

OPEN ACCESS

Physico-chemical and biological characteristics of water and sediment in the mariculture zone park of Sto. Tomas, La union

Sancho V. Bilog^{*1,2}, Cjay B. Soliven¹, Victoria N. Malaya³, Rosario Segundina P. Gaerlan²

¹Iloilo State University of Fisheries Science and Technology College of Fisheries and Aquatic Sciences, Tiwi, Barotac Nuevo, Philippines

²Bureau of Fisheries and Aquatic Resources, Sevilla, San Fernando City, La Union, Philippines ³Don Mariano Marcos Memorial State University, Bacnotan, La Union, Philippines

Article published on March 08, 2025

Key words: Physico-chemical, Biological, Mariculture zone park

Abstract

Mariculture Park Program launched by DA-BFAR in the early 2000 primary aimed to promote sustainable supply of food, ensure job generation, and strengthen local development. In La Union, one of the two mariculture parks established is the Sto. Tomas MZP situated in Sto. Tomas Cove, Sto. Tomas, La Union. Formally launched in October 2002 with 60 HDPE fish cages maintained, the MZP has been a source of significant volume of quality milkfish. This study was designed to assess the effect of the mariculture activity on water quality and sediment. Specifically, it aimed to determine the physical, chemical, and biological characteristics of water and sediment in MZP Sto. Tomas and correlate water and sediment parameters with chlorophyll and phytoplankton. Samplings were conducted in the three established stations in MZP Sto. Tomas from February to May 2020. Water quality parameters (temperature, dissolved oxygen, salinity, pH, ammonia, nitrite, arsenic, cadmium, lead, mercury, fecal and total coliform count, chlorophyll, and phytoplankton density and composition) were analyzed for three months and sediment quality (color, organic matter, phosphorous, potassium, moisture content, pH, arsenic, mercury, lead, mercury) was analyzed for one month. Results showed spatial and temporal variability among selected water quality and no spatial variability among sediment quality from different stations. Of the twelve physico-chemical and biological parameters, ten passed the Philippine Standards. Twenty-eight species belonging to five phytoplankton groups were recorded. Sediment quality of the MZP Sto. Tomas is within range of the different International environmental standards. Correlation analysis revealed significant differences (p>0.05) in phytoplankton density and temperature, and strong correlation between temperature and chlorophyll.

*Corresponding Author: Sancho V. Bilog 🖂 sancho_bilog@yahoo.com

Introduction

Aquaculture has been developing rapidly for the previous decades and now dominates the fisheries sector. It contributes a significant supply of quality food for the consuming public and considered at the front line of the national government's programs to sustain food security and alleviate poverty. In the Philippines, top two (2) commodities produced which are commercially important are seaweeds and milkfish, among other aquaculture commodities such as shrimp, grouper, tilapia, siganids, and mussels. In 2020, the estimated total volume of fisheries production was 4,403.71 metric tons (MT), with total aquaculture production recorded at 2,324 MT contributing 52.7% in the total fisheries production (Philippine Statistics Authority, 2020).

The intensification of aquaculture was conceptualized by the government in partnership with private sectors due to the declining catch of marine fishes from small-scale or municipal fisheries and commercial fishing boat operators. From 2002 to 2010, commercial fisheries grew slightly from 1.042 million MT in 2002 to 1.24 million MT in 2010; however, it declined thereafter to 1.094 million MT in 2015 and 0.975 million MT in 2020. Aquaculture on the other hand, constantly and significantly contributes to fisheries production, similarly in terms of volume and value. Production from aquaculture since 2005 has been contributing 46%-52% to the total fisheries production, and accounts to 34%-39% to total production value (Ferrer et al., 2017). As such, the importance of aquaculture as a major source of food and generates employment today and in the future was given emphasis. It then became as one of the main programs of the government to increase the incremental fisheries production in the country. Nevertheless, owing to various aspects of planning, implementation, and monitoring, along with weak local fishery regulations, aquaculture practices particularly in fish cages in some areas, faced environmental degradation thereby resulting to fish kill occurrences. The coastal environment and the socio-economic welfare of the marginalized group still remain as major concerns.

Among the sectors of aquaculture, marine fish farming or mariculture of finfish in pens and cages, shellfish particularly oyster and mussel, and seaweed, contributes to about 73% of aquaculture production, and about 63% shares of seaweed farming (PSA, 2020). Fresh, brackish, and marine water cultures using fish pens and cages contribute 10%–12% in the total aquaculture production, but it has potential to increase rapidly. In the year 2010 for instance, only 2,700ha from the 50,150 ha of the 62 mariculture parks in the country was developed by a 2,199 investors and the Bureau of Fisheries and Aquatic Resources (BFAR) (Salayo *et al.*, 2012).

Philippine mariculture is defined as the culture or rearing of finfishes, shellfish, seaweeds and other commodities in marine environment using different structures such as cages, pens, stakes and rafts (Salayo et al., 2012). Usually, cages or pens are used for fin fishes adopting either monoculture (single species) or polyculture (multiple species) system. In the country, mariculture has been practiced for more than 600 years, but only in the year 1965 when fish cages were first introduced for the culture of common carp in Laguna de Bay seconded by milkfish culture in fish pens in the early 1970s. Milkfish culture in fish pens spread to Lingayen Gulf in the early 1990s whereas fish cages proliferate in Taal Lake. HDPE cages otherwise known as Norwegian cages were introduced in Sual Bay, Pangasinan in 1996 purposely for salmon culture (Ferrer et al., 2017). However, the development of mariculture has been difficult to monitor, and at times unregulated, especially on the aspect of stocking and feeding practices.

Fish farmers are not monitored during the installation of structures, thus, cages of any number and size are installed simply anywhere. The situation inevitably led to mass fish kills in cages and other environmental problems in some areas (Ferrer *et al.*, 2017). But with technological innovations, mariculture of seafood in coastal and open marine areas is now reliably supplying increasing amounts of protein. Based on PSA Report (2020), marine aquaculture production has

J. Bio. & Env. Sci. 2025

reached 81,850.56 MT and 102,294.57 MT in 2019 and 2020 respectively.

In the early 2000, the DA-BFAR introduced the Program on Mariculture Park so as to promote sustainable supply of food, ensure job generation, and improved income in order to encourage development in local level. It is a community-based marine type of project with the involvement of the organized fisherfolk within a coastal community through the assistance of the Municipal Fisheries and Aquatic Resources Management Councils (MFARMC). In La Union, there are two (2) mariculture parks established, one in Rosario and one in Sto. Tomas. Rosario Mariculture Zone Park is located in Barangay Bani and established in 2007. The Sto. Tomas MZP is situated in Sto. Tomas Cove located along the western side of Sto. Tomas, La Union.

The Cove was chosen to be the site of the Mariculture Zone Park (MZP) because it is a sheltered area that could protect the mariculture structures from strong winds, waves and currents during the southwest monsoon. Formally launched in October 2002, the mariculture zone is complimentary to make the area attractive to tourists. The ten (10) hectare mariculture area is covered by the Agoo-Damortis Protected Landscapes and Seascape (ADPLS), hence, it is managed by the Protected Area Management Board (PAMB) of the Department of Environment and Natural Resources (DENR).

