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RESEARCH PAPER

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Chemical composition and bioefficacy of *Dennettia tripetala* and *Uvariodendron angustifolium* leaves essential oils against the angoumois grain moth, *Sitotroga cerealella*

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Key words: Fumigation, toxicity, essential oil, D. tripetala, U. angustifolium, S. cerealella.

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Abstract

The essential oils of the leaves of two aromatic species collected in Benin, *Dennettia tripetala* and *Uvariodendron angustifolium* were analyzed by GC and GC / MS. The major components of the *D. tripetala* oil were 2-Phenylnitroethane (52.6%), linalol (26.8%) and methyl eugenol (5.6%). That *U. angustifolium* was dominated by geranial (44.9%), neral (32.1%) and geraniol (2.0%). The evaluation of the toxicity on *S. cerealella* was performed in the laboratory by a fumigation method in a closed glass jar at a temperature of 29 ± 2 °C and natural photoperiod with a relative humidity of $70 \pm 10\%$. The results show an insecticidal effect on the samples for the two dose 0.5μ l.ml⁻¹ 24 h after exposure, with an effect significantly higher in the case of *D. tripetala* (LC₅₀ = 0.253μ l.ml⁻¹ and LC₉₉ = 2.685μ l.ml⁻¹) efficiency. This toxicity of the essential oils was also illustrated by the significant inhibition of emergence of insects compared to control groups, without affecting the germination of rice seeds treated.

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Introduction

Rice is an important food product in the world economy. During the phase just prior to harvest and especially after harvest during storage, this product is attacked by insects stocks including Angoumois grain moth, Sitotroga cerealella (Olivier, 1789). It is considered a dangerous pest for stored grains and difficult to combat. Today, the infestation of rice stocks by Angoumois grain moth emerges as a serious problem in the rice-growing areas in Benin (Togola et al., 2010). Under conditions of heavy infestation, the stored products can suffer 100% loss. S. cerealella attacks result in the reduction of the weight of products, lower germination of seeds and the loss of nutritional value and market value. Control of this pest of many grains revealed the use of ionizing radiation from gamma source of cobalt 60 and resistant varieties of grain in the case of rice, wheat or corn; parasites, pathogens or predators (Trichogramma spp Blattisocius tarsalis Cotesia ruficrus, and Bracon hebetor Pteromalus cerealella) as a biological insecticide on different developmental stages of S. cerealella; frequent use of synthetic chemicals such as deltamethrin, malathion and phosphine fumigation alone or combined treatments and mostly based powders or extracts of plants or insecticides potential repellents such as Cymbopogon citratus, Chenopodium ambrosioides, Azadirachta indica, and Khaya ivorensisaux were studied. (Adjalian et al., 2014). However, unlike pests such as Sitophilus spp, Rhizopertha dominica, , Tribolium sp, little work is done on the fight against the Angoumois grain moth out of volatile extracts.

In Some essential oils are Traditionally Farming through fumigant or touch actions to protect grain storage pests Against, a suitable method to preserve products Stored in warehouses and on small farms (Shaaya et al., 1997, Bell, 1994). According Alzouma et al. (1994) fumigation is the most cost effective tool for managing pests stored. Activities fumigation *S. cerealella* were evaluated from the essential oil of garlic (*Allium sativum*). Fumigation toxicity of the essential oil of neem seeds in doses of 25-200 pi caused 100% mortality of adults and larvae. The

toxicity of extracts of *Eugenia aromatica* (L) in the protection of six varieties of NERICA rice paddy *infested S. cerealella* showed that the extract produced a low adult emergence. Also these treatments did not affect the viability and capacity of water absorption of grains compared to the control treatment (Aringbangba, 2011). The toxicity of essential for stored product insects oils is influenced by the chemical composition of the oil and used part (Don-Pedro, 1966; Lee *et al.*, 2001).

The search for new biologically active molecules against populations Sitotroga cerealella has explored two Annonaceae flora of Benin. Uvariodendron angustifolium, syn. Uvaria angustifolia. This species, in the south find of Benin, is farming in traditional medicine to treat rheumatism and stomach ache malaria, or for flavoring local dishes (leaves) (Noudogbessi et al., 2014). Dennettia tripetala (G.) (Baker f.) GE Schatz where the leaves and fruits are Farming in combination with herbs for the treatment other of cough, infantile convulsions, and worm (Ejechi infestation and Akpomedaye, 2005) Dennettia tripetala extracts have been reported about also to exhibit insecticidal properties (Egwunyenga et al., 1998) and antifungal (Nwachukwu and Osuji, 2008). In the present study, the chemical constituents of essential oils from Dennettia tripetala and Uvariodendron angustifolium were determined, and the insecticidal activity of these essential oils was tested through toxicity fumigation against the adult stages and the emergence of the F1 generation of the stored-products pest, Sitotroga cerealella and also the effect of treatments on the germination of rice grains. No study has been reported previously concerning the activity of these compounds as fumigants against this stored product insect. The essential oils were applied primarily we adults to prevent prevention egg mass output and further damages from larvae and for the protection of rice grains.

