



RESEARCH PAPER

OPEN ACCESS

Cassia sophera L.: Antioxidant potential profiling of different extracts of leaf, flower, seed and stem

Shahin Aziz^{*1}, M. Mahboob Ali Siddiqi², Md. Sajal Sorowar³

¹Chemical Research Division, BCSIR Laboratories Dhaka, Bangladesh Council of Scientific and Industrial Research, Dr. Quadrat-I-Khuda Road, Dhanmondi, Dhaka, Bangladesh

²Institute of Natural Sciences, United International University, Dhaka, Bangladesh

³Department of Applied Chemistry and Chemical Engineering, Islamic University, Kushtia, Bangladesh

Key words: *Cassia sophera* L., Antioxidant activity, DPPH, Calsalpiniaceae

<http://dx.doi.org/10.12692/ijb/25.4.76-84>

Article published on October 04, 2024

Abstract

Cassia sophera L. is a predominant traditional medicinal shrub as well as considered as a remedy in folk arena due to its antidiarrhoeal, antipyretic, antinociceptive, anthelmintic and antioxidant effect. In the present study, the antioxidant effect of n-hexane, chloroform, ethyl acetate and methanol extract of leaf, flower, seed and stem of *C. sophera* L. was evaluated by DPPH (2,2-diphenyl-1-picrylhydrazyl) method. The assay of free radical scavenging activity of four different solvent extracts revealed that the ethyl acetate extract of leaf and stem exhibited good antioxidant activity with IC₅₀ of 61.296 and 121.289 µg/ml respectively as compared to ascorbic acid considered as positive control. In addition, the antioxidant potential of the solvent extracts of leaf, flower, seed and stem of *C. sophera* L. is firstly reported in the present study except methanolic extract of leaf.

* **Corresponding Author:** Shahin Aziz ✉ shaziz2408@yahoo.com

Introduction

Plants are a great repository of phytochemicals rendering with health aid and development due to a large area of therapeutic actions as medicines from primitive period till today. Besides, modern analytical technology has opened a large window of tremendous investigations on the plants and herbs used in folk medicine all over the world (Mulaudzi *et al.*, 2022; Mehari *et al.*, 2021; Anokwuru *et al.*, 2022; Ahmed *et al.*, 2022). In fact, bioactive compounds obtained from herbs are beneficial against the undesirable side effects of chemotherapy and promote general health and longevity (Yuan *et al.*, 2022). This is why an intense inclination on the investigation of the medicinal potential of plants is currently growing. In connection to this, one of the most significant potential phytochemicals is natural antioxidants like ascorbic acid, tocopherol, flavonoids, phytoestrogens, phenolic acids, phytic acids, etc (Bendich and Langseth *et al.*, 1995) which show magnificent therapeutic efficacy to lessen and combat the risk of chronic diseases, cancers, and heart diseases caused by reactive oxygen species (ROS) in the body (George *et al.*, 2021; Majumder and Afia *et al.*, 2019; Miller *et al.*, 2000).

Cassia sophera L. originates from the subfamily of Caesalpinioideae under Leguminosae family spreading from Africa to India and South East Asia. *C. sophera* which is assumed as an indigenous species of South America is a shrub (Gulzar *et al.*, 2014). In Bangladesh the plains and hills of Chittagong, Sylhet are the abundantly growing place of *C. sophera* which is locally known as Kalkasunda, Kasaundi, Kasunda and Baner (Yusuf *et al.*, 1994). Moreover, it also grows at a large extent in India as well as other subtropical and tropical Asian countries such as Pakistan, Sri Lanka and Myanmar. Consequently its use is evidently found as traditional medicines in Japan, Korea and China as well as the Indian subcontinent (Drever *et al.*, 2008; Guo *et al.*, 1998). It can grow more than three meters tall. Local people sometimes consume the soft leaves and the pods. The Ayurveda and Unani traditions reported its potential and effective use for treating piles, jaundice, psoriasis,

