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RESEARCH PAPER

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Phytochemical analysis and antioxidant activity of ethanolic leaves extract of *Psidium guajava*

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ABSTRACT

Psidium guajava, or guava, is used in traditional medicine for a wide range of conditions, including diarrhea, diabetes, high blood pressure, respiratory issues like coughs, and various infections. This study was conducted to assess the phytochemical constituents in *Psidium guajava* leaf extracts using standard methods. The ethanolic leaves extract of *Psidium guajava* was analaysed by HPLC and GC to determine various phytochemicals. The free radicals scavenging activity of extract by using DPPH, NO and super oxide radicals generated *in vitro*. The ethanolic leaves extract of *Psidium guajava* was found to contain alkaloids (6.18 mg/gm), tannins (1.82mg/g), phenols (7.76 mg/gm) and flavonoids (5.15 mg/gm). The major flavonoids detected were kaempherol. The ethanolic extract of *Psidium guajava* was found to possess significant free radical scavenging activity against DPPH, NO and SOD anions the IC₅₀ value of 39.0 μg/ml, 34.25 μg/ml and 30.05μg/ml respectively and comparable to that of their corresponding IC₅₀ value. The medicinal property of *Psidium guajava* may be attributed to the presence of flavonoids and phenolic compounds with rich antioxidant potential.

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INTRODUCTION

Plants have been major source of medicine in all cultures from ancient times. In the traditional system, various indigenous plants are being used in the diagnosis, prevention and elimination of physical, mental or social imbalance. Phenolic compounds, ubiquitous in plants, are of considerable interest and have received more and more attention in recent years due to their bioactive functions. Polyphenols are amongst the most desirable phytochemicals because of their antioxidant activity. Natural therapy for various human ailments purified with plant products has gained much attention now days, due to various side effects associated with allopathic medicine these can be derived from any part of the plant like bark, leaves, stem, flowers, roots, seeds, etc., (Cragy and David, 2001). Medicinal plants are believed to be an important source of chemical substances with potential therapeutic effects (Farnsworth, 1989).

Free radicals play an important role in various pathological conditions such as tissue injury, inflammation, neurodegenerative diseases, cancer and aging. The Compound that can scavenge free radicals has great potential in ameliorating these diseases (Coban *et al.*, 2003).

Inflammation is a disorder characterized by invasion of leucocytes and production of proinflammatory cytokines (Mantri and Witiak, 1994).

Psidium guajava (guava) exhibits a wide range of pharmacological activities including antioxidant, anti-inflammatory, antimicrobial, anti-diabetic, antispasmodic, anti-ulcer, anti-allergic, anti-cancer, and cardioprotective effects. These activities are attributed to its rich content of bioactive compounds such as flavonoids, terpenoids, polyphenols and ascorbic acid. These properties support its traditional use for various ailments, including diarrhea, diabetes, and gastrointestinal issues.

Psidium guajava Linn. is commonly called guave, goyave in French; guave, guavenbaum, in German; banjiro in Japanese; goiaba, in Portu gal; arac, guaiaba in Brazil; and guava in English (Killion, 2000). P. guajava used as an important food as well

as a medicinal plant in tropical and subtropical countries, therefore its nickname as the poor man's apple. The scientific evidences of the medicinal uses of *P. guajava* began in 1940's and reports, maintain a tradition of repeating the data each decade.

Many people habitually take *P. guajava* leaf decoction for its antispas modic and antimicrobial properties for the treatment of dysentery and diarrhea (Gutierrez, 2008).

Therefore, the efficacy and safety of *P. guajava* leaves have empirically been confirmed (Hamada and Kitanaka, 1999). P. guajava leaf contains plenty of phenolic compounds which inhibit the peroxidation reaction in the body, and so it can be expected to prevent various chronic diseases such diabetes, cardiovascular disease and cancer. Furthermore, decreasing of free radicals in the body, means that the polyphenols in the leaf of P. guajava can prevent atherosclerosis, cataract and also inhibits biological aging of the body and skin (Okuda, 1982)

P. quajava leaves contains triterpenes, cineol and tannins. Additionally, three lavonoids (avicularin, guaijaverin, and quercetin) have been isolated from the leaves (Khadem and Mohammed, 1959). In mature the concentrations leaves, greatest flavonoids of are found Ouercetin>Myricetin>Kaempferol>Luteolin (Vargas et al., 2006). Most of the medicinal activities of P. guajava are credited to the flavo noids and these phytoconstituents are well-known for their multi-di rectional medicinal activities. Therefore the present study quantifies the secondary metabolites and the antioxidant potential of ethanolic extract of P. quajava. Considering all these facts, the present study was designed to investigate the presence of various phytochemicals in the different extracts of P. guajava, a plant which evokes various therapeutic effects.

