# INNSPLIR

### International Journal of Biosciences | IJB |

ISSN: 2220-6655 (Print); 2222-5234 (Online)

Website: https://www.innspub.net

Email contact: info@innspub.net Vol. 27, Issue: 3, p. 1-11, 2025

#### RESEARCH PAPER

OPEN ACCESS

Physicochemical, microbiological and sensory quality changes in *Brama* orcini fillets during 90 days of frozen storage at -25 °C

Hannah Abigail R. Daita\*1,2

'Undergraduate Program in Bachelor of Science in Fisheries, University of the Philippines Visayas, Miagao, Iloilo, Philippines

<sup>2</sup>Faculty of the College of Fisheries and Allied Sciences, Cebu Technological University-Moalboal Campus, Moalboal, Cebu, Philippines

**Key words:** Bycatch utilization, Frozen storage stability, Post-harvest quality, Shelf-life extension, Value-added seafood

DOI: <a href="https://dx.doi.org/10.12692/ijb/27.3.1-11">https://dx.doi.org/10.12692/ijb/27.3.1-11</a>
Published: September 03, 2025

#### **ABSTRACT**

Freezing is one of the most effective methods for preserving fish quality, yet gradual deterioration can occur during storage. Extending the shelf life of underutilized bycatch species such as  $Brama\ orcini$  can enhance resource utilization and market potential. This study evaluated the effects of frozen storage at  $-25^{\circ}$ C for 90 days on the physicochemical, microbiological, and sensory quality of skinless  $B.\ orcini$  fillets. Samples were assessed at 10-day intervals for drip loss (%), pH, total volatile basic nitrogen (TVB-N), thiobarbituric acid reactive substances (TBARS), total viable count (TVC), and sensory attributes of raw and steamed fillets. Drip loss increased slightly (p > 0.05) from 0.35% to 0.42%, while pH significantly rose from 5.27 to 6.36. TVB-N and TBARS increased significantly, reaching 25.97 mg N/100 g and 0.97 mg MDA/kg, respectively, both below spoilage thresholds. TVC remained well under the  $10^5$  CFU/g safety limit. Sensory evaluation revealed gradual declines in most attributes, with juiciness showing the most pronounced reduction in both raw and steamed samples. General acceptability decreased significantly but remained within the "like slightly" range by Day 90. Overall,  $B.\ orcini$  fillets maintained acceptable quality for up to 90 days at  $-25^{\circ}$ C. These findings support the potential of freezing as a practical strategy to extend shelf life and improve market availability of this underutilized species.

\*Corresponding author: Hannah Abigail R. Daita ⊠ hannahdaita@gmail.com \*• https://orcid.org/0009-0004-1724-288X

#### INTRODUCTION

Fish, consumed either fresh or frozen, is valued worldwide for its high nutritional content but is highly perishable, with quality deterioration beginning immediately after harvest due to enzymatic activity, microbial growth, and oxidative reactions, influenced by factors such as species, handling, and storage temperature (Karami *et al.*, 2013; Dawson *et al.*, 2018; Duarte *et al.*, 2020; Muzamail *et al.*, 2024).

Freezing, a widely used and cost-effective preservation method, inhibits microbial activity and slows spoilage reactions, allowing fish to retain nutritional and sensory qualities comparable to fresh products for months (Karami *et al.*, 2013; Wangtueai *et al.*, 2014; Hematyar *et al.*, 2017). However, quality degradation can still occur during frozen storage through protein denaturation, residual enzymatic activity, and lipid oxidation (Dawson *et al.*, 2018; Christophe *et al.*, 2024).

Quality changes in frozen fish are generally assessed through a combination of sensory, physicochemical, biochemical, and microbiological analyses (Duarte et al., 2020). Common indicators include pH, which signals enzymatic breakdown of muscle proteins (Aberoumand, 2020; Phu et al., 2022); total volatile basic nitrogen (TVB-N), reflecting degradation from microbial activity (Popelka et al., 2016); thiobarbituric acid reactive substances (TBARS), which measure secondary products of lipid oxidation (Leygonie et al., 2012; Phu et al., 2022); and total viable count (TVC), indicating overall microbial load (Lakshmisha et al., 2008). Sensory evaluation-encompassing appearance, color, odor, texture, and taste-remains a practical and consumerrelevant approach for determining product acceptability (Yasin et al., 2018; Daita et al., 2025).