Materials and methods

Plant material and extraction of essential oils The leaves of Dennettia tripetala Baker f. and

Uvariodendron angustifolium (Engl. & Diels) RE Fries, family Annonaceae, were harvested in the Ewe-Adakplamè locality in the municipality of Ketou in the south of Benin in 2013. They were identified and certified at the National Herbarium of the University of Abomey-Calavi. In the laboratory, they were spread on the bench away from the light at 20 ° C. Essential oils were obtained by hydro distillation from the leaves (200 to 250g) for 3 hours using Clevenger-type extractor. The less dense than water species are collected by simple decantation and dried over anhydrous sodium sulfate. The extracted oils were stored at 4 ° C and protected from light in amber vials. Oil yields were calculated using the following formula:

Yield (%) =
$$\frac{\text{weight of oil (g)}}{\text{Mass of plant materiel (g)}} \times 100$$

Insects

Strains Sitotroga cerealella used for mass rearing for this study came from the reserve West Africa Rice Development Association, ex International Institute of Tropical Agriculture (Benin). They were reared in the laboratory at the temperature of $29 \pm 2^{\circ}$ C with relative humidity of $70 \pm 10\%$ and natural photoperiod in glass jars or plastic on paddy rice as a substrate.

Analysis of the volatile constituents GC/MS

The essential oils were analyzed on a Hewlett-Packard gas chromatograph Model 7890, coupled with a Hewlett-Packad MS model 5875, equipped with a DB5 MS column (30m x 0.25mm; 0.25μm), programming from 50°C (5 min) to 300°C at 5°C/min, 5 min hold. Helium as carrier gas (1.0 ml/min); injection in split mode (1:30); injector and detector temperature: 250 and 280°C respectively. The MS working in electron impact mode at 70 eV; electron multiplier: 2500eV; ion source temperature: 180°C; mass spectra data were acquired in the scan mode in *m/z* range 33-450.

GC/FID

The essential oils were analyzed on a Hewlett-Packard gas chromatograph Model 6890, equipped with a DB₅ MS column (30m x 0.25mm; 0.25μm), programming from 50°C (5min) to 300°C at 5°C/min, 5min hold. Hydrogen was used as carrier gas (1.0 ml/min); injection in split mode (1:60); injector and detector temperature, 280 and 300°C respectively. The essential oil is diluted in hexane: 1/30. The compounds assayed by GC in oils different essential were identified by comparing their retention indices with those of reference compounds in the literature and confirmed by GC-MS by comparison of their mass spectra with those of reference substances (Rösch et al., 1999; Adams, 1989); Swigar and Silverstein, 1981).

TEST

All tests were performed in the laboratory at a temperature of 29 \pm 2 $^{\circ}$ C and natural photoperiod with relative humidity of 70 \pm 10%.

Fumigation toxicity

The device used consists of glass jars containing 50g capacity of 1 liter of paddy rice (Oryza sativa L.) variety of IR841, cotton mass was suspended in 0.3 g using a wire attached to the inner face of the lid of the jars. Concentrations (0, 0.2, 0.5, 1 and 3 μl.ml⁻¹) were selected after several preliminary tests of each essential oil dissolved in absolute ethanol were tested. The control was carried out with pure 96% ethanol. A 50µl volume of each solution thus prepared was taken and applied onto the cotton. Three replicates were performed for each dose and were introduced into each jar containing ten (10) couples of Sitotroga cerealella adult's aged o to 24 or 10 males and 20 females from the breeding ground, all sealed. Mortality in populations of S. cerealella exposed to the insecticidal activity by fumigation with different treatments was observed for 24h, 48h, 72h and 96h. The number of dead individuals was counted after each exposure time. If no movement of the wings or legs is observed, the insect is considered dead. There are, in fact in any population treated natural mortality adds to the mortality caused by this toxic, the mortality percentages were corrected by Abbott's formula. The experimental units were then observed

at regular intervals of time (24 h) for the emergence of young insects from the 20th day until the 45th day after treatment.

Effect of treatments on seed germination of rice paddy

The effect of essential oils on seed germination of paddy rice was evaluated. Paddy was treated with different concentrations of essential oils mentioned before. After 11 days of treatment (sufficient for mortality of all insects exposed time) with different concentrations of the tested oil, rice seeds were transferred to plates containing kneaded wet cotton with water to obtain their seeds. The percentage of germination was computed Ogendo *et al.* (2004) as follows:

Germinations (%) = (number of seed germinated) / (Total grain sampled) × 100%

Statistical analysis

The raw data from the experiments performed were processed statistically by the method of analysis of variance (ANOVA) using SAS software (Statistical Analysis System) Version 9. 1 (Dagnelie, 1975). They underwent the following changes: 2Arcsin ($\sqrt{\frac{X}{n}}$),

X being the number of dead insects under the effect of the essential oil and n denotes the total number of insects added to each jar. $\sqrt{X+0.5}$ (X is the number of young *S. cerealella* having emerged from the

substrate).

The masses of being attacked quantitative and continuous data, observing the conditions of normalization and equal variance seeds have undergone no statistical transformation.