The actual and possible effects of marine fish farming in cages on the environment, especially in water quality, are of utmost concern of the government agency in particular, to develop an ecologically responsible industry. A suitable water quality is required to sustain a feasible aquaculture production and the growing aquaculture industry. Unsuitable water quality may have led to less profit from poor quality of the product and potential risks to human health. There is a reduced production when the water contains contaminants which may lead to impaired growth and development, reproduction, or even mortality to the cultured species. Some of the contaminants can accumulate up to the point whereit threatens human health even in low quantities and cause no evident adverse effects. The environmental quality of water and the associate components of an aquatic ecosystem cannot be evaluated without learning the bottom sediment characteristics (Avramidis *et al.*, 2013). The sediment stratum is an important habitat for the benthic macro invertebrates whose metabolic activities contribute to aquatic productivity.

Recognizing the importance of Sto. Tomas Cove as a resource area and the critical role of its water quality in sustaining intensive aquaculture enterprises in the mariculture park, it is the paramount concern of the researcher to analyze the current status of the water and sediment quality in terms of its physical, chemical, and biological characteristics, and to correlate these parameters to the primary productivity of the MZP.

Materials and methods

Research design

This study is an experimental type of research using Complete Randomized Design (CRD) to determine the different variables of water and sediment. Likewise, a correlation research design was applied to determine the relationship between primary productivity (chlorophyll-a and phytoplankton density) and water and sediment quality.

Collection site

Water and sediment samples were collected in identified sampling sites. The first station situated in 16°14' 54.43"N and 120°22'47.24"E has Norwegian cage stocked with milkfish which is approximately 760meters away from the next site. The second station has the same structure with the first site situated in 16°14'35.28"N and 120°23'3.60"E. The third station has no cages in the area, which is approximately 1.1km away from the second site, has coordinates 16°14'11.61"N and 120°23'3.162"E (Figs 1&2).

Station	GPS coordinates		Description
	Latitude (N)	Longitude (E)	
1	16°14' 54.43"N	120°22'47.24"E	Norwegian cage owned by Mr. Medriano, stocked with milkfish
			Approximately 760 meters away from Station 2
2	16°14'35.28"N	120°23'3.60"E	Norwegian cages owned by Mr. Hilario, stocked with milkfish
	(a) 1 ())		Approximately 105 meters away from BFAR Station
3	16°14'11.61"N	120°23'31.62"E	No cages in the area, approximately 1.1 km away from Station 2

Table 1. Location and description of sampling station in Mariculture Zone Park of Sto. Tomas, La Unio



Fig. 1. Map showing the sampling stations



Fig. 2. Bathymetric profile of MZP Sto Tomas, La Union

Sample collection

Three (3) liters of water samples were collected in the sub surface of every sampling stations using water sampler. The collected samples were properly labelled and placed in a cooler box with coolant and brought to the laboratory for chemical (ammonia and nitrite), biological (chlorophyll-a), and heavy metal (arsenic, mercury, lead, cadmium) analysis. Separate water samples for microbiological analysis were collected in the sub surface using a sterilized glass sampling bottles covered with aluminum foil. Water samples were collected using the water sampler. The water samples were placed in a separate cooler box with coolant to avoid contamination and brought to the laboratory for the analysis of total and fecal coliform bacteria. A 20 μ m mesh size plankton net with water collector was used in the collection of water sample for plankton density and composition.

A total of three (3) samples of sediment were gathered using a soil grabber to about 10 meters depth. The grab has a hinge and lockable inspection flaps constructed with 0.5 mm mesh. The grab was slowly released to the water until it reaches the bottom part of the sites. Once it reaches the seabed, the rope connecting the grab was carefully pulled up allowing the grab to enclose with the collected sediment inside. One-two grab samples were taken at each site to complete the desired quantity for laboratory analysis and was placed in a plastic container with wide opening and labelled with site identification.

Data gathered

Physico-chemical characteristics of the water such as temperature, dissolved oxygen, salinity, and pH were gathered in situ at sub-surface level. The water ammonia and nitrite was analyzed using BFAR-NIFTDC's procedures, whereas chlorophyll-a was analyzed using the extraction fluorescence method of Guo et al. (2008). Total and fecal coliform content of collected water samples were analyzed using the Most Probable Number (MPN) method of bacteriological analysis. Water samples collected and submitted to CRL Environmental Corporation Laboratory were analyzed using Inductively Coupled Plasma - Optical Emission Spectroscopy (ICP-OES) for arsenic, cadmium and lead determination and Manual Cold Vapor Atomic Absorption Spectrophotometry for mercury determination. Plankton density was computed based on the description of the Photo Guide to cyanobacteria (IOC- IOC-Danida Advanced

J. Bio. & Env. Sci. 2025

Workshop on HAB. Workshop IV: Cyanobacteria, 2004) using the formula of: D= (N×V1)/Vs.

Sediment samples were analyzed following the Walkley and Black Spectrophotometric method for organic matter, Olsen method for available phosphorous, Cold Sulfuric Acid Extraction Flame Atomic Emission Spectroscopy method for available potassium, Feel method for texture, Gravimetric method for moisture content, and Potentiometric/Conductimetric method for pH and electrical conductivity for heavy texture samples. The method for arsenic determination followed the procedure of the Inductively Coupled Plasma -Optical Emission Spectroscopy, Flame Atomic Absorption Spectrophotometry (AAS) for cadmium and lead, and Manual Cold Vapor Atomic Absorption Spectrophotometry for mercury.

Data analysis

Data recorded on physico-chemical and biological characteristics of water and sediment quality were analyzed using Kruskal-Wallis Test and Pairwise Comparisons at 5% level of significance. Likewise, correlation analysis on primary productivity (chlorophyll-a and phytoplankton density) between water and sediment quality was done using Bivariable Analysis of data using Inter-Basal Statistical Product and Service Solutions (version 27).

Results and discussion

Water physico-chemical and biological characteristics

There was no observed spatial variability among water quality like temperature, dissolved oxygen, salinity, pH, ammonia, arsenic, cadmium, lead, fecal coliform, chlorophyll, mercury, and phytoplankton density, from the different monitoring stations which means that fewer but more evenlyspaced monitoring stations are recommended for the MZP (Table 2a-b, Figs 3, 4&5). Dissolved oxygen and pH have exceeded the DENR-DAO directive for Class SC waters, while others (temperature, salinity, ammonia, arsenic, cadmium, lead, mercury, fecal and total coliform, chlorophyll, and phytoplankton density) are all within desirable level as compared to standards. Fecal coliform was not detected in all stations, whereas total coliform was detected only at Station 1.