skin diseases, eye inflammation and even snake bites (Drever *et al.*, 2008; Bilal *et al.*, 2005; Arijit *et al.*, 2012; Aminabee and Lakshmana *et al.*, 2012). The seed, flower, root and bark of *C. sophera* are well known for its astringency, laxative and carminative (Drever *et al.*, 2008; Kirtikar and Basu *et al.*, 2006; Lee *et al.*, 2001; Gupta *et al.*, 2010). In addition the ethnobotanical literature states that the leaves are used medicinally for its anti-inflammatory, anti-rheumatic, laxative properties. The leaves were also considered as expectorant for coughs, colds, bronchitis, asthma and also effective for liver disorder. Previous studies showed that analgesic, anticonvulsants, antidiabetic, herbicidal, bactericidal activity of seed were investigated (Majumder and Afia *et al.*, 2019). Moreover alcoholic extract obtained from leaves is a potential relaxant for intestinal and bronchial muscle. On the other hand, phytochemical investigation revealed that a flavone glycoside and senna glycoside from leaves and anthraquinones, chrysophanol, physcion and beta-sitosterol from root were reported (Bilal *et al.*, 2005). It is also noted that a good number of bioactive compounds such as 1,2,7-trihydroxy-3-methylanthraquinone, sopheranin, beta-sitosterol, chrysophanol, physcion, emodin, 1-octadecanol and quercetin were isolated from heartwood (Khare *et al.*, 2009).

Although several studies have been performed on *C. sophera* so far, extensive research on the phytochemical and pharmacological aspects of *C. sophera* cultivated in Bangladesh is inadequate. Therefore, a deeper attention should be given to evaluate several biological and chemical potential of *C. sophera*. This is why, we have assessed and compared the antioxidant activities of n-hexane, chloroform, ethyl acetate and methanol extract of leaf, flower, seed and stem of *C. sophera* by using standard *in vitro* assays for substantiating its ethnobotanical medicinal use in the present study for the first time except methanolic extract of leaf.

Materials and methods

Fully matured plant parts like fresh leaves, stems, flowers and seeds of *C. sophera* were collected from

the hilly areas of Sylhet district in Bangladesh. The specimen of the samples was authenticated by the taxonomist of the Bangladesh national Herbarium, Dhaka where a voucher (No.43734) has been deposited.

The leaves, stems, flowers and seeds were separately dried in an oven at 38°C. These dried samples of leaves, stem, flower and seed were ground using a Wiley mill up to a particle size of 20 mesh. Under the cold extraction process, the powdered material of leaves (250 g) was extracted with n-hexane for 5 days at room temperature. Afterwards the solid residue obtained from the first extraction step was successively soaked with chloroform, ethyl acetate and methanol. In this process the polarity of the solvent was gradually increased. The similar cold extraction process applied on the powdered material of leaves was also implemented for the solvent extraction of the powdered material of flower (175 g), seed (110 g) and stem (100 g) respectively. Each extract obtained from the respective solvent was subjected to evaporation by Rotary evaporator under reduced pressure. Consequently, a gummy mass of n-hexane extract (9 g), chloroform extract (7.5 g), ethyl acetate extract (7 g) and methanol extract (6 g) was obtained.

UV-Visible Spectrophotometer (Perkin Elmer Shelton, CT 06484 USA, Lambda 25) was used to measure UV absorbance measurements. In order to evaporate solvents, a rotary evaporator (BUCHI, Rotavapor R-210, Switzerland) was used. All the solvents used in this experiment were analytical grade (Sigma-Aldrich, St. Louis, MO, USA).