Finally, it was performed a structuring medium using the Newman-Keuls test (Dagnelie, 1975). Statistical results were considered significantly different when the null hypothesis probability is less than or equal to 5%. For more accurate results, the effectiveness of the toxicity of these oils was assessed, and the LC₅₀ and LC₉₉ calculated. They were deduced from the plot of the regression by the method of Finney. For this, the corrected mortality percentages are converted into probit.

Results

Average yields of essential oil were obtained on three replicates. The essential oil yield of D. tripetala is relatively better $(0.95\% \pm 0.03)$ than U. angustifolium and respectively $(0.92 \pm 0.02\%)$. Fiftyone (51) compounds have been identified in the leaves essential oil $Dennettia\ tripetala$, representing 98% of the oil. These main components are 1-phenyl-2-nitroethane (52.6%), methyl eugenol (5.6%) and linalool (26.8%). The essential oil of U. angustifolium leaves consists of forty-three (43) compounds dominated by geranial (44.9%), neral (32.1%) and geraniol (2.0%).

Table 1. Yields and chemical composition of the essential oil from *D. tripetala* leaves of *D. tripetala*.

N°	Names of the compound	RI	(%)
1	Cis-3-Hexenol	846	t
2	2E-Hexenol	857	0.1
3	Hexanol	860	0.1
4	α-Thujene	918	t
5	α-Pinene	926	0.8
6	Camphene	942	0.2
7	Isomer triethylbenzene	953	t
8	Isomer triethylbenzene	956	t
9	Sabinene	965	1.3
10	β-Pinene	970	0.6
11	6-Methyl-5-Heptene-2-ol	978	t
12	Myrcene	982	0.2
13	α-Phellandrene	998	t
14	δ-3-Carene	999	t
15	α-terpinene	1009	t

16	Para-Cymene	1017	0.3
17	Limonene	1022	0.5
18	β-Phellandrene	1023	0.1
19	Eucalyptol	1025	0.2
20	(Z)- β-Ocimene	1029	0.1
21	(E)- β-Ocimene	1039	1.0
22	δ-Terpinene	1051	0.2
23	Cis Oxide de Linalol	1064	0.1
24	Terpinolene	1078	t
25	Trans Oxide de Linalol	1080	0.1
26	Linalol	1095	26.8
27	cis-para-Menth-2-en-1-ol	1118	0.1
28	Benzeneacetonitrile	1131	0.1
29	Borneol	1166	0.3
30	Terpinen-4-ol	1174	0.4
31	Naphthalene	1178	0.2
32	α-Terpineol	1188	1.0
33	Bornyl acetate	1277	0.1
34	2-Phenylnitroethane	1296	52.6
35	α-Cubebene	1339	t
36	Eugenol	1343	1.0
37	α-Copaene	1369	0.1
38	β-Elemene	1382	0.8
39	Methyl eugenol	1390	5.6
40	β-Caryophyllene	1414	0.5
41	Neryl acetone	1438	0.1
42	α-humulene	1450	0.1
43	Germacrene D	1475	0.7
44	α-selinene	1489	0.2
45	E,E-α-Farnesene	1490	0.1
46	Germacrene A	1501	0.1
47	Elemol	1541	0.2
48	Germacrene B	1553	t
49	Spathulenol	1571	0.2
50	Oxide de Caryophyllene	1577	0.3
51	Guaiol	1589	0.1
Total			97.6%
Essential oil yield (%)			0.95%
Monoterpenic hydrocarbons			5.5%
Oxygenated monoterpenes			30.1%
Sesquiterpenic hydrocarbons			2.6%
Oxygenated sesquiterpenes		0.8%	
Aromatic oxygenated compounds			58.2%
oxygenated aliphatic compounds			0.4%
	≤ 0.1%; RI = Retention Index		
,			

 $\textit{Effectiveness of essential oils tested against S. } \\ \text{cerealella}$

Mean mortality caused by the influence of two different concentrations of essential oils tested on adult S. cerealella populations are shown in Tables 3 and 4. Toxicity of essential oils depends on the concentration and duration exposure. They caused a highly significant mortality (P <0.001) of adult S. cerealella from the lowest dose. It reached 100%

mortality at a dose of 0.2 μ l.ml⁻¹ essential oil *D. tripetala* whereas this rate is reached after 48 hours at a dose of 0.5 μ l.ml⁻¹. This toxicity has been also shown 45 days after infestation by the total inhibition of the emergence of young insects from *S. cerealella* 0.5 μ l.ml⁻¹ for both tested unlike in the controls oils. Thus, a highly significant difference (p <0.001) was also noted for medium emergences regarding both treatments.

Table 2. Yield and chemical composition of the essential oil from U. angustifolium leaves.