Brama orcini, commonly known as bigbelly pomfret, is a marine epipelagic species widely distributed across tropical and subtropical Indo-Pacific waters (Bos and Gumanao, 2013; Motomura et al., 2017). In Panay, Philippines, it is landed regularly as purse seine bycatch and is predominantly marketed fresh in

local markets, thereby restricting its broader distribution (Babaran et al., 2009; Bos and Gumanao, 2013). Taxonomically, B. orcini belongs to the order Scombriformes and family Bramidae, and its IUCN Red List status is currently "Not Evaluated" (Froese and Pauly, 2025). It is regarded as a locally important food fish in certain Philippine regions, including Panay and Southern Mindanao (Cantabaco et al., 2015). Despite its potential as raw material for surimi (Endoma et al., 2022), research on its preservation and processing is scarce. To the best of the researcher's knowledge, this is the first investigation to assess the frozen storage quality of B. orcini, a locally abundant tropical bycatch species. Effective frozen storage could enhance year-round availability. reduce post-harvest losses, and open markets for small-scale processors (Aberoumand, 2020; Zuanazzi et al., 2020; Tahiluddin and Kadak, 2022).

B. orcini is a highly migratory tropical species (Cantabaco et al., 2015) that is a locally abundant peak bycatch during seasons yet remains underexploited. Most research on freezing effects has focused on commercially important species, particularly temperate fish (Turan et al., 2003; Popelka et al., 2016; Dawson et al., 2018); migratory marine species (Aubourg et al., 2004; Lakshmisha et al., 2008; Seki et al., 2016); freshwater species (Liu et al., 2010; Gandotra et al., 2012; Karami et al., 2013; Wangtueai et al., 2014; Hematyar et al., 2017; Zuanazzi et al., 2020; Malik et al., 2021); species commonly used for surimi production (Benjakul et al., 2005); and various commercially sold fillets (Muzamail et al., 2024).

Similar to the present study, Aberoumand *et al.* (2018) examined the physicochemical properties of fillets from seasonally abundant species such as *Sparidentex hasta* and *Pampus argenteus* in Iran, highlighting the importance of research on underutilized yet locally significant fish.

This study examined the physicochemical, microbiological, and sensory changes in skinless B. orcini fillets during 90 days of frozen storage at -25 °C, providing baseline data for optimal storage protocols.

Findings aim to support bycatch utilization, market expansion, and sustainable fisheries aligned with the United Nations Sustainable Development Goals—specifically SDG 2 (food security), SDG 12 (responsible consumption), and SDG 14 (sustainable marine resource use).

#### MATERIALS AND METHODS

#### Preparation of B. orcini fillets

The study was conducted in 2018 at the Post-harvest Laboratory, Institute of Fish Processing Technology, University of the Philippines Visayas, Miagao, Iloilo. Whole *B. orcini* (average body weight ~563 g) were purchased from a local wet market, washed, eviscerated, filleted, deboned, and skinned. A total of 7.2 kg skinless fillets was obtained. Paired fillets (average combined weight 240.99g  $\pm$  20.13g) were individually packed in low-density polyethylene (LDPE) bags, heat-sealed, and stored at -25 °C for 90 days. Each fish was processed within approximately 10 minutes at room temperature (28–29 °C), while the samples were held on crushed ice (0–4 °C) during handling to maintain chill conditions until freezing.

## Quality evaluation of *B. orcini* fillets during frozen storage

Sampling was performed at Day o and at 10-day intervals until Day go. All analyses were conducted in triplicate (n=3) using three bags of paired fillets. From each bag, one fillet was halved for physicochemical analyses, the other half used for microbial testing, and the paired fillet reserved for sensory evaluation. All reagents used in the study were laboratory-grade.

#### **Drip loss**

Drip loss was determined following Wangtueai *et al.* (2014). Frozen samples were weighed, thawed under running water while sealed in LDPE bags, reweighed, and drip loss (%) calculated.

$$Drip \ loss(\%) = \frac{(weight \ of \ before \ thawing - weight \ of \ after \ thawing)}{weight \ of \ before \ thawing} X \ 100$$

#### pH

The pH was measured by homogenizing 10 g muscle with 20 mL distilled water for 1 min and reading with a calibrated pH meter.

#### Total volatile basic nitrogen

TVB-N content was analyzed by the Conway microdiffusion method (Conway and Byrne, 1933) using 2 g muscle homogenized with 8 mL 4% (w/v) trichloroacetic acid (TCA), filtered, and assayed.

#### Thiobarbituric acid reactive substances

Lipid oxidation was measured as described by Lemon (1975): 10 g sample was homogenized with 16 mL extracting solution, filtered, mixed with 5 mL 0.2 M TBA reagent, heated in boiling water for 40 min, cooled, and read at 530 nm with a calibrated spectrophotometer to quantify lipid oxidation.

#### Microbiological analysis

Microbial quality was assessed by total viable count (TVC). Twenty-five grams of sample was homogenized in 225 mL sterile peptone diluent (10<sup>-1</sup> dilution), serially diluted, plated on nutrient agar using the spread plate method, and incubated at 35 °C for 24 h before enumeration (CFU/g).