The DPPH method is a facile spectrophotometric technique for the determination of antioxidant activity (Braca *et al.*, 2001; Angeli *et al.*, 2021). An unpaired electron on the DPPH (2, 2-diphenyl-1-picrylhydrazyl) free radical imparts an intense violet due to its solution. The free radical scavenging activity was easily measured by the reduction of absorbance in the 517 nm region using a spectrophotometer. In the procedure of DPPH

method, firstly 1 ml of each of ascorbic acid solutions of different strengths in methanol (5, 10, 25, 50, 100, 200, 400 µg/ml in methanol) was mixed with 3 ml of 0.4 mM DPPH solution. Afterwards, 1 ml of each of the extracts was also mixed with 3 ml of 0.4 mM DPPH solution. As per the standard procedure, the mixture was kept totally enclosed for protection from exposure to light for 30 minutes to measure the absorbance at 517 nm under the UV-Visible Spectrophotometer. In this study, ascorbic acid solution was used as a positive control. The higher the free radical terminating potential of the sample, the lower the absorbance obtained. Under the action of the antioxidants, the DPPH solution turned to yellow from purple and the degree of this transformation indicates the radical quenching power of the analyte.

The free radical scavenging potential against DPPH was computed according to the formula:

$[(A-B)/A] \times 100$. Where "A" is the absorbance of the reference (DPPH solution free from any antioxidant) and "B" is the absorbance of the free radical source solution in the presence of a sample (DPPH plus extract/ascorbic acid). The radical quenching power (in percent) was then plotted against concentration and from the graph, the IC₅₀ (Concentration at 50% inhibition) value was estimated through linear regression analysis.

For the assessment of antioxidant activity, each experiment was repeated in triplicate and the mean value of the results was calculated.

Results and discussion

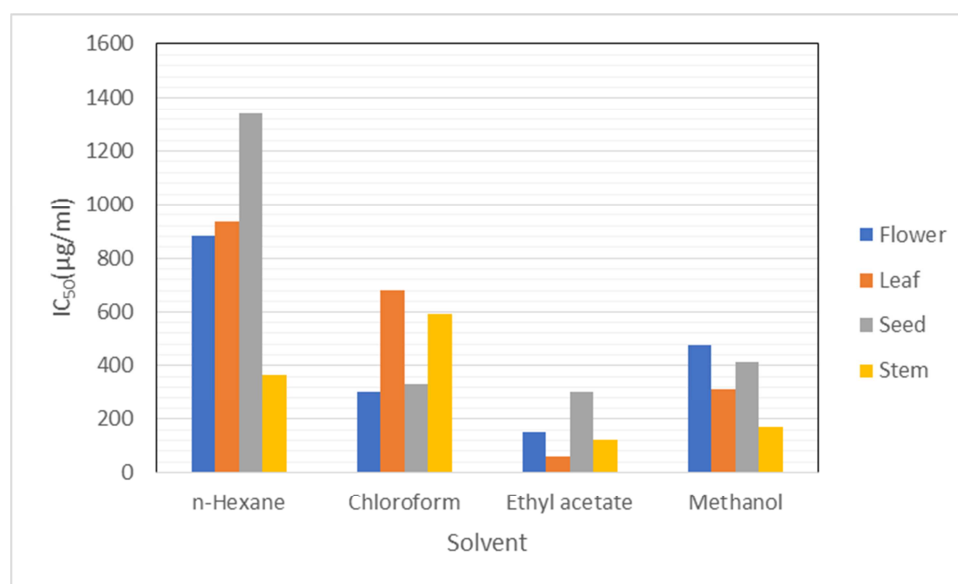
The antioxidant activity of the various extracts collected from leaf, flower, seed and stem of the plant was assayed by using the DPPH method.

The n-hexane extracts of stem, flower, leaf and seed showed diminutive free radical scavenging effect on DPPH radicals where these four parts exhibited a range of IC₅₀ from 368.175 to 1342.413 µg/ml shown in Table 1.

Table 1. IC₅₀ values of solvent extracts of leaf, flower, seed and stem of *Cassia sophera* L.