1	Nº	Names of the compound	RI	(%)
3 β-pinene 978 0.8 4 6-methylhept-5-en-2-one 984 1.3 5 myrcene 989 0.4 6 dehydro-1,8-cincole 990 t 7 δ-3-carene 1007 t 8 α-terpinene 1018 t 10 limonene 1028 0.2 11 8-terpinene 1057 0.2 12 cis-linalool oxide 1069 t 13 trans-linalool oxide 1073 t 14 linalool 1098 1.7 15 (Z)-isocitral 1162 0.2 16 p-mentha-1,-dien-8-ol 1166 0.2 17 (E)-Isocitral 1180 1.4 18 trans-carveol 1191 0.1 19 cis-carveol 1226 0.1 20 neral 1244 32.1 21 gerania 1244 32.1 22	1	α-pinene	934	1.4
4 6-methylhept-5-en-2-one 984 1.3 5 myrcene 989 0.4 6 dehydro-1,8-cincole 990 t 7 8-3-carene 1007 t 8 α-terpinene 1028 0.2 10 limonene 1028 0.2 11 8-terpinene 1057 0.2 12 cés-linalool oxide 1069 t 13 trans-linalool oxide 1073 t 14 linalool 1098 1.7 15 (Z)-isocitral 1166 0.2 16 p-mentha-1,5-dien-8-ol 1166 0.2 17 (E)-Isocitral 1180 1.4 18 trans-carveol 1191 0.1 19 cé-carveol 1226 0.1 20 neral 1244 32.1 21 geraniol 1253 2.0 22 geranic 133 0.8 24	2	camphene	949	t
5 myrcene 989 0.4 6 dehydro-1,8-cincole 990 t 7 δ-3-carene 1007 t 8 α-terpinene 1018 t 9 p-cymene 1024 0.2 10 limonene 1028 0.2 11 8-terpinene 1057 0.2 12 cks-linalol oxide 1069 t 13 trans-linalol oxide 1073 t 14 linalool 1098 1.7 15 (Z)-Isocitral 1166 0.2 16 p-mentha-15-dien-8-ol 1166 0.2 17 (E)-Isocitral 1180 1.4 18 trans-carveol 1191 0.1 19 cis-carveol 1226 0.1 20 neral 1244 32.1 21 gerania 1244 32.1 21 geranic 1233 0.8 24 α-copaen	3	β-pinene	978	0.8
6 dehydro-1,8-cineole 990 t 7 δ-3-carene 1007 t 8 α-terpinene 1018 t 9 p-cymene 1024 0.2 10 limonene 1028 0.2 11 8-terpinene 1057 0.2 12 c/s-linalool oxide 1069 t 13 trans-linalool oxide 1098 1.7 15 (Z)-isocitral 1162 0.2 16 p-mentha-1,5-dien-8-ol 1166 0.2 17 (E)-Isocitral 1180 1.4 18 trans-carveol 1191 0.1 19 c/s-carveol 1226 0.1 20 neral 1244 32.1 21 geranial 1274 44.9 23 geranic acid 1353 0.8 24 α-copaene 1379 0.5 25 β-demene 1390 0.1 26		6-methylhept-5-en-2-one	984	1.3
7 δ-3-carene 1007 t 8 α-terpinene 1018 t 9 p-cymene 1024 0.2 10 limonene 1028 0.2 11 8-terpinene 1057 0.2 12 cis-linalool oxide 1069 t 13 trans-linalool oxide 1073 t 14 linalool 1098 1.7 15 (2.)-isocitral 1162 0.2 16 p-mentha-1,5-dien-8-ol 1166 0.2 17 (B.)-Isocitral 1180 1.4 18 trans-carveol 1191 0.1 19 cis-carveol 1226 0.1 20 neral 1244 32.1 21 geranial 1274 44.9 23 geranial 1274 44.9 23 geranial 1274 44.9 23 geranic 1353 0.8 24 α-co	5	myrcene	989	0.4
8 α-terpinene 1018 t 9 p-cymene 1024 0.2 10 limonene 1028 0.2 11 8-terpinene 1057 0.2 12 cis-linalool oxide 1069 t 13 trans-linalool oxide 1073 t 14 linalool 1098 1,7 15 (Z)-isocitral 1162 0.2 16 p-mentha-1,5-dien-8-ol 1166 0.2 17 (E)-Isocitral 1180 1.4 18 trans-carveol 1191 0.1 19 cis-carveol 1226 0.1 20 neral 1244 32.1 21 geraniol 1253 2.0 22 gerania cid 1333 0.8 24 α-copaene 1379 0.5 25 β-elemene 1390 0.1 26 β-caryophyllene 1425 1.8 27		dehydro-1,8-cineole	990	t
9 p-cymene 1024 0.2 10 limonene 1028 0.2 11 8-terpinene 1057 0.2 12 cis-linalool oxide 1069 t 13 trans-linalool oxide 1073 t 14 linalool 1098 1,7 15 (Z)-isocitral 1162 0.2 16 p-mentha-1,5-dien-8-ol 1166 0.2 17 (E)-Isocitral 1180 1.4 18 trans-carveol 1191 0.1 19 cis-carveol 1226 0.1 20 neral 1244 32.1 21 geraniol 1253 2.0 22 geranial 1274 44.9 23 geranic acid 1353 0.8 24 α-copaene 1379 0.5 25 β-clemene 1390 0.1 26 β-caryophyllene 1425 1.8 27	7	δ-3-carene	1007	t
