INNSPUB

International Journal of Biosciences | IJB |

ISSN: 2220-6655 (Print); 2222-5234 (Online)

Website: https://www.innspub.net

Email contact: info@innspub.net Vol. 27, Issue: 2, p. 280-286, 2025

RESEARCH PAPER

OPEN ACCESS

Phytochemical and antibacterial screening of different solvent extracts of *Mentha spicata*

V. R. Rakhi, T. S. Dhanaraj, V. Ramamurthy*

P.G & Research Department of Biochemistry, Maruthupandiyar College, Thanjavur, Affiliated to Bharathidasan University, Tiruchirappalli, Tamil Nadu, India

Key words: Mentha spicata, Antimicrobial activity, Agar diffusion method, Phytochemical analysis

DOI: https://dx.doi.org/10.12692/ijb/27.2.280-286

Published: August 26, 2025

ABSTRACT

The aim of the present study was to investigate antibacterial activity of the leaf of *Mentha spicata* plant against common pathogenic bacteria. The agar diffusion method was used to examine the antibacterial activity of different solvent extracts of *Mentha spicata* tested against *Escherichia coli*, *Pseudomonas aeruginosa*, *Bacillus subtilis* and *Staphylococcus aureus*. For qualitative phytochemical investigation, several solvent extracts of the complete *M. spicata* plant were investigated. The minimum inhibitory concentration (MIC) of *M. spicata* ethanol extract had significant antimicrobial effectiveness against *Escherichia coli* (489 μ g/ml), and *Pseudomonas aeruginosa* (449 μ g/ml). The ethanolic extract showed (437 μ g/ml and 410 μ g/ml) antimicrobial activity against *Bacillus subtilis* and *Staphylococcus aureus* respectively. The lowest antibacterial activity was found in acetone extract with MIC (216 μ g/ml) of *M. spicata*. Various parts of the plant have the presence of variety of phytoconstituents such as flavonoids, saponins, tannins, terpenes, steroids, amino acids, essential oil, polysaccharides and pectin.

*Corresponding author: V. Ramamurthy 🖂 v.ramamoorthy07@gmail.com

INTRODUCTION

Due to increasing consumers concerns regarding processed and ready-to-eat foods containing antibiotics, pesticides, hormones, and synthetic additives and also increasing demand to replace artificial antimicrobial agents with alternatives, the usage of natural and organic foods has been experiencing explosive market growth (Lv et al., 2011; Moosavy and Shavisi, 2013; Sepahvand et al., 2014). However, the untreated products and natural foods may be more susceptible to growth of food-borne pathogens than the conventional food version (Jay et al., 2005). The most important foodborne pathogenic bacteria that have survived and grow in these products include Staphylococcus aureus, Bacillus spp., Listeria monocytogenes, Salmonella spp. Escherichia coli, Yersinia spp., and Clostridium spp. (Oroojalian et al., 2010; Friedman et al., 2002). These bacteria cause a great proportion of food-borne outbreaks in different foods such as dairy products, vegetables, and meat and fish products (Warriner and Namvar, 2009). In this context, plant essential oils are attracting interest as natural food preservatives in order to ensure the safety of food (Burt, 2004).

Mentha spicata L., commonly called spearmint, belongs to the Lamiaceae family, genus Mentha, which comprises about 25–30 species originating in Europe. It is one of Brazil's most cultivated varieties of spearmint and is well adapted to the subtropical climate.

The interest in cultivating Mentha is mainly related to the commercial importance of its essential oil, which is among the 10 most traded in the world. The oil is used in many industries, including pharmaceuticals, cosmetics, food, and chemicals. Spearmint is also known for its ability to improve memory. Besides being a stimulant, it has several biological uses, such as in insecticides, antimicrobials, antioxidants, antispasmodics, and antiplatelets (Scherer *et al.*, 2013).

M. spicata L. (spearmint) is an herbaceous rhizomatous perennial plant growing 30–100 cm tall,

with variably hairless to hairy stems and foliage, and a wide-spreading fleshy underground rhizome. The leaves are 5–9 cm long and 1.5–3 cm broad, with a serrated margin. The stem is square-shaped, a trademark of the mint family of herbs. Spearmint produces flowers in slender spikes, each flower pink or white and 2.5–3 mm long and broad.