Plant parts	IC ₅₀ (µg/ml)			
	n-Hexane	Chloroform	Ethyl acetate	Methanol
Flower	884.595 ± 4.47	304.98 ± 1.175	152.224 ± 3.30	476.95 ± 1.99
Leaf	937.048 ± 2.32	681.915 ± 1.91	61.296 ± 1.55	313.99 ± 1.03
Seed	1342.413 ± 3.08	333.724±2.22	305.26 ± 1.25	415.787 ± 2.50
Stem	368.175 ± 2.53	590.972 ± 3.29	121.289 ± 0.67	170.45 ± 1.59
Ascorbic acid	6.344±0.0175			

Values are the means ± standard deviations (n = 3).

**Fig. 1.** IC₅₀ values of solvent extracts of four different parts of *Cassia sophera* L.

In addition, Table 4 shows that the lowest free radical scavenging effect obtained from n-hexane extract of seed was 32.83% inhibition at a concentration of 400 µg/ml whereas the highest effect found from the extract of stem was 51.40% inhibition at the same concentration compared to positive control given in the Table 5.

Moreover, from the Table 1, it is seen that the chloroform extract of leaf, flower, seed and stem showed moderate scavenging effect on DPPH radicals and their activity based on IC₅₀ was in the range from 304.98 to 681.915 µg/ml.

In assessment of chloroform extract, the leaf extract had lowest free radical scavenging effect that was 40.26% inhibition at a concentration of 400 µg/ml given in the Table 3 while the flower extract had the highest effect was 56.59% inhibition at the same concentration compared to standard ascorbic acid shown in the Table 2.

Table 2. Free radical scavenging activity of different solvent extracts of flower

Extracts	Concentration (µg/ml)	Absorbance	%Inhibition
n-Hexane	5	1.053	23.25
	10	1.041	24.12
	25	1.033	24.70
	50	1.017	25.87
	100	0.997	27.33
	200	0.956	30.32
	400	0.888	35.27
	blank	1.372	
Chloroform	5	0.852	27.05
	10	0.851	27.14
	25	0.841	27.99
	50	0.820	29.79
	100	0.764	34.58
	200	0.665	43.06
	400	0.507	56.59
	blank	1.168	
Ethyl acetate	5	0.846	27.56
	10	0.825	29.36
	25	0.804	31.16
	50	0.726	37.84
	100	0.620	46.91
	200	0.430	63.18
	400	0.245	79.02

	blank	1.168	
Methanol	5	0.983	27.13
	10	0.974	27.79
	25	0.958	28.98
	50	0.943	30.09
	100	0.898	33.43
	200	0.847	37.21
	400	0.728	46.03
	blank	1.349	

Furthermore, the Table 1 & Fig. 1 reveals that the ethyl acetate extract leaf manifested the most significant free radical scavenging at IC₅₀ of 61.296 µg/ml while the stem and flower extract showed moderate antioxidant activity at IC₅₀ of 121.289 and 152.224 µg/ml respectively. On the other hand, the lowest free radical scavenging effect at IC₅₀ of 305.26µg/ml was obtained from ethyl acetate extract of seed shown in the Table 3.

Table 3. Free radical scavenging activity of different solvent extracts of Leaf

Extracts	Concentration (µg/ml)	Absorbance	%Inhibition
n-Hexane	5	0.883	26.84
	10	0.878	27.25
	25	0.873	27.67
	50	0.864	28.41
	100	0.851	29.49
	200	0.804	33.38
	400	0.769	36.28
	blank	1.207	
Chloroform	5	0.879	27.17
	10	0.871	27.83
	25	0.864	28.41
	50	0.847	29.82
	100	0.827	31.48
	200	0.783	35.12
	400	0.741	40.26
	blank	1.207	
Ethyl acetate	5	0.669	44.57
	10	0.652	45.98
	25	0.643	46.72
	50	0.615	49.04
	100	0.552	54.26
	200	0.445	63.13
	400	0.334	72.32
	blank	1.207	
Methanol	5	0.869	28.66
	10	0.853	29.32
	25	0.834	30.90
	50	0.775	35.79
	100	0.732	39.35
	200	0.682	43.49
	400	0.554	54.10
	blank	1.207	

