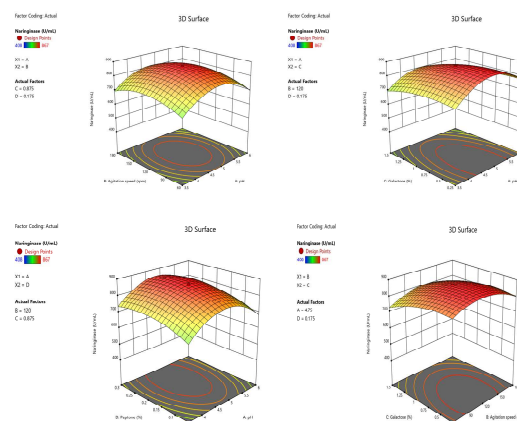
**Table 3.** ANOVA Table.

Source	Sum of Squares	Df	Mean Square	F-value	p-value	
Model	4.454E+05	14	31817.30	292.69	< 0.0001	significant
A-Ph	590.04	1	590.04	5.43	0.0353	
B-Agitation speed	155.04	1	155.04	1.43	0.2522	
C-Galactose	26600.04	1	26600.04	244.69	< 0.0001	
D-Peptide	20126.04	1	20126.04	185.14	< 0.0001	
AB	410.06	1	410.06	3.77	0.0725	
AC	2425.56	1	2425.56	22.31	0.0003	
AD	150.06	1	150.06	1.38	0.2596	
BC	2093.06	1	2093.06	19.25	0.0006	
BD	12712.56	1	12712.56	116.94	< 0.0001	
CD	1743.06	1	1743.06	16.03	0.0013	
A <sup>2</sup>	3.334E+05	1	3.334E+05	3067.34	< 0.0001	
B <sup>2</sup>	94152.84	1	94152.84	866.11	< 0.0001	
C <sup>2</sup>	25320.95	1	25320.95	232.93	< 0.0001	
D <sup>2</sup>	42012.30	1	42012.30	386.47	< 0.0001	
Residual	1521.92	14	108.71			
Lack of Fit	1521.92	10	152.19			Not significant
Pure Error	0.0000	4	0.0000			
Cor Total	4.470E+05	28				

The model F-value of 292.69 and the p-value less than 0.0001 implied that model was significant and there is only 0.01% chance that the mode F-value could occur due to noise. For the good statistical model  $R^2$  value should be close to 1.0. Adequate precision measure the signal to noise of ratio, a ratio of 61.74 indicates adequate signal. The value of the adjusted determination co-efficient ( $Adj R^2 = 0.9932$ ) was also high to advocate for the high significance of the model. All these considerations indicate good adequacy of the regression model.

The 3 dimensional diagram and plots are given by regression model are plotted below in the fig. 4. The 3 dimensional images is obtained in a combination i.e. 2 variables in a centre and 2 variables are independent. From the curve of three-dimensional plots optimum concentration of the media is recognized. The counterpart pinnacles the character played variables and their interactive effect. From the fig 4, it is observed that increase in the variable concentration increases the naringinase activity upto an optimal level and there after significant decrease in naringinase activity is observed. The counter plots showed a broad plateau region in which the activities change relatively little when there is concentration

change in the nutrient is observed. Thus it is observed that optimal solution concentration can accommodate small errors.



**Fig. 4.** Three-dimensional plot of Naringinase activity of *Aspergillus flavus* KLA-80 a) agitation speed and pH. b) Galactose and pH. c) Peptone and pH. d) Galactose and agitation speed.

For the determination of optimum values of the variable for the maximum production of naringinase point prediction method was used as per the equation given by (Mukund *et al.*, 2013). In the present investigation galactose of 1.25%, peptone-0.15%, pH of 4.75 and agitation speed of 120rpm showed highest

activity of 867 U/mL i.e 20% increment in the activity is observed when compared with the activity obtained by one-factor-at-a-time method.

### Conclusion

Naringinase was known to be one of the important enzymes as it converts bitter naringin to no-bitter naringenin. As this enzyme is one the budding approaches for the food industries. In this research previously isolated *Aspergillus flavus* KLA-80 were taken to optimize the production of naringinase. As this isolate produce 559U/mL. It is further optimized by one-factor-at-a-time followed by statistical experimental design was applied by RSM CCD method and there 20.41% is increase in the activity of the enzyme was observed i.e the activity increased to 780U/mL from 559U/mL at an galactose of 1.25%, peptone of 0.15%, naringin of 0.1%, agitation speed of 120rpm, pH of 4.75 and temperature of 35°C.