The leaves are popularly used as a tea flavoring agent and the whole plant is used as a carminative. The fresh and dried plants and their essential oils are widely used in the food, cosmetic, confectionary, chewing gum, toothpaste, and pharmaceutical industries (Bensabah et al., 2013). This species is also often used in Indian and Italian cuisine and usually added fresh or dried to fish and shellfish plates before or after cooking. M. spicata possesses several biological activities and is used in folkloric medicine as a carminative, antispasmodic, diuretic, antibacterial, antifungal, and antioxidant agent, and for treatment of colds and flu, respiratory tract gastralgia, hemorrhoids, problems, stomachache (Tawaha et al., 2007; Kizil et al., 2010; Tetika et al., 2013).

Numerous medications have caused microbes to become resistant, which presents a substantial therapeutic issue in the management of infectious diseases. The overuse of commercially available antimicrobials, which are routinely used to treat illnesses, led to the development of the bacteria' tolerance (Lewis and Ausubel, 2006). In order to find new antimicrobial chemicals, researchers were motivated to investigate in other sources, particularly herbal resources.

MATERIALS AND METHODS

Collection and identification of plant material

For the study, the whole plant of *Mentha* spicata belongs to Lamiaceae family was collected from Kerala, South India. The whole plant were identified taxonomically and authenticated according to various literatures, Flora of Madras Presidency and Wealth of India including other pertinent taxonomic literature.

Preparation of plant materials and extract

The leaves were carefully cleaned, shade dried and powdered. The powdered material was stored in a closed air-tight plastic container at low temperature. The powdered plant material (50 g) was extracted with 300 mL of each solvent ethanol by maceration (3×24 h) at room temperature. The collected solvents were concentrated by rotary vacuum evaporator at 45°C and then dried using a freeze dryer. All extracts and acyclovir (extracted from commercial tablet) were dissolved in dimethyl sulphoxide (DMSO). The final concentration of DMSO was 0.1% v/v in cell culture environment.

Phytochemical analysis

The preliminary phytochemical evaluation of leaves was carried on extract prepared by successive extraction method in Soxhlet. The resultant extracts were evaporated to dryness under vacuum. These extract were subjected to chemical test for different phytoconstituents viz. alkaloids, carbohydrates, phenolics, flavonoids, proteins, amino acids, saponins, mucilage and resins etc. Chemical tests were identifying the phytochemicals as described (Trease and Evans, 1983; Harborne, 1973). Alkaloids, carbohydrates, tannins and phenols, flavonoides, gums and mucilage, fixed oils and fats and saponins were qualitatively analyzed.

Test microorganisms

Escherichia coli, Pseudomonas aeruginosa, Bacillus subtilis, and Staphylococcus aureus were used as test organisms in the current study. For the current experiment, the obtained cultures were repeatedly subcultured.

Antimicrobial activity by agar diffusion method

The antibacterial efficacy of several Mentha spicata solvent extracts was assessed using the agar diffusion method. For spreading agar media, a subcultured microbial suspension (100 µl) was prepared. Various concentrated varied extracts were used to measure antimicrobial activity (Magaldi et al., 2004). The plates were filled with the sample and then left to allow for an hour to enable the extract to disperse. The plates were maintained in an incubator for 24 hours at 37°C, and the inhibitory zone was measured in millimeters (mm). Results are compared with those of conventional antibacterial drugs.

RESULTS AND DISCUSSION

Phytochemical studies of Mentha spicata showed that it contains a number of phenolic and polyphenolic compounds, saponin, essential oil, polysaccharides and pectin. The main bioactive constituents found are bioflavonoid, amentoflavone with minute amount of cupressoflavone (Abinash et al., 2012). All of the extracts from the Mentha spicata contained saponin, phenols, tannins, glycosides, terpenoids, flavonoids, alkaloids, and coumarins, according to a preliminary phytochemical examination. With the exception of the chloroform extract's lack of saponins, glycosides, and coumarins and the extract from ethyl acetate's absence of saponin. The results of the phytochemical analysis are displayed in Table 1. A higher degree of biological activity derives from the presence of a high concentration of phytochemicals in the plant.

This plants growing under natural conditions contain the spectrum of secondary metabolites such as phenols, flavanoids, quinones, coumarins, tannins and their glycosides, alkaloids, essential oils etc., the importance of these substance as microbial agents against the pathogen has been emphasized (Sofowora, 1993). In the present study, it was clearly understood that the ethanolic extracted maximum amount of the different type of metabolites present in the Mentha spicata. Boominathan and Ramamurthy (Boominathan and Ramamurthy, 2009) reported that the phytochemical analysis of the *H. indicum* and *C.* procumbens extracts showed the presence of tannins, alkaloids, flavonoids and phenolic compounds. Tannins have been found to form irreversible complexes with proline-rich proteins.