Table 4. Free radical scavenging activity of different solvent extracts of seed

Extractives	Concentration (µg/ml)	Absorbance	%Inhibition
n-Hexane	5	0.846	25.91
	10	0.849	25.65
	25	0.849	25.65
	50	0.838	26.61
	100	0.827	27.58
	200	0.808	29.24
	400	0.767	32.83
	blank	1.142	
Chloroform	5	0.840	28.44
	10	0.829	29.38
	25	0.814	30.66
	50	0.795	32.28
	100	0.730	37.81
	200	0.663	43.52
	400	0.554	52.81
	blank	1.174	
Ethyl acetate	5	0.849	27.68
	10	0.843	28.19
	25	0.828	29.47
	50	0.801	31.77
	100	0.754	35.77
	200	0.677	42.33
	400	0.509	56.64
	blank	1.174	
Methanol	5	0.835	28.87
	10	0.822	29.98
	25	0.826	29.64
	50	0.803	31.60
	100	0.774	34.07
	200	0.717	38.92
	400	0.598	49.06
	blank	1.174	

From the deeper studies of other researchers on different plants, we can predict that this significant antioxidant potential may be due to the phenolic phytochemical species extracted in this polar fraction (Semwal *et al.*, 2015; Cosa *et al.*, 2019; Yao *et al.*, 2013). It is clearly noted that phenolic acids and flavonoids widely occurred in the plant kingdom, especially in fruits and vegetables, are the chief categories of phenolic forming an important category of phenolic phyto-compounds that manifest antioxidant activity (Wojdyło *et al.*, 2007).

Moreover the medicinal and physiological function of the phenolic phyto-compounds possess benevolent promises for human well-being. Besides being radical scavengers, these are known to be able to prevent inflammations and cancers (Ghiselli *et al.*, 1998;

Visioli *et al.*, 1998). The origin of the antioxidant property of phenolic phytochemicals is their oxidation-reduction property which impart to exterminate free radicals, quench reactive oxygen (singlet and triplet), or to neutralize peroxy-compounds (Galalto *et al.*, 2001).

In addition, Table 1 also shows that methanol extracts of stem showed moderate activity with IC₅₀ of 170.45µg/ml among its four extracts. On the contrary, the lowest free radical scavenging effect at IC₅₀ of 476.95µg/ml was obtained from methanol extract of flower given in the Table 1.

Table 5. Free radical scavenging activity of different solvent extracts of stem

Extractives	Concentration (µg/ml)	Absorbance	%Inhibition
n-Hexane	5	0.898	25.53
	10	0.885	26.72
	25	0.824	29.63
	50	0.787	32.79
	100	0.766	34.58
	200	0.704	39.88
	400	0.589	51.40
	blank	1.171	
Chloroform	5	0.838	28.43
	10	0.830	29.12
	25	0.825	29.54
	50	0.796	32.02
	100	0.771	34.15
	200	0.736	37.14
	400	0.673	42.52
	blank	1.171	
Ethyl acetate	5	0.850	27.41
	10	0.819	30.05
	25	0.759	35.18
	50	0.682	41.75
	100	0.594	49.27
	200	0.387	66.95
	400	0.129	88.98
	blank	1.171	
Methanol	5	0.835	28.69
	10	0.823	29.71
	25	0.790	32.53
	50	0.730	37.66
	100	0.643	45.08
	200	0.495	57.72
	400	0.313	73.27
	blank	1.171	

In connection to this point, from previous research of other investigators, we can surmise that these highly polar extracts probably contain phenolic

phytochemicals known as anti-tumor nutraceuticals (Ahmed *et al.*, 2022; Li *et al.*, 2012). Reactive oxygen species (ROS) play a significant role in cell differentiation, aging and various other cellular processes related to overall health (Mates *et al.*, 2000). The phyto-compounds in the highly polar extract of the stem may have the potential to fight against the ROS levels in human tissue. Therefore, we can clearly see that the ethyl acetate and methanol fractions of various parts of the plant contained more potent antioxidants in the bioassays employed.