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11 8-terpinene 1057 0.2 12 cis-linalool oxide 1069 t 13 trans-linalool oxide 1073 t 14 linalool 1098 1.7 15 (Z)-isocitral 1162 0.2 16 p-mentha-1,5-dien-8-ol 1166 0.2 17 (E)-Isocitral 1180 1.4 18 trans-carveol 1191 0.1 19 cis-carveol 1226 0.1 20 neral 1244 32.1 21 geraniol 1253 2.0 22 geranial 1274 44.9 23 geranic acid 1333 0.8 24 α-copaene 13379 0.5 25 β-elemene 1390 0.1 26 β-caryophyllene 1425 1.8 27 α-humulene 1459 0.3 28 8-muurolene 1490 0.1 30 <td>9</td> <td>p-cymene p-cymene</td> <td>1024</td> <td>0.2</td>	9	p-cymene p-cymene	1024	0.2
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13 trans-linalool oxide 1073 t 14 linalool 1098 1.7 15 (Z)-isocitral 1162 0.2 16 p-mentha-1,5-dien-8-ol 1166 0.2 17 (E)-Isocitral 1180 1.4 18 trans-carveol 1191 0.1 19 cis-carveol 1226 0.1 20 neral 1244 32.1 21 geraniol 1253 2.0 22 geranial 1274 44.9 23 geranic acid 1333 0.8 24 α-copaene 1379 0.5 25 β-elemene 1390 0.1 26 β-caryophyllene 1425 1.8 27 α-humulene 1459 0.3 28 8-muurolene 1480 0.1 29 germacrene-D 1486 1.1 30 β-selinene 1492 1.3 31	11	8-terpinene	1057	0.2
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14 linalool 1098 1.7 15 (Z)-isocitral 1162 0.2 16 p-mentha-1,5-dien-8-ol 1166 0.2 17 (E)-Isocitral 1180 1.4 18 trans-carveol 1191 0.1 19 cis-carveol 1226 0.1 20 neral 1244 32.1 21 geraniol 1253 2.0 22 geranial 1274 44.9 23 geranic acid 1353 0.8 24 α-copaene 1379 0.5 25 β-clemene 1390 0.1 26 β-caryophyllene 1425 1.8 27 α-humulene 1459 0.3 28 8-muurolene 1486 1.1 30 β-selinene 1492 1.3 31 α-selinene 1492 1.3 31 α-selinene 1526 0.2 32 δ-c	13	trans-linalool oxide	1073	t
16 p-mentha-1,5-dien-8-ol 1166 0.2 17 (E)-Isocitral 1180 1.4 18 trans-carveol 1191 0.1 19 cis-carveol 1226 0.1 20 neral 1244 32.1 21 geraniol 1253 2.0 22 geranial 1274 44.9 23 geranic acid 1353 0.8 24 α-copaene 1379 0.5 25 β-elemene 1390 0.1 26 β-caryophyllene 1425 1.8 27 α-humulene 1459 0.3 28 8-muurolene 1486 1.1 29 germacrene-D 1486 1.1 30 β-selinene 1492 1.3 31 α-selinene 1499 0.5 32 δ-cadinene 1526 0.2 33 elemol 1554 0.5 35 spathu		linalool		1.7
16 p-mentha-1,5-dien-8-ol 1166 0.2 17 (E)-Isocitral 1180 1.4 18 trans-carveol 1191 0.1 19 cis-carveol 1226 0.1 20 neral 1244 32.1 21 geraniol 1253 2.0 22 geranial 1274 44.9 23 geranic acid 1353 0.8 24 α-copaene 1379 0.5 25 β-elemene 1390 0.1 26 β-caryophyllene 1425 1.8 27 α-humulene 1459 0.3 28 8-muurolene 1486 1.1 30 β-selinene 1492 1.3 31 α-selinene 1499 0.5 32 δ-cadinene 1526 0.2 33 elemol 1526 0.2 33 elemol 1552 0.3 34 germacrene-	15	(Z)-isocitral	1162	0.2
17 (E)-Isocitral 1180 1.4 18 trans-carveol 1191 0.1 19 cis-carveol 1226 0.1 20 neral 1244 32.1 21 geraniol 1253 2.0 22 geranial 1274 44.9 23 geranic acid 1353 0.8 24 α-copaene 1379 0.5 25 β-elemene 1390 0.1 26 β-caryophyllene 1425 1.8 27 α-humulene 1459 0.3 28 8-muurolene 1480 0.1 29 germacrene-D 1486 1.1 30 β-selinene 1499 0.5 31 α-selinene 1499 0.5 32 δ-cadinene 1526 0.2 33 elemol 1552 0.3 34 germacrene-B 1564 0.5 35 spathulenol		p-mentha-1,5-dien-8-ol	1166	0.2
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32 δ-cadinene 1526 0.2 33 elemol 1552 0.3 34 germacrene-B 1564 0.5 35 spathulenol 1586 0.1 36 caryophyllene oxide 1591 1.2 37 8-eudesmol 1637 0.1 38 epi-α-muurolol 1647 0.1 39 β-eudesmol 1658 1.0 40 selin-11-en-4-α-ol 1668 1.9 41 intermedeol 1667 0.3 42 (2Z, 6Z)-farnesol 1676 0.1 43 (2E, 6Z)-farnesal 1719 0.1 Total 99.6% Essential oil yield (%) 0.92% Monoterpenic hydrocarbons 3.2% Oxygenated monoterpenes 84.8% Sesquiterpenic hydrocarbons 6.4% Oxygenated sesquiterpenes 5.2%		α-selinene		0.5