For instance, the presence of flavonoids suggest that the plant have been reported to exert multiple biological effects including, anti-allergic, inflammatory, anti- microbial antioxidant,

cancer activity (Kunle and Egharevba, 2009). It also suggests that the plant might have diuretic properties (Jayvir *et al.*, 2002). The presence of tannins shows that the plant is astringent as documented and suggests that it might have antiviral and anti-bacterial activities and can relief in wound healing and burns (Haslem, 1989). Saponins and glycoside are also very important classes of secondary metabolites as some

are cardio-active and used in treatment of heart conditions (Oloyode, 2005). Some researchers have also investigated that some saponins have anti-cancer and immune modulatory properties (Evans, 2002). Volatile oils are used in the industries for various purposes, both as a pharmaceutical/ cosmetic raw material for production of emollients and active ingredient for the respiratory tract infections.

Table 1. Qualitative phytochemical screening on extracts of Mentha spicata

Name of test	Test applied / Reagent used	Ethanol	Water	Chloroform	Hexane	Acetone	Ethyl acetate
Alkaloids	A] Mayer's	+++	++	++	++	+++	++
	B] Wagner's	+++	++	++	++	+++	++
	C] Hagner's	+++	++	++	+++	+++	++
	D] Dragndorff's test	++	++	++	++	++	+
Flavonoids	HCl and magnesium turnings	+++	++	+	++	+	++
Carbohydrate	Molisch's test	+	+	+	+	+	+
Tannins & phenols	A] 10% Lead acetate	+++	+	++	++	++	++
	B] Fecl ₃	+++	+	++	++	++	++
Test for steroids	A] Salkowski's test	++	++	++	++	++	++
	B] Libermann-Burchard's test	++	++	++	++	++	++
Gums & mucilages	Alcoholic precipitation	-	-	-	-	-	-
Fixed oil & fats	Spot test	+	-	+	+	-	-
Saponins	Foam test	+	+	+	+	+	+
Phytosterols	LB test	+	+	+	+	+	+
Volatile oils	Hydro distillation method	+	+	+	+	+	+
Protein & free amino	A] Biuret test	++	++	++	++	++	++
acids.	B] Ninhydrin test	+++	++	++	++	++	++
	C] Xanthoprotein test	+++	++	++	++	++	++

Table 2. Antimicrobial activity of the plant extracts Mentha spicata

Microorganism	Minimum inhibitory concentration (MIC)									
		Plant extract of <i>Mentha spicata</i> (μg/ml)								
	Ethanol	Water	Chloroform	Hexane	Acetone	Ethyl	Gentamicin	Cephalosporins		
						acetate	(μg/ml)	(μg/ml)		
P. aeruginosa	489	386	287	263	227	312	ND	49		
B. subtilis	437	385	294	242	209	274	ND	47		
E. coli	446	392	323	257	230	302	ND	55		
S. aureus	410	336	258	224	216	255	35	ND		

The antibacterial properties of various solvent-based extracts of *Mentha spicata* are displayed in Table 2. The ethanol extract of *Mentha spicata* had the highest antimicrobial activity with MIC (489 µg/ml) against the *Escherichia coli*, (449 µg/ml) against the *Pseudomonas aeruginosa*, (437 µg/ml) against the *Bacillus subtilis* and (410 µg/ml) against the *Staphylococcus aureus*. The various extracts of *Mentha spicata* tested against *Escherichia coli* and showed considerable MIC results in water extract (392 µg/ml), chloroform extract (323 µg/ml), ethyl acetate extract (312 µg/ml), acetone extract (230 µg/ml), hexane extract (257 µg/ml). The results were

compared with standard Cephalosporins as reference compounds with MIC (55 μ g/ml). The different extracts of *Mentha spicata* were checked against the *Pseudomonas aeruginosa* and exhibited significant MIC values in water extract (386 μ g/ml), chloroform extract (287 μ g/ml), ethyl acetate extract (302 μ g/ml), acetone extract (227 μ g/ml), hexane extract (263 μ g/ml). The obtained results were compared with Cephalosporins with MIC (49 μ g/ml).

The individual extract of *Mentha spicata* was checked against *Bacillus subtilis* and found impressive MIC values in water extract (385 µg/ml), chloroform

extract (294 μ g/ml), ethyl acetate extract (274 μ g/ml), acetone extract (209 μ g/ml), hexane extract (242 μ g/ml). The different solvent extract of *Mentha spicata* was evaluated against *Staphylococcus aureus* and found impressive MIC values in water extract (336 μ g/ml), chloroform extract (258 μ g/ml), ethyl acetate extract (255 μ g/ml), acetone extract (216 μ g/ml), hexane extract (224 μ g/ml).