Table 2a. Mean values of water parameters in MZP Sto. Tomas

Water parameters	Station 1	Station 2	Station 3	Mean	DENR standard*
Temperature (°C)	28.89±1.39	28.93±1.48	28.92±1.52	28.84±1.29	25-31
DO (ppm)	5.03 ± 0.43	4.73 ± 0.52	5.32 ± 0.42	5.07 ± 0.47	5
Salinity (ppt)	36.00±2.36	35.83 ± 2.04	35.00±3.162	35.70 ± 2.39	None
pH	8.64±0.15	8.68±0.09	8.71±0.10	8.68 ± 0.11	6.5-8.5
Ammonia (mg/L)	0.032 ± 0.024	0.027 ± 0.03	0.04 ± 0.03	0.037 ± 0.027	0.05
Nitrite (mg/L)	0.013±0.009 ^b	0.009±0.006 ^a	0.012 ± 0.007^{b}	0.011±0.006	<0.1***
Arsenic (mg/L)	< 0.008	< 0.008	< 0.008	0.008 ± 0.00	0.02
Cadmium (mg/L)	< 0.001	< 0.001	< 0.001	0.001 ± 0.00	0.005
Lead (mg/L)	< 0.005	< 0.005	< 0.005	0.005 ± 0.00	0.05
Mercury (mg/L)	< 0.0002	< 0.0002	< 0.0002	0.0002 ± 0.00	0.002
Fecal Coliform	0.00 ± 0.00	0.00 ± 0.00	0.00 ± 0.00	0.00 ± 0.00	200**
(MPN/100ml)					
Total Coliform	0.79±1.36 ^a	0.00 ± 0.00^{b}	0.00 ± 0.00 b	$0.26 \pm .851$	5,000**
(MPN/100ml)					
Chlorophyll (mg/m ³)	0.006±0.008	0.006±0.006	0.005 ± 0.008	0.005 ± 0.007	None
Phytoplankton density	3.8945 × 103	2.494 × 10 ³	4.6 × 10 ³	3.655×10^{3}	None
(cells/Li)	± 6608.05	±3947.23	± 8724.53	± 6278.82	

References: *DAO 2016-08, ** DAO 1990-34 (Class SB & SC), ***PHILMINAQ, 2006

^a Mean values in a column having superscripts differ significantly at .05 level of significance.

Phytoplankton density is most abundant at Station 3 and least at Station 2 which is composed mainly of diatoms (Bacillariophyceae) and dinoflagellates (Dinophyceae). Twenty eight (28) phytoplankton species were identified from the three (3) stations, covered by five (5) classes namely, Bacillariophyceae, Dinophyceae, Oligotrichea, Mediophyceae, and Raphidophyceae.

		Samplii	ng		
Initial	First	Second	Third	Fourth	Final
(February 18)	(March 3)	(March 17)	(March 31)	(April 14)	(May 14)
27.53	27.30	28.85	29.13	29.26	31.38
$\pm 0.13^{ab}$	±0.084 ^a	±0.093 ^b	±.0.04 ^{bc}	$\pm .0.05$ ^{cd}	$\pm .0.15^{d}$
5.00±0.25 ^a	5.35±0.18 ^a	4.99±0.487 ^b	5.46±0.312 ^a	5.17 ± 0.225^{b}	4.28 ± 0.29^{bc}
34.29±2.45 ^a	39.33±0.48 ^b	37.36±0.496 ^b	35.00 ± 0.00^{a}	35.00 ± 0.00 a	32.64±0.501 ^a
8.70 ± 0.027^{ab}	8.66±0.060 ^b	8.66 ± 0.022^{b}	8.819±0.062 ^{cd}	8.70±0.084 a	8.52±0.133 ^e
0.048±0.004 ^a	0.052±0.0067 ^a	0.043±0.038 ^a	No data	No data	0.00±0.00 ^b
0.016 ± 0.003^{a}	0.013±0.003 ^a	0.0014 ± 0.0012^{1}	^o No data	No data	0.016 ± 0.005^{a}
0.008 ± 0.00	-	-	-	-	-
0.001±0.00	-	-	-	-	-
0.005 ± 0.00	-	-	-	-	-
0.0002 ± 0.00	-	-	-	-	-
0.00	0.00	No data	0.00	No data	0.016
±0.00	±0.00		±0.00		±0.0046
1.05	0.00	No data	0.00 ± 0.00	No data	0.016
±1.48	±0.00				±0.0046
0.017	0.0016	0.001	No data	No data	0.003
$\pm 0.001^{a}$	±0.0004 ^{bc}	±0.000 ^b			±0.002 ^{cd}
1.6937×10^{4}	5.856×10^{2}	7.7749×10^{2}	7.812×10^{2}	1.4409×10^{3}	1.6632×10^{3}
±4963.24 ^a	±86.014 ^b	± 302.354 ^{bc}	±608.11 ^b	± 240.012 ^{cd}	±228.611 ac
	$\begin{tabular}{ l l l l l l l l l l l l l l l l l l l$	$\begin{tabular}{ c c c c } \hline Initial & First & (March 3) \\ \hline (February 18) & (March 3) \\ \hline (27.53 & 27.30 & & & & & & & & & & & & & & & & & & &$	$\begin{tabular}{ c c c c } \hline Samplin \\ \hline Initial & First & Second \\ \hline (February 18) & (March 3) & (March 17) \\ \hline 27.53 & 27.30 & 28.85 \\ \pm 0.13^{ab} & \pm 0.084^{a} & \pm 0.093^{b} \\ 5.00\pm 0.25^{a} & 5.35\pm 0.18^{a} & 4.99\pm 0.487^{b} \\ 34.29\pm 2.45^{a} & 39.33\pm 0.48^{b} & 37.36\pm 0.496^{b} \\ 8.70\pm 0.027^{ab} & 8.66\pm 0.060^{b} & 8.66\pm 0.022^{b} \\ 0.048\pm 0.004^{a} & 0.052\pm 0.0067^{a} & 0.043\pm 0.038^{a} \\ 0.016\pm 0.003^{a} & 0.013\pm 0.003^{a} & 0.0014\pm 0.0012^{b} \\ 0.008\pm 0.00 & - & - \\ 0.001\pm 0.00 & - & - \\ 0.000\pm 0.00 & - & - \\ 0.000\pm 0.00 & - & - \\ 0.000 & 0.00 & No data \\ \pm 0.00 & \pm 0.00 \\ 1.05 & 0.00 & No data \\ \pm 1.48 & \pm 0.00 \\ 0.017 & 0.0016 & 0.001 \\ \pm 0.001^{a} & \pm 0.004^{bc} & \pm 0.000^{b} \\ 1.6937 \times 10^{4} & 5.856 \times 10^{2} & 7.7749 \times 10^{2} \\ \pm 4963.24^{a} & \pm 86.014^{b} & \pm 302.354^{bc} \\ \hline \end{tabular}$	$\begin{array}{c c c c c c } \hline Sampling \\ \hline Initial & First & Second & Third \\ \hline (February 18) & (March 3) & (March 17) & (March 31) \\ \hline 27.53 & 27.30 & 28.85 & 29.13 \\ \pm 0.13^{ab} & \pm 0.084^{a} & \pm 0.093^{b} & \pm .0.04^{bc} \\ 5.00\pm 0.25^{a} & 5.35\pm 0.18^{a} & 4.99\pm 0.487^{b} & 5.46\pm 0.312^{a} \\ 34.29\pm 2.45^{a} & 39.33\pm 0.48^{b} & 37.36\pm 0.496^{b} & 35.00\pm 0.00^{a} \\ 8.70\pm 0.027^{ab} & 8.66\pm 0.060^{b} & 8.66\pm 0.022^{b} & 8.819\pm 0.062^{cd} \\ 0.048\pm 0.004^{a} & 0.052\pm 0.0067^{a} & 0.043\pm 0.038^{a} & No data \\ 0.016\pm 0.003^{a} & 0.013\pm 0.003^{a} & 0.0014\pm 0.0012^{b} No data \\ 0.008\pm 0.00 & - & - & - \\ 0.001\pm 0.00 & - & - & - \\ 0.000\pm 0.00 & - & - & - \\ 0.000\pm 0.00 & - & - & - \\ 0.000 & 0.00 & No data & 0.00 \\ \pm 0.00 & \pm 0.00 & & \pm 0.00 \\ 1.05 & 0.00 & No data & 0.00\pm 0.00 \\ \pm 1.48 & \pm 0.00 & & \\ 1.6937\times 10^{4} & 5.856\times 10^{2} & 7.7749\times 10^{2} & 7.812\times 10^{2} \\ \pm 4963.24^{a} & \pm 86.014^{b} & \pm 302.354^{bc} & \pm 608.11^{b} \\ \end{array}$	$\begin{array}{ c c c c c } \hline Sampling \\ \hline Initial & First & Second & Third & Fourth \\ \hline (February 18) & (March 3) & (March 17) & (March 31) & (April 14) \\ \hline 27.53 & 27.30 & 28.85 & 29.13 & 29.26 \\ \pm 0.13^{ab} & \pm 0.084^a & \pm 0.093^b & \pm 0.04^{bc} & \pm 0.05^{cd} \\ 5.00\pm0.25^a & 5.35\pm0.18^a & 4.99\pm0.487^b & 5.46\pm0.312^a & 5.17\pm0.225^b \\ 34.29\pm2.45^a & 39.33\pm0.48^b & 37.36\pm0.496^b & 35.00\pm0.00^a \\ 8.70\pm0.027^{ab} & 8.66\pm0.060^b & 8.66\pm0.022^b & 8.819\pm0.062^{cd} & 8.70\pm0.084^a \\ 0.048\pm0.004^a & 0.052\pm0.0067^a & 0.043\pm0.038^a & No data & No data \\ 0.016\pm0.003^a & 0.013\pm0.003^a & 0.0014\pm0.0012^bNo data & No data \\ 0.008\pm0.00 & - & - & - & - \\ 0.001\pm0.00 & - & - & - & - \\ 0.000\pm0.00 & - & - & - & - \\ 0.000\pm0.00 & - & - & - & - \\ 0.000 & 0.00 & No data & 0.00 & No data \\ \pm 0.00 & 10.00 & & 10.00 & & \pm 0.00 \\ 1.05 & 0.00 & No data & 0.00\pm0.00 & No data \\ \pm 1.48 & \pm 0.00 & & & \pm 0.00 \\ 1.05 & 0.00 & No data & 0.00\pm0.00 & No data \\ \pm 1.48 & \pm 0.00 & & & & \\ \pm 1.48 & \pm 0.00 & & & & \\ \pm 0.001^a & \pm 0.004^{bc} & \pm 0.000^b & & \\ \pm 1.6937 \times 10^4 & 5.856 \times 10^2 & 7.7749 \times 10^2 & 7.812 \times 10^2 & 1.4409 \times 10^3 \\ \pm 4963.24^a & \pm 86.014^b & \pm 302.354^{bc} & \pm 608.11^b & \pm 240.012^{cd} \\ \hline \end{array}$