#### Sensory evaluation

Sensory evaluation was performed by seven semitrained panelists on thawed raw and steamed (unseasoned) fillets. Raw samples were evaluated for color, odor, firmness, and juiciness; steamed samples for color, odor, flavor, mouthfeel, and juiciness. General acceptability was rated for both. Attribute scores used a 5-point hedonic scale; general acceptability used a 9point scale (Table 1).

#### Statistical analyses

Data were analyzed by one-way ANOVA to assess effects of storage duration on physicochemical (pH, drip loss, TBARS, TVB-N), microbiological (TVC), and sensory attributes, followed by Tukey's HSD (p < 0.05). Pearson's correlation coefficients were calculated among quality parameters and sensory scores (p < 0.05, 0.01, 0.001). Analyses were performed using JASP (Jeffrey's Amazing Statistics Program) v0.19.3.0 (University of Amsterdam, Netherlands). JASP is an accessible and user-friendly software package suitable for both basic and advanced statistical analyses, and has been recommended for use in research, including medical and biostatistics studies (Ashour, 2024).

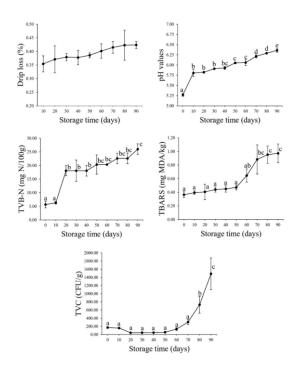
**Table 1.** Sensory attributes and hedonic scales used for evaluating raw and steamed frozen *Brama orcini* fillets

Sample type	Attributes evaluated	Scale used	Scale description
Raw (Thawed)	Color, odor, firmness, juiciness	5-point hedonic scale	5 – Like very much
			4 – Like moderately
			3 – Like slightly
			2 – Neither like nor dislike
			1 – Dislike
Steamed	Color, odor, flavor, mouthfeel,	5-point hedonic scale	5 – Like very much
	juiciness		4 – Like moderately
			3 – Like slightly
			2 – Neither like nor dislike
			1 – Dislike
Raw (Thawed) and	General acceptability	9-point hedonic scale	9 – Like extremely
Steamed			8 – Like very much
			7 – Like moderately
			6 – Like slightly
			5 – Neither like nor dislike
			4 – Dislike slightly
			3 – Dislike moderately
			2 – Dislike very much
			1 – Dislike extremely

#### RESULTS AND DISCUSSION

#### Physicochemical and microbiological changes

The variations in physicochemical and microbiological quality of B. orcini fillets during 90 days of frozen storage at -25 °C are presented in Fig. 1.



**Fig. 1.** Changes in physicochemical and microbiological quality parameters of *Brama orcini* fillets during frozen storage for 90 days. Values with similar superscripts are statistically not significantly different based on one-way analysis of variance

(ANOVA) and Tukey's honestly significant difference (HSD) test (p<0.05). Vertical bars indicate the standard deviation of the mean (n=3).

Drip loss increased slightly from 0.35% (Day 0) to 0.42% (Day 90), with no significant differences (p > 0.05). This minor increase aligns with the known effects of freezing on fish muscle where ice crystal formation causes muscle membrane disruption, resulting in protein denaturation and reduced water-holding capacity, ultimately leading to thaw exudation and product dehydration (Duarte et al., 2020; Christophe et al., 2024). Similar patterns have been reported in Nile tilapia fillets (Zuanazzi et al., 2020), common carp fillets (Hematyar et al., 2017), Atlantic mackerel fillets (Cropotova et al., 2019), and rainbow trout (Turan et al., 2003). In this study, drip loss was strongly correlated with pH and TVB-N (r = +0.72, p < 0.001) (Table 2), suggesting a link between moisture loss and biochemical changes rather than microbial proliferation, as no significant association with TVC was found. Notably, the recorded drip loss values were lower than those reported by Zuanazzi et al. (2020), who observed >1% drip loss in Nile tilapia fillets after only two days at -18 °C.

The initial low post-mortem pH of 5.27 in *B. orcini* fillets is consistent with lactic acid accumulation from anaerobic glycogen breakdown post-harvest (Islami *et al.*, 2014). Such low values generally reflect good

nutritional condition at harvest and potential for extended shelf life by slowing microbial proliferation (Kayim and Can, 2010; Aberoumand, 2020; Daita et al., 2025). A comparable pH (5.76) was reported for  $B.\ orcini$  minced meat by Endoma  $et\ al.$  (2022). During frozen storage, pH rose significantly (p <

o.o5) to 6.36 by Day 90, consistent with the accumulation of alkaline volatile bases from protein degradation and microbial metabolism (Gil and Barbosa, 2011). Despite this increase, values remained below the spoilage threshold of 7.9 (Aberoumand *et al.*, 2018).