The gentamicin (35 µg/ml) was used as a standard compound. A considerable inhibitory zone may also be caused by the variety of phytochemicals present in the extract. The presence of different flavonoids, alkaloids, terpenoids, phenols, saponins, coumarins has bactericidal properties (Kalidindi et al., 2015). According to various scientific studies, high concentrations of phytochemicals and bioactive compounds are thought to have a stronger potential for treating a variety of pathogenic bacteria. Numerous plants and their various portions of them have historically been used to treat a variety of chronic illnesses, such as gastrointestinal problems, urinary tract infections, skin conditions, and various respiratory issues., etc. (Alzoreky and Nakahara, 2003). Several chronic illnesses caused by various bacteria may be prevented and managed with the use of plant-based remedies. Many societies still employ ethnomedicines to treat illnesses and overcome obstacles without creating negative side effects. The inclusion of several phytoconstituents, including alkaloids, flavonoids, saponins, polyphenols, tannins, and terpenoids, is what gives herbal preparations their therapeutic effects (Bhalodia and Shukla, 2011). The presence of secondary metabolites prevents the growth of harmful microorganisms causing serious diseases (George et al., 2016). The microorganisms are resistant to many antibiotics that is very harmful to humans. The researchers are finding an alternative to commercial antibiotics to prevent harmful infections against a variety of microorganisms using plant-based medicines (Khan et al., 2013). The higher concentration of crude extracts sometimes may cause cytotoxicity in humans hence the dose-dependent values are determined using in vitro cell cytotoxicity assay (George *et al.*, 2016). As compared to commercial antibiotics, plant-based medicines have very small side effects if they are consumed in excess quantity (Talib and Mahasneh, 2010). In the world, 80 % of different pharmaceuticals are prepared from plant-based medicines and which are effective to cure any chronic disease.

CONCLUSION

Mentha spicata used for its wide therapeutic potential of antimicrobial agents. According to the studies, ethanol extract has the most potential, which may be because it includes the majority of the phytochemical compounds and bioactive compounds that have antibacterial activity. The complete plant extract of Mentha spicata has to be further studied in order to identify and purify chemicals that might be used as natural medicinal alternatives to synthetic commercial ones. The future aspects of the plant can be anti-microbial as it contains many of the phytochemicals and work has not been performed yet.

REFERENCES

Abinash C, Bharati A, Sahu AN. 2012. Ethnobotany, phytochemistry and pharmacology of *Biophytum sensitivum* DC. Pharmacognosy Reviews **6**(11), 68–73.

Alzoreky NS, Nakahara K. 2003. Antibacterial activity of extracts from some edible plants commonly consumed in Asia. International Journal of Food Microbiology **80**, 223–230.

Bensabah F, Houbairi S, Essahli M, Lamiri A, Naja J. 2013. Chemical composition and inhibitory effect of the essential oil from *Mentha spicata* irrigated by wastewater on the corrosion of aluminum in 1 molar hydrochloric acid. Portugaliae Electrochimica Acta 31, 195–206.

Bhalodia NR, Shukla VJ. 2011. Antibacterial and antifungal activities from leaf extract of *Cassia fistula*, an ethnomedicinal plant. Journal of Advanced Pharmaceutical Technology & Research **2**, 104–109.

Boominathan M, Ramamurthy V. 2009. Antimicrobial activity of *Heliotropium indicum* and *Coldenia procumbens*. Journal of Ecobiology **24**(1), 11–15.

Burt S. 2004. Essential oils: Their antibacterial properties and potential applications in foods: A review. International Journal of Food Microbiology **94**(3), 223–253.

Evans WC. 2002. Trease and Evans pharmacognosy. 15th edition. W.B. Sanders, London, 183–184 and 191–393.

Friedman M, Henika PR, Mandrell RE. 2002. Bactericidal activities of plant essential oils and some of their isolated constituents against *Campylobacter jejuni, Escherichia coli, Listeria monocytogenes* and *Salmonella enterica*. Journal of Food Protection **65**(10), 1545–1560.

Harborne JB. 1973. Phytochemical methods: A guide to modern techniques of plant analysis. Chapman and Hall, London, 33–41.

Haslem E. 1989. Plant polyphenols: Vegetable tannins revisited – chemistry and pharmacology of natural products. Cambridge University Press, Cambridge, 169.