Several investigations in the past ascribe the antioxidant potential of phyto-extracts to the occurrence of flavonoids, phenolics, terpenoids as well as saponins, tannins, alkaloids etc and we believe that different solvent extracts of our study contain similar class of compounds due to the physico-chemical reasons like solvent polarity (Dangles *et al.*, 2000; Moura *et al.*, 2007; Gülçin *et al.*, 2004; Grabmann *et al.*, 2005). The antioxidative property of polar mixtures like the ethyl acetate and the methanol extract of leaf, stem and flower of *C. sophera* L. imply that this traditional herb may impart health benefits in human beings by reducing oxidative stress. Our findings provide the herbal physician another natural source of strong antioxidants for treatment.

It can be summarized that the antioxidant activity of the different extracts of leaf, flower, seed and stem of *C. sophera* L. was found to be consistent with the folk uses of this plant by local people. This is the first report of antioxidant activity of four separate parts of this shrub. In the present study, we have found significant reactive free radical extinguishing potentials in the most highly polar media extractions, namely ethyl acetate and methanol of the four parts of the plant. We strongly believe that deeper studies at the molecular and pharmacological level on the plant may bring greater human benefits.

Acknowledgements

We thankfully acknowledge the generous help provided by the Department of Pharmacy, Dhaka

University of Dhaka, for the assessment of antioxidant of the crude extracts of plant materials. We are also thankful to the Director, BCSIR Laboratories, Dhaka, Bangladesh for facilitating the research related procurements.