**Table 2.** Correlation matrix among physicochemical and microbiological quality parameters of *B. orcini* fillets during frozen storage for 90 days

Variable	Drip loss	pН	TVBN	TBARS
pН	+0.87***			
TVB-N	+0.72***	+0.85***		
TBARS	+0.44*	+0.75***	+0.63***	
TVC	+0.24	+0.55**	+0.47**	+0.70***

Note: \*\*\*Correlation is significant at the 0.001 level (2-tailed). \*\*Correlation is significant at the 0.01 level (2-tailed). \*Correlation is significant at the 0.05 level (2-tailed).

**Table 3.** Correlation matrix among sensory characteristics of raw (thawed) *Brama orcini* fillets during frozen storage for 90 days

Variable	Color	Odor	Firmness	Juiciness
Odor	+0.20			
Firmness	+0.24*	+0.32**		
Juiciness	+0.29*	+0.44***	+0.34**	
General acceptability	+0.30*	+0.51***	+0.47***	+0.72***

Note: \*\*\*Correlation is significant at the 0.001 level (2-tailed). \*\*Correlation is significant at the 0.01 level (2-tailed). \*Correlation is significant at the 0.05 level (2-tailed).

TVB-N increased from 5.65 mg N/100 g to 25.97 mg N/100 g, with significant rises from Day 50 onward (p < 0.05). This trend reflects proteolysis and residual bacterial activity, consistent with previous findings in other species (Phu *et al.*, 2022). The final value remained within the generally accepted limit of 30–35 mg N/100 g (Huss, 1988; Connell, 1995).

TBARS rose gradually from 0.36 mg MDA/kg to 0.65 mg MDA/kg by Day 60, followed by significant increases, reaching 0.97 at Day 90 (p < 0.05). This pattern aligns with previous reports in tilapia fillets and Indian mackerel, where secondary oxidation progresses slowly under low-temperature conditions (Lakshmisha *et al.*, 2008; Liu *et al.*, 2010). The final TBARS value remained far below the rancidity threshold of 5–8 mg MDA/kg (Phu *et al.*, 2022), consistent with findings by Benjakul *et al.* (2005) who found that TBARS levels in tropical species-including threadfin bream, bigeye snapper, lizardfish, and croaker—stored at –18 °C for 15 weeks increased

throughout storage but remained below 5 mg MDA/kg.

Microbiological analysis showed an initial TVC decline from 171.11 CFU/g to 43.33 CFU/g by Day 20, followed by an increase at Day 60 (133.33 CFU/g), reaching 1478.78 CFU/g at Day 90 (p < 0.05). The early reduction likely reflects the sensitivity of many microorganisms to freezing leading to inactivation (Xia et al., 2025), while the subsequent rise likely resulted from the recovery and slow growth of psychotropic bacteria (Li et al., 2023). Although only total viable counts were measured, it is important to note that the composition and abundance of microbial communities in raw fish can vary considerably depending on factors such as geographic origin, season, and post-harvest handling practices (Popelka et al., 2016). Final TVC values remained well below the accepted limit of 105 CFU/g for fishery products (Huss, 1993), indicating microbiological safety at the end of the storage period.

Correlation analysis (Table 2) revealed strong positive associations between pH and TVB-N (r = +0.85, p < 0.001), TBARS (r = +0.75, p < 0.001), and TVC (r = +0.55, p < 0.001). TVB-N also correlated with TBARS (r = +0.63, p < 0.001), supporting the established link between lipid and protein oxidation with malondialdehyde from lipid oxidation reacting with proteins to form carbonyl compounds and promote denaturation, thereby accelerating quality deterioration (Benjakul *et al.*, 2005; Leygonie *et al.*, 2012).

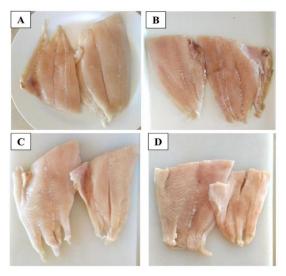
The moderate correlation between TVB-N and TVC (r = +0.47, p < 0.01) and the strong correlation between TVC and TBARS (r = +0.70, p < 0.001) indicate that biochemical degradation, oxidation, and microbial activity progressed concurrently. These findings align with Dawson *et al.* (2018), who emphasized that both biological and chemical reactions—particularly enzymatic activity and lipid oxidation—have a significant impact on the long-term quality stability of frozen fish. Collectively, these interrelationships highlight the multifactorial nature of spoilage in *B. orcini* fillets under prolonged frozen storage.

#### Sensory quality changes

Sensory evaluation was conducted to determine sensory changes in the quality of *B. orcini* fillets during 90 days of frozen storage. For raw fillets, the organoleptic attributes evaluated included color, odor (fishy aroma), firmness (resistance to pressure and resilience), juiciness (perceived moisture based on visual and tactile cues), and general acceptability. For steamed fillets, the parameters assessed were color, odor, flavor (initial and aftertaste), mouthfeel (texture perception when eaten), and general acceptability. Representative images of the thawed raw fillets are shown in Fig. 2. Attributes were evaluated using the hedonic scales described in Table 1.