### Acknowledgement

The author is very much thankful to the Karnataka Fund for Infrastructure Strengthening in Science and Technology in Higher Educational Institutions (K-FIST) and Karnataka Science and Technology Promotion Society (KSTePS), Department of Science and Technology, Karnataka, in helping to develop lab infrastructure and providing financial support to carry out this research work.

### Funding

This study was supported by K-FIST and KSTePS

### Conflicts of interest

The authors report no financial or any other conflicts of interest in this work

### References

**Adami R, Russo P, Amante C, De Soricellis C, Della Porta G, Reverchon E, Del Gaudio P.** 2022. Supercritical Antisolvent Technique for the Production of Breathable Naringin Powder. *Pharmaceutics* **14(8)**, 1623. <https://doi.org/10.3390/pharmaceutics14081623>

**Alam MA, Subhan N, Rahman MM, Uddin SJ, Reza HM, Sarker SD.** 2014. Effect of Citrus Flavonoids, Naringin and Naringenin, on Metabolic Syndrome and Their Mechanisms of Action. *Advances in Nutrition* **5(4)**, 404-417.

<https://doi.org/10.3945/an.113.005603>

**Amiri MJ, Bahrami M, Dehkodaie F.** 2019. Optimization of Hg(II) adsorption on bio-apatite based materials using CCD-RSM design: characterization and mechanism studies. *Journal of Water and Health* **17(4)**, 556-567.

<https://doi.org/10.2166/wh.2019.039>

**Borkar V, Chakraborty S, Gokhale JS.** 2020. Fermentative Production of Naringinase from *Aspergillus niger* van Tieghem MTCC 2425 Using Citrus Wastes: Process Optimization, Partial Purification, and Characterization. *Applied Biochemistry and Biotechnology* **193(5)**, 1321-1337. <https://doi.org/10.1007/s12010-020-03385-9>

**Chen R, Qi QL, Wang MT, Li QY.** 2016. Therapeutic potential of naringin: an overview. *Pharmaceutical Biology* **54(12)**, 3203-3210. <https://doi.org/10.1080/13880209.2016.1216131>

**Cook N.** 1996. Flavonoids--Chemistry, metabolism, cardioprotective effects, and dietary sources. *The Journal of Nutritional Biochemistry* **7(2)**, 66-76. [https://doi.org/10.1016/0955-2863\(95\)00168-9](https://doi.org/10.1016/0955-2863(95)00168-9)

**Dutt K, Gupta P, Saran S, Misra S, Saxena RK.** 2009. Production of Milk-Clotting Protease from *Bacillus subtilis*. *Applied Biochemistry and Biotechnology* **158(3)**, 761-772. <https://doi.org/10.1007/s12010-008-8504-9>

**Gupta AK, Yumnam M, Medhi M, Koch P, Chakraborty S, Mishra P.** 2021. Isolation and characterization of naringinase enzyme and its application in debittering of Pomelo juice (*Citrus grandis*): A comparative study with macroporous resin. *Journal of Food Processing and Preservation* **45(5)**. Portico. <https://doi.org/10.1111/jfpp.15380>