MATERIALS AND METHODS Collection of plant and extraction

The leaves of *Psidium guajava* were collected in the MPC garden, Thanjavur. The leaves were air dried

under natural conditions for five days and dried leaves were powdered by using home blender and aqueous extract was prepared and stored in refrigerator under 4°C for further analysis.

Qualitative phytochemical analysis

The extract was subjected to qualitative test for the identification of various phytochemical constituents as per standard procedures (Sofowora, 1993).

Quantitative phytochemical analysis

HPLC-UV analysis (Total phenols)

Ethanolic extract of *Punica granatum* was subjected to solid phase extraction using column 5mm (4.6.mm), and peptides, small molecules were removed; fractionation of neutral and acidic phenolic acids was also carried out simultaneously. The resulting fraction was then subjected to reverse phase high performance liquid chromatography (RP-HPLC). The total phenolics in ethanolic extract of *P. granatum* was detected using, Stationary phase octadecylsil. Silica and mobile phase (A phosphoric acid: water (0.5: 99.5v/v) B acetonitrile). The UV detector was set at 220 nm with the flow rate adjusted to 1.0ms / min. The major peaks were identified and the retention times were compared with these of standards.

Fractionation of total alkaloids

Ethanolic extract of *P. granatum* was detected using monobasic phosphate as mobile phase (270ml. of Acetonitril). The liquid chromatography is equipped with 235 nm detector and 4.6nm x 150 mm column. The flow rate was adjusted to 1.8ml / minute. The major peaks were identified and the total alkaloids concentration was determined.

Fractionation of total flavonoids

HPLC chromatography (System Name: LACKROM L-7000 MERCK, Proc Method- HITECHI) total flavonoids. The total flavonoids in the extract was determined by using octadecysil silica gel as stationary phase and acetonitril, sodium dihydrogen phosphate with dilute orthophosphoric acid as mobile phase. UV detector was set at 350nm with flow rate of 0.5ml/min. The major peaks in ethanolic extract of *P. granatum* were determined in comparison to the

retention time of standards run at identical conditions.

Free radical scavenging activity

Diphenyl-2- picrylhydrazyl (DPPH) radical scavenging activity

DPPH radical scavenging assay is a commonly recommended method for assessment of antioxidant potential of plant extracts. The assay is based on the ability of DPPH, a free radical which get decolorized in the presence of antioxidants. To 200ml. of ethanolic solution of DPPH (1 μ g/ml) various concentration of (20mg- 100 μ g/ml) in water were added and incubated at 37°C for 30 min in dark and the absorbance was measured at 517nm. Ascorbic acid was used as the reference standard. The percentage scavenging of DPPH free radical was calculated and compared with that of the standard ascorbic acid. The IC50 value also determined.

Superoxide anion scavenging activity

The method of Nishkimi et~al.~(1972) was applied for the measurement of MIT superoxide anion scavenging activity, Briefly 312 μ m Nitroblue tetarzolium in 120 μ m phosphate buffer pH 7.4 were added to an aliquots of MIT (20-100 μ g/ml) the reaction was started by adding 100ml of phenazine metho sulphate (120mm preaperd in phosphate buffer pH 7.4) and the colour change was monitored at 560nm against water blank querceutin was used as the positive control.

Nitric oxide scavenging activity

The nitric oxide scavenging activity of the aqueous extract was measured by taking various concentrations of MIT and standard. Ascorbic acid (20-100µg/ml) dissolved in phosphate buffer (0.025m, pH 7.4) and incubated with sodium nitroprusside (5mm) in standard phosphatebuffer at 25°C for 5 hrs. After the incubation, 0.5ml of the reaction mixture was added with 0.5ml of Griess reagent (equal volume of 1% sulphanilamide in 2% phosphoric acid and 0.1% napthlthyl ethylene diamine dihydrochloride in water). The absorbance of the chromophore formed was read at 540nm. The activity was compared with that of similar concentration of Ascorbic acid (Sreejavan Rao, 1997).

RESULTS AND DISCUSSION

Phytochemicals are naturally occurring biochemical compounds that plants developed, in order to protect themselves from oxidation, insect disease and other hazards in their environment. These phytochemicals give their characteristic colour, flavour, smell and texture. Epidemiological studies indicate that populations consuming high levels of plant derived foods have low incidence rates of various cancers.