References

- Ahmed ME, Senabe J, Yahaya ES, Fouche G, Steenkamp P, Steenkamp V.** 2022. Isolation and antioxidant activity of 5-methyl-dihydroflavasperone from ethanol leaf extract of *Guiera senegalensis* JF Gmel. *Journal of Medicinal Plants for Economic Development* **6**(1), 137.
<https://doi.org/10.4102/jomped.v6i1.137>.
- Aminabee SK, Lakshmana RA.** 2012. A plant review of *Cassia sophera* Linn. *International Journal of Pharmacy, Chemistry and Biological Sciences* **2**(3), 408-414. ISSN: 2249-9504.
- Angeli L, Imperiale S, Ding Y, Scampicchio M, Morozova K.** 2021. A novel stoichiometric model for the DPPH• assay: The importance of the side reaction and application to complex mixtures. *Antioxidants* **10**(7), 1019.
DOI: 10.3390/antiox10071019.
- Anokwuru CP, Makolo FL, Sandasi M, Tankeu SY, Elisha IL, Agoni C, Combrinck S, Viljoen A.** 2022. Cannabigerol: a bibliometric overview and review of research on an important phytocannabinoid. *Phytochemistry Reviews* **21**(5), 1523-1547.
DOI: 10.1007/s11101-021-09794-w.
- Arijit M, Sanjay KK, Tanushree S, Rajalingam D.** 2012. Evaluation of hepatoprotective effect of leaves of *Cassia sophera* Linn. *Evidence-Based Complementary and Alternative Medicine*, 2012:436139.
DOI: 10.1155/2012/436139.
- Bendich A, Langseth L.** 1995. The health effects of vitamin C supplementation: a review. *Journal of the American College of Nutrition* **14**(2), 124-136.
DOI: 10.1080/07315724.1995.10718484.
- Bilal A, Khan NA, Ghufuran A, Inamuddin H.** 2005. Pharmacological investigation of *Cassia sophera*, Linn. var. *purpurea*, Roxb. *Medical Journal of the Islamic World Academy of Sciences* **15**, 105-109.
<https://www.researchgate.net/publication/256090640>.
- Braca A, De Tommasi N, Di Bari L, Pizza C, Politi M, Morelli I.** 2001. Antioxidant principles from *Bauhinia tarapotensis*. *Journal of Natural Products* **64**(7), 892-895.
DOI: 10.1021/np0100845.
- Cosa S, Chaudhary SK, Chen W, Combrinck S, Viljoen A.** 2019. Exploring common culinary herbs and spices as potential anti-quorum sensing agents. *Nutrients* **11**(4), 739.
DOI: 10.3390/nu11040739.
- Dangles OG, Fargeix C, Dufourb C.** 2000. Antioxidant properties of anthocyanins and tannins: A mechanistic investigation with catechin and the 3', 4', 7'-trihydroxyflavylium ion. *Journal of the Chemical Society, Perkin Transactions* **2**, 1653-1663.
<https://doi.org/10.1039/B003260N>.
- Drever BD, Anderson WG, Riedel G, Kim DH, Ryu JH, Choi DY, Platt B.** 2008. The seed extract of *Cassia obtusifolia* offers neuroprotection to mouse hippocampal cultures. *Journal of Pharmacological Sciences* **107**, 380-392.
DOI: 10.1254/jphs.08034fp.
- Galalto D, Ckless K, Susin MF, Giacomelli C, Ribeiro do Valle RM, Spinelli A.** 2001. Antioxidant capacity of phenolic and related compounds: Correlation among electrochemical, visible spectroscopy methods, and structure-antioxidant activity. *Redox Report* **6**, 243-250.
DOI: 10.1179/135100001101536391.
- George BP, Chandran R, Abrahamse H.** 2021. Role of phytochemicals in cancer chemoprevention: Insights. *Antioxidants* **10**(9), 1455.
DOI: 10.3390/antiox10091455.