- Jitolis JL, Awang Ali AN, Saad I, Taha NA, Idris J, Bolong N.** 2021. Utilization of Response Surface Methodology and Regression Model in Optimizing Bioretention Performance. 2021 IEEE International Conference on Artificial Intelligence in Engineering and Technology (IICAET). <https://doi.org/10.1109/iicaiet51634.2021.9573644>
- Kalil SJ, Maugeri F, Rodrigues MI.** 2000. Response surface analysis and simulation as a tool for bioprocess design and optimization. *Process Biochemistry* **35(6)**, 539-550.
- Karuppaija S, Kapilan R, Seevaratnam V.** 2016. Optimization of naringinase production by *Rhizopus stolonifer* in solid state fermentation media using paddy husk as support. *Scholars Academic Journal of Biosciences* **4(6)**. <https://doi.org/10.21276/sajb>.
- Kranz P, Adler P, Kunz B.** 2010. Sorption of citrus flavour compounds on XAD-7HP resin during the debittering of grapefruit juice. *International Journal of Food Science & Technology* **46(1)**, 30-36. <https://doi.org/10.1111/j.1365-2621.2010.02442.x>
- Lee HS, Kim JG.** 2003. Effects of debittering on red grapefruit juice concentrate. *Food Chemistry* **82(2)**, 177-180. [https://doi.org/10.1016/s0308-8146\(02\)00](https://doi.org/10.1016/s0308-8146(02)00)
- Lee WC, Yusof S, Hamid NSA, Baharin BS.** 2006. Optimizing conditions for enzymatic clarification of banana juice using response surface methodology (RSM). *Journal of Food Engineering* **73(1)**, 55-63. <https://doi.org/10.1016/j.jfoodeng>.
- Manish Kumar, Veenya Pareek.** 2022. Minimization of Limonin and Naringin Content in Kinnow Juice Using Response Surface Methodology. *Annals of Biology* **38(1)**, 88-95.
- Mishra P, Kar R.** 2003. Treatment of Grapefruit Juice for Bitterness Removal by Amberlite IR 120 and Amberlite IR 400 and Alginate Entrapped Naringinase Enzyme. *Journal of Food Science* **68(4)**, 1229-1233. <https://doi.org/10.1111/j.1365-2621.2003>
- Mukesh Kumar DJ, Saranya GM, Suresh K, Andal Priyadharshini D, Rajakumar R, Kalaichelvan PT.** 2012. Production and Optimization of Pectinase from *Bacillus* sp. MFW7 using Cassava Waste. *Asian Journal of Plant Science and Research*, 2012, **2(3)**, 369-375. [https://www.researchgate.net/profile/Puthupalayam-Kalaichelvan/publication/264851241\\_Production\\_and\\_Optimization\\_of\\_Pectinase\\_from\\_Bacillus\\_sp\\_MFW7\\_using\\_Cassava\\_Waste/links/540c60140cf2f2b29a377e64/Production-and-Optimization-of-Pectinase-from-Bacillus-sp-MFW7-using-Cassava-Waste.pdf](https://www.researchgate.net/profile/Puthupalayam-Kalaichelvan/publication/264851241_Production_and_Optimization_of_Pectinase_from_Bacillus_sp_MFW7_using_Cassava_Waste/links/540c60140cf2f2b29a377e64/Production-and-Optimization-of-Pectinase-from-Bacillus-sp-MFW7-using-Cassava-Waste.pdf)
- Mukund P, Belur PD, Saidutta MB.** 2013. Production of naringinase from a new soil isolate, *Bacillus methylotrophicus*: Isolation, optimization and scale-up studies. *Preparative Biochemistry and Biotechnology* **44(2)**, 146-163. <https://doi.org/10.1080/10826068.2013.797910>
- Muñoz M, Holtheuer J, Wilson L, Urrutia P.** 2022. Grapefruit Debittering by Simultaneous Naringin Hydrolysis and Limonin Adsorption Using Naringinase Immobilized in Agarose Supports. *Molecules* **27(9)**, 2867. <https://doi.org/10.3390/molecules27092867>
- Oskouie SFG, Tabandeh F, Yakhchali B, Eftekhari F.** 2008. Response surface optimization of medium composition for alkaline protease production by *Bacillus clausii*. *Biochemical Engineering Journal* **39(1)**, 37-42. <https://doi.org/10.1016/j.bej.2007>
- Puri M.** 2011. Updates on naringinase: structural and biotechnological aspects. *Applied Microbiology and Biotechnology* **93(1)**, 49-60. <https://doi.org/10.1007/s00253-011-3679-3>
- Puri M, Kalra S.** 2005. Purification and characterization of naringinase from a newly isolated strain of *Aspergillus niger* 1344 for the transformation of flavonoids. *World Journal of Microbiology and Biotechnology* **21(5)**, 753-758. <https://doi.org/10.1007/s11274-004-5488-7>