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35 spathulenol 1586 0.1 36 caryophyllene oxide 1591 1.2 37 8-eudesmol 1637 0.1 38 epi-α-muurolol 1647 0.1 39 β-eudesmol 1658 1.0 40 selin-11-en-4-α-ol 1668 1.9 41 intermedeol 1667 0.3 42 (2Z, 6Z)-farnesol 1676 0.1 43 (2E, 6Z)-farnesal 1719 0.1 Total 99.6% Essential oil yield (%) 0.92% Monoterpenic hydrocarbons 3.2% Oxygenated monoterpenes 84.8% Sesquiterpenic hydrocarbons 6.4% Oxygenated sesquiterpenes 5.2%		germacrene-B		0.5
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37 8-eudesmol 1637 0.1 38 epi-α-muurolol 1647 0.1 39 β-eudesmol 1658 1.0 40 selin-11-en-4-α-ol 1668 1.9 41 intermedeol 1667 0.3 42 (2Z, 6Z)-farnesol 1676 0.1 43 (2E, 6Z)-farnesal 1719 0.1 Total 99.6% Essential oil yield (%) 0.92% Monoterpenic hydrocarbons 3.2% Oxygenated monoterpenes 84.8% Sesquiterpenic hydrocarbons 6.4% Oxygenated sesquiterpenes 5.2%			1591	1.2
38 epi-α-muurolol 1647 0.1 39 β-eudesmol 1658 1.0 40 selin-11-en-4-α-ol 1668 1.9 41 intermedeol 1667 0.3 42 (2Z, 6Z)-farnesol 1676 0.1 43 (2E, 6Z)-farnesal 1719 0.1 Total 99.6% Essential oil yield (%) 0.92% Monoterpenic hydrocarbons 3.2% Oxygenated monoterpenes 84.8% Sesquiterpenic hydrocarbons 6.4% Oxygenated sesquiterpenes 5.2%		8-eudesmol		0.1
39 β-eudesmol 1658 1.0 40 selin-11-en-4-α-ol 1668 1.9 41 intermedeol 1667 0.3 42 (2Z, 6Z)-farnesol 1676 0.1 43 (2E, 6Z)-farnesal 1719 0.1 Total 99.6% Essential oil yield (%) 0.92% Monoterpenic hydrocarbons 3.2% Oxygenated monoterpenes 84.8% Sesquiterpenic hydrocarbons 6.4% Oxygenated sesquiterpenes 5.2%		epi-α-muurolol	1647	0.1
40 selin-11-en-4-α-ol 1668 1.9 41 intermedeol 1667 0.3 42 (2Z, 6Z)-farnesol 1676 0.1 43 (2E, 6Z)-farnesal 1719 0.1 Total 99.6% Essential oil yield (%) 0.92% Monoterpenic hydrocarbons 3.2% Oxygenated monoterpenes 84.8% Sesquiterpenic hydrocarbons 6.4% Oxygenated sesquiterpenes 5.2%		β-eudesmol		1.0
41 intermedeol 1667 0.3 42 (2Z, 6Z)-farnesol 1676 0.1 43 (2E, 6Z)-farnesal 1719 0.1 Total 99.6% Essential oil yield (%) 0.92% Monoterpenic hydrocarbons 3.2% Oxygenated monoterpenes 84.8% Sesquiterpenic hydrocarbons 6.4% Oxygenated sesquiterpenes 5.2%		selin-11-en-4-α-ol	1668	1.9
42 (2Z, 6Z)-farnesol 1676 0.1 43 (2E, 6Z)-farnesal 1719 0.1 Total 99.6% Essential oil yield (%) 0.92% Monoterpenic hydrocarbons 3.2% Oxygenated monoterpenes 84.8% Sesquiterpenic hydrocarbons 6.4% Oxygenated sesquiterpenes 5.2%		intermedeol	1667	
43 (2E, 6Z)-farnesal 1719 0.1 Total 99.6% Essential oil yield (%) 0.92% Monoterpenic hydrocarbons 3.2% Oxygenated monoterpenes 84.8% Sesquiterpenic hydrocarbons 6.4% Oxygenated sesquiterpenes 5.2%		(2Z, 6Z)-farnesol	1676	
Total 99.6% Essential oil yield (%) 0.92% Monoterpenic hydrocarbons 3.2% Oxygenated monoterpenes 84.8% Sesquiterpenic hydrocarbons 6.4% Oxygenated sesquiterpenes 5.2%				0.1
Essential oil yield (%) Monoterpenic hydrocarbons Oxygenated monoterpenes 84.8% Sesquiterpenic hydrocarbons Oxygenated sesquiterpenes 5.2%				99.6%
Monoterpenic hydrocarbons3.2%Oxygenated monoterpenes84.8%Sesquiterpenic hydrocarbons6.4%Oxygenated sesquiterpenes5.2%				0.92%
Oxygenated monoterpenes 84.8% Sesquiterpenic hydrocarbons 6.4% Oxygenated sesquiterpenes 5.2%				
Sesquiterpenic hydrocarbons 6.4% Oxygenated sesquiterpenes 5.2%				
Oxygenated sesquiterpenes 5.2%				
	Oxygenated sesquiterpenes			