- Ghiselli A, Nardini M, Baldi A, Scaeni C.** 1998. Antioxidant activity of different phenolic fractions separated from an Italian red wine. *Journal of Agricultural and Food Chemistry* **46**, 361-367. DOI: 10.1021/jf970486b.
- Grabmann J.** 2005. Terpenoids as plant antioxidants. *Vitamins & Hormones* **72**, 505-535. DOI: 10.1016/S0083-6729(05)72015-X.
- Gülçin İ, Mshvildadze V, Gepdiremen A, Elias R.** 2004. Antioxidant activity of saponins isolated from ivy: α -hederin, hederasaponin-C, hederacolchiside-E, and hederacolchiside-F. *Planta Medica* **70**(6), 561-563. DOI: 10.1055/s-2004-827158.
- Gulzar A, Siddiqui MB, Bi S.** 2014. Assessment of allelopathic potential of *Cassia sophera* L. on seedling growth and physiological basis of weed plants. *African Journal of Biotechnology* **13**(9), 1037-1046. DOI: 10.5897/AJB2013.13512.
- Guo H, Chang Z, Yang R, Guo Dm, Zheng J.** 1998. Anthraquinones from hairy root cultures of *Cassia obtusifolia*. *Phytochemistry* **49**, 1623-1625. DOI: 10.1016/S0031-9422(98)00325-2.
- Gupta RK.** 2010. Medical and aromatic plants. CBS Publishers and Distributors, 151-152.
- Khare CP.** 2009. Indian Medicinal Plants. Springer Science and Business Media, LLC, 223 Spring Street, New York, NY 10013, USA.
- Kirtikar KR, Basu BD.** 2006. Indian Medicinal Plants, 2, 856-860.
- Lee CK, Lee PH, Kuo YH.** 2001. The chemical constituents from the aril of *Cassia fistula* L. *Journal of the Chinese Chemical Society* **48**(6A), 1053-1058. <https://doi.org/10.1002/jccs.200100154>.
- Li HF, Guan XY, Yang WZ, Liu KD, Ye M, Sun C, Lu S, Guo DA.** 2012. Antioxidant flavonoids from *Epimedium wushanense*. *Fitoterapia* **83**(1), 44-48. DOI: 10.7717/peerj.8361.
- Majumder S, Afia IJ.** 2019. A comparative study on the antioxidant activity of *Cassia sophera* L. leaf and bark extracts. *Journal of Drug Delivery and Therapeutics* **9**(4-s), 177-181. <https://doi.org/10.22270/jddt.v9i4-s.3241>.
- Mates JM.** 2000. Effects of antioxidant enzymes in the molecular control of reactive oxygen species toxicology. *Toxicology* **153**, 83-104. DOI: 10.1016/S0300-483X(00)00306-1.
- Mehari B, Chandravanshi BS, Redi-Abshiro M, Combrinck S, McCrindle R, Atlabachew M.** 2021. Polyphenol contents of green coffee beans from different regions of Ethiopia. *International Journal of Food Properties* **24**(1), 17-27. <https://doi.org/10.1080/10942912.2020.1858866>.
- Miller HE, Rigelhof F, Marquart L, Prakash A, Kanter M.** 2000. Antioxidant content of whole grain breakfast cereals, fruits and vegetables. *Journal of the American College of Nutrition* **19**(3), 312s-319s. DOI: 10.1080/07315724.2000.10718966.
- Moura DJ, Richter MF, Boeira JM, Pêgas Henriques JA, Saffi J.** 2007. Antioxidant properties of β -carboline alkaloids are related to their antimutagenic and antigenotoxic activities. *Mutagenesis* **22**(4), 293-302. DOI: 10.1093/mutage/gem016.
- Mulaudzi N, Combrinck S, Vermaak I, Joubert E, Viljoen A.** 2022. High performance thin layer chromatography fingerprinting of rooibos (*Aspalathus linearis*) and honeybush (*Cyclopia genistoides*, *Cyclopia intermedia* and *Cyclopia subternata*) teas. *Journal of Applied Research on Medicinal and Aromatic Plants* **30**, 100378. DOI: 10.1016/j.jarmap.2022.100378.

Semwal RB, Semwal DK, Vermaak I, Viljoen

A. 2015. A comprehensive scientific overview of *Garcinia cambogia*. *Fitoterapia* **102**, 134-148.

DOI: 10.1016/j.fitote.2015.02.012.

Visioli F, Bellosta S, Galli CO. 1998. The bitter principles of olives enhance nitric oxide production by mouse macrophages. *Life Sciences* **62**, 541-546.

<https://doi.org/10.1002/med.1028>.

Wojdyło A, Oszmiański J, Czemerys R.

2007. Antioxidant activity and phenolic compounds in 32 selected herbs. *Food Chemistry* **105**(3), 940-949.

<https://doi.org/10.1016/j.foodchem.2007.04.038>.

Yao XH, Zhang DY, Zu YG, Fu YJ, Luo M, Gu CB, Li CY, Mu FS, Efferth T. 2013. Free radical scavenging capability, antioxidant activity, and chemical constituents of *Pyrola incarnata* Fisch.

leaves. *Industrial Crops and Products* **49**, 247-255.

DOI: 10.1016/j.indcrop.2013.04.058.

Yuan M, Zhang G, Bai W, Han X, Li C, Bian S.

2022. The role of bioactive compounds in natural products extracted from plants in cancer treatment and their mechanisms related to anticancer effects.

Oxidative Medicine and Cellular Longevity.

<https://doi.org/10.1155/2022/1429869>.

Yusuf M, Chowdhury JU, Wahab Ma, Begum

J. 1994. Medicinal plants of Bangladesh. BCSIR, Dhaka, Bangladesh, 149.