The phytochemical analysis of *Psidium guajava* was shown in Table 1. The phytochemical analysis showed the presence of alkaloids, flavonoids, phenols, steroids, tannins, saponins sugar and aminoacids with the absences of terpenoids and cardiac glycosides. Phytochemicals in the plants are non-nutritive chemicals that have protective or disease preventive properties. It is well documented that the presence of these chemicals is responsible for various medicinal properties and there are many reports available to support the role of phytochemical constituents and their activity against specific disease (Suresh and Nagarajan, 2009).

Table 1. Qualitative phytochemical analysis

Phytochemicals	Observation
Alkaloids	+
Flavonoids	+
Phenols	+
Steroids	+
Tannins	+
Saponins	+
Terpenoids	-
Cardiac glycosides	-
Sugar	+
Aminoacids	+

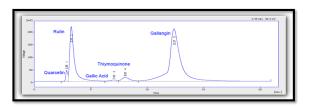
In this investigation, presence of quantitative phytochemicals in ethanolic extract of *Psidium guajava* expressed the value in mg/g phenols was (7.76 mg/g), tannins (1.82 mg/g), flavonoids (5.15 mg/g) and alkaloids (6.18). Finding the natural substance of medicinal plant that decrease the inflammation and reduce oxidative stress and there by counteracting the macromolecular damage. Flavonoids and phenols in general are highly effective in scavenging free radical and providing antioxidant defense in living cells. Quantitative analysis of ethanolic extract of *Psidium guajava* was given in Table 2.

Table 2. Quantitative phytochemical analysis of *Psidium guajava*

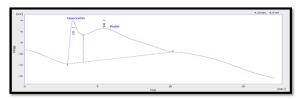
Phytochemical	Values in (mg/g)
Phenols (mg/g)	7.76
Tannins (mg/g)	1.82
Flavonoids (mg/g)	5.15
Alkaloids (mg/g)	6.18

HPLC analysis of ethanolic leaves extract of Psidium guajava

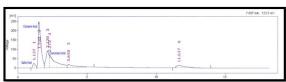
HPLC analysis reveals that the extract was found to be rich in Alkaloids (6.18 mg/g) terpenoids (1.82 mg/g) and phenols (7.76 mg/g). Ethanolic leaves extract of *Psidium guajava* also contain flavonoids such as Rutin (1.12 mg/g) and quercetin (1.19 mg/g).



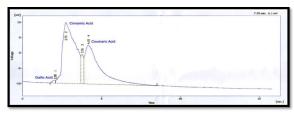
A. HPLC finger prints of standard flavonoids



B. HPLC finger print of flavonoids present in *Psidium guajava*



C. HPLC finger print of standard phenolics compound



D. HPLC finger prints of phenols present in *Psidium* guajava

Fig. 1 (A) to (D) many reports demonstrate that antioxidant principle present in medicinal plants are responsible for their therapeutic potential (Larson, 1988) flavonoic compound such as quercetin and

Rutin are formed to be responsible for antiinflammatory and anticancer properties proliferates by their terminating action of free radicals. Alkaloids have many pharmacological activities including anticancer and anti-arthythmic effect (Cordell, 1983).

Alkaloids are known to reduce the inflammation level significantly. These results shows that ethanolic leaves extract of *Psidium guajava* containing which could be accounted for the antioxidant and anti-inflammatory effects. In the present investigation, HPLC chromatrographics pattern of the ethanolic extract of *Psidium guajava* showed 2 peaks of flavonoids and 3 peaks of phenolic compounds. In HPLC analysis of ethanolic extract of *P. guajava* was found to be rich in flavonoids such as quercetin, Rutin and phenols such as gallic acid, cinnamic acid and coumaric acid. The natural phytonutrients presents in fruits

and vegetables scavenge the free radicals and protect the cells from oxidative damages. The phytonutrients present in ethanolic leaves extract of *P. guajava* migrates the responsible for the traditional claim by the test drug.

In vitro antioxidant assays

Antioxidants are known to exhibit their biochemical effects through numerous mechanisms, including the prevention of chain initiation, reductive capacity and radical scavenging mechanisms. Several methods have been used to measure the antioxidant activity of biological materials. It is essential to use more than one method to evaluate antioxidant capacity of plant materials simply because of the complex nature of phytochemicals present in them. Therefore, in the present study, DPPH free radical scavenging activity and nitric oxide scavenging activity assessment were done.