At Day o, raw fillets were rated "like moderately" for color (4.43), odor (4.57), firmness (4.14), and juiciness (4.57), while general acceptability was "like very much" (8.29). Over the storage period, all parameters declined (Fig. 3). Color, odor, and firmness decreased to 3.71, 3.57, and 3.57, respectively, by Day 90, without significant differences (p > 0.05). Juiciness declined

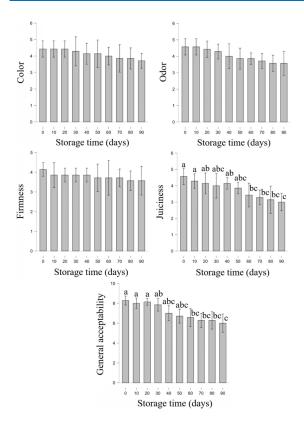
significantly (p < 0.05) from 4.57 to 3.00 ("like slightly"), and general acceptability also fell significantly from 8.29 to 6.00 (p < 0.05).



**Fig. 2.** Thawed *Brama orcini* fillet samples at various frozen storage duration: A) o-day (fresh fillets), B) 30 days, C) 60 days, D) 90 days

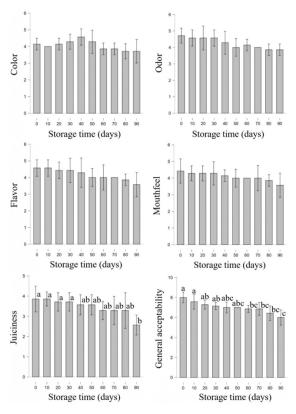
This trend is consistent with reports of progressive declines in sensory properties such as color, odor, texture, and general acceptability in red tilapia fillets at -18 °C (Karami *et al.*, 2013), chilled tilapia fillets at 0 °C (Liu *et al.*, 2010), and *Oreochromis niloticus*, *Labeo rohita*, and *Cirrhinus mrigala* (Yasin *et al.*, 2018). The strong correlations between general acceptability and juiciness (r = +0.72, p < 0.001), odor (r = +0.51, p < 0.001), and firmness (r = +0.47, p < 0.001) suggest that moisture retention, aroma and perceived texture were key determinants of panelist preference for raw fillets (Table 3).

Initial scores for steamed fillets were "like moderately" for color (4.14), odor (4.71), flavor (4.57), and mouthfeel (4.43), with juiciness at 3.86 ("like slightly") and general acceptability at 8.00 ("like very much"). Similar to raw samples, most attributes declined gradually over storage (Fig. 4). Changes in color, odor, flavor, and mouthfeel were not significant (p > 0.05), ending at 3.71, 3.86, 3.57, and 3.57, respectively, by Day 90. Juiciness decreased significantly (p < 0.05) to 2.57 ("neither like nor dislike"), and general acceptability dropped significantly (p < 0.05) to 6.00.



**Fig. 3.** Changes in sensory attributes of raw (thawed) *Brama orcini* fillets during frozen storage for 90 days. Values with similar superscripts are statistically not significantly different based on one-way analysis of variance (ANOVA) and Tukey's honestly significant difference (HSD) test (p < 0.05). Vertical bars indicate the standard deviation of the mean (n=7).

Juiciness loss was more pronounced in steamed fillets, consistent with literature showing that heat treatment causes fish to lose about 20 % of their initial weight due to protein denaturation and fat melting, which release chemically bound water (Leygonie et al., 2012; Nsoga et al., 2021). Similar moisture losses, which also diminish flavor by removing amino acids and nucleotides (Zuanazzi et al., 2020), have been documented in Cyprinus carpio, Arius parkii, and Ethmalosa fimbriata after smoking and boiling (Christophe et al., 2024). For steamed fillets, general acceptability correlated moderately with odor (r = +0.44, p < 0.001) and juiciness (r = +0.39, p < 0.001) (Table 4). These relationships matched panelist feedback, where dryness was the most frequent comment for both raw and steamed samples.



**Fig. 4.** Changes in sensory attributes of steamed *Brama orcini* fillets during frozen storage for 90 days. Values with similar superscripts are statistically not significantly different based on one-way analysis of variance (ANOVA) and Tukey's honestly significant difference (HSD) test (p<0.05). Vertical bars indicate the standard deviation of the mean (n=7).

Panelists also noted gradual lightening and slight yellowing of raw fillets from Day 80 onward, consistent with Hematyar *et al.* (2017), who observed similar color shifts in common carp fillets during frozen storage at –20 °C for 24 weeks. Samples were perceived as fresh until Day 70 for raw fillets and Day 80 for steamed fillets, after which slight fishy odors and very mild sourness were detected. These offodors in fish are often linked to microbial activity producing volatile compounds such as ammonia and sulfur derivatives (Duarte *et al.*, 2020).