Table 2b. Mean values of wate	parameters in the MZP Sto. Tomas	per sampling (February to May 20	20
-------------------------------	----------------------------------	----------------------------------	----

* Mean values in a column having different superscripts differ significantly at .05 level of significance. Arsenic, Cadmium, Lead, and Mercury were sampled only once, during the initial sampling



Fig. 3. Plankton density (a), over all species composition (b), and representative genera (c) in MZP Sto.Tomas

Significant differences (p<0.05) were noted on nitrite and total coliform in all stations. Temporal variations in water quality parameters like temperature, dissolved oxygen, salinity, pH, ammonia, nitrite, chlorophyll, and phytoplankton density were detected. The highest temperature was recorded during the final sampling on May 2020, whereas the lowest temperature was recorded during the first sampling on the month of March.



Fig. 4. Plankton density (a), composition (b,c), and abundant species (d) per sampling

The range of dissolved oxygen during the sampling was 4.28 ppm to 5.46 ppm, with the highest mean value recorded during the third sampling and lowest during the final sampling on May. Highest salinity

93 | Bilog et al.

value of 39.33 ppt was recorded during the first sampling period, and lowest value of 32.64 ppt during the final sampling. The third sampling had the highest pH mean value of 8.81, and lowest value of 8.52 during the final sampling on May 2020. Ammonia level ranged from 0.00 mg L⁻¹ to 0.052 ± 0.03 mg L⁻¹, highest level was recorded during the first sampling period and lowest during the final sampling. Highest nitrite level is recorded during the initial and final sampling (0.016 mg L⁻¹) and lowest during the second sampling. Fecal coliform is not detected in all sampling. On the other hand, total coliform was only recorded during the initial sampling on February.



Fig. 5. Water quality (a-temperature, b-DO, c – Salinity, d-pH, e-Phytoplankton density, f-species composition) of Station 2, MZP Sto. Tomas during diurnal sampling

Chlorophyll mean values were recorded at a range of 0.0016 mg m³ to 0.017 mg m³, wherein the highest value was recorded during the initial sampling and lowest on the first sampling on the month of March. Phytoplankton was most abundant during the initial sampling on a hot, sunny day of February and lowest during the first sampling. *Skeletonema* sp. belonging to Class Bacillariophyceae is highest at 20,851 cells L⁻¹ during the initial sampling, whereas Pseudonitzschia sp. was lowest during the first sampling at 11 c cells L-¹. Statistical analysis using Krukal-Wallis test showed that temperature, DO, salinity, pH, ammonia, nitrite, chlorophyll, and phytoplankton density are statistically significant (p<0.05). These variations are attributed to natural causes such as tidal fluctuations and rainfall. As regard to the diurnal sampling in Station 2, temporal variability (6 hours interval) was not observed among water quality (temperature, dissolved oxygen, salinity, pH, and phytoplankton density) from different monitoring time. However, temporal variability was detected across sampling of water quality (temperature, salinity, pH) for the 3month diurnal sampling. Highest temperature was noted in 8:00 AM sampling on the final sampling and lowest on 8:00 PM of the initial sampling on March. Significant differences (p<0.05) was observed in temperature across the sampling periods from March to May. DO was lowest in 8:00 AM of the final sampling, and highest during 2:00 PM of the first sampling on April. Same salinity was recorded during sampling in the morning (8:00 AM), evening (8:00 PM), and dawn (2:00 AM) at 35.00±2.00 ppt. pH value is highest at 8:00AM and lowest in the evening and dawn. A significant difference (p<0.05) was observed in pH (p=0.048) across the sampling periods from March to May. Highest density of phytoplankton (1.201 × $10^3 \pm 494.54$) is recorded in the morning (8:00 AM) sampling period and lowest in the evening (8:00 PM) sampling (1.016 \times 10² ± 87.85). Phytoplankton density was highest when the temperature is highest at the same time of 8:00 AM, during the final sampling. Twenty three (23) phytoplankton species were identified from the diurnal sampling. These species cover three (3) classes namely, Bacillariophyceae, Dinophyceae, and Oligotricheae.