Treatment (μl.ml ⁻¹)	Mean (\pm SE) mortality of <i>S. cerealella</i>			
	24h	48h	72h	96h
0	0.47±0.05(1.15)e	0.52±0.00(1.15)b	0.56±0.04(2.34)b	0.67± 0.03(3.62)b
0.2	1.43±0.03(42.67)d	3.14±0.00(100)a	3.14±0.00(100)a	3.14±0.00(100)a
0.5	2.01±0.04(70.78)c	3.14±0.00(100)a	3.14±0.00(100)a	3.14±0.00(100)a
1	2.80± 0.18(95.50)b	3.14±0.00(100)a	3.14±0.00(100)a	3.14±0.00(100)a
3	3.14 ±0.00(100)a	3.14±0.00(100)a	3.14±0.00(100)a	3.14±0.00(100)a
Probability	<0.0001***	<0.0001***	<0.0001***	<0.0001***

Table 3. Rate of *S. cerealella* death provoked by *D. tripetala* essential oil in fumigation_method.

o: ethanol treatment corrected with the control without treatment; *** = very highly significant difference (0.1%). The averages in brackets arose from raw data. The averages followed by the same letter were not significantly different at the beginning of 5% (Newman and Keuls test).

1.42

To further assess the effectiveness of the toxicity of these oils, we calculated LC_{50} and LC_{99} . It is clear from this table that the essential oil of *D. tripetala* appears to have a relatively higher efficiency. These results are confirmed by the values of LC_{50} and LC_{99}

7.15

CV(%)

obtained from a function of the regression line and which correspond to $0.253\mu l.ml^{-1}$ and $2.685\mu l.ml^{-1}$ (Table 6). Indeed, the LC_{50} is very close to the first dose, so that the LC_{99} is between the second and third doses.

1.43

1.79

Table 4. Rate of *S. cerealella* death provoked by *U. angustifolium* essential oil in fumigation method.

	Mean (±SE) mortality of S. cerealella			
Treatment (µl.ml ⁻¹)	24h	48h	72h	96h
0	0.47±0.05(1.15)e	0.52±0.00(1.15)c	0.56±0.04(2.34)b	0.67± 0.03(3.62)b
0.2	1.39 ±0.04(40.43)d	2.84±0.15(96.63)b	3.14±0.00(100)a	3.14±0.00(100)a
0.5	1.88±0.06(65.15)c	3.14±0.00(100)a	3.14±0.00(100)a	3.14±0.00(100)a
1	2.36±0.08(85.39)b	3.14±0.00(100)a	3.14±0.00(100)a	3.14±0.00(100)a
3	2.96±0.17(97.75)a	3.14±0.00(100)a	3.14±0.00(100)a	3.14±0.00(100)a
Probability	<0.001***	<0.001***	<0.001***	<0.001***
CV(%)	7.97	4.56	1.79	1.43

o: ethanol treatment corrected with the control without treatment; *** = very highly significant difference (0.1%). The averages enter brackets arise raw data. The averages followed by the same letter were not significantly different at the beginning of 5% (Newman and Keuls test).

Table 5. Efficacy of *Dennettia tripetala* or *Uvariodendron angustifolium* essential oil on *Sitotroga cerealella* progeny emergence at 45 days post treatment.

Dose (µl.ml ⁻¹)	F1 progeny emergence (mean ± SE)		
	D. tripetala	U. angustifolium	
0	2.60 ± 0.04 (401.33)a	2.60 ± 0.04 (401.33)a	
0.2	0.10± 0.10(0.33)b	0.41± 0.06(1.66)b	
0.5	0.00±0.00(0.00)b	0.00±0.00(0.00)c	
1	d(00.0)00.0±00.0	0.00±0.00(0.00)c	
3	0.00±0.00(0.00)b	0.00±0.00(0.00)c	
Probability	<0.001***	<0.001***	
CV(%)	9.67	6.38	

o: ethanol treatment corrected with the control without treatment; *** = very highly significant difference (0.1%). The averages in brackets arose from raw data. The averages followed by the same letter were not significantly different at the beginning of 5% (Newman and Keuls test).

Effect on the germination of seeds of paddy rice. Fig 1 shows the graph illustrating the percentage of sprouted grains of paddy rice according to two different doses of the essential oils. Both essential oils have greatly reduced adult populations *S.cerealella* by fumigation without affecting the germination of grains of paddy rice processed. The germination rate change from 90% to 100% from the low dose of essential oil while it is less than 80% in the treated only with ethanol. In fact, according to statistical analysis of data there is a highly significant difference (p <0.001) for both treatments as regards the percentages of seed germination after the paddy rice test.