However, biochemical analyses indicated that spoilage thresholds were not reached. At Day 60, TVC was only 133.33 CFU/g; by Day 70 it reached 303.33 CFU/g, coinciding with most sensory parameters in the "like slightly" ratings for raw fillets.

At Day 80, steamed fillets had a TVC of 727.78 CFU/g, with most sensory scores also in the "like slightly" range. These values remained far below the typical sensory rejection threshold of 10<sup>7</sup>–10<sup>8</sup> CFU/g for fish fillets (Duarte *et al.*, 2020).

While microbial activity remained low, prolonged frozen storage can still lead to oxidative deterioration, which may contribute to flavor and aroma changes. Secondary lipid oxidation products are known to impart rancid or pungent flavors (Zuanazzi *et al.*, 2020; Christophe *et al.*, 2024), and later stages of lipid peroxidation can also alter both color and nutritional quality (Karami *et al.*, 2013).

Overall, the combined physicochemical, microbiological, and sensory results of this study confirm that while  $B.\ orcini$  fillets remain safe and acceptable after 90 days at  $-25\ ^{\circ}$ C.

**Table 4.** Correlation matrix among sensory characteristics of steamed *Brama orcini* fillets during frozen storage for 90 days

Variable	Color	Odor	Flavor	Mouth feel	Juiciness
Odor	+0.28*				
Flavor	+0.25*	+0.29*			
Mouth feel	+0.21	+0.02	+0.24*		
Juiciness	+0.12	+0.39***	+0.14	+0.08	
General acceptability	+0.24*	+0.44***	+0.35**	+0.21	+0.29*

Note: \*\*\*Correlation is significant at the 0.001 level (2-tailed). \*\*Correlation is significant at the 0.01 level (2-tailed). \*Correlation is significant at the 0.05 level (2-tailed).

#### CONCLUSION

Based on the results of this study, B. orcini fillets stored at -25 °C retained acceptable physicochemical, microbiological, and sensory quality for 90 days. Juiciness was the most affected sensory attribute. Future research should explore longer storage periods and profile specific spoilage organisms. Complementary analyses of nutritional quality and consumer preference could support broader utilization of this underexploited species.

#### **ACKNOWLEDGEMENTS**

The author gratefully acknowledges Mrs. May Flor S. Muegue for her valuable guidance, insightful feedback, and encouragement throughout the conduct of this study.

#### REFERENCES

**Aberoumand A, Ziaei Nejad S, Baesi F, Kolyaee Z.** 2018. Comparative studies of the effect of freezing on the physico chemical properties of the fillets of two fish species in Iran. Food Science and Technology **12**(1), 31–35.

http://dx.doi.org/10.15673/fst.v12i1.835

**Aberoumand A.** 2020. Effects of traditional fish processing methods on the proximate composition and pH of fish black pomfret (*Parastromateus niger*). Potravinarstvo Slovak Journal of Food Sciences **14**, 271–276. https://doi.org/10.5219/1294

**Ashour L.** 2024. A review of user-friendly freely-available statistical analysis software for medical researchers and biostatisticians. Research in Statistics **2**(1), 2322630.

https://doi.org/10.1080/27684520.2024.2322630

**Aubourg SP, Piñeiro C, González MJ.** 2004. Quality loss related to rancidity development during horse mackerel (*Trachurus trachurus*) frozen storage. Journal of the American Oil Chemists' Society **81**(7), 671–678.

**Babaran RP, Selorio Jr CM, Anraku K, Matsuoka T.** 2009. Comparison of the food intake and prey composition of payao-associated and free swimming bigtooth pomfret *Brama orcini*. Fisheries Research **95**(1), 132–137.

https://doi.org/10.1016/j.fishres.2008.07.011

Benjakul S, Visessanguan W, Thongkaew C, Tanaka M. 2005. Effect of frozen storage on chemical and gel-forming properties of fish commonly used for surimi production in Thailand. Food Hydrocolloids **19**(2), 197–207.

https://doi.org/10.1016/j.foodhyd.2004.05.004

**Bos AR, Gumanao GS.** 2013. Seven new records of fish (Teleostei: Perciformes) from coral reefs and pelagic habitats in southern Mindanao, the Philippines. Marine Biodiversity Records **6**, e95, 6 pp. https://doi.org/10.1017/S1755267213000614

Cantabaco JKO, Celedio SF, Gubalani CMB, Sialana RJ, Torres MAJ, Requieron EA, Martin TTB. 2015. Determining sexual dimorphism in bigtooth pomfret, *Brama orcini*, in Tuka Bay, Kiamba, Sarangani Province. AACL Bioflux **8**(6), 1009–1018.