Sediment physico-chemical and biological characteristics

In terms of sediment quality, Station 2 has black-colored sediment as compared to the other two (2) Stations wherein mariculture is minimal or absent. Data obtained for some parameters like organic matter content, phosphorous, and potassium are baseline data (Table 3). The highest organic matter content (%) is recorded at Station 2 with 3.05, followed by Stations 1 and 2. Station 1 has the highest available phosphorous with 37.93 ppm, followed by Station 2 and Station 3 with 33.66 ppm and 28.22 ppm, respectively. The highest available potassium was obtained at Station 2 with 1,375.87 ppm, followed by Station 1 with 968.65 ppm and Station 3 with 936.09 ppm. The highest recorded moisture content is at Station 3 with 9.06% and lowest moisture content of 7.64% in Station 2. Station 1 has the lowest pH values of 6.73 and highest pH values of 8.31 in Station 3. Stations 1 and 2 have the same electrical conductivity levels of 0.02 mS cm⁻¹ while Station 3 is the lowest at 0.02mS cm⁻¹. Results of the initial analysis of composite sediment samples on 2016 shows the concentrations of Arsenic have greatly increased from 1.6 ppm to 3.58 ppm (mean of 2020). Cadmium (Cd) content of the surface sediment is highest at Station 2 with 1.2 ppm and lowest at Station 3 with 1 ppm. Stations 1 and 3 have the same Lead content of 11 ppm, Station 2 having the highest content of 12 ppm. The mercury content of sediment is not detected in the samples collected for this study. There was no observed spatial variability of different parameters such as organic matter, phosphorous, potassium, moisture content, pH, electrical conductivity, arsenic, mercury, lead, and mercury) from the three stations.

Table 3. Mean values of sediment parameters in MZP Sto. To	nas
--	-----

Sediment parameters	Station 1	Station 2	Station 3	Mean	Guidelines
Color	Gray	Black	Gray		
Organic matter (%)	1.62	3.05	1.44	2.0 ± 0.731	None
Phosphorous (ppm)	37.93	33.66	28.22	33.29±4.182	30-60ppm*
Potassium (ppm)	968.65	1,375.87	936.09	1083.75±203.14	None
Moisture Content (%)	8.02	7.64	9.06	8.25±0.619	None
pH	6.73	7.56	8.31	7.52 ± 0.679	None
Electrical conductivity (mS/cm)	0.02	0.02	0.01	0.016±0.0049	None
Arsenic (mg/kg)	4.2	2.2	4.2	3.58 ± 0.957	8.2**
Cadmium (mg/kg)	1.1	1.2	1	1.097±0.083	1.2^{**}
Lead (mg/kg)	11	12	11	11.31±0.479	46.7**
Mercury (mg/kg)	ND	ND	ND	0.00 ± 0.00	0.15^{**}
ND – Not Detected	* After Ban	erjea (1967)	**NOAA	ERL, 1999	

	. 1	•	1 1 1	
Table 4 Correlation between water	narameters and	nrimarv i	roductivity	٧7
rubic 4. correlation between water	purumeters unu	primary	nouucuiin	y.

Parameters	Station	Mean Ranges	Chlorophyll	Phytoplankton density
Temperature (°C)	1	28.81±1.25	-0.358	-0.372 ^a
	2	28.84±1.33	-0.169	-0.377 ^a
	3	28.87±1.36	-0.407	-0.432 ^a
DO (ppm)	1	5.06 ± 0.38	0.216	0.163
	2	4.78±0.48	-0.069	-0.195
	3	5.35 ± 0.38	-0.163	-0.329
Salinity (ppt)	1	36.12 ± 2.25	-0.172	-0.076
	2	35.91 ± 1.883	-0.384	-0.057
	3	35.12±2.89	0.713	-0.668
pH	1	8.65±0.13	0.242	0.081
	2	8.68±0.09	0.667	-0.044
	3	8.72±0.097	0.735	0.060
Ammonia (mg/L)	1	0.03 ± 0.02	0.217	0.170
	2	0.029 ± 0.027	0.368	0.441
	3	0.047±0.032	0.033	0.024
Nitrite (mg/L)	1	0.01±0.01	0.398	0.363
	2	0.009 ± 0.005	0.359	0.201
	3	0.012±0.006	0.730	0.736
Total Coliform	1	0.79±1.357	1.000	0.998
	2	N/A	N/A	N/A
	3	N/A	N/A	N/A

* Mean values in a column having the same superscripts differ significantly at .05 level of significance. N/A Not analyzed

Correlation of water parameters and sediment quality on the productivity

Overall results revealed that temperature and primary productivity are inversely related. Temperature significantly influenced productivity in terms of chlorophyll and phytoplankton density (Table 4). In Station 1, dissolved oxygen is directly related with chlorophyll and phytoplankton density while in Stations 2 and 3, dissolved oxygen and chlorophyll and phytoplankton density are inversely related. In all weak associations and insignificant stations, correlation between the parameters were found. DO did not significantly influenced the primary productivity in the mariculture area. Weak negative correlation was noted between salinity and chlorophyll at Stations 1 and 2, and a strong positive correlation at Station 3. Phytoplankton density in all stations is negatively correlated to salinity, with weak correlation at Station 1 and Station 2, and moderate correlation at Station 3. pH has a positive weak correlation to chlorophyll at Station 1, moderate correlation at Station 2 and strong correlation at Station 3. Phytoplankton density in Stations 1 and 3 have direct correlation with pH, however, negatively correlated in Station 2. Phytoplankton density and pH have weak associations and insignificant correlation in all stations.

Ammonia is directly related with primary productivity indicators but weak association was found between the two parameters. In terms of correlation to chlorophyll, a weak association was recorded in all stations. Ammonia has positive weak to moderate correlation to phytoplankton density in all stations. Nitrite is positively correlated to chlorophyll and phytoplankton. A weak to strong correlation of nitrite to chlorophyll was observed. Phytoplankton has a positive weak and strong correlation recorded. Total coliform in Station 1 has a strong to perfect correlation to phytoplankton and chlorophyll.

Conclusion

Based on the results obtained in the study, the following conclusions are drawn:

- 1. Of the 12 physico-chemical and biological water quality parameters tested, 10 passed the standards (DENR-DAO PHILMINAQ), and namely: temperature, salinity, ammonia, nitrite, arsenic, cadmium, lead, mercury, fecal coliform, total coliform; while DO and pH exceeded the limit of criteria for water quality standard for Class SC waters. Spatio-temporal and diurnal variations on water quality in MZP Sto. Tomas were noted in some sites and parameters. Diurnal sampling revealed that DO level starts to decrease at dawn (2:00 AM) until 8:00 AM onwards. Desirable level of DO (5 ppm) was recorded at 2:00 PM sampling. Water quality and sediment are in good condition, thus, are able to sustain the intensive aquaculture enterprises in the Mariculture Zone Park Sto. Tomas.
- 2. There was an accumulation of heavy metals (arsenic, cadmium, lead) in the sediment of the MZP Sto. Tomas for the four (4) years of mariculture operations, but still within range of the different international environmental standards. Cadmium level in the sediments of MZP Sto. Tomas is classified under the Effect Range Low (ERL) of NOAA (1999).
- 3. Temperature significantly influenced the primary productivity (chlorophyll and phytoplankton density) of the MZP Sto. Tomas. Dissolved oxygen, salinity, pH, ammonia, nitrite, and total coliform has insignificant correlation with primary productivity variables, thus did not influence the chlorophyll and phytoplankton density in the MZP.

Recommendations

Based on the conclusions obtained in the study, the following recommendations are drawn:

- Sustain continuous monitoring of water quality parameters (temperature, dissolved oxygen, salinity, pH, ammonia, nitrite), heavy metals, and coliform content. Intensive monitoring should be done during marketable size of the stocks to prevent mortalities.
- 2. Further study on the microbiota of the sediment to determine its possible impacts in the mariculture

area brought about by feeding practices and nutrient loading.

- Sustain phytoplankton species identification to determine presence of harmful algal blooms (HABs) that may cause water quality deterioration in MZP Sto. Tomas.
- 4. Further study on the sediment quality to include other parameters such as grain size, hydrogen sulphide, carbon, nitrogen, etc. within MZP to investigate nutrient loading.
- 5. Strict implementation of good aquaculture practices of the mariculture investors to sustain their operations without causing environmental degradation.