Table 3. Free radicals scavenging activity in *Psidium guajava*

Free radicals	Samples	Conce	Concentration of standard and extract in (µg/ml)					
		20	40	60	80	100		
DPPH	Ascorbic acid	35.6	41.7	62.8	72.4	81.8		
	P. guajava	25.5	32.4	45.6	56.5	65.2		
Nitric oxide	Ascorbic acid	29.5	35.8	42.9	55.7	67.1		
	P. guajava	26.8	31.7	36.8	47.9	57.5		
Superoxide	Gallic acid	29.8	32.7	47.9	53.9	78.6		
	P. guajava	18.8	24.8	35.3	42.2	58.4		

The percent DPPH scavenging activity ethanolic extract of Psidium guajava were depicted (Table 3). Ascorbic acid was used as a positive control for comparison of plant materials. The IC_{50} value of ascorbic acid was found to be 52.25µg/ml. The IC₅₀ value of DPPH scavenging activity was found to be ethanolic extract of Psidium guajava was 39.0 µg/ml. DPPH is a stable free radical and accepts an electron or hydrogen radical to become a stable diamagnetic molecule. The degree of discoloration indicated the scavenging potential of the antioxidants in the sample. The extract significantly inhibited the activities of DPPH radicals in a dose-dependent manner. The comparison of scavenging effects of ethanolic extract on DPPH radical showed consistently higher radical scavenging activity observed in seabuckthorm Hippophae rhamnoidesl seeds extracts of phenolic compounds (Heim et al., 2002), Flavonoids (Apati et al., 2003) and terpens

(Saranya Panneerselvam *et al.*, 2010) might be taken in to account.

Activity evaluation of Psidium quajava extract showed the capacity of the plant towards nitric oxide scavenging in a dose dependent manner. Ascorbic acid, a natural antioxidant was used as a positive control for comparison. Nitric oxide or reactive nitrogen species, formed during their reaction with oxygen or with super oxides. These compounds are responsible for altering the structural and functional behaviour of many cellular components. Incubation of solutions of sodium nitroprusside in PBS at 25°C for 2 hours resulted in linear time dependent nitrite production, which is reduced by the ECH. The IC₅₀ of ascorbic acid was found to be 39.35µg/ml. The order of the scavenging activity was found to be ethanolic extract of Psidium guajava was IC₅₀ 34.25 μg/ml. Nitric oxide scavenging activity of ethanolic leaves

extract of P. granatum is on important chemical mediator generated by endothelial macrophages, neuron and it is involved in the regulation of various physiological process like control of arthritis, cytotoxice effects alzhemer's disease (Sainani et al., 1997). Thirunavukkarasu et al. (2011) previously studied higher free radicals compared to the results of 73.89±4.22 at 2mg/ml concentration of ethanol extracts of mangrove A. officinalis in Cuddalore region. Higher nitric oxide scavenging activity was reported by Athiperumalsami et al. (2010) in the methanol and water extracts of H. ovalis but the IC50 values were much lower than that of standard tocopherol.

Superoxide anion is a free radical created from the normal process of energy generation in the human body. Superoxide anion is toxic to cells and tissues and plays an important role in the formation of other reactive oxygen species such as hydrogen peroxide, hydroxyl radical or singlet oxygen. The concentration of standard gallic acid to inhibit 50% of superoxide formation was found to be 40.3µg/ml. The scavenging activity of Psidium quajava extract was found to be IC_{50} 30.05µg/ml. The superoxide onions are toxic intermediates formed during inflammatory process and found to enhance the risk of inflammation related disorders such as arthritis and atherosclerosis. Super oxide anion is a free radio that plays an important role in the formation of reactive oxygen species such as hydrogen periode, hydroxyl / radicals, or singlet oxygen in living organism. Korycka et al. (1978) reported that the therapeutic activity of medicinal plants can be determined by superoxide activity. Superoxide anion is a free radical created from the normal process of energy generation in the human body. Superoxide anion is toxic to cells and tissues and plays an important role in the formation of other reactive oxygen species such as hydrogen peroxide, hydroxyl radical or singlet oxygen (Nohl et al., 2003).

CONCLUSION

Nowadays herbs are extensively used for the research purpose and it possesses more than one chemical entity so it has been widely used for the research investigations. The plant based compounds have the effective dosage response and minimal side effects when compared to the synthetic compounds Phytochemical screening of *Psidium guajava* leaves reveals it as a valuable medicinal plant with numerous medicinal properties. Since the ethanolic extract of *Psidium guajava* leaves contains more constituents it can be considered beneficial for further investigation. A typical research and developmental work needs to be carried out for its better therapeutic and commercial utilization. The free radical scavenging activity of *Psidium guajava* revealed that they can be used for the prevention or treatment of human diseases such as cancer, arthritis, diabetes mellitus which are associated with oxidative stress.