Discussion

Uvariodendron angustifolium, Uvaria svn. angustifolia (Annonaceae) is a tree in the forests of West Africa that can reach 15-40 m high (Hutchinson et al., 1954). This species, find in the south of Benin, is used in traditional medicine to treat rheumatism and stomach ache malaria, or for flavoring local dishes (leaves) (Analytical Flora of Benin, 2006). The essential oils obtained from the leaves of U. angustifolium were characterized by a high proportion of oxygenated monoterpenes (84.8%). Essential oils have been mainly dominated by citral (geranial: 44.9% neral and 32.1%). These results are similar with the only chemical study reported by Noudogbessi et al. (2014). For essential oil of D. tripetala, the present results are different to those obtained by Adeoti et al. (2000) and Gbolade et al. (2009) on the same plant harvested respectively in Benin and Nigeria. Variability levels recorded could be related to the importance of the secretory cells in the leaves of our sample, their physiology, the place or the harvest period. Dennettia tripetala (G.) (Baker f.) GE Schatz (Annonaceae) is a woody spicy vegetable and forest, where the leaves and fruits are used in combination with other herbs for the treatment of cough, infantile convulsion, and worm infestation (Ejechi and Akpomedaye, 2005). Dennettia tripetala extracts have also been reported to exhibit insecticidal properties (Egwunyenga et al., 1998) and antifungal (Nwachukwu and Osuji, 2008). The essential oils obtained from leaves of *D. tripetala* harvested in Benin in this study were characterized by a high proportion of aromatic oxygen compounds (58.2%) and oxygenated monoterpenes (30.1%).

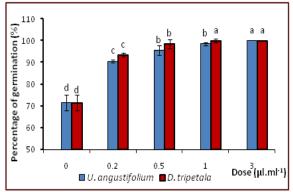


Fig. 1. Percentage of germination of rice according to the essential oil doses o: ethanol treatment corrected with the control without treatment. The averages followed by the same letter were not significantly different at the beginning of 5% (Newman and Keuls test).

Regarding the insecticidal activities of two essential oils, our findings corroborate the work of several researchers who have demonstrated the toxicity of essential oils by inhalation or fumigation against stored product pests (Keita et al., 2001; Lee et al., 2001; Kim et al., 2003; Shaaya et al., 1997). The major advantage of fumigation is to facilitate the penetration of gases inside the grain and thus destroy eggs, larvae and pupae that develop (Benayad, 2008). The toxicity of the fumigant components of essential oils of plants against adult insects were significantly (P <0.001) influenced by the dose and time. The cumulative rate of insect mortality was highest 48 hours after the treatment. Levels of fumigant activities observed could be explained by variations in the structure of the complex relationships of insecticidal activity that influenced their degree of penetration into the insect cuticle and neurotoxicity (Ogendo et al., 2010). Insects undergoing treatment with a dose of 0.2µl.ml⁻¹ showed a small resistance which did not last more than a day since mortality could reach over 90% after the second day. The very low mortality level indicator shows that our test remains reliable for the study of the insecticidal effect of essential oils tested. The insecticidal activity of the

essential oil of D. tripetala did not need much time to occur, since the maximum 100% mortality was recorded the first day post treatment with a dose of 3 μl.ml⁻¹. In addition, LC₅₀ and LC₉₉ values (0.253μl.ml⁻¹ ¹ and 2.685µl.ml⁻¹) confirm its high toxicity to insects in respect of fumigation method. Indeed, the remarkable presence of high aromatic oxygen compounds (58.2%) and monoterpene oxygenates (30.1%) could explain its pronounced insecticidal effect. Reducing the emergence of F1 progeny in the treated groups could be due to increased adult mortality, ovicidal and larvicidal properties of essential oils confirming the findings of Selase and Getu (2009); Bamaiyi et al. (2007); Tapondjou et al. (2002). The emergence of high levels recorded in the control plots also confirm the effectiveness of essential oils tested. Note that it does not exist in the literature work on insecticidal activities of essential oil Uvariodendron angustifolium. Germination tests showed that the plant materials tested against S. cerealella showed no visible adverse effects on the germination capacity of seeds. Also, according to Ketoh et al. (2002) and Glitho et al. (2008), the presence of residues in treated seeds does not affect their ability to germination. Unlike the current results, Paranagama et al. (2003) study showed that the treatment of C. citratus oil reduced the germination capacity of paddy compared to noninfested lot. All tests have confirmed that the treatment of food with essential oil of the two aromatic and medicinal plants of Benin can be very effective against the pest of these commodities Regnault-Roger et al. (2008); Philogène et al. (2008); Vincent et al. (2000); Vincent et al. (2003); Foua Bi (1993).

Conclusion

The present study assessed the insecticidal properties of essential oils of *Dennettia tripetala* and *Uvariodendron angustifolium leaves*. Volatile extracts from two Annonaceae studied proved to be very effective by way of fumigation and do not alter the germination quality of paddy grains developed. The use of these two essential oils appears to be a promising method for the protection of stored rice

against *Sitotroga cerealella*. However, since the plant products evaporate quickly in the environment and do not persist longer unlike synthetic pesticides, pesticide efficacy of herbs could be enhanced when dissolved or mixed with a material fixer for slow release or carrier such as starch or liquid paraffin, and incorporated as part of the integrated pest management especially at the small-scale farmers.

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