Christophe MKJ, Marlène YT, François NVJ, Merlin NN, Inocent G, Mathieu N. 2024. Assessment of cooking methods and freezing on the nutritional value and health risks of heavy metals in four fish species consumed in Douala, Cameroon. Heliyon 10(7), e28316.

http://dx.doi.org/10.1016/j.heliyon.2024.e28316

**Connell JJ.** 1995. Control of fish quality (4th ed.). Fishing News Book, Scotland, 256 pp.

**Conway EJ, Byrne A.** 1933. An absorption apparatus for the micro-determination of certain volatile substances. Biochemical Journal **27**(2), 419–429.

Cropotova J, Mozuraityte R, Standal IB, Grøvlen MS, Rustad T. 2019. Superchilled, chilled and frozen storage of Atlantic mackerel (*Scomber scombrus*) fillets – changes in texture, drip loss, protein solubility and oxidation. International Journal of Food Science and Technology **54**(6), 2228–2235.

https://doi.org/10.1111/ijfs.1413

**Daita HAR, Amihan Jr RS, Redoble JLS.** 2025. Effects of post-mortem chilling duration on recovery yield and sensory characteristics of manually deboned cage-cultured milkfish (*Chanos chanos*) butterfly fillets using a modified technique. AACL Bioflux **18**(4), 1881–1898.

**Dawson P, Al-Jeddawi W, Remington N.** 2018. Effect of freezing on the shelf life of salmon. International Journal of Food Science **2018**(1), 1686121. https://doi.org/10.1155/2018/1686121

**Duarte AM, Silva F, Pinto FR, Barroso S, Gil MM.** 2020. Quality assessment of chilled and frozen fish-mini review. Foods **9**(12), 1–26.

Endoma LF, Gabo JHC, Sargadillos RM, Condrillon CG, Francisco WA, Daet IP, Silaya FC, Monaya KJM, Muegue MFS. 2022. Physicochemical properties and sensory quality of surimi from bigtooth pomfret (*Brama orcini*) at different washing cycles. Food Research **6**(5), 266–274.

https://doi.org/10.26656/fr.2017.6(5).597

Froese R, Pauly D (Eds.). 2025. Brama orcini Cuvier, 1831. FishBase.

https://www.fishbase.se/summary/Brama-orcini, accessed 15 August 2025.

**Gandotra R, Koul M, Gupta S, Sharma S.** 2012. Change in proximate composition and microbial count by low temperature preservation in fish muscle of Labeo rohita (Hambuch). IOSR Journal of Pharmacy and Biological Sciences **2**(1), 13–17.

**Gil MM, Barbosa AL.** 2011. Microorganisms and safety. In: Cruz RMS (Ed.) Practical food and research. CRC Press, USA. p.195–217.

Hematyar N, Masilko J, Mraz J, Sampels S. 2017. Nutritional quality, oxidation, and sensory parameters in fillets of common carp (*Cyprinus carpio* L.) influenced by frozen storage (–20°C). Journal of Food Processing

and Preservation **42**(5), e13589. https://doi.org/10.1111/jfpp.13589

**Huss HH.** 1988. Fresh fish quality and quality changes. FAO Fisheries Series No. 29. FAO, Rome.

**Huss HH.** 1993. Assurance of seafood quality. Fisheries Technical Paper No. 334. FAO, Rome.

Islami SN, Reza MS, Mansur MA, Hossain MI, Shikha FH, Kamal M. 2014. Rigor index, fillet yield and proximate composition of cultured striped catfish (*Pangasianodon hypophthalmus*) for its suitability in processing industries in Bangladesh. Journal of Fisheries **2**(3), 157–162.

**Karami B, Moradi Y, Motallebi AA, Hosseini E, Soltani M.** 2013. Effects of frozen storage on fatty acids profile, chemical quality indices and sensory properties of red tilapia (*Oreochromis niloticus* × *Tilapia mosambicus*) fillets. Iranian Journal of Fisheries Sciences **12**(2), 378–388.

**Kayim M, Can E.** 2010. The pH and total fat values of fish meat in different iced storage period. Asian Journal of Animal and Veterinary Advances **5**(5), 346–348.

**Lakshmisha IP, Ravishankar CN, Ninan G, Mohan CO, Gopal TKS.** 2008. Effect of freezing time on the quality of Indian mackerel (*Rastrelliger kanagurta*) during frozen storage. Journal of Food Science **73**(7), S345–S353.

https://doi.org/10.1111/j.1750-3841.2008.00876.x

**Lemon DW.** 1975. An improved TBA test for rancidity. New Series Circular No. 51. Fisheries and Marine Service, Halifax, Nova Scotia, Canada. 4 pp.