REFERNCES

Apati P, Szentmihalyi K, Kristo SZT, Papp I, Vinkler P, Szoke E, Kery A. 2003. Herbal remedies of Solidago, correlation of phytochemical characteristics and antioxidative properties. Journal of Pharmacological and Biomedical Analysis 32, 1045–1053.

Athiperumalsami T, Devi RV, Hastha PS, Kumar V, Louis JL. 2010. Antioxidant activity of seagrasses and seaweeds. Botanica Marina 53, 251–257.

Coban T, Citoglu GS, Sever B, Iscan M. 2003. Antioxidant activities of plants used in traditional medicine in Turkey. Pharmaceutical Biology **41**, 608–613.

Cordell GA. 1983. Introduction to alkaloids: A biogenic approach. Wiley, New York.

Cragg GM, David J. 2001. Natural products drug discovery in the next millennium. Journal of Pharmaceutical Biology **39**, 8–17.

Farnsworth NR. 1989. Screening plants for new medicines. In: Wilson EO (ed.) Biodiversity Part II. National Academy Press, Washington, 83–97.

Gutierrez RM, Mitchell S, Solis RV. 2008. *Psidium guajava*: A review of its traditional uses, phytochemistry and pharmacology. Journal of Ethnopharmacology **117**(1), 1–27.

Hamada S, Kitanaka S. 1999. Method of treatment of atopic dermatitis with dried guava leaves. United States Patent **5**, 942231.

Heim KE, Taigliaferro AR, Bobilya DJ. 2002. Flavonoid antioxidants: Chemistry, metabolism and structure-activity relationships. Journal of Nutritional Biochemistry **13**, 572–584.

Khadem EHE, Mohammed YS. 1959. Constituents of the leaves of *Psidium guajava* L. Journal of the Chemical Society **11**, 3320–3323.

Killion KH. 2000. The review of natural products. 3rd ed. Facts and Comparisons, USA, 250–251.

Korycka DM, Richardson T. 1978. Phytogeneration of superoxide anion in serum of bovine milk and in model systems containing riboflavin and amino acids. Journal of Dairy Science **61**, 400–407.

Larson RA. 1988. The antioxidants of higher plants. Phytochemistry **27**, 96.

Mantri P, Witiak DT. 1994. Inhibition of cyclooxygenase and 5-lipoxygenase. Current Medicinal Chemistry 1, 328–355.

Nishkimi M, Appaji N, Yagi K. 1972. The occurrence of superoxide anion in the reaction of reduced phenazine methosulfate and molecular oxygen. Biochemical and Biophysical Research Communications **46**, 849–854.

Nohl H, Kozlov AV, Gille L, Staniek K. 2003. Cell respiration and formation of reactive oxygen species: Facts and artifacts. Biochemical Society Transactions **31**, 1308–1311.

Okuda T, Yoshida T, Hatano T, Yakazi K, Ashida M. 1982. Ellagitannins of the Casuarinaceae, Stachyuraceae and Myrtaceae. Phytochemistry 21(12), 2871–2874.

Panneerselvam S, Geetha A, Karthikeyan SMN, Selvamathy N. 2010. The antioxidant and H+K+ ATPase inhibitory effect of *Andrographis paniculata* and andrographolide – *In vitro* and *in vivo* studies. Pharmacologyonline 1, 356–376.

Sainani GS, Manika JS, Sainani RG. 1997. Oxidative stress – A key factor in pathogenesis of chronic disease. Medical Update 1, 1.

Sofowara A. 1993. Screening plants for bioactive agents. In: Medicinal plants and traditional medicine in Africa. 2nd ed. Spectrum Book Ltd, Sunshine House, Ibadan, Nigeria, 134–156.

Sreejayan, Rao MN. 1997. Nitric oxide scavenging by curcuminoids. Journal of Pharmacy and Pharmacology **49**, 105–107.

Suresh SN, Nagarajan N. 2009. Preliminary phytochemical and antimicrobial activity analysis of *Begonia malabarica* Lam. Journal of Basic & Applied Biology **3**(1–2), 59–61.

Thirunavukkarasu P, Ramanathan T, Ramkumar L, Shanmugapriya R, Renugadevi G. 2011. The antioxidant and free radical scavenging effects of *Avicennia officinalis*. Journal of Medicinal Plants Research 5(19), 4754–4758.

Trease GE, Evans WC. 2002. Pharmacognosy. 15th ed. Saunders Publishers, London, 42–44, 222–229, 246–249, 304–306, 331–332, 391–393.

Vargas AD, Soto HM, Gonzalez HVA, Engleman EM, Martinez GA. 2006. Kinetics of accumulation and distribution of flavonoids in guava (*Psidium guajava*). Agrociencia 40, 109–115.