References

A'yun Q, Takarina D. 2017. Ambient temperature effects on growth of milkfish (*Chanos chanos*) at aquaculture scale in Blanakan, West Java. International Symposium on Current Progress in Mathematics and Sciences 2016.

https://doi.org/10.1063/1.4991221.

Adesuyi A, Ngwoke M, Akinola M, Njoku K, Jolaoso A. 2016. Assessment of physicochemical characteristics of sediment from Nwaja Creek, Niger Delta, Nigeria. Journal of Geoscience and Environment Protection **4**, 16-27. http://dx.doi.org/10.4236/gep.2016.41002.

Ajithamol A, Venkadesh B, Babu M, Saraswathy S. 2016. Impact of nutrients nitrogen, phosphorus, potassium and their role in the sediment of Manakudy Estuary, Tamilnadu, South India. International Journal of Current Research **8**(4), 29248-29251.

Albelda R, Purganan DJ, Gomez NC, Narvarte BC, Calalang PC, Genovia TG, Gernato EG, Bondoc KG, San Diego-McGlone ML, Onda DF. 2019. Summer phytoplankton community structure and distribution in a mariculture-affected coastal environment. Philippine Science Letters 12(2), 157-166. Algul F, Beyhan M. 2020. Concentrations and sources of heavy metals in shallow sediments in Lake Bafa, Turkey. Scientific Reports 10, 11782. https://doi.org/10.1038/s41598-020-68833-2.

Ali H, Khan E, Ilahi I. 2019. Environmental chemistry and ecotoxicology of hazardous heavy metals: Environmental persistence, toxicity, and bioaccumulation. Journal of Chemistry **2019**, 1-14. https://doi.org/10.1155/2019/6730305.

Altinok I, Ozturk R. 2017. Adverse effects of mariculture activities and practices on marine environment. Oceanography and Fisheries **4**(1), 1-8.

Andleeb S, Maryam M, Aslam A. 2014. Quantitative analysis of a fish pond for coliform bacterial content. Punjab University Journal of Zoology **29**(1), 33-39.

Argente F, Palla H, Narido C, Celedonio M, Dy D. 2013. Spatial distribution of organic matter in the surface sediments of Calape, Bohol, Central Philippines. Philippine Journal of Science. Retrieved on August 11, 2020.

Ashour M, Abo-Taleb H, Abou-Mahmoud M, El-Feky M. 2018. Effect of the integration between plankton natural productivity and environmental assessment of irrigation water, El-Mahmoudia Canal, on aquaculture potential of *Oreochromis niloticus*. Turkish Journal of Fisheries and Aquatic Sciences **18**, 1163-1175. https://doi.org/10.4194/1303-2712-v18_10_03.

Aslam S, Venzi M, Venkatraman V, Mikkelsen O. 2020. Chemical assessment of marine sediments in vicinity of Norwegian fish farms – A pilot study. Science of the Total Environment **732**, 139130.

https://doi.org/10.1016/j.scitotenv.2020.139130.

Avramidis P, Samiotis A, Kalimani E, Papoulis D, Lampropoulou P, Bekiari V. 2013. Sediment characteristics and water physicochemical parameters of the Lysimachia Lake, Western Greece. Environmental Earth Sciences **70**, 383-392.

https://doi.org/10.1007/s12665-012-2134-9.

Baleta MJ, Bolaños JM. 2016. Phytoplankton identification and water quality monitoring along the fish-cage belt at Magat Dam Reservoir, Philippines. International Journal of Fisheries and Aquatic Studies 4(3), 254-260.

Beldowski J, Szubska M, Beldowska M, Jankowska K, Kotlarska E, Graca B. 2018. Seasonal changes of mercury speciation in the coastal sediments. Journal of Soils and Sediments **18**, 3424-3436. https://doi.org/10.1007/s11368-018-1993-4.

Bisht A, Singh U, Pandey N. 2014. Comparative study of seasonal variation in bacterial flora concomitant with farm-raised fingerlings of *Cyprinus carpio* at Tarai Region of Uttarakhand. Journal of Environmental Biology **35**(2), 363-367.

Boyd CE. 2000. Bacteria, phytoplankton, and water quality. In: Water Quality. Springer, Boston, MA. https://doi.org/10.1007/978-1-4615-4485-2_9.

Bureau of Fisheries and Aquatic Resources – National Integrated Fisheries Technology Development Center (BFAR NIFTDC). 2019. MZP Sto. Tomas Water Current Profiling.

Bureau of Fisheries and Aquatic Resources – **National Integrated Fisheries Technology Development Center (BFAR NIFTDC).** Undated. Laboratory procedure for ammonia and nitrite determination.

Cam P, Dalsgaard A, Mara D. 2007. Microbiological quality of fish grown in wastewaterfed and non-wastewater-fed fishponds in Hanoi, Vietnam: influence of hygiene practices in local retail markets. Journal of Water and Health **5**(2), 209-218.

Chatta A, Khan M, Mirza Z, Ali A. 2016. Heavy metal (cadmium, lead, and chromium) contamination in farmed fish: a potential risk for consumers' health. Turkish Journal of Zoology **40**, 1-10.

https://doi.org/10.3906/zoo-1506-1.

Chen H, Zhou W, Chen W, Xie W, Jiang L, Liang Q, Huang M, Wu Z, Wang Y. 2017. Simplified, rapid, and inexpensive estimation of primary productivity based on chlorophyll fluorescence parameter Fo. Journal of Plant Physiology **128**, 128-135. https://doi.org/10.1016/j.jplph.2016.12.015.

Daniel S, Larry W, Joseph H. 2005. Comparative oxygen consumption and metabolism of striped bass (*Morone saxatilis*) and its hybrid. Journal of World Aquaculture Society **36**(4), 521-529.

Demir N, Kirkagac M, Pulatsü S, Bekcan S. 2001. Influence of trout cage culture on water quality, plankton, and benthos in an Anatolian dam lake. The Israeli Journal of Aquaculture – Bamidgeh **53**(3-4), 115-127.

DENR Administrative Order No. 2016-08. Water Quality Guidelines and General Effluent Standards of 2016.

Devi P, Padmavathy P, Aanand S, Aruljothi K. 2017. Review on water quality parameters in freshwater cage fish culture. International Journal of Applied Research **3**(5), 114-120.

Dias J, Takahashi E, Santana N, Bonecker C. 2011. Impact of fish cage culture on the community structure of zooplankton in a tropical reservoir. Iheringia, Série Zoologia **101**(1-2).

https://doi.org/10.1590/S0073-47212011000100011.

Echapare E, Pacala FA, Mendaío A, Araza J. 2019. Physico-chemical and microbial analysis of water in Samar mussel farms. Egyptian Journal of Basic and Applied Sciences **45**, 225-230.

https://doi.org/10.1016/j.ejar.2019.05.007.

Ferrer AJ, Francisco H, Predo C, Carmelita B, Hopanda J. 2017. The first 15 years of mariculture parks in the Philippines: Challenges and the way forward. EEPSEA Research Report No. 2017-RR9. Economy and Environment Program for Southeast Asia, Laguna, Philippines. **Garcia LM.** Undated. Technical Paper on Fisheries Biology of Milkfish (*Chanos chanos* Forskal). Proceedings of the Regional Workshop on Milkfish Culture Development in the South Pacific.