**Leygonie C, Britz TJ, Hoffman LC.** 2012. Impact of freezing and thawing on the quality of meat: Review. Meat Science **91**(2), 93–98.

https://doi.org/10.1016/j.meatsci.2012.01.013

Li B, Liu S, Chen X, Su Y, Pan N, Liao D, Qiao K, Chen Y, Liu Z. 2023. Dynamic changes in the microbial composition and spoilage characteristics of refrigerated large yellow croaker (*Larimichthys crocea*) during storage. Foods 12, 3994.

https://doi.org/10.3390/foods12213994

Liu S, Fan W, Zhong S, Ma C, Li P, Zhou K, Peng Z, Zhu M. 2010. Quality evaluation of tray-packed tilapia fillets stored at o°C based on sensory, microbiological, biochemical and physical attributes. African Journal of Biotechnology **9(5)**, 692–701. https://doi.org/10.5897/AJB09.1369

Malik IA, Elgasim EA, Adiamo OQ, Ali AA, Mohamed Ahmed IA. 2021. Effect of frozen storage on the biochemical composition of five commercial freshwater fish species from River Nile, Sudan. Food Science and Nutrition 9(7), 3758–3767. https://doi.org/10.1002/fsn3.2340

Motomura H, Alama UB, Muto N, Babaran RP, Ishikawa S. 2017. Commercial and bycatch market fishes of Panay Island, Republic of the Philippines. Kagoshima: The Kagoshima University Museum, Iloilo: University of the Philippines Visayas, and Kyoto: Research Institute for Humanity and Nature. 246 pp.

Muzamail S, Batool R, Paras A, Ahmad T, Javaria S, Syal M, Mukhtar A, Aslam M, Anwar M, Rabia. 2024. Quality and safety characteristics of different commercially available frozen fish fillets. Journal of Xi'an Shiyou University, Natural Science Edition 20(3), 442–449.

Nsoga JVF, Ndomou M, Koule JCM, Melong CSM, Ngafon MN, Ngangue RJEM, Njok PRN, Dama AR, Tang CDN, Tegueu MY, Gouado I, Tchoumbougnang F. 2021. Effects of two smoking and storage methods on the sensory and bromatological quality of *Ilisha africana* (Bloch, 1795). International Journal of Fisheries and Aquatic Sciences 9(2), 280–284.

Phu TM, Duyen HTK, Dao NLA, Ha NTN, Thinh NQ. 2022. Effect of *Camellia sinensis* and *Euphorbia hirta* extracts on the quality of cobia (*Rachycentron canadum*) fillets during ice storage. AACL Bioflux **15(1)**, 350–364.

**Popelka P, Jevinova P, Marcinčák S.** 2016. Microbiological and chemical quality of fresh and frozen whole trout and trout fillets. Potravinarstvo Scientific Journal for Food Industry **10**(1), 431–436. https://doi.org/10.5219/599

**Seki H, Nakazato K, Kobayashi K, Lee TS, Sakurada M, Hamada-Sato N.** 2016. Effect of freezing and thawing on the quality of northern bluefin tuna *Thunnus thynnus* (Linnaeus 1758). Asian Fisheries Science **29**, 232–244.

https://doi.org/10.33997/j.afs.2016.29.4.006

**Tahiluddin AB, Kadak AE.** 2022. Traditional fish processing techniques applied in the Philippines and Turkey. Menba Kastamonu University Faculty of Fisheries Journal **8**(1), 50–58.

**Turan H, Kaya Y, Erkoyuncu İ.** 2003. Effects of glazing, packaging and phosphate treatments on drip loss in rainbow trout (*Oncorhynchus mykiss* W., 1792) during frozen storage. Turkish Journal of Fisheries and Aquatic Sciences **3**, 105–109.

Wangtueai S, Tongsiri S, Maneerote J, Supaviriyakorn W. 2014. Effect of phosphate on frozen Nile tilapia fillets. Food and Applied Bioscience Journal 2(3), 203–215.

**Xia L, Zhou S, Lian K, Chen S.** 2025. Integrated metabolomic and microbial analysis of quality dynamics in channel catfish (*Ictalurus punctatus*) under refrigerated and frozen storage. Foods **14**, 1089. https://doi.org/10.3390/foods14071089

Yasin R, Samiullah K, Hafeez-Ur-Rehman M, Malik IU, Mubarik MS, Draz O, Naz S, Gilani M. 2018. Organoleptic assessment after different processing techniques (drying, smoking, freezing and salting) of exotic and indigenous fish species of Pakistan. International Journal of Biosciences 12(4), 207–215. http://dx.doi.org/10.12692/ijb/12.4.207-215

**Zuanazzi JSG, Goes ESDR, Almeida FLAD, Goes MD, Lara JAFD, Ribeiro RP.** 2020. Effects of freezing and thawing cycles on the quality of Nile tilapia fillets. Food Science and Technology, Campinas **40**(Suppl. 1), 300–304.

https://doi.org/10.1590/fst.11119