- Puri M, Banerjee A, Banerjee UC.** 2005. Optimization of process parameters for the production of naringinase by *Aspergillus niger* MTCC 1344. *Process Biochemistry* **40(1)**, 195-201. <https://doi.org/10.1016/j.procbio.2003.12.009>
- Puri M, Kaur A, Barrow CJ, Singh RS.** 2010. Citrus peel influences the production of an extracellular naringinase by *Staphylococcus xylosus* MAK2 in a stirred tank reactor. *Applied Microbiology and Biotechnology* **89(3)**, 715-722. <https://doi.org/10.1007/s00253-010-2897-4>
- Puri M, Kaur A, Singh RS, Singh A.** 2009. Response Surface Optimization of Medium Components for Naringinase Production from *Staphylococcus xylosus* MAK2. *Applied Biochemistry and Biotechnology* **162(1)**, 181-191. <https://doi.org/10.1007/s12010-009-8765-y>
- Ribeiro M, Silveira D, Ferreira-Dias S.** 2002. Selective adsorption of limonin and naringin from orange juice to natural and synthetic adsorbents. *European Food Research and Technology* **215(6)**, 462-471. <https://doi.org/10.1007/s00217-002-0592->
- Sandon F.** 1958. *Experimental Designs*. By W. G. Cochran and G. M. Cox. 2nd ed. Pp. xiv, 611. 82s. 1957. (John Wiley and Sons, New York; Chapman and Hall, London). *The Mathematical Gazette* **42(342)**, 334-334. <https://doi.org/10.2307/3610494>
- Sanjay Kumar, Vijay Kumar, Pankaj Gautam, Dinesh Puri.** 2020. Isolation, screening and characterization of fungi from rotten pomelo (citrus maxima) for the production of debittering enzyme naringinase **11(5)**, pp.1152-1160, Article ID: IJARET\_11\_05\_125.10.34218/IJARET.11.5.2020.
- Shivalee A, Lingappa K, Mahesh D.** 2018. Influence of bioprocess variables on the production of extracellular chitinase under submerged fermentation by *Streptomyces pratensis* strain KLSL55. *Journal of Genetic Engineering and Biotechnology* **16(2)**, 421-426. <https://doi.org/10.1016/j.jgeb.2017.12.006>
- Srikantha K, Kapilan R, Seevaratnam V.** 2017. Kinetic Properties and Metal Ion Stability of the Extracellular Naringinase Produced By *Aspergillus Flavus* Isolated From Decaying Citrus Maxima Fruits. *International Journal of Scientific Research in Environmental Sciences* **5(3)**, 71-81. <https://doi.org/10.12983/ijres-2017-p0071-0081>
- Sukhvir Kaur, Harjot Pal Kaur, Bhairav Prasad, Tejaswani Bharti.** 2016. Production and optimization of pectinase by bacillus sp. isolated from vegetable waste soil. *Indo American Journal of Pharmaceutical Research*, **6(01)**, 2016. [https://www.researchgate.net/profile/BhairavPrasad/publication/327644739\\_Production\\_and\\_optimization\\_of\\_pectinase\\_by\\_Bacillus\\_sp\\_isolated\\_from\\_vegetable\\_waste\\_soil/links/5d01e33ba6fdccd13096a619/Production-and-optimization-of-pectinase-by-Bacillus-sp-isolated-from-vegetable-waste-soil.pdf](https://www.researchgate.net/profile/BhairavPrasad/publication/327644739_Production_and_optimization_of_pectinase_by_Bacillus_sp_isolated_from_vegetable_waste_soil/links/5d01e33ba6fdccd13096a619/Production-and-optimization-of-pectinase-by-Bacillus-sp-isolated-from-vegetable-waste-soil.pdf)
- Supriya Chatla, Devalarao Garikapati, Abdul Rahaman, Iswarya Obilineni.** 2022. Production of Naringinase by using Amla on Solid State Fermentation. *Research J. Pharm and Tech* **15(3)**, 1225-1229. DOI: 10.52711/0974-360X.2022.00204.
- Tran AM, Nguyen TB, Nguyen VD, Bujna E, Dam MS, Nguyen QD.** 2020. Changes in bitterness, antioxidant activity and total phenolic content of grapefruit juice fermented by *Lactobacillus* and *Bifidobacterium* strains. *Acta Alimentaria* **49(1)**, 103-110. <https://doi.org/10.1556/066.2020.49.1.13>
- Vinoth Kumar V, Kayambu P, Revathi Babu S.** 2010. Optimization of fermentation parameters for enhanced production of naringinase by soil isolate *Aspergillus niger* VBO7. *Food Science and Biotechnology* **19(3)**, 827-829. <https://doi.org/10.1007/s10068-010-0116-9>
- Xia XK, Zhang YE, Lei SJ, Hu B, Fu CX.** 2021. Optimization of process parameters for naringinase production by *Aspergillus tubingensis* UA13 and pilot scale-up study. *Preparative Biochemistry & Biotechnology* **52(2)**, 226-233. <https://doi.org/10.1080/10826068.2021.1925914>

**Zhang J, Zhang J, Shan Y, Guo C, He L, Zhang L, Ling W, Liang Y, Zhong B.** 2022. Effect of harvest time on the chemical composition and antioxidant capacity of *Gannan navel* orange (*Citrus sinensis* L. Osbeck 'Newhall') juice. *Journal of Integrative Agriculture* **21(1)**, 261-272. [https://doi.org/10.1016/s2095-3119\(20\)63395-0](https://doi.org/10.1016/s2095-3119(20)63395-0)

**Zhu Y, Jia H, Xi M, Li J, Yang L, Li X.** 2017. Characterization of a naringinase from *Aspergillus oryzae* 11250 and its application in the debitterization of orange juice. *Process Biochemistry* **62**, 114-121. <https://doi.org/10.1016/j.procbio.2017.07.012>