Guo P, Baum M, Varshney R, Graner A, Grando S, Ceccarelli S. 2008. QTLs for chlorophyll and chlorophyll fluorescence parameters in barley under post-flowering drought. Euphytica **163**, 203-214. https://doi.org/10.1007/s10681-007-9629-6.

Han J, Park H, Kim J, Jeong D, Kang J. 2019. Toxic effects of arsenic on growth, hematological parameters, and plasma components of starry flounder (*Platichthys stellatus*) at two water temperature conditions. Fisheries and Aquatic Sciences **22**(3). https://doi.org/10.1186/s41240-019-0116-5.

Hansson K, Nicolle A, Graneli W, Hallgren P. 2013. Food-chain length alters community responses to global change in aquatic systems. Nature Reports Climate Change **3**(3), 228-233. https://doi.org/10.1038/nvlimate1689.

Ismail I, Saleh I. 2012. Analysis of heavy metals in water and fish (*Tilapia* sp.) samples from Tasik Mutiara, Puchong. The Malaysian Journal of Analytical Sciences **16**(3), 346-352.

Jamshidi S, Bakar N. 2011. A study on distribution of chlorophyll-a in the coastal waters of Anzali port, South Caspian Sea. Ocean Science Discussions **8**, 435-451. https://doi.org/10.5194/osd-8-435-2011.

Jedruch A, Beldowski J, Beldowska M. 2015. Long-term changes and distribution of mercury concentrations in surface sediments of the Gdansk Basin (Southern Baltic Sea). Journal of Soils and Sediments 15, 2487-2497.

Kadim M, Pasisingi N, Kasim F. 2018. Spatial and temporal distribution of phytoplankton in the Gorontalo Bay, Indonesia. AACL Bioflux **11**(3), 833-845.

Khalifa K, Hamil A, Al-Houni A, Ackacha M. 2010. Determination of heavy metals in fish species of the Mediterranean Sea (Libyan coastline) using Atomic Absorption Spectrometry. International Journal of ChemTech Research **2**(2).

Kromkamp J, Capuzzo E, Philippart C. 2017. Measuring phytoplankton primary production: Review of existing methodologies and suggestions for a common approach. Report for the EcApRHA project (Applying an Ecosystem Approach to (sub) Regional Habitat Assessment).

Kumar P, Jetani K, Yusuzai S, Sayani A, Dar S, Rather M. 2012. Effect of sediment and water quality parameters on the productivity of coastal shrimp farm. Pelagia Research Library **3**(4), 2033-2041.

Lee H, Park H, Lee J, Park A, Cheon S. 2016. Coliform pollution status of Nakdong river and tributaries. J Korean Soc Water Environ **32**, 271-280.

Longphuirt S, McDermott G, O'Boyle S, Wilkes R, Stengel D. 2019. Decoupling abundance and biomass of phytoplankton communities under different environmental controls: A new multi-metric index. Frontiers in Marine Science 6, 312. https://doi.org/10.3389/fmars.2019.00312.

Mallasen M, Barros H, Traficante D, Camargo A. 2012. Influence of a net cage tilapia culture on the water quality of the Nova Avanhandava reservoir, São Paulo State, Brazil. Acta Scientiarum Biological Sciences **34**(3), 289-296.

Mangaliso J, Guilford S, Hecky R. 2011. Physicalchemical measurements in the water column along a transect through a tilapia cage fish farm in Lake Malawi, Africa. Journal of Great Lakes Research **37**, 102-113.

Mohanty A, Bramha S, Satpathy K, Padhi R, Panigrahi S, Samantara M, Kumar S, Sarkar S, Prasad M. 2018. Geochemical distribution of forms of phosphorus in marine sediment of Bay of Bengal, southeast coast of India. Indian Journal of Geo Marine Sciences 47(6), 1132-1141. Moncada C, Hassenruck C, Gardes A, Conaco C. 2019. Microbial community composition of sediments influenced by intensive mariculture activity. FEMS Microbiology Ecology **95**. https://doi.org/10.1093/femsec/fiz006.

Mondal K, Samanta S. 2015. A review on arsenic contamination in freshwater fishes of West Bengal. Journal of Global Biosciences **4**(5), 2369-2374.

Moritsch M, Szendrenyi A, Leger C, Frossard B. 2010. Associations among plankton abundance, water quality, and sediment quality in the San Francisco Bay: Nitrogen and phosphorus. Berkeley Scientific Journal **14**, 45-54.

Muñiz O, Revilla M, Rodriguez JG, Laza-Martinez A, Fontan A. 2018. Annual cycle of phytoplankton community through the water column: Study applied to the implementation of bivalve offshore aquaculture in the southeastern Bay of Biscay. Elsevier **61**, 114-130.

https://doi.org/10.1016/j.oceano.2018.08.001.

National Oceanic and Atmospheric Administration (NOAA). 1999. Screening quick reference tables (SQuiRTs).

Neil W, Bryan J. 1991. Responses of fish to temperature and oxygen and response integration through metabolic scope. In: Brune DE, Tomasso J, editors. Aquaculture and water quality (Advances in World Aquaculture vol. 3). The World Aquaculture Society, 30-57.

Neori A, Holm-Hansen O. 1982. Effect of temperature on rate of photosynthesis in Antarctic phytoplankton. Polar Biol **1**, 33-38.

Omura T, Iwataki M, Borja V, Takayama H, Fukuyo Y. 2012. Marine phytoplankton of the Western Pacific. 1-160. **Panhwar Q, Ali A, Naher U, Memon M.** 2019. Fertilizer management strategies for enhancing nutrient use efficiency and sustainable wheat production. Research Gate **6**, 151-174.

https://doi.org/10.1016/B978-0-12-813272-2.00002-1.

Parlevliet D, Moheimani N. 2014. Efficient conversion of solar energy to biomass and electricity. Aquatic Biosystem **10**, 1-9.

Pazi I. 2011. Assessment of heavy metal contamination in Candarli Gulf sediment, Eastern Aegean Sea. Environ Monit Assess **174**, 199-208.

Perelonia KB, Abendanio C, Raña J, Opinion AG, Villeza J, Cambia F. 2017. Heavy metal contamination in water and fishery resources in Manila Bay aquaculture farms. The Philippine Journal of Fisheries **24**(2), 74-97. https://doi.org/10.31398/tpjf/24.1.2016A0014.

Philippine Statistics Authority. 2019. Fisheries Situation Report January-December 2019.

Philippine Statistics Authority. 2020. Fisheries Situation Report January-December 2020.

PHILMINAQ. 2006. Mitigating impact from aquaculture in the Philippines. Annex 2. Water Quality Criteria and Standards for Freshwater and Marine Aquaculture.

Pleto JV, Migo V, Arboleda MD. 2020. Preliminary water and sediment quality assessment of the Meycauayan River segment of the Marilao-Meycauayan-Obando River System in Bulacan, the Philippines. Journal of Health and Pollution **10**(26), 1-9.

Prangnell D, Staresinic N. 2019. Sustainable biofloc systems for marine shrimp. Elsevier, 37-58. https://doi.org/10.1016/B978-0-12-818040-2.00004-6.

Price C, Black K, Hargrave B, Morris J Jr. 2015. Marine cage culture and the environment: Effects on water quality and primary production. Aquaculture Environment Interactions **6**, 151-174.

Raña J, Domingo J, Opinion AG, Cambia F. 2017. Contamination of coliform bacteria in water and fishery resources in Manila Bay aquaculture farms. The Philippine Journal of Fisheries **24**(2), 98-126. https://doi.org/10.31398/tpjf/24.2.2016A0015.