Jay JM, Loessner MJ, Golden DA. 2005. Modern food microbiology. 7th edition. Springer Science+Business Media, New York, USA.

Jayvir A, Minoo P, Gauri B, Ripal K. 2002. Nature heals: A glossary of selected indigenous medicinal plants of India. 2nd edition. SRIST Innovations, India, 22.

Kalidindi N, Thimmaiah NV, Jagadeesh NV, Nandeep R, Swetha S, Kalidindi B. 2015. Antimicrobial and antioxidant activities of organic and aqueous extracts of *Biophytum sensitivum* Linn. leaves. Journal of Food and Drug Analysis 23, 795–802.

Khan UA, Rahman H, Niaz Z, Qasim M, Khan J, Tayyaba. 2013. Antibacterial activity of some medicinal plants against selected human pathogenic bacteria. European Journal of Microbiology and Immunology **3,** 272–274.

Kizil S, Hasimi N, Tolan V, Kilinç E, Yüksel U. 2010. Mineral content, essential oil components and biological activity of two *Mentha* species (*M. piperita* L., *M. spicata* L.). Turkish Journal of Field Crops **15**, 148–153.

Kunle OF, Egharevba HO. 2009. Preliminary studies on *Vernonia ambigua*: Phytochemistry and antimicrobial screening of whole plant. Ethnobotanical Leaflets **13**, 1216–1221.

Lewis K, Ausubel FM. 2006. Prospects for plant-derived antibacterials. Nature Biotechnology **24**(12), 1504–1507.

Lv F, Liang H, Yuan Q, Li C. 2011. In vitro antimicrobial effects and mechanism of action of selected plant essential oil combinations against four food-related microorganisms. Food Research International 44(9), 3057–3064.

Magaldi S, Mata-Essayag S, Hartung DE, Capriles C, Perez C, Colella MT, Olaizola C, Ontiveros Y. 2004. Well diffusion for antimicrobial susceptibility testing. International Journal of Infectious Diseases 8(1), 39–45.

Mathew G, Joesph L, Kumar U. 2016. *Biophytum sensitivum* chemical constituents and medicinal properties: A review. International Journal of Current Research and Academic Review 4(7), 57–67.

Moosavy MH, Shavisi N. 2013. Determination of antimicrobial effects of nisin and *Mentha spicata* essential oil against *Escherichia coli* O157:H7 under various conditions (pH, temperature and NaCl concentration). Journal of Faculty of Pharmacy **19**(2), 61–67.

Oloyode OI. 2005. Chemical profile of unripe pulp of *Carica papaya*. Pakistan Journal of Nutrition **4**(6), 379–381.

Oroojalian F, Kasra-Kermanshahi R, Azizi M, Bassami MR. 2010. Phytochemical composition of the essential oils from three Apiaceae species and their antibacterial effects on food-borne pathogens. Food Chemistry **120**(3), 765–770.

Scherer R, Lemos MF, Lemos MF, Martinelli GC, Martins JDL, Silva AG. 2013. Antioxidant and antibacterial activities and composition of Brazilian spearmint (*Mentha spicata* L.). Industrial Crops and Products **50**, 408–413.

Sepahvand R, Delfan B, Ghanbarzadeh S, Rashidipour M, Veiskarami GH, Ghasemian-Yadegari J. 2014. Chemical composition, antioxidant activity and antibacterial effect of essential oil of the aerial parts of *Salvia sclareoides*. Asian Pacific Journal of Tropical Medicine 7(1), S491–S496.

Sofowora EA. 1993. Medicinal plants and traditional medicine in Africa. John Wiley and Sons Ltd, Nigeria, 1–3.

Talib WH, Mahasneh AM. 2010. Antimicrobial, cytotoxicity and phytochemical screening of Jordanian plants used in traditional medicine. Molecules **15**, 1811–1824.

Tawaha K, Alali FQ, Gharaibeh M, Mohamed M, El Elimat T. 2007. Antioxidant activity and total phenolic content of selected Jordanian plant species. Food Chemistry **104**, 1372–1378.

Tetika F, Civelek S, Cakilcioglu U. 2013. Traditional uses of some medicinal plants in Malatya (Turkey). Journal of Ethnopharmacology **146**, 331–336.

Trease GE, Evans WC. 1983. Textbook of pharmacognosy. 12th edition. Balliere, Tindall, London, 57–59.

Warriner K, Namvar A. 2009. What is the hysteria with *Listeria*? Trends in Food Science & Technology **20**(6–7), 245–254.