Rauf A, Javed M, Ubaidullah M. 2009. Heavy metal levels in three major carps (*Catla catla, Labeo rohita,* and *Cirrhina mrigala*) from the river Ravi, Pakistan. Pak Vet J **29**, 24-26.

Roa E, Prado G, Quiao M, Dela Peña G, Gorospe J. 2017. Seasonal variation of water quality of the selected mariculture parks in Northern Mindanao, Philippines. AACL Bioflux **10**(5), 1266-1280.

Rzetala M. 2016. Cadmium contamination of sediments in the water reservoirs in Silesian upland (southern Poland). J Soils Sediments **16**, 2458-2470. https://doi.org/10.1007/s11368-016-1477-3.

Salayo N, Perez M, Garces L, Pido M. 2012. Mariculture development and livelihood diversification in the Philippines. Marine Policy **36**(4), 867-881.

Salem Z, Ghobara M, Nahrawy A. 2017. Spatiotemporal evaluation of the surface water quality in the middle Nile Delta using Palmer's algal pollution index. Egyptian Journal of Basic and Applied Sciences **4**, 219-226.

http://dx.doi.org/10.1016/j.ejbas.2017.05.003.

Saoud P, Naamani S, Ghanawi J, Nasser N. 2014. Effects of acute and chronic nitrite exposure on rabbitfish *Siganus rivulatus* growth, haematological parameters, and gill histology. Journal of Aquaculture **5**(6), 1-9. DOI: 10.4172/2155-9546.1000263. **Seo M, Lee H, Kim Y.** 2019. Relationship between coliform bacteria and water quality factors at Weir Stations in the Nakdong River, South Korea. Water **11**, 1-16. DOI: 10.3390/w11061171.

Sharifuzzaman S, Rahman H, Ashekuzzaman S, Islam M, Chowdhury S, Hossain M. 2016. Heavy metals accumulation in coastal sediments. Springer Japan Book Chapter, 21-42. DOI: 10.1007/978-4-431-55759-3_2.

Shinohara R, Kiroki M, Kohzu A, Imai A, Inoue T, Furusato E, Komatsu K, Satou T, Tomioka N, Shimotori K, Miura S. 2017. Role of organic phosphorous in sediment in a shallow eutrophic lake. Water Resources Research **53**, 7175-7189. DOI: 10.1002/2017WR020486.

Simon N, Cras AL, Foulon E, Lemée R. 2009. Diversity and evolution of marine phytoplankton. C.R. Biologies **332**, 159-170.

Srivastava S, Vaih B, Sing A, Singh P. 2020. Nutrient recovery from municipal water stream: Status and prospects. Urban Ecology, 265-297. DOI: 10.1016/B978-0-12-820730-7.00015-X.

Ssanyu G, Rasowo J, Auma E, Ndunguru M. 2011. Evaluation of plankton community structure in fish refugia acting as *Oreochromis niloticus* propagation and nursery units for rice/fish trials. J Aquacult Res Dev **2**(4).

Sthanadar I, Begum B, Sthanadar A, Nasir M, Ahmad I, Muhammad A, Ullah S. 2015. Bioaccumulation of heavy metals in intestine of mulley (*Wallago attu*, Bloch & Scheneider, 1801): A case study of Kalpani river at district Mardan, Khyber Pakhtunkhwa, Pakistan. Journal of Biodiversity and Environmental Sciences 6(2), 74-80.

Storelli M, Storelli A, D'ddaabbo R, Morano C, Bruno R, Marcotrigiano G. 2005. Trace elements in loggerhead turtle (*Caretta caretta*) from the Eastern Mediterranean; Overview and Evaluation. Environmental Pollution **135**, 163-170.

J. Bio. & Env. Sci. 2025

Striebel M, Schabhüttl S, Hodapp D, Hingsamer P, Hillebrand H. 2016. Phytoplankton responses to temperature increases are constrained by abiotic conditions and community composition. Ocealogia 182, 815-827. DOI: 10.1007/s00442-016-3693-3.

Sukumaran M, Muthukumaravel K, Sivakami R. 2013. Seasonal variation in physico-chemical characteristics of Agniar Estuary, Southeast Coast of India. Asia Pac J Res **2**(8), 108-120.

Sundararajan S, Kamalakannan B, Karthikeyan R, Khadanga M, Jena B. 2018. Diurnal variation and water quality parameters of three different ecosystems in Gulf of Mannar, Southeast Coast of India. Journal of Marine Science: Research & Development 8(3), 1-6. DOI: 10.4172/2155-9910.1000252.

Swingle H. 1969. Methods of analysis for water, organic matter and pond bottom soils in fisheries and research. Alabama, USA, 119.

Tabinda A, Bashir S, Yasar A, Munir S. 2013. Heavy metals concentrations in water, sediment and fish in River Sutlej at Sulemanki Headworks. Pakistan Journal of Zoology **45**(6), 1663-1668.

Takarina N, Nurliansyah W, Wardhana W. 2019. Relationship between environmental parameters and the plankton community of the Batuhideung Fishing Grounds, Pandeglang, Banten, Indonesia. Biodiversitas **20**(1), 171-180.

Tamot P, Mishra R, Samdutt. 2008. Water quality monitoring of Halali Reservoir with reference to cage aquaculture as a modern tool for obtaining enhanced fish production. In: Proceedings of Taal 2007: the 12th World Lake Conference, 318-324.

Türkmen M, Türkmen A, Tepe Y, Töre Y, Ates A. 2009. Determination of metals in fish species from Aegean and Mediterranean seas. Food Chemistry **113**, 233-237. **Ukoha P, Ekere N, Udeogu U, Agbazue V.** 2014. Potential health risk assessment of heavy metals (Cd, Cu and Fe) concentrations in some imported frozen fish species consumed in Nigeria. International Journal of Chemical Sciences **12**(2), 366-374.

United States Environmental Protection Agency (US EPA). Ecological Soil Screening Level (Eco-SSL) Guidance and Documents.

Vajravelu M, Martin Y, Ayyappan S, Mayakrishnan M. 2018. Seasonal influence of physico-chemical parameters on phytoplankton diversity, community structure and abundance at Parangipettai coastal waters, Bay of Bengal, South East Coast of India. Science Direct **60**, 114-127.

Vallius H. 2014. Heavy metals concentrations in sediment cores from the Northern Baltic Sea: declines over the last two decades. Marine Pollution Bulletin **79**, 359-364.

Vallius H. 2015. Quality of surface sediments of the northern coast of the Gulf of Finland, Baltic Sea. Marine Pollution Bulletin **99**, 250-255.

Vinodhini R, Narayanan M. 2008. Bioaccumulation of heavy metals in organs of freshwater fish *Cyprinus carpio* (common carp). International Journal of Environmental Science and Technology **5**, 179-182.

Wan J, Yuan X, Han L, Ye H, Yang X. 2020. Characteristics and distribution of organic phosphorous fractions in the surface sediments of the inflow rivers around Hongze Lake, China. International Journal of Environmental Research and Public Health 17, 648.

DOI: 10.3390/ijerph17020648.

Yunus K, Zuraidah M, John A. 2020. A review on the accumulation of heavy metals in coastal sediment of Peninsular Malaysia. Ecofeminism and Climate Change 1(1), 21-35.

DOI: 10.1108/EFCC-03-